Supplemental data



Fig S1. A schematic graphic of the genomic locus of LERFS.



**Fig. S2. RACE assay of LERFS. (A)** Amplification products of 5' and 3' ends of LERFS. **(B)** Representative pictures of sequencing results of RACE products.



**Fig.S3. Analysis of coding capacity of LERFS.** Full-length LERFS was cloned into pcDNA3.1 downstream of the T7 promoter and translated *in vitro* using fluorescently labeled transfer RNA (tRNA). Luciferase mRNA was used as a positive translation control, and a mock-translated sample (no RNA template) was used as NC. The fluorescent translation product was resolved by SDS-PAGE and then visualized.



**Fig. S4. Localization of LERFS by nuclear/cytoplasm fractionation.** RT-qPCR analysis of cytoplasmic and nuclear fraction of HC FLSs revealed that LERFS expression was enriched in cytoplasm. Values are expressed relative to expression level of nucleus. Actin served as a cytoplasmic marker; U6 served as a nuclear marker.



Fig. S5. Efficiency of LERFS knockdown. RA FLSs were transfected with LncRNA Smart Silencer for 48h, and silencing efficiency was determined by RT-qPCR. Data show the Silencing efficiency of LERFS siRNA from 3 independent experiments. Ct values are normalized to GAPDH. \*P < 0.05 vs. control Smart Silencer (siC) by Student's t test.



**Fig. S6. Infected efficiency of LERFS overexpression lentivirus in RA FLSs.** RA FLSs infected with control lentivirus (VECTOR) or LERFS OE for 5 days were analyzed for LERFS expression by RT-qPCR assay. Ct values are normalized to GAPDH. \* P < 0.05 vs. VECTOR by Student's t test.



**Fig. S7. Inhibitory effect of LERFS overexpression on migration and invasion of RA FLSs induced by monocyte chemoattractant protein-1 (MCP-1).** (A) Chemotaxis migration of RA FLSs was evaluated using a Transwell assay. MCP-1 (50 ng/ml) was used as a chemoattractant in the lower wells. (B) In vitro invasion was determined using inserts coated with Matrigel Basement Membrane Matrix. Relative invasion rate was calculated by counting invaded cells and normalized to VECTOR. Data are shown as mean ± SEM from 5 independent experiments of 5 different RA patients. \*\*\* P < 0.001 vs. VECTOR by Student's t test.



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**Fig. S8. Growth rate of RA FLSs in serum-free medium at 0, 24 and 48 hours.** Cell number was evaluated by absorbance at 490 nm using an MTT assay at the indicated time points after lentivirus infection (D0 indicates day of infection). Values are expressed relative to D0 as mean ± SEM from 3 independent experiments of 3 different patients with RA.



Fig. S9. Effect of a proliferation inhibitor mitomycin on the migration and invasion of RA FLSs. Cells were pretreated with or without mitomycin (MMC,  $10 \mu g/ml$ ) for 6 hours. (A) Chemotaxis migration of RA FLSs was evaluated using a Transwell assay. (B) In vitro invasion was determined using inserts coated with Matrigel Basement Membrane Matrix. Relative invasion rate was calculated by counting invaded cells and normalized to VECTOR. Data are shown as mean ± SEM from 3 independent experiments of 3 different RA patients.



**Fig. S10. Effect of LERFS overexpression on cell cycle by RA FLS cell line MH7A**. Cells were stained with DAPI and detected by flow cytometry to analyze the cell cycle. Percentages of cells in the different phases of the cell cycle were calculated, and data are shown as mean  $\pm$  SEM from 5 independent experiments. \*\*\* *P* < 0.001 vs. VECTOR by Student's t test.



**Fig.S11. Silencing efficiency of hnRNP Q siRNA**. RA FLSs were transfected with hnRNP Q siRNA Q1–3 for 48 hours and subjected to RT-qPCR analysis. Data shown are mean  $\pm$  SEM from 3 independent experiments. \**P* < 0.05, \*\**P* < 0.01, \*\*\**P* < 0.001 vs. control siRNA (siC) by 1-way ANOVA.



**Fig.S12. Effect of hnRNP Q1 on mRNA expression of RhoA, Rac1 and CDC42 in RA FLSs.** RA FLSs were infected with control lentivirus (VECTOR) or hnRNP Q1 OE, and mRNA expression of indicated targets was analyzed by RT-qPCR. Ct values are normalized to GAPDH. \*\* P < 0.01 vs. VECTOR by Student's t test.



**Fig.S13.** Effect of translation inhibitor cycloheximide on hnRNP Q knockdown-induced increase of protein expression of CDC 42 and RhoA in LERFS overexpressed-RA FLSs. Cells were transfected with hnRNP Q siRNA or siC for 24 hours, followed by infection of control lentivirus (VECTOR) or LERFS OE for three days, and then treated with or without cycloheximide (10  $\mu$ M) for 24 hours. Representative WB images are shown in left panel. The semi-quantification of protein levels (right panel) is expressed as mean ± SEM of 3 independent experiments from 3 different RA patients. \**P*<0.05, \*\**P* < 0.01, \*\*\**P*<0.001 vs. VECTOR + siC, <sup>###</sup> *P* < 0.001 vs. LERFS OE + siC, <sup>&&</sup> *P* < 0.001 vs. sihnRNPQ +LERFS OE by Student's t test



**Fig.S14.** Effect of hnRNP Q knockdown on migration, invasion and proliferation of LERFS-overexpressed RA FLSs. Cells were transfected with hnRNP Q siRNA or siC for 24 hours, followed by infection of control lentivirus (VECTOR) or LERFS OE for three days. (A) Chemotaxis migration of RA FLSs was evaluated using a Transwell assay. (B) In vitro invasion was determined using inserts coated with Matrigel Basement Membrane Matrix. Relative invasion rate was calculated by counting invaded cells and normalized to VECTOR. (C) An EdU incorporation assay was performed to evaluate cell proliferation. Proliferating RA FLSs were labeled with EdU (red), and nuclei were stained with Hoechst 33342 (blue). Representative images (left panel) are shown (original magnification 200×). Data (right panel) are expressed as mean  $\pm$  SEM of 3 independent experiments from 3 different RA patients. \**P* < 0.05, \*\**P*<0.01 vs. VECTOR + siC, <sup>#</sup>*P* < 0.05 vs. LERFS OE + siC by Student's t test.

## Tables

Table S1. Demographic and clinical features of patients with active RA				
51.50±7.37				
32 (34)				
4.00±1.86				
26 (34)				
22 (34)				
5.69±2.26				
12 (34)				
18 (34)				
10 (34)				
7 (34)				
15 (34)				

## Table S2. Comparison of RA patients and HC subjects in age and gender

_			Gender	(Number)
Group	Number	Age (years) (mean $\pm$ SD)	Female	Male
RA	34	51.50±7.37	32	2
НС	29	52.45±8.08	27	2

Table S3. Sequences of primers, siRNA and ASO.

		RT-qPCR primers			
CADDU	Forward	GCACCGTCAAGGCTGAGAAC			
GAPDH	Reverse	TGGTGAAGACGCCAGTGGA			
	Forward	CGACTGGTGCTGAAGTGTTG			
LERFS	Reverse	CCCTACTCCTGCCTGCTCTA			
CDC42	Forward	CCATCGGAATATGTACCGACTG			
CDC42	Reverse	CTCAGCGGTCGTAATCTGTCA			
Rac1	Forward	ATGTCCGTGCAAAGTGGTATC			
RACI	Reverse	CTCGGATCGCTTCGTCAAACA			
RhoA	Forward	GGAAAGCAGGTAGAGTTGGCT			
KIIOA	Reverse	GGCTGTCGATGGAAAAACACAT			
	Forward	CTGGTCTCAATAGAGGTTATGCG			
hnRNP Q	Reverse	TCCGGTTGGTGGTATAAAATGAC			
MMP-1	Forward	CTCTGGAGTAATGTCACACCTCT			
	Reverse	TGTTGGTCCACCTTTCATCTTC			
	Forward	CGGTTCCGCCTGTCTCAAG			
MMP-3	Reverse	CGCCAAAAGTGCCTGTCTT			
	Forward	TCCTGATGTGGGTGAATACAATG			
MMP-13	Reverse	GCCATCGTGAAGTCTGGTAAAAT			
		RACE primers			
LERFS 5' RACE	CGCAGGTAAAGTCCATCCGT				
LERFS 3' RACE	AAACCAAGATTACACAAAAGCG				
UPM	CTAATACGACTC	ACTATAGGGCAAGCAGTGGTATCAACGCAGAGT			
		Clone primers			
LERFS	Forward	CTCTATTGAATGCTGGGTATCTTCCA			
	Reverse	AGTGAGGTCGGCGCCC			
		AGTGAGGTCGGCGCCC RiboTM IncRNA Smart Silencer			
	I	RiboTM IncRNA Smart Silencer			
	siRNA-1	RiboTM IncRNA Smart Silencer CTTCTGCCCTGAATTGAAA			
LERFS	siRNA-1 siRNA-2	RiboTM IncRNA Smart Silencer CTTCTGCCCTGAATTGAAA CTGATTGCTTTGCCAGTTA			
LERFS	siRNA-1 siRNA-2 siRNA-3	RiboTM IncRNA Smart Silencer CTTCTGCCCTGAATTGAAA CTGATTGCTTTGCCAGTTA GCCAATGACCTGAAGTTAA			
LERFS	siRNA-1 siRNA-2 siRNA-3 ASO-1	RiboTM IncRNA Smart Silencer CTTCTGCCCTGAATTGAAA CTGATTGCTTTGCCAGTTA GCCAATGACCTGAAGTTAA AGGACACTGAGGACTGGGAC			
LERFS	siRNA-1 siRNA-2 siRNA-3 ASO-1 ASO-2	RiboTM IncRNA Smart Silencer CTTCTGCCCTGAATTGAAA CTGATTGCTTTGCCAGTTA GCCAATGACCTGAAGTTAA AGGACACTGAGGACTGGGAC GACCTGCCAATGACCTGAAG			
LERFS	siRNA-1 siRNA-2 siRNA-3 ASO-1 ASO-2	RiboTM IncRNA Smart Silencer CTTCTGCCCTGAATTGAAA CTGATTGCTTTGCCAGTTA GCCAATGACCTGAAGTTAA AGGACACTGAGGACTGGGAC GACCTGCCAATGACCTGAAG AGAAGGCTGTAGAGGGATGA			
LERFS hnRNP Q	siRNA-1 siRNA-2 siRNA-3 ASO-1 ASO-2 ASO-3	RiboTM IncRNA Smart Silencer CTTCTGCCCTGAATTGAAA CTGATTGCTTTGCCAGTTA GCCAATGACCTGAAGTTAA AGGACACTGAGGACTGGGAC GACCTGCCAATGACCTGAAG AGAAGGCTGTAGAGGGATGA siRNA			