

IRE1 α RNase-dependent lipid homeostasis promotes survival in Myc-transformed cancers

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Supplementary Figure Legends

Figure S1. ER stress and IRE1 α -XBP1 signaling are increased in human and murine c-Myc transformed cancers. (A) Correlation between *HSPA5* and an established Myc signature (see Methods for description). (B) Comparison of RIDD targets between BL and CB. Data were obtained from the Oncomine database. $P < 0.05$ was regarded as significant (two tailed Student t test). (C) mRNA was extracted from liver tumors and paired normal liver tissues from the *LAP/MYC* mouse model. *MYC* and *Xbp1s/Xbp1t* examined with qRT-PCR. *Actb* was used as an endogenous control gene. Relative mRNA expression was determined by normalizing to levels in each normal sample. Three technical triplicates were used in each sample.

Figure S2. c-Myc engages ER stress and regulates the IRE1 α -XBP1 pathway. (A) Growth curve and immunoblot analysis for P493 cells with different levels of c-Myc expression (annotated as “No Myc”, “Low Myc”, and “High Myc”, $n=3$). (B) Quantification of relative cell size from **Figure 2A** ($n=3$). (C) Total RNA content per million cells determined by Nanodrop 1000 ($n=3$). (D) c-Myc target *LDHA* mRNA expression analyzed in P493 cells with different c-Myc levels ($n=3$). (E) qRT-PCR analysis of *MYC* and *LDHA* from experiment in **Figure 2F**. (F) Western blot comparing protein levels between P493 cells and 4 bona fide BL cell lines: Raji, Daudi, Ramos, and EB-2. (G) ChIP-seq analysis shows binding of c-Myc to the promoters of *ERN1*, *HSPA5*, and *XBP1* in P493 cells, canonical (CACGTG) or non-canonical (CACGCG) E-boxes were found in the binding regions. (H) Immunoblots of IRE1 α decay in P493 cells with different c-Myc levels after treatment with 100 μ g/mL cycloheximide for the indicted times. The graph represents the quantification of IRE1 α protein levels. (I and J) Immunoblots for DDRGK1 (I), SEL1L1 and HRD1 (J) in P493 cells. (K) Protein expression of SEL1L and HRD1 in experiment of **Figure 2G**. (L and M) qRT-PCR (L) and Western blot (M) analysis of BL cell lines treated with 500nM JQ1 for 24 hours or 48 hours. (N) qRT-PCR analysis of Ramos cells treated with 0.1 μ g/mL CHX for 4 hours. For qRT-PCR, *ACTB* was utilized as the endogenous control gene; data are representative of 3 independent experiments, if not specified elsewhere. Two-way ANOVA with Bonferroni correction was used to determine significance. *, $P < 0.05$, **, $P < 0.01$, ***, $P < 0.001$.

Figure S3. Synthetic lethality between c-Myc overexpression and IRE1 α RNase inhibition.

(A) Chemical structure of B-I09. (B) Western blot shows IRE1 α phosphorylation (phos-tag SDS-PAGE) of P493 High Myc cells treated with indicated concentrations of B-I09, in the presence of DMSO or 5 μ g/mL tunicamycin for 6 hours. (C) Representative contour plots of P493 High Myc, Low Myc, and No Myc cells treated with indicated concentrations of B-I09 for 48 hours. (D) Immunoblots of XBP1s and c-Myc in P493 cells treated with 10 μ M B-I09 for 48 hours. (E) P493 cells treated with indicated concentrations of Doxorubicin or JQ1 for 48 hours. Viability was examined and relative viability determined by normalizing to cells treated with Control or DMSO. (F) Growth curve of CLL cell lines (MEC1, MEC2, and WaC3) (n=3); Immunoblots show expression of c-Myc and XBP1s in each. (G) CLL cells treated with 20 μ M B-I09 for 72 hours, relative cell growth (n=3) and viability were determined. Immunoblots show expression of PARP cleavage upon B-I09 treatment. (H) Cells cultured with different concentrations of 4 μ 8c, and counted at indicated times (n=3). (I) Cells treated with indicated concentrations of 4 μ 8c for 48 hours. Viability was examined and relative viability determined by normalizing to viability of cells treated with DMSO. (J) Body weight of mice bearing P493 High Myc xenografts treated with Control or B-I09 (n=5 for Control, n=4 for B-I09). (E and I) Results are representative of 3 independent experiments. *, "High Myc" vs. "Low Myc"; #, "High Myc" vs. "No Myc". Two-way ANOVA with Bonferroni correction was used to determine significance. **, $P < 0.01$; ***, $P < 0.001$. ###, $P < 0.001$.

Figure S4. B-I09 has no effect on IRE1 α phosphorylation. (A) Western blot shows IRE1 α phosphorylation (phos-tag SDS-PAGE) of Ramos cells treated with indicated concentrations of B-I09, in the presence of DMSO or 5 μ g/mL tunicamycin for 6 hours.

Figure S5. B-I09 treatment responses are dependent on SCD1 loss in P493 cells. (A) P493 High Myc cells treated with 10 μ M B-I09 for 24 hours and 48 hours. Phospho-JNK and total JNK protein levels analyzed. (B) P493 cells treated with 10 μ M B-I09 for 48 hours. Autophagy markers

including p62 and LC3 analyzed. **(C)** Schematic model of U-¹³C-glucose tracing and fatty acid labeling. **(D)** Western blot analysis of P493 cells treated with 20μM 4μ8c for 48 hours. **(E)** CLL cell lines treated with 20μM B-I09 for 48 hours; c-Myc and SCD1 expression monitored. **(F)** P493 High Myc cells transfected with non-targeting (NT) siRNA or siRNA targeting XBP1 (siXBP1) for 48 hours, XBP1s and SCD1 were analyzed. **(G)** Immunoblots of experiment from **Figure 5F**. **(H)** P493 High Myc cells treated with DMSO or 20μM B-I09 for 48 hours, with OA, or Palm, or the combination (1:1 ratio). **(I)** P493 High Myc cells cultured in media with normal lipid concentration (100%) or delipidated condition (10%), treated with 10μM B-I09, and rescued with BSA or OA for 48 hours. Contribution ratios of viability restoration of BSA and OA in 100% or 10% lipid media were calculated (see Methods). **(J and K)** P493 cells treated with 0.5μM SCDi for 72 hours, cell growth (n=3) and viability were determined. For viability assays, data are representative of 3 independent experiments. Two-way ANOVA with Bonferroni correction was used to determine significance. ***, $P < 0.001$. n.s., not significant.

Figure S6. B-I09 treatment responses are dependent on SCD1 loss in BL cells. (A)

Immunoblots of experiment from **Figure 6B**. **(B)** Ramos cells treated with DMSO or 20μM B-I09 for 48 hours, with OA, or Palm, or the combination (1:1 ratio). **(C and D)** Ramos or Daudi cells cultured in media with normal lipid concentration (100%) or delipidated condition (25%), treated with 10μM B-I09, and rescued with BSA or OA for 48 hours. Contribution ratios of viability restoration of BSA and OA in 100% or 25% lipid media were calculated (see Methods). **(E)** Body weight of mice bearing Ramos xenografts treated with Control or SCDi. **(F)** mRNA was extracted from Ramos cells in experiment of **Figure 6D**; genes representing three UPR pathways examined with qRT-PCR. ATF6 pathway: *HERPUD1*; IRE1α-XBP1 pathway: *XBP1s*, *DNAJB9*; PERK-ATF4 pathway: *ATF3*, *DDIT3*. *TBP* and *ACTB* were utilized as endogenous control genes. *, comparison of SCDi and DMSO treatment. #, comparison of SCDi+OA and SCDi. For qRT-PCR and viability assay, data are representative of 3 independent experiments. Two-way ANOVA with Bonferroni correction was used to determine significance. **, $P < 0.01$, ***, $P < 0.001$. #, $P < 0.05$; ###, $P < 0.001$. n.s., not significant.

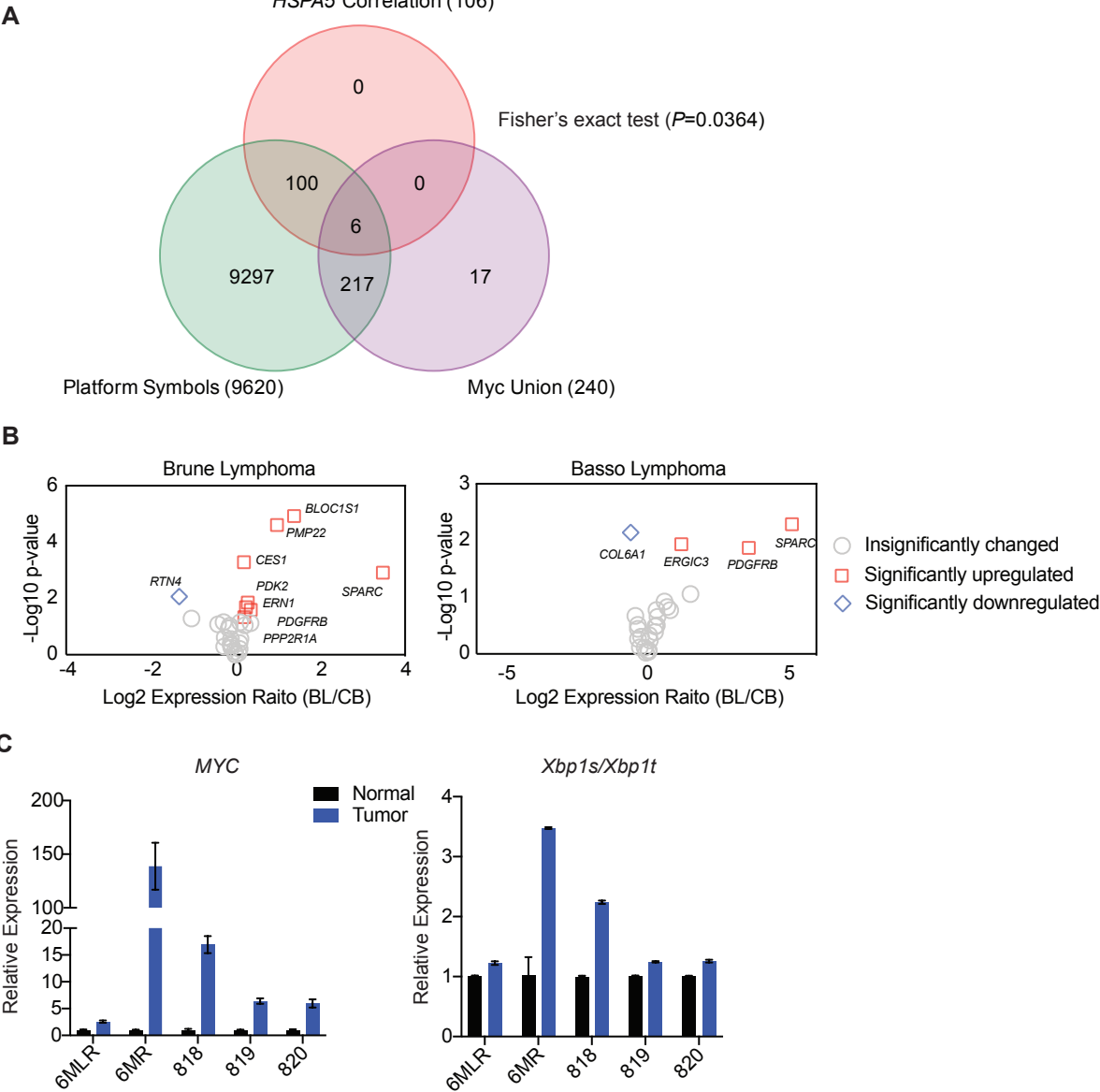
Figure S7. N-Myc activation engages the IRE1 α -XBP1 pathway, rendering cells vulnerable to XBP1s loss. (A) N-Myc target *ODC1* examined with qRT-PCR (n=3). (B) Control and 4-OHT SHEP cells were exposed to 100, 50, 25, 12.5, 6.25, 3.125, 1.5625 μ M B-I09 for 72 hours, IC₅₀ was then determined (n=3). (C and D) Control and 4-OHT SHEP cells treated with indicated concentrations of B-I09 for different times, viability was then measured. Relative viability was determined by normalizing to viability of cells with DMSO treatment. (E) Control or N-Myc SHEP cells treated with DMSO or 30 μ M B-I09, rescued with BSA or OA for 72 hours (n=6). Absorbance was measured using WST-1 reagents. *, comparison of B-I09 and DMSO treatment. #, comparison of B-I09+BSA or B-I09+OA and B-I09 treatment. (F) Control or N-Myc SHEP cells treated with 0.5 μ M SCDi for indicated times, viability was examined. Relative viability was determined by normalizing to viability of cells with DMSO treatment. (G) Control or N-Myc SHEP cells treated with siNT or siSCD for 72 hours, and viability was determined. Relative viability was determined by normalizing to viability of cells with siNT treatment. (H) N-Myc SHEP cells treated with 0.5 μ M SCDi, rescued with BSA control or OA for 72 hours, and viability was determined. Relative viability was determined by normalizing to viability of cells with DMSO treatment. For viability assays, data are representative of 3 independent experiments. Two-way ANOVA with Bonferroni correction was used to determine significance. **, $P < 0.01$, ***, $P < 0.001$. ####, $P < 0.001$. n.s., not significant.

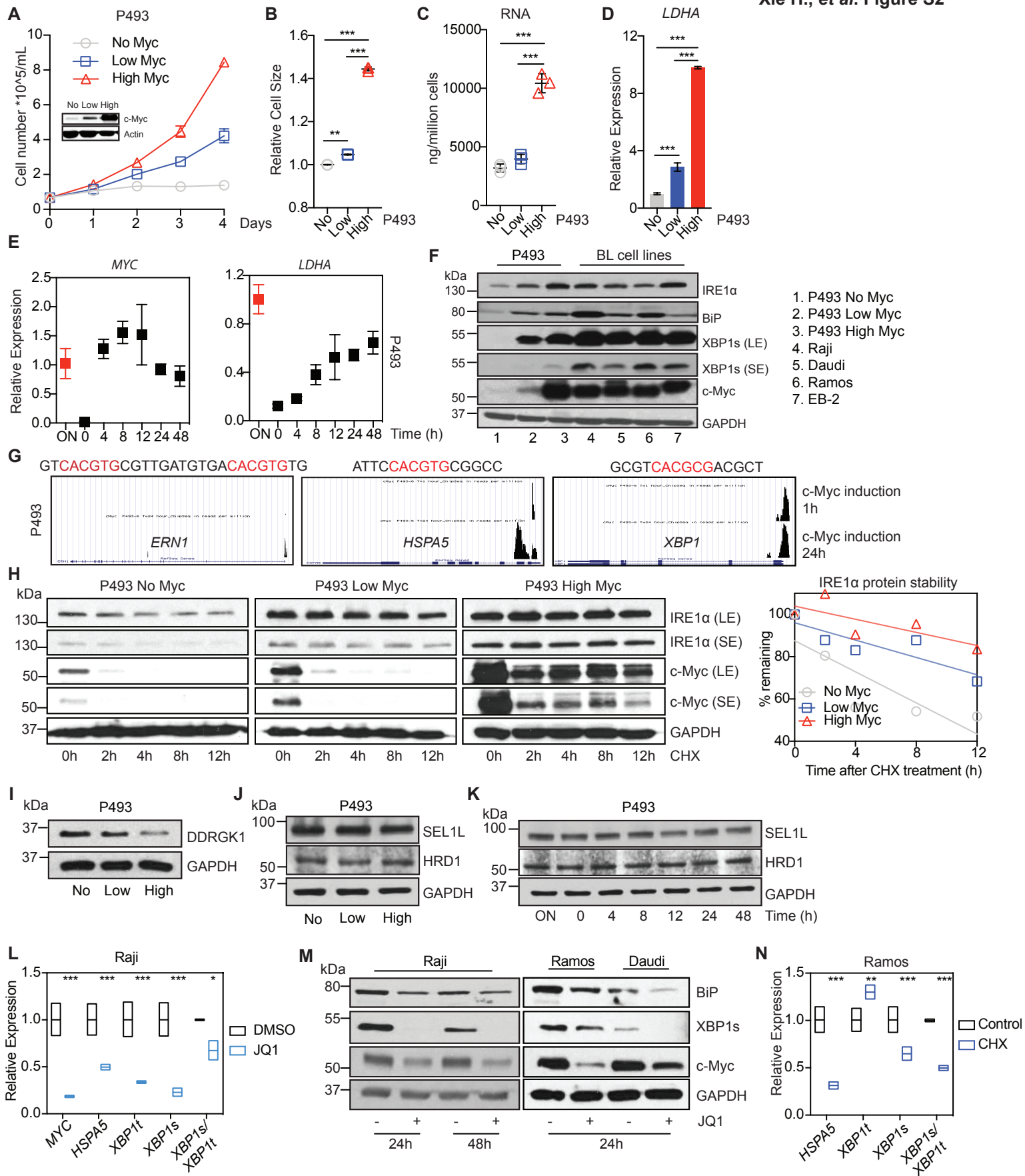
Figure S8. XBP1 depletion in neuroblastoma cells. (A) qRT-PCR comparing 3 neuroblastoma cell lines SK-N-AS, BE2C, and Kelly in terms of expressions of *MYCN*, *ODC1*, *HSPA5*, and *XBP1s*. (B) Three neuroblastoma cell lines were tested for IC₅₀ of B-I09 treatment for 72 hours. (C) Validation of *XBP1* knockdown in Kelly xenograft tumors by qRT-PCR (two-tailed paired t test). (D) Body weight of mice bearing Kelly xenograft tumors (n=10). For qRT-PCR, *TBP* and *ACTB* were used as endogenous control genes, 3 technical triplicates were used in each sample, and data are representative of 3 independent experiments. ***, $P < 0.001$.

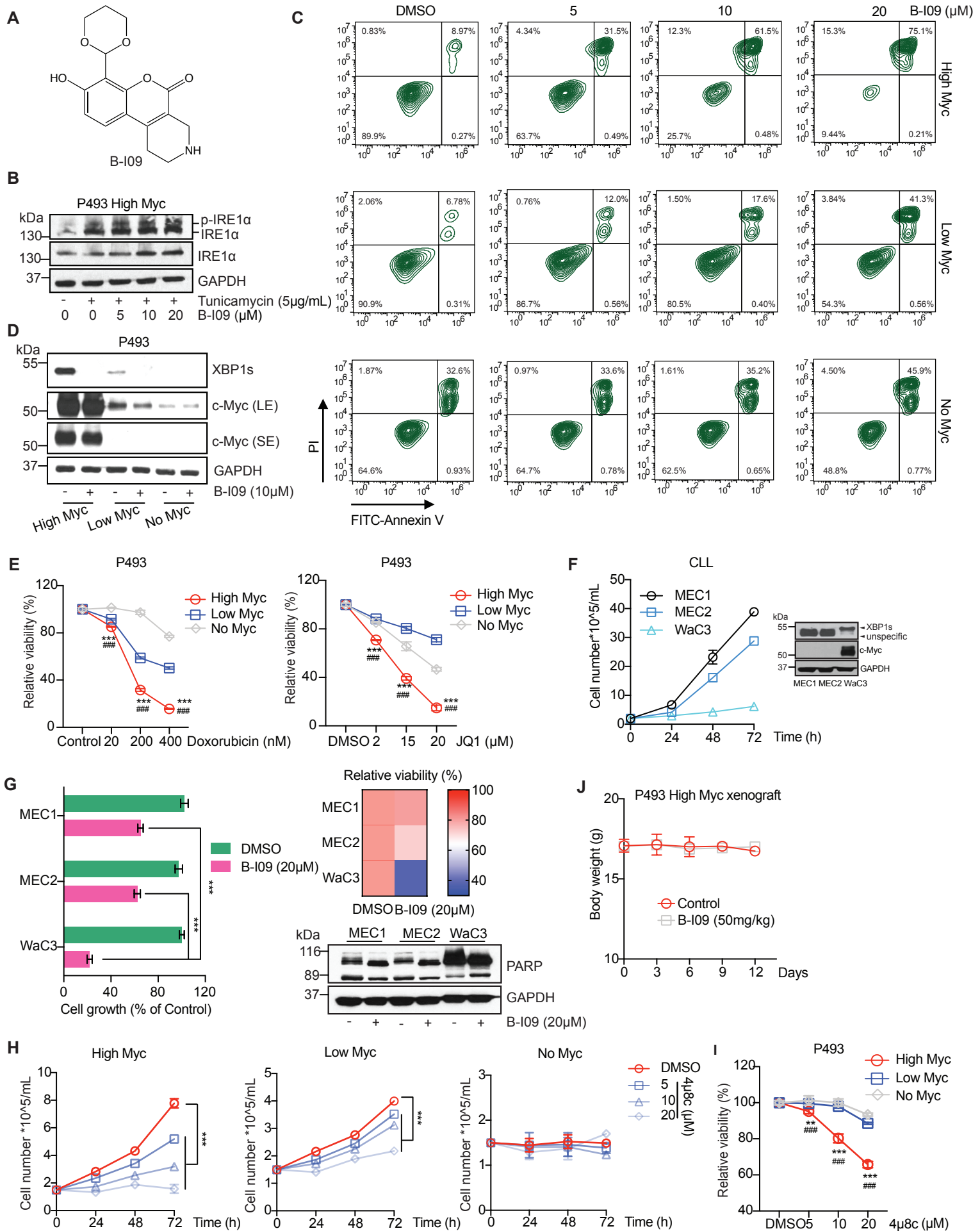
Figure S9. Combinational treatment between B-I09 and Vincristine. (A) Western blot examining SCD1 expression after treatment with 10 μ M B-I09 or 0.5 μ M Vincristine alone, or the combination of both in Daudi and Ramos cells after 48 hours. (B) Daudi or Ramos cells treated with 10 μ M B-I09 or 0.5 μ M Vincristine alone, or the combination of both, rescued with OA for 48 hours, viability was determined. Data are representative of 3 independent experiments.

Supplementary Table 1. Fisher's exact test to determine correlation between *HSPA5* and Myc signature.

	Myc Union	Not Myc Union	Total
<i>HSPA5</i> correlation	6	100	106
<i>HSPA5</i> no correlation	217	9297	9514
Total	223	9397	9620







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