The Journal of Clinical Investigation

Research Article

Mutations in the netrin-1 gene cause congenital mirror movements

Aurélie Méneret, 1,2 Elizabeth A. Franz, 3 Oriane Trouillard, 1 Thomas C. Oliver, 4 Yvrick Zagar, 5 Stephen P. Robertson, 6 Quentin Welníczk, 1,7 R.J. MacKinlay Gardner, 5 Cécile Gallea, 5 Myriam Smour, 5,6 Christel Depienne, 1,8,9 Christine L. Jasoni, 6 Caroline Dubacz, 7 Florence Riant, 7,14 Jean-Charles Lamy, 1 Marie-Pierre Morel, 7 Raphael Guérois, 15 Jessica Andreani, 15 Coralie Fouquet, 9 Mohamed Doulazmi, 9 Marie Vidalhiet, 1,2 Guy A. Rouleau, 8,17,18 Alexis Brice, 1,19 Alain Chédotal, 5 Isabelle Dusart, 7 Emmanuel Roze, 1,2 and David Markie 4

1 INSERM U1127, CNRS UMR 7225, Sorbonne Universités, UPMC Université Paris 06, UMR S1127, CIC-1422, Institut du Cerveau et de la Moelle épinière (ICM), Paris, France. 2 AP-HP, Hôpital de la Pitié-Salpêtrière, Département de Neurologie, Paris, France. 3 Department of Psychology and FMRIotogo, University of Otago, Dunedin, New Zealand. 4 Pathology Department, Dunedin School of Medicine, University of Otago, Dunedin, New Zealand. 5 Sorbonne Universités, UPMC Université Paris 06, INSERM, CNRS, Institut de la Vision, Paris, France. 6 Department of Women’s and Children’s Health, Dunedin School of Medicine, University of Otago, Dunedin, New Zealand. 7 Sorbonne Universités, UPMC Université Paris 06, INSERM, CNRS, Institut de Biologie Paris Seine, Neuroscience Paris Seine, Paris, France. 8 Department of Neurology and Neurosurgery, and 9 Department of Paediatrics, McGill University, Montreal, Quebec, Canada. 9 Institut de Génétique et de Biologie moléculaire et cellulaire (IGBMC), CNRS UMR 7104, INSERM U564, Université de Strasbourg, Illkirch, France. 10 Laboratoires de génétique, Institut de génétique médicale d’Alsace, Hôpitaux Universitaires de Strasbourg, Strasbourg, France. 11 Department of Anatomy, School of Biomedical Sciences, University of Otago, Dunedin, New Zealand. 12 AP-HP, Groupe hospitalier Laïboisié-Fernand Widal, Laboratoire de Génétique, Paris, France. 13 INSERM, UMR 5740, Université Paris 7 Denis Diderot, Paris, France. 14 Institute for Integrative Biology of the Cell (I2BC), CEA, CNRS, Université Paris Sud, Université Paris-Saclay, Gill sur Yvette, France. 15 Sorbonne Universités, UPMC Université Paris 06, INSERM, CNRS, Institut de Biologie Paris Seine, Adaptation Biologique et Vieillissement, Paris, France. 16 Montreal Neurological Institute, Montreal, Quebec, Canada. 17 Department of Human Genetics, McGill University, Montreal, Quebec, Canada. 18 AP-HP, Hôpital de la Pitié-Salpêtrière, Fédération de Génétique, Département de Génétique et de Cytogénétique, Paris, France.

Netrin-1 is a secreted protein that was first identified 20 years ago as an axon guidance molecule that regulates midline crossing in the CNS. It plays critical roles in various tissues throughout development and is implicated in tumorigenesis and inflammation in adulthood. Despite extensive studies, no inherited human disease has been directly associated with mutations in NTN1, the gene coding for netrin-1. Here, we have identified 3 mutations in exon 7 of NTN1 in 2 unrelated families and 1 sporadic case with isolated congenital mirror movements (CMM), a disorder characterized by involuntary movements of one hand that mirror intentional movements of the opposite hand. Given the diverse roles of netrin-1, the absence of manifestations other than CMM in NTN1 mutation carriers was unexpected. Using multimodal approaches, we discovered that the anatomy of the corticospinal tract (CST) is abnormal in patients with NTN1-mutant CMM. When expressed in HEK293 or stable HeLa cells, the 3 mutated netrin-1 proteins were almost exclusively detected in the intracellular compartment, contrary to WT netrin-1, which is detected in both intracellular and extracellular compartments. Since netrin-1 is a diffusible extracellular cue, the pathophysiology likely involves its loss of function and subsequent disruption of axon guidance, resulting in abnormal decussation of the CST.

Introduction

Netrins are a family of extracellular proteins that regulate cell migration and survival during development and adulthood. The first member, netrin-1, was identified as a secreted protein mediating axon guidance in the spinal cord (1). Netrin-1 is expressed at the midline of the developing CNS and acts as an attractive or repulsive cue for different populations of crossing/commissural axons (2). The classical view that attraction of commissural axons is mediated by a gradient of floor-plate–derived netrin-1 has been recently challenged: netrin-1 primarily acts locally by promoting growth cone adhesion (3, 4). In netrin-1–KO mice, many commissural axons fail to cross the midline in the corpus callosum, the hippocampal and anterior commissures, the corticospinal tract (CST), and the spinal cord (5–7). Netrin-1 binds to several transmembrane receptors including deleted in colorectal carcinoma (DCC) (8) and UNC5s, which respectively mediate attractive or repulsive axonal responses to netrin-1.

Netrin-1 exerts a variety of functions beyond axon guidance and is involved in synaptogenesis, apoptosis, cell migration, and angiogenesis (9, 10). There is also evidence suggesting that netrin-1 influences remyelination and tumorigenesis (11, 12). Netrin-1 is a potential therapeutic target and/or biomarker in cancer, cardiovascular disease, neuroinflammation, and kidney disease (12–14). However, except for a possible association with cleft lip in GWAS studies (15), the 20th anniversary of the discovery of netrin-1 has passed without the identification of a single human disease caused by inherited mutations in NTN1, the gene coding for netrin-1. In contrast, variants in the genes coding for receptors of netrin-1 such as UNC5C have been linked to Alzheimer’s disease and colorectal cancer (16), while mutations in DCC have...
CMM is characterized by involuntary movements of one side of the body that accompany and mirror intentional movements on the opposite side (Online Mendelian Inheritance in Man [OMIM] #157600)(22, 23). CMM mostly involves the hands. Affected subjects are unable to perform pure unimanual movements and have difficulty with skills requiring dissociated movements of the 2 hands (24, 25). It has been described as a familial disorder with autosomal dominant inheritance and incomplete penetrance, but sporadic cases also exist. The pathophysiology of CMM is probably related to developmental abnormalities of the CST, which conveys motor outputs from the motor cortex to the contralateral side of the body, and of the corpus callosum, which underlies communication between motor cortices. CMM usually involves abnormal decussation of the CSTs and altered interhemispheric inhibition with bilateral activation of the primary motor cortices during unimanual movements (26–28). To date, the established culprit genes are DCC and RAD51, which encodes a DNA repair protein, and possibly DNAL4, which encodes an axonemal dynein light chain (19, 21, 29–31). Recently, RAD51 has been described as a negative regulator of the netrin-1 signaling pathway (32). Only 35% of affected individuals or families carry a pathogenic variant in 1 of these 3 genes (22). This study has been undertaken to investigate whether mutations in other axon guidance genes, particularly those interacting with DCC or belonging to the netrin-1/DCC pathway, might cause CMM.

Results

Three netrin-1 variants are associated with CMM. Exome sequencing was conducted in 25 index cases of CMM (8 familial and 17 sporadic) without mutations in DCC, RAD51, or DNAL4. Patients’ characteristics and genetic findings are summarized in Supplemental Table 1; supplemental material available online with this article; https://doi.org/10.1172/JCI95442DS1.

Figure 1. Pedigrees of the CMM families and distribution of the mutations in NTN1. (A) Family 1 (from France) with a C601R mutation (the unusual family tree is a result of individual 1.1, who had children with 4 different women), (B) family 2 (from the United Kingdom [data were collected in New Zealand], which was previously identified as Family C in ref. 33), with an I518del mutation, and (C) a Canadian sporadic case with a C601S mutation. Black symbols represent individuals with CMM, symbols with an embedded black circle indicate asymptomatic carriers, white symbols indicate unaffected individuals, and symbols with a diagonal line represent deceased individuals. Squares represent males and circles represent females. m, mutated allele; +, WT allele. Electrophoregrams confirm the 3 NTN1 mutations obtained by Sanger sequencing. Red arrows point to sequence changes. Electrophoregrams were obtained with SeqScape software, version 2.6 (Applied Biosystems).
than 35,000 variants were identified per individual (Supplemental Tables 2 and 3). The flow chart of exome data analysis is summarized in Supplemental Figure 1. We initially filtered the data to remove variants that are too common to explain the low prevalence of the disorder (allele frequency greater than 0.1% in the Exome Variant Server [EVS] and 1000 Genomes databases; see Methods), those unlikely to affect protein-coding genes (intronic and synonymous), and low-confidence variant calls owing to poor coverage in our data (read depths of less than 10× in the French exomes and less than 5× in the New Zealand exomes) (Supplemental Table 4). Genes were then prioritized as candidates by known connection to DCC and RAD51 function by the number of unrelated individuals with variants, the functional prediction scores, and the rarity of variants in the Exome Aggregation Consortium (ExAC) database (comprising exome data on 60,706 unrelated individuals; see Methods). A total of 17 genes, with variants in at least 3 index cases or in 2 index cases but with a known function in axon guidance or known interaction with DCC or RAD51, were selected (Supplemental Figure 1). Interestingly, the NTN1 gene, coding for netrin-1, was among them, with 2 individuals carrying heterozygous variants. One missense variant (c.1801T>C/p.Cys601Arg) of NTN1 segregated with CMM in all three affected members of a French family (Figure 1A) was predicted to be pathogenic by Sorting Intolerant From Tolerant (SIFT), MutationTaster, and PolyPhen-2 algorithms and was absent from the ExAC database (Table 1). An in-frame deletion (c.1552_1554del/p.Ile518del) of NTN1 segregated with CMM in all 3 affected members of a family from the United Kingdom (data collected in New Zealand; Family C in ref. 33) (Figure 1B) and was also predicted to be pathogenic (Table 1). To confirm the role of NTN1 in CMM, we sequenced the NTN1 gene in 22 additional CMM index cases (20 sporadic and 2 familial cases from a Canadian cohort, all negative for DCC, RAD51, and DNAL4) and found another missense variant of NTN1 (c.1802G>C/p.Cys601Ser) in a sporadic case (Figure 1C). This variant was also predicted to be pathogenic (Table 1). These 3 variants alter amino acids in the NTR (netrin) domain (Figure 2A). ExAC data showed a rare variant in Ile518 (Ile518Val, 2 of 119,708 alleles), but valine has physicochemical properties similar to those of isoleucine. No variant affecting the 6 conserved cysteines (C472, C476, C491, C544, C546, and C601) of the NTR domain was present in ExAC, except for a synonymous variant in C472. ExAC data showed several rare NTN1 variants (allele frequency <0.1%) predicted to be pathogenic by both SIFT and PolyPhen-2 (Supplemental Table 5) and 2 rare loss-of-function variants, which have not been confirmed. Statistical analysis showed that the frequency of NTN1 variants predicted to be deleterious was significantly different between patients with CMM and the ExAC controls (3 of 47 vs. 63 of 60,706; P = 2.306 × 10^-5). In addition, all 3 variants described here in CMM patients (I518del, C601R, C601S) alter amino acids that are highly conserved among vertebrates (Figure 2C). Because the precise structure of the NTR domain of netrin-1 has not yet been determined experimentally through crystallography, we predicted its structure by using Rosetta comparative modeling software (34). This structural model of the NTR domain revealed that Cys601 is probably involved in a conserved disulfide bridge with Cys491 found in many members of

Table 1. Summary of the NTN1 mutations identified in patients with CMM

| Family | Patient origin | No. of patients | No. of asymptomatic carriers | Variant protein | Variant type | Gene domain | Mutation type | In silico predictions: SIFT | In silico predictions: MutationTaster | In silico predictions: PolyPhen-2 | ExAC | CADD score | CADD score raw score | PhD | ExAC score phen | ExAC score raw score | CADD score (http://cadd.gs.washington.edu/); ExAC (http://exac.broadinstitute.org). Reference sequences used: NM_004822 and NP_004813.

<table>
<thead>
<tr>
<th></th>
<th></th>
<th>1</th>
<th>2</th>
<th>3</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>France</td>
<td>3</td>
<td>2</td>
<td>c.1801T&gt;C/p.Cys601Arg</td>
</tr>
<tr>
<td></td>
<td>UK</td>
<td>0</td>
<td>0</td>
<td>c.1552_1554del/p.Ile518del</td>
</tr>
<tr>
<td></td>
<td>Canada</td>
<td>0</td>
<td>0</td>
<td>c.1802G&gt;C/p.Cys601Ser</td>
</tr>
</tbody>
</table>

the superfamily of NTR domains and that Ile518 has a key position in a β-strand (Figure 2B) (35–37). Altogether, 3 NTN1 variants from 3 independent families were associated with CMM and predicted to be pathogenic by the SIFT, MutationTaster, and PolyPhen-2 algorithms, the combined annotation-dependent depletion (CADD) score, and by the structural model.

Patients harboring NTN1 variants have abnormal crossing of the CST. Clinically, the NTN1 patients (including an 81-year-old asymptomatic carrier) had normal eyesight, no oculomotor abnormalities, no cardiovascular or respiratory disease, no inflammatory disease, and no cancer. Two of the patients had childhood-onset constipation-dominant irritable bowel syndrome (IBS), and two had peripheral vasoconstriction when exposed to cold (Supplemental Table 1). Except for mirror movements, neurological examination was normal. MRI analysis was performed for 1 patient (patient 1.9) and 20 control subjects. A brain MRI of this patient was normal, showing no corpus callosum hypoplasia or visible pontine abnormalities (Supplemental Figure 2). Tractography can provide individualized volumes of interest for the investigation of white matter microstructural characteristics in the context of disease. Tracts are personalized to each individual, and their positions are subject to interindividual anatomical variability. The variation of fractional anisotropy (FA) (the basic diffusion tensor imaging [DTI] metric) throughout the white matter is informative, for instance, with low values at the transition between gray and white matter or at the crossing of 2 main pathways. Therefore, we performed tract-based analyses that allowed between-subjects comparisons and optimized the localization of within-tract alterations (Figure 3, A and B). First, for each subject, we calculated the CST laterality coefficient on the basis of ratio between the number of streamlines crossing the midline and the number of streamlines projecting to the ipsilateral side after the pyramidal decussation. Comparison of the CST laterality coefficient of patient 1.9 with that of a group of 20 controls using the Crawford-Howell t test showed a significant group difference (P = 0.001; Figure 3C) (34). Second, we performed between-subjects analysis of FA values along the length of the crossed CST, focusing on the pyramidal decussation. We investigated the projection of the CST in 1 patient and 1 control subject (age and sex matched, Figure 3D). Mean FA values increased along the crossed CST in the patient compared with the control subject at the level of the pyramidal decussation (Figure 3D). The relative increased FA value at this location would suggest that fewer axons are crossing the midline in the patient. These results suggested that a higher proportion of fibers was located in the crossed versus the uncrossed CST in the control subject, whereas a higher proportion of fibers was located in the uncrossed CST in the patient (Figure 3D). We then used single-pulse transcranial...
netrin-1–AP proteins could be detected in HEK293 cell lysates, but not in the supernatant, contrary to the WT netrin-1–AP constructs (Figure 5, A and B). Similar results were obtained using untagged human WT netrin-1 and a netrin-1C601R variant (Supplemental Figure 3, A and B). In addition, the initial rate of reaction in the AP assay was higher in the supernatant of cells transfected with WT netrin-1–AP constructs than in the supernatant of nontransfected cells, whereas there was no difference between the nontransfected cells and cells transfected with each of the mutated netrin-1–AP constructs (Figure 5C). To further demonstrate the lack of netrin-1 in the extracellular space and quantify the expression of the different netrin-1 constructs, we conducted transfection experiments in Flp-In TRex tetracycline transactivator stable HeLa cell lines.

Magnetic stimulation (TMS) to study the propagation of neural signals along the CST in 2 patients (patients 1.9 and 2.3) and 2 control subjects (age and sex matched). We found that unilateral stimulation of the primary motor cortex frequently elicited bilateral motor-evoked potentials (MEPs) in patients compared with strictly unilateral MEPs, contralateral to the stimulation, in controls (Figure 4 and Table 2). From this multimodal approach, we showed that patients with CMM due to NTN1 mutations have an increased proportion of ipsilateral CST projections.

The 3 mutated netrin-1 proteins were almost exclusively detected in the intracellular compartments. To assess the pathogenicity of the 3 variants, mutations (I518del, C601R, and C601S) were introduced into mouse and human netrin-1 cDNAs fused to an alkaline phosphatase (AP) reporter. We showed that the mutated netrin-1–AP proteins could be detected in HEK293 cell lysates, but not in the supernatant, contrary to the WT netrin-1–AP constructs (Figure 5, A and B). Similar results were obtained using untagged human WT netrin-1 and a netrin-1C601R variant (Supplemental Figure 3, A and B). In addition, the initial rate of reaction in the AP assay was higher in the supernatant of cells transfected with WT netrin-1–AP constructs than in the supernatant of nontransfected cells, whereas there was no difference between the nontransfected cells and cells transfected with each of the mutated netrin-1–AP constructs (Figure 5C). To further demonstrate the lack of netrin-1 in the extracellular space and quantify the expression of the different netrin-1 constructs, we conducted transfection experiments in Flp-In TRex tetracycline transactivator stable HeLa cell lines. This transfection strategy allows expression cassettes to be inte-
HeLa cells, the mutated proteins were almost exclusively detected in the intracellular compartment, contrary to WT netrin-1. Since netrin-1 is a diffusible extracellular cue, the pathophysiology probably involves loss of function. As a consequence, CST midline axonal guidance was likely disrupted in the patients.

Given the numerous roles of netrin-1 during development and beyond, the absence of manifestations other than mirror movements is quite surprising. None of the subjects in our study had vision or hearing impairment (39), oculomotor abnormalities (40), hypogonadism (41), cardiovascular diseases (13), cancers (12), or inflammatory diseases (14). Constipation-dominant IBS and temperature regulation issues (10) have each been observed in 2 patients. The occurrence of IBS is especially interesting. Considering its high prevalence in the general population and the lack of specificity of the core IBS manifestations, it could be a coincidence. However, it is noteworthy that its onset was in infancy in our 2 patients instead of in young adulthood, as is the case in most patients with IBS. In addition, netrins and DCC have been linked to the distribution of neurons in the enteric nervous system, specifically to the migration of neurons to the submucosal plexus and to the pancreas (42). Abnormalities of the submucosal plexus are likely to affect gastrointestinal secretion of chloride and thus water, which could cause defects in motility. Submucosal neuronal abnormalities could also lead to defects in intrinsic motor reflexes, with a similar outcome. These might result in changes that are interpreted as IBS. No data were available concerning the secretion of pancreatic enzymes or hormones in our patients.

The lack of other phenotypes might be explained first by the fact that DCC has other ligands outside netrin-1 and that it also binds to other secreted netrins (43–45), cerebellin-4 (46), and other receptors such as ROBOs and UNC5s (47). Therefore, DCC mutations are expected to lead to more severe brain phenotypes than NTN1 mutations. In addition, the patients only carry heterozygous mutations, and it is therefore difficult to extrapolate from

gated into only 1 predefined site in the genome, thus eliminating differences in expression levels that may arise because of different sites of integration (38). After induction by doxycycline, mutant protein levels in cell lysates were either comparable (C601S) or lower (I518del and C601R) than WT protein levels, indicating that the intracellular stability of the mutant proteins might be slightly reduced. In contrast to WT netrin-1, the mutated netrin-1 proteins could not be detected in the supernatants (Figure 5E). However, by concentrating the supernatants, we were able to detect very low levels of the 3 mutant proteins, indicating a minimal presence of the mutant proteins in the extracellular space (Figure 5F). Finally, we studied cellular localization through immunofluorescence. In the absence of doxycycline, no netrin-1 was detected (Figure 6). After induction by doxycycline, none of the constructs was detected in the nucleus (Figure 6). We found that mutant netrin-1 proteins were mostly confined to the cytoplasm (double labeling with actin), whereas WT netrin-1 was mainly detected in the extracellular compartment (Figure 7). In both HEK293 and HeLa cells, all 3 mutated proteins found in patients with CMM were almost exclusively detected in the cellular compartment, whereas WT netrin-1 was detected in both cellular and extracellular compartments.

Discussion

We have shown that heterozygous mutations in exon 7 of NTN1 caused autosomal dominant CMM in 2 unrelated families and 1 sporadic case. NTN1-CMM patients have an increased proportion of ipsilateral CST projections compared with that observed in control subjects. When expressed in HEK293 or HeLa cells, the mutated proteins were almost exclusively detected in the intracellular compartment, contrary to WT netrin-1. Since netrin-1 is a diffusible extracellular cue, the pathophysiology probably involves loss of function. As a consequence, CST midline axonal guidance was likely disrupted in the patients.

Given the numerous roles of netrin-1 during development and beyond, the absence of manifestations other than mirror movements is quite surprising. None of the subjects in our study had vision or hearing impairment (39), oculomotor abnormalities (40), hypogonadism (41), cardiovascular diseases (13), cancers (12), or inflammatory diseases (14). Constipation-dominant IBS and temperature regulation issues (10) have each been observed in 2 patients. The occurrence of IBS is especially interesting. Considering its high prevalence in the general population and the lack of specificity of the core IBS manifestations, it could be a coincidence. However, it is noteworthy that its onset was in infancy in our 2 patients instead of in young adulthood, as is the case in most patients with IBS. In addition, netrins and DCC have been linked to the distribution of neurons in the enteric nervous system, specifically to the migration of neurons to the submucosal plexus and to the pancreas (42). Abnormalities of the submucosal plexus are likely to affect gastrointestinal secretion of chloride and thus water, which could cause defects in motility. Submucosal neuronal abnormalities could also lead to defects in intrinsic motor reflexes, with a similar outcome. These might result in changes that are interpreted as IBS. No data were available concerning the secretion of pancreatic enzymes or hormones in our patients.

The lack of other phenotypes might be explained first by the fact that DCC has other ligands outside netrin-1 and that it also binds to other secreted netrins (43–45), cerebellin-4 (46), and other receptors such as ROBOs and UNC5s (47). Therefore, DCC mutations are expected to lead to more severe brain phenotypes than NTN1 mutations. In addition, the patients only carry heterozygous mutations, and it is therefore difficult to extrapolate from

Table 2. Frequency and amplitude of the mirror MEPs in 2 NTN1 patients

<table>
<thead>
<tr>
<th>Patient</th>
<th>Stimulated hemisphere</th>
<th>Frequency of ipsilateral MEPs</th>
<th>Relative amplitude (ipsi-/contralateral MEPs)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.3</td>
<td>Dominant (left)</td>
<td>19%</td>
<td>0.36</td>
</tr>
<tr>
<td></td>
<td>Nondominant (right)</td>
<td>57%</td>
<td>1.27</td>
</tr>
<tr>
<td>1.9</td>
<td>Dominant (left)</td>
<td>17%</td>
<td>0.02</td>
</tr>
<tr>
<td></td>
<td>Nondominant (right)</td>
<td>98%</td>
<td>0.31</td>
</tr>
<tr>
<td>Controls</td>
<td>Dominant (left)</td>
<td>0%</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Nondominant (right)</td>
<td>0%</td>
<td></td>
</tr>
</tbody>
</table>

The frequency of ipsilateral MEPs is the percentage of trials in which bilateral muscle responses were elicited from the unilateral stimulation of the dominant or nondominant hemisphere.
what has been described in Ntn1-null or hypomorphic-mutant mice. Other human secreted netrins (NTN2/3, NTN4, and NTN5) could partially compensate for the lack of netrin-1. Whether these netrins act on CST axons is unknown, and neither netrin-5–KO (45) nor netrin-4–KO (48) mice display locomotor defects. Furthermore, penetrance was incomplete, at least in family 1. Incomplete penetrance has already been observed for RAD51 and DCC mutations (19, 21, 29). Interestingly, despite the small number of patients, both tractography and TMS results converged to support the existence of an abnormal uncrossed CST in NTN1 patients. The presence of an abnormal uncrossed CST has also been observed in RAD51-CMM and DCC-CMM patients (26, 49). Altogether, these results suggest that the CST midline guidance function of netrin-1, as with RAD51 and DCC, is the most sensitive to loss of function, with abnormalities occurring when only a subfraction of the translated protein is fully functional, whereas alterations of other functions might require greater deficits.

The presence of numerous uncrossed CST fibers in NTN1 and DCC-CMM patients is consistent with findings in severe hypomorphic Ntn1-mutant mice, which show much-reduced decussation of the CST (6). These mutants also exhibit corpus callosum agenesis (7), an absence of forebrain commissures, and defects in spinal commissural axon projections (5). Ntn1 hypomorphic mutants usually die shortly after birth. Dcc-KO mice also display commissural axon guidance defects and are not viable (50). Electrophysiological recordings in embryonic spinal cord preparations showed that neuronal circuits controlling locomotion are disorganized in Dcc and Ntn1 mutants (51). In addition, Dcc mutants carrying the “kanga” mutation (which results in a deletion of the P3 domain in the cytoplasmic tail of DCC) have a hopping gait (6). These mice...
also present with corpus callosum agenesis and defects in the decussation of the CST. This is partly consistent with the abnormalities found in CMM patients that involve both altered inter-hemispheric inhibition and abnormal decussation of the CST (26, 27). The study of Ntn1-conditional KO mice (3, 10) might allow for a better understanding of the pathogenic underpinnings of CMM in NNT1 patients. In addition, Ntn-1+/− mice showed normal neural anatomy of the colon but heightened colonic inflammation in experimental colitis (52). This is in keeping with the hypothesis that NNTN1 mutations may induce intestinal disease.

NNT1 mutations probably alter netrin-1 function and/or stability through deleterious structural modifications that result in a lack of netrin-1 in the extracellular compartment. The 3 reported variants are 2 missense mutations and 1 in-frame deletion located in a single domain, the NTR domain, at the C-terminal part of the protein. The NTR domain is not necessary for secretion (53) and not required for DCC binding (54–56). The 3 mutations are predicted to be pathogenic by different algorithms (Table 1), whereas very few NNT1 variants in the ExAC database are predicted to be damaging. The DECIPHER database (Database of genomic variation and Phenotype in Humans using Ensembl Resources, https://decipher.sanger.ac.uk/) lists 3 deletions encompassing NNTN1, including, respectively, 28, 65, and 1,299 genes that have not been associated with CMM. In the latter, the reported phenotype is severe, including, among others, developmental delay. These individuals might have CMM that has gone undetected, in the presence of more obvious comorbidities associated with the deletion. Incomplete penetrance and difficulty to diagnose CMM cloud the picture as well, as they do for potentially pathogenic alleles that are present in online databases like ExAC. The frequency of NNTN1 variants predicted to be damaging was significantly higher in individuals with CMM than in the ExAC population. The 6 conserved cysteines located in the NTR module of netrin-1 form disulphide bonds that are probably very important for the structure of netrin-1 (35). The cysteine in position 601 seems to be of particular importance, as it was mutated in 2 of our 3 families. The structural model suggests that C601R and C601S mutants most likely induce the disruption of a disulfide bridge and therefore alter the folding of the NTR domain. Besides, mutation of 1 cysteine involved in a disulfide bridge leaves the other cysteine (C491) free, which is potentially highly detrimental for proper expression in the extracellular medium. In parallel, the I518 deletion induces a deletion of 1 amino acid in a β-strand that is predicted to be deleterious and may be highly detrimental for the structure of the NTR domain. Altogether, our data strongly support the pathogenicity of these 3 variants. More demonstrative, we show that the mutated proteins are present in extremely small amounts in the extracellular space in contrast to WT netrin-1. Since netrin-1 is an extracellular cue, a decrease of its quantity in the extracellular space might explain the pathophysiology through loss of function. As mutant protein quantity was either comparable or smaller than WT proteins in the cellular compartments, the defect of netrin-1 in the extracellular space might be due to a secretion defect or a slightly reduced stability of the mutant proteins. Consistent with reports of the existence of a nucleolus localization signal in the C-terminal part of netrin-1 (57), we considered the alternative hypothesis of an abnormal relocation of the protein to the nucleus. However, our experiments showed that the mutant netrin-1 proteins were not detected in the nucleus but were located throughout the cytoplasm. The quasi-absence of mutated netrin-1 in the extracellular space might thus be due to impairment of the secretion process or to protein instability, particularly in the extracellular space. Although the NTR, per se, is thought not to be
required for netrin-1 secretion (46) but might be involved in the control of the activity of metalloproteases (35), our main hypothesis is that its structural disruption might prevent the presence of secreted netrin-1 in the extracellular space. Further studies are needed to decipher the exact mechanisms by which NTNI mutations result in a lack of netrin-1 in the extracellular compartment. Since netrin-1 is considered a potential therapeutic target for cancer and inflammatory diseases, identification of these mechanisms might open new therapeutic avenues.

Methods

Patient selection
Data on international cases were collected over a period of 5 years in France and New Zealand. Twenty-five index cases without a mutation in either DCC or RAD51 (rearrangements being excluded in 14 of them) were selected for exome sequencing. Each patient as well as available family members underwent a standardized neurologic assessment and DNA sampling. The severity of mirror movements (MM) was scored with the Woods and Teuber scale. Familial history, MM location, associated disorders, and reported functional disability were noted.

Exome sequencing and analysis
With regard to the exome sequencing performed in France, genomic DNA was captured using Agilent in-solution enrichment methodology (SureSelect Human All Exon Kits V5; Agilent Technologies) with their biotinylated oligonucleotide probes library (SureSelect Human All Exon V5, 50 Mb; Agilent Technologies), followed by paired-end 75-base massive parallel sequencing on an Illumina HiSEQ 2000. Sequence capture, enrichment, and elution were performed according to the manufacturer’s instructions and protocols, without modification except for library preparation, which was performed with the NEB-Next Ultra Kit (New England BioLabs). For library preparation, 600 ng of each genomic DNA was fragmented by sonication and purified to yield fragments of 150 to 200 bp. Paired-end adaptor oligonucleotides from the NEB kit were ligated on repaired, A-tailed fragments and then purified and enriched by 8 PCR cycles. Purified libraries (1.2 μg) were then hybridized to the SureSelect oligonucleotide probe capture library for 72 hours. After hybridization, washing, and elution, the eluted fraction was PCR amplified with 9 cycles and then purified and enriched by 8 PCR cycles. Purified libraries (1.2 μg) were then hybridized to the SureSelect oligonucleotide probe capture library for 72 hours. After hybridization, washing, and elution, the eluted fraction was PCR amplified with 9 cycles and then purified and quantified by quantitative PCR (qPCR) to obtain an adequate DNA template for downstream applications. Each elution-enriched DNA sample was then sequenced on an Illumina HiSEQ 2000 as paired-end 75-base reads. Image analysis and base calling were performed using an Illumina Real Time Analysis (RTA 2.1.3) system with default parameters. Bioinformatics analysis of sequencing data was based on the Illumina pipeline (CASAVA 1.8.2). CASAVA performs alignment of a sequencing run to a reference genome (hg19), calls the SNPs on the basis of the allele calls and read depth, and detects variants (SNPs and indels). The alignment algorithm used was ELANDv2 (CASAVA),

Figure 7. Localization of WT and mutated netrin-1 expressed in stable HeLa cell lines. Stable HeLa stable cell lines were grown for 24 hours in the presence of doxycycline. The experiments were replicated 3 times. Confocal microscopy (original magnification, ×63) showed that WT netrin-1 was present in the extracellular compartment, whereas the 3 mutants were mainly confined to the cytoplasm in the presence of doxycycline. None of the constructs was detected in the nucleus. Hoechst staining (blue), phalloidin (actin, red), and anti-c-Myc (netrin-1, green) were used. For each condition, asterisks are in the same place at low (third row) and high (last row) magnification. Scale bars: 30 μm in the top 3 rows and 7 μm in the bottom row.
which performs multisise and gapped alignments. Only the positions included in the bait coordinates were conserved. Genetic variation annotation was realized from the IntegraGen in-house pipeline. It consists of gene annotations (RefSeq, https://www.ncbi.nlm.nih.gov/ refseq/) and known polymorphisms (dbSNP 132, Single Nucleotide Polymorphism Database, https://www.ncbi.nlm.nih.gov/projects/SNP/). 1000 Genomes, http://www.1000genomes.org/; and EVS, http://evs.gs.washington.edu/EVS/), followed by a mutation characterization (exonic, intronic, silent, nonsense). For each position, the exonic frequencies (Homo and HTZ) were determined from all the exomes already sequenced at IntegraGen, and the exome results were provided by 1000 Genomes, EVS, and HapMap (haplotype map of the human genome, ftp://ftp.ncbi.nlm.nih.gov/hapmap/). Algorithms. The minimum average depth on the exome was approximately 50×. When coverage was below 50× for an exon and/or bp in NTN1 (which was the case for exon 6 and the first bp of exon 2 in most patients), the analysis was completed by Sanger sequencing.

Concerning the exome sequencing performed in New Zealand, genomic DNA was collected and extracted from saliva samples using Oragene DNA kits (DNA Genotek Inc.). Exome sequencing was provided by Otogenetics Corporation using Agilent SureSelect Human All Exon V4 or Agilent SureSelect Human All Exon V5+UTR for capture and Illumina sequencing platforms for data generation (paired end, 100 cycles). Sequence alignment was performed with the Burrows-Wheeler Aligner (BWA) mem using human genome assembly GRCh37 as the reference. Further processing was done with Picard tools and the Genome Analysis Toolkit following the Broad Institute’s best practices guidelines (https://www.broadinstitute.org/gatk/) to produce called variants with HaplotypeCaller. Estimates for population allele frequencies of individual variants were obtained from the National Heart, Lung, and Blood Institute’s (NHLBI) Gene Ontology (GO) Exome Sequencing Project (ESP6500) database, the 1000 Genomes Project, and the ExAC (http://exac.broadinstitute.org/) database. The predicted effect of individual variants on gene function was identified through annotation with SnpEff (58).

We then conducted the analysis with Variant Studio (Illumina). Variants present at an allele frequency greater than 0.1% in the general population (1000 Genomes and EVS databases), synonymous variants, and intronic variants that were not predicted to alter splicing were discarded. Variants with a read depth below 10× in the French exomes and 5× in the New Zealand exomes were also filtered out. A lower threshold was chosen for the New Zealand exomes, as coverage was poorer because the DNA was extracted from saliva rather than blood.

We listed all sequence variants detected in NTN1 in 60,706 controls on the ExAC algorithm. We used Fisher’s exact test to compare the frequency of truncating and rare (frequency <1%) missense variants predicted to be pathogenic by either the PolyPhen-2 or SIFT algorithm in our patients and control subjects.

Sanger sequencing and quantitative multiplex PCR of short fluorescent fragments

The coding and flanking intronic regions of NTN1 were amplified using the primers listed in Supplemental Table 6. Forward and reverse sequencing reactions were performed with the Big Dye Terminator Cycle Sequencing Ready Reaction Kit (PE Applied Biosystems), and the products were analyzed on an ABI 3730 automated sequencer (PE Applied Biosystems). Quantitative multiplex PCR of short fluorescent fragments (QMPSF) analysis was performed for all exons of NTN1 in 14 patients, and results were analyzed using GeneMapper analysis software, version 4.0 (Applied Biosystems).

Structural analysis

On the basis of the alignment obtained using the HHpred method (59), a structural model was generated using Rosetta comparative modeling software (34). The template structure used to build the model is the complement C3C structure sharing 23% of sequence identity with the NTR domain of netrin-1 (Protein Data Bank [PDB] code 2a74).

Site-directed mutagenesis

To generate human and mouse netrin-1 AP fusion proteins in the C-terminal, human and mouse NTN1 cDNAs were amplified by PCR and cloned in pAP-Tag-5 (GenHunter, no. Q202) between NheI and BglII sites. Plasmids encoding human netrin-1 and mouse netrin-1 were provided by Patrick Mehlen (Centre de Recherche en Cancérologie de Lyon [CRCL], Lyon, France). Mutations were introduced by site-directed mutagenesis (QuickChange II Site-Directed Mutagenesis Kit; Agilent Technologies) with primers (listed in Supplemental Table 7) containing the mutations and verified by Sanger sequencing. Clones were then selected and purified (ZR Plasmid Miniprep Classic from Zymo Research and NucleoBondXtra Midi/Maxi from Macherey-Nagel). We ensured that no other mutation was introduced by sequencing the entire cDNA of NTN1.

Stable cell lines

Flp-In T-Rex HeLa cells were provided by Stephen Taylor (University of Manchester, Manchester, United Kingdom) and cultured (not authenticated, tested for mycoplasma contamination with a negative result) as described previously (38), and pcDNA5.2/FRT/TO-6cMyc was provided by Alain Joliot (Collèg de France, Paris, France). NTN1-Δ20cDNA was amplified with Phusion DNA polymerase (Thermo Fisher Scientific) and cloned into pcDNA5.2/FRT/TO-6cMyc between the NheI and BsrGI sites using the NEBuilder HiFi DNA Assembly Cloning Kit (New England BioLabs). Mutations were introduced by site-directed mutagenesis using the QuikChange Lightning Site-Directed Mutagenesis Kit with primers (listed in Supplemental Table 7) containing the mutations and verified by Sanger sequencing. Plasmids pcDNA5.2/FRT/TO/NTN1-6cMyc (WT and mutants) were cotransfected with pOG44, a plasmid expressing the Flp recombinase (Invitrogen, Thermofisher Scientific), into Flp-In T-Rex HeLa cells using Lipofectamine 2000 Reagent (Invitrogen, Thermofisher Scientific) according to the manufacturer’s instructions. Cells were selected in 5 μg/ml blasticidin (Life Technologies, Thermofisher Scientific) and 250 μg/ml hygromycin B (Sigma-Aldrich). Netrin-1 expression was induced by the addition of 1 μg/ml doxycycline (Sigma-Aldrich) for 24 hours.

Immunofluorescence labeling

Twenty-four hours after induction, cells grown on coverslips were fixed with 4% (v/v) paraformaldehyde for fifteen minutes, permeabilized in 0.1% Triton X-100/PBS for ten minutes, and blocked in PBS containing 1% BSA (Sigma-Aldrich). Cells were incubated for 45 minutes with Phalloidin-TRITC (Sigma-Aldrich) to detect the cytoplasm by labeling filamentous actin and then overnight at 4°C with
monoclonal anti-c-Myc antibody (1:100; Santa Cruz Biotechnology Inc.; sc-40, RRID:AB_627268) (60) to detect netrin-1. Alexa Fluor 488 donkey anti-mouse (1:1,000; Jackson ImmunoResearch Laboratories; 715-545-150) was used as a secondary antibody. Nuclei were counterstained with Hoechst (Sigma-Aldrich). Noninduced cells were treated in the same way. Cellular localization of proteins was examined by confocal microscopy and replicated 3 times.

Western blot analysis
HEK293 cells (cell line from human embryonic kidney, Ad5 DNA transformed; ATCC, not authenticated, tested for mycoplasma contamination with a negative result) were transfected with pAPtag5, pAPtag5-netrin-1 human or pAPtag5-netrin-1 mouse, WT, and mutated plasmids using Lipofectamine 2000 Reagent (Invitrogen, Thermo Fisher Scientific) and grown for 48 hours. Cells were separated from medium by centrifugation, and total cellular proteins were extracted in lysis buffer (10 mM HEPES [pH 7], 100 mM NaCl, 2 mM EDTA, 0.5% NP-40).

For the experiments on stable lines, 24 hours after induction, supernatants were collected and centrifuged to pellet cell debris. The resulting supernatants were directly used for protein analysis or concentrated using Amicon Ultra-15 centrifugal filters (Ultracel 30K; EMD Millipore). Proteins from cells were extracted in lysis buffer (150 mM NaCl, 25 mM Tris-HCl [pH 7.5], 5% glycerol, 1% NP-40, 0.5% sodium deoxycholate, 1 mM EDTA and protease inhibitor cocktail).

The samples were then loaded on 4% to 15% Mini-PROTEAN TGX Tris-glycine buffer SDS-PAGE and transferred onto a 0.2-μm Trans-Blot Turbo nitrocellulose membrane (Bio-Rad). Membranes were blocked for 1 hour at room temperature in 1× TBS (10 mM Tris-HCl [pH 8], 150 mM NaCl) supplemented with 5% (w/v) dried skim milk powder. Primary antibody incubation was carried out overnight at 4°C. The presence of netrin-1 in total lysates and in the supernatants at the expected molecular weight was confirmed by Western blotting with anti-netrin-1 antibodies (1:500; R&D Systems; catalog MAB1109, RRID:AB_2154710) (61). Anti-actin (1:1,000; EMD Millipore; catalog MAB1501, RRID:AB_2223041) (62) and anti-α-tubulin (1:4,000; Sigma-Aldrich; catalog T5168, RRID:AB_477579) (63) antibodies were used as controls. The secondary HRP-coupled antibodies used were donkey anti-rat for detection of netrin-1, goat anti-mouse for actin (Jackson ImmunoResearch Laboratories; 712-036-153 and 115-035-003, respectively), and sheep anti-mouse for α-tubulin (GE Healthcare). In between and after antibody incubations, membranes were extensively washed in TBS containing 2.5% Tween-20 (TBS-T). Western blots were visualized using the ECL method (Pierce ECL Western Blotting Substrate, and to better detect netrin-1, the SuperSignal West Femto Maximum Sensitivity Substrate; Thermo Fisher Scientific). To exclude an effect of the AP tag on netrin-1 secretion, we removed it from human WT and C601R-mutated netrin-1–AP plasmids by BglII/ XbaI digestion. We then conducted the same experiments described above. Western blot quantification for 3 independent experiments was performed using ImageJ software (NIH).

Determination of AP activity in the supernatant
Forty-eight hours after transfection of HEK293 cells, supernatants were cleared of cells by centrifugation. Supernatants (100 μl, 1:100) were dispended in 96-well plates. Three replicates were tested for each sample. Plates were incubated for 1 hour with 100 μl AP substrate according to the manufacturer’s instructions (AttoPhos AP Fluorescent Substrate System; Promega). Fluorescence intensity was measured at excitation 430/emission 570, every 5 minutes, using TECAN Infinite M1000. Kinetics activity was calculated using GraphPad Prism 7 (GraphPad Software).

MRI
MRI data were acquired using a Siemens VERIO 3T scanner with a 32-channel head coil. The MRI protocol included anatomical 3D TI-weighted magnetization-prepared rapid gradient echo (MP RAGE) imaging (repetition time, TR = 2.3 s; echo time, TE = 4.18 ms; flip angle = 9°; inversion time, TI = 900 ms; voxel size = 1 × 1 × 1 mm³; 176 slices) and spin-echo echo-planar diffusion tensor imaging (TR = 10 s, TE = 87 ms, voxel size = 2 × 2 × 2 mm³, 60 slices, 60 gradient-encoding directions, with a b-value of 1,500 s/mm² and 11 nondiffusion-weighted volumes).

Tractography analysis
Tractography analysis was performed for 1 patient (patient 1.9) and 20 control subjects. Diffusion images were preprocessed according to previous methods (64–66) and processed for probabilistic diffusion tractography with MRtrix (http://www.mrtrix.org/). Raw diffusion-weighted data were corrected for motion and geometric distortions secondary to eddy currents using a registration technique based on the geometric model of distortions (67). The fiber orientation distribution function (ODF) was estimated using the constrained spherical deconvolution (CSD) method in Mrtrix (66, 68). The sufficient angular resolution allowed high-order fiber orientation estimation algorithms (68–70). The ODF information obtained from CSD was used with a suitable fiber-tracking algorithm to infer connectivity of crossing fibers. We used a probabilistic streamlines algorithm with the entire ODF as a probability density function (ODF threshold = 0.1; step size = 0.2 mm as 1/10 of the voxel size; radius of curvature = 1 mm; up-sampling of diffusion-weighted images [DWI], data to 1 mm). In the native individual space, we performed a seed-to-target analysis from regions of interest (ROI) defined along the CST (see ROI section in ref. 26). These regions included the anterior bundle of the CST in the upper part of the brainstem, the lower part of the brainstem, and the lateral horn of the spinal cord (see Figure 3A). We used the following probabilistic tractography algorithm: the number of fibers connecting a seed voxel to a target voxel was calculated by sampling 1 million draws for each fiber connecting the seed to the target. The CST tracts (the normally crossed CST and the abnormally uncrossed CST) were reconstructed for each subject. The normally uncrossed CST passes through the anterior medial horn of the spinal cord. After the tracts were reconstructed, 2 complementary measures were considered. First, the number of fibers reaching the target at the level of the lateral horn of the spinal cord (the crossed and uncrossed CST separately) was calculated. We analyzed the proportion of crossed versus uncrossed portions of the CST using a laterality coefficient (NF crossed – NF uncrossed)/(NF crossed + NF uncrossed) (NF, number of fibers, as in ref. 71). Ratio values closer to 1 indicated more crossed than uncrossed CST fibers, whereas ratio values closer to –1 indicated more uncrossed than crossed CST. Second, mean FA values were measured along the CST. Along-tract measures of FA were obtained on the basis of B-spline resampling of the fibers and averaging the FA values for each individ-
Electrophysiological experiments

Electrophysiological experiments were conducted in 2 patients (patients 1.9 and 2.3).

Electromyography recordings. Electromyographic (EMG) signals were recorded bilaterally from the first dorsal interosseous (FDI) muscle (active electrode over the motor point and reference electrode on the metacarpophalangeal joint), with disposable surface Ag/AgCl electrodes placed 3 cm apart on the muscle belly.

Single-pulse TMS and corticospinal excitability. MEPs induced by single monophasic pulses delivered with a TMS device connected to a figure-of-eight coil (Magstim) were recorded from EMG signals. TMS pulses were applied over the cortical representations (hot spots) of the FDI muscles in MI (primary motor cortex). The resting motor threshold was defined as the minimum stimulus intensity that resulted in MEPs of 50 μV in at least 5 of 10 trials. Thirty to sixty MEPs evoked by a calibrated stimulation (1.2 – 1.3 × resting motor threshold) of the dominant and nondominant hemispheres were recorded bilaterally in the FDI muscle to compare the frequency, latency, and amplitude of the normal contralateral MEPs with those of potential mirror MEPs recorded in the hand ipsilateral to the stimulation site.

Statistics

Data were analyzed with SPSS statistical software, version 24.0 (IBM). Normality in variable distributions and homogeneity of variances across groups were assessed with Shapiro-Wilk and Levene tests, respectively. Variables that passed the normality test were analyzed using a 1-way ANOVA followed by Bonferroni’s post-hoc test for comparisons or by a Crawford-Howell t test for comparison of patients with control subjects. A P value of less than 0.05 was used as a cutoff for statistical significance.

Study approval

In France, the study was approved by the ethics committee of the Hôpital de la Pitié-Salpêtrière and the Comités de Protection des Personnes (CPP), Ile-de-France 6, Paris. In New Zealand, all procedures were approved by the Health and Disability Ethics Committees, Multi-region Ethics Committee of New Zealand. Written informed consent was obtained from all patients (or parents of minors) before genetic analyses. All experiments were performed according to the respective approved protocols.

Author contributions

AM, AC, ID, OT, C. Dubacq, ER, and DM wrote and revised the manuscript. ER, DM, and ID designed and supervised the study. AM, EAF, OT, TCO, YZ, RJMQ, QW, SPR, CG, C. Depienne, MS, CLJ, C. Dubacq, FR, JCL, MMP, RG, JA, CF, MD, MV, ID, GAR, and AB acquired, analyzed, and interpreted data.

Acknowledgments

The authors would like to thank the patients for their participation in this study; the DNA and cell bank of the ICM (in particular Christelle Dussert and Sylvie Forlani) for DNA extraction; Stephen Taylor (University of Manchester, Manchester, United Kingdom); and Alain Joliot (Collège de France, Paris, France) for providing the reagents for the stable cell line studies; and the ExAC and other groups that provided exome variant data for comparison. The work undertaken in New Zealand was supported by a Royal Society of New Zealand Marsden Fund Grant. The work undertaken in France was supported by the Fondation Desmar est, Merz-Pharma, Elivie, the Journées de Neurologie de Langue Française (JNFL), and Djillali Mehri. This work was also supported by a grant from the Agence Nationale de la Recherche (ANR) (ANR-14-CE13-0004-01) and was performed within the framework of LABEX LIFESENSES (ANR-10-LABX-65) supported by French state funds managed by the ANR within the Investissements d’Avenir program (ANR-11-IDEX-0004-02).

Address correspondence to: Isabelle Dusart, Université Pierre et Marie Curie (UPMC), Neuroscience Paris Seine, Bat B, 6ème, Case 16, 9 Quai Saint Bernard, 75005 Paris, France. Phone: 331.44.27.21.29; Email: isabelle.dusart@upmc.fr.


33. Bányai L, Pathy L. The NTR module: domains...
RESEARCH ARTICLE