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Article

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VEGF enhances angiogenesis and promotes blood-brain barrier leakage in the ischemic brain

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VEGF is a secreted mitogen associated with angiogenesis and is also a potent vascular permeability factor. The biological role of VEGF in the ischemic brain remains unknown. This study was undertaken to investigate whether VEGF enhances cerebral microvascular perfusion and increases blood-brain barrier (BBB) leakage in the ischemic brain. Using magnetic resonance imaging (MRI), three-dimensional laser-scanning confocal microscope, and functional neurological tests, we measured the effects of administering recombinant human VEGF₁₆₅ (rhVEGF₁₆₅) on angiogenesis, functional neurological outcome, and BBB leakage in a rat model of focal cerebral embolic ischemia. Late (48 hours) administration of rhVEGF₁₆₅ to the ischemic rats enhanced angiogenesis in the ischemic penumbra and significantly improved neurological recovery. However, early postischemic (1 hour) administration of rhVEGF₁₆₅ to ischemic rats significantly increased BBB leakage, hemorrhagic transformation, and ischemic lesions. Administration of rhVEGF₁₆₅ to ischemic rats did not change BBB leakage and cerebral plasma perfusion in the contralateral hemisphere. Our results indicate that VEGF can markedly enhance angiogenesis in the ischemic brain and reduce neurological deficits during stroke recovery and that inhibition of VEGF at the acute stage of stroke may reduce the BBB permeability and the risk of hemorrhagic transformation after focal cerebral ischemia.

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Introduction

Functional neurological recovery from stroke commonly occurs, but is often incomplete in patients who survive a stroke (1, 2). Stroke patients with a higher density of blood vessels appear to have reduced morbidity and survive longer (3). Functional imaging of stroke patients shows increased cerebral blood flow and metabolism in tissue surrounding focal brain infarcts (4, 5). These data suggest that restoration of cerebral microvascular circulation is important for functional recovery after a stroke. Late restoration of cerebral microcirculatory flow may involve increases in the collateral circulation and angiogenesis in the penumbra (6, 7). The substances responsible for inducing formation of new blood vessels in the ischemic brain have not been identified conclusively.

VEGF is an angiogenic growth factor that binds to two high-affinity receptors, *fms*-like tyrosine kinase (*flt-1*) and the kinase domain region (KDR) (8, 9). Administration of recombinant human VEGF₁₆₅ (rhVEGF₁₆₅) improves myocardial perfusion in patients with coronary ischemia and enhances angiogenesis in animal models of myocardial and limb ischemia (10–14). Transient and permanent middle cerebral artery (MCA) occlusion in rats evokes expression of VEGF in the ischemic brain (15–17), suggesting that after a stroke VEGF may be involved in

angiogenesis. However, whether administration of rhVEGF₁₆₅ to the ischemic brain has the potential to promote angiogenesis and thereby improve functional neurological outcome has not been tested. In addition, VEGF is a potent vascular permeability factor that increases microvascular permeability to blood plasma proteins within minutes after exposure (18). Administration of rhVEGF₁₆₅ after focal cerebral ischemia may exacerbate blood-brain barrier (BBB) permeability. After a stroke, disruption of the BBB occurs acutely whereas regeneration of cerebral microvessels develops relatively late in ischemic brain (6, 19–24). Using a model of focal embolic cerebral ischemia in rats, we therefore tested the hypotheses that administration of VEGF at a late stage of stroke improves cerebral microvascular perfusion in the ischemic penumbra and that administration of VEGF early after a stroke increases BBB leakage.

Methods

All experimental procedures have been approved by the Care of Experimental Animals Committee of Henry Ford Health Sciences Center.

General experimental procedures. Male Wistar rats weighing 320–400 g were used in the experiments. Animals were anesthetized with 0.5–3.5% halothane in 70% N₂O

and 30% O₂ during the surgical procedure and magnetic resonance imaging (MRI) measurements. Rectal temperature was maintained at 37 ± 0.5°C throughout the experiments (25, 26). The right femoral artery and vein were cannulated with a PE-50 catheter for continuous monitoring of blood pressure, measurement of blood gases, and for drug administration, respectively.

Animal model. The MCA was occluded by placement of a single, intact, fibrin-rich, 24-hour-old, homologous clot (~1 µl) at the origin of the MCA (25). This model is most relevant to human stroke (25, 26).

Administration of rhVEGF₁₆₅. One hour or 48 hours after embolization, using a Harvard pump (Harvard Apparatus; South Natick, Massachusetts, USA), rhVEGF₁₆₅ (Genentech Inc., San Francisco, California, USA) was infused intravenously to rats at a dose of 1 mg/kg over a 4-hour interval (5 µl/min) to reduce the hemodynamic side effects of VEGF (27, 28). The dose of rhVEGF₁₆₅ was chosen based on previous studies in the same species (27, 28). An equal volume of 0.9% saline was administered to rats as a control group.

MRI measurements. MRI measurements were performed using a 3.0 T whole-body magnet (MagneX Scientific, Abingdon, United Kingdom) equipped with actively shielded gradients and a SMIS (Surrey, United Kingdom) console. MRI images were recorded using a birdcage radial frequency coil.

The apparent diffusion coefficient of water (ADC_w) was measured using the method described by LeBihan et al. (29). The diffusion-weighting gradient was increased, in a nonlinear manner, from 0 to approximately 2.8 gauss/cm to obtain a series of five images with gradient b values of 0, 200, 400, 600, 800 s/mm².

Perfusion estimates employed methods of Williams et al. that were further refined in our laboratory (30, 31). This technique is based on the selective inversion of blood-water protons at the level of the carotid arteries before ¹H MRI measurement in the brain (31). Two images were obtained for perfusion measurement with parameters: repetition time (TR) = 1.055 seconds; echo time (TE) = 30 milliseconds, 64 × 64 image matrix, 2-mm slice thickness, and a 40-mm field of view (FOV). The duration of the inversion pulse was 1 second and 0.3 kHz in amplitude.

A gadolinium-chelate (Gd-DTPA) contrast material (Omniscan Gadodiamide; Nycomed Inc., Princeton, New Jersey, USA; 0.1 mmol/kg) was injected (intravenous bolus) at the end of the infusion of rhVEGF₁₆₅ or saline to measure the BBB leakage. Contrast-enhanced MRIs were acquired in a multislice mode (nine slices) with a 2-mm slice thickness, a 40-mm FOV, and a 256 × 256 image matrix. The contrast-enhanced MRI images were obtained using multislice T₁-weighted images with spin-echo sequences (TR = 500 milliseconds, TE = 30 milliseconds).

MRI analyses of ADC_w and cerebral blood flow (CBF) maps were performed using Eigentool software (32). ADC_w maps were produced on a pixel-by-pixel basis using a least-squares fit to a straight-line plot of the

natural logarithm of normalized image intensity versus gradient b value (ADC_w). CBF maps were calculated according to the method published previously (31).

Accelerating rotarod test. An accelerating rotarod test was employed to measure motor function of rats (33, 34). Each rat received a test trial on the accelerating rotarod at 2, 7, 14, and 28 days after embolic MCA occlusion, and the time the animal remained on the device was recorded. Data are presented as the time the rat remained on the rotating spindle normalized to the preischemia duration.

Adhesive removal test. The adhesive removal test, which measures somatosensory deficit, was performed (35). Briefly, two small pieces of adhesive-backed paper dots (of equal size, 113.1 mm²) were used as bilateral tactile stimuli occupying the distal-radial region on the wrist of each forelimb. The time required to remove the stimuli from each limb was recorded. Each animal received five trials at 2, 7, 14, and 28 days after embolic MCA occlusion, and the mean time required to remove both stimuli from limbs was recorded.

Tissue preparation. For paraffin sections, brains were fixed in 10% buffered formalin and embedded in paraffin. Six-micrometer-thick coronal sections were cut and stained with hematoxylin and eosin (H&E) for histopathological evaluation. For vibratome sections, brains were fixed in 4% paraformaldehyde at 4°C for 48 hours. Coronal sections (100 µm) were cut on a vibratome.

Immunohistochemistry. A mouse mAb against microtubule-associated protein 2 (MAP2; Serotec Ltd., Oxford, United Kingdom), the neuronal cytoskeletal protein, was used as a marker for early ischemic neuronal damage (36). For immunofluorescence, 100-µm thick vibratome sections were incubated with primary mAb's against MAP2 (1:200) and with Cy 5-conjugated secondary Ab's.

Three-dimensional image acquisition. The vibratome sections were analyzed with a Bio-Rad MRC 1024 (argon and krypton) laser-scanning confocal imaging system (Bio-Rad, Cambridge, Massachusetts, USA). With the Evans blue- and FITC-perfused tissue samples, vibratome sections corresponding to the MRI coronal sections (interaural 6.20 mm to interaural 1.00 mm) (37) were scanned throughout the entire coronal section in 512 × 512 pixel (1042.4 × 1042.4 µm) format in the x-y direction using a 4× frame-scan average, and 16 optical sections along the z-axis with a 5-µm step-size under a ×10 objective were acquired with two separate photomultiplier tubes. Each image was assembled into a coronal section using Adobe Photoshop software (Adobe Systems Inc., Mountain View, California, USA). For FITC-dextran-perfused tissue stained with anti-MAP2 Ab (Cy5), green (FITC-dextran-perfused microvessels), and far red (Cy5, MAP2 immunoreactivity) fluorochromes on the sections were excited by a laser beam at 488 nm and 647 nm, respectively, and emissions were acquired simultaneously with two separate photomultiplier tubes through 522-nm and 680-nm emission filters, respectively.

Three-dimensional image analysis. Measurements of vascular diameters are limited in two dimensions and are based on the assumption that vessel contours are round (38, 39). However, this is not the case in vivo in which cerebral microangiarchitecture has a highly irregular pattern of tortuous, anastomosing capillaries. To overcome most limitations of previous methods, we developed a new computerized system, MIRAGE, to examine cerebral microvessels perfused in vivo by FITC-dextran (K. Davies et al., manuscript submitted for publication). The methodology used by MIRAGE for measurement of blood vessel diameter involves a linearly applied spherical inflation technique. For each vessel under analysis, the system follows the center of the vessel and determines the long-axis path of flow. Vessel diameter is continuously measured as the center of the vessel is traced by using the length of 24 different divergent vectors, each perpendicular to this axis of flow. In a preliminary study, we measured 8,930 microvascular diameters in a nonischemic rat cortex, and the average microvessel diameter was $4.15 \pm 1.24 \mu\text{m}$ (3–8 μm), which is comparable to cortical microvessel diameters of $4.4 \pm 1 \mu\text{m}$ (2.5–8.7 μm) obtained from intravital microscopy in vivo (39), $4.26 \pm 0.07 \mu\text{m}$ from manually measured fixed coronal sections stained with toluidine blue (38), and 3–8 μm measured by scanning electron microscopy on corrosion casts (40). We therefore employed this program to analyze cerebral microvessels in the present study. Images obtained under a $\times 40$ objective from a laser-scanning confocal microscope (LSCM) had a resolution of $0.5 \mu\text{m}/\text{pixel}$ in the x and y dimensions and $1 \mu\text{m}/\text{pixel}$ along the z axis. All quantitative measurements were performed blindly.

Statistical analysis. Repeated measures analysis was conducted using a generalized estimating equation to test the treatment effect on the rotarod and the adhesive removal tests, respectively. Student's *t* tests were performed to compare cerebral microvascular plasma perfusion and vascular diameters in control and VEGF-treated groups. A paired *t* test was used to analyze the effects of VEGF and saline on measurement of perfusion- and diffusion-weighted MRIs. All data are presented as mean plus or minus SE, and a *P* value of 0.05 was considered to be statistically significant.

Results

The effects of administration of rhVEGF₁₆₅ on cerebral microvascular perfusion and neurological outcome. To determine whether administration of rhVEGF₁₆₅ enhances cerebral microvascular perfusion, embolic ischemic rats were treated with rhVEGF₁₆₅ at a dose of 1 mg/kg 48 hours after a stroke to avoid the peak of brain edema in this stroke model and were sacrificed at 9 days after ischemia. The time point of 9 days after ischemia was chosen based on data from a previous study using a cranial window: newly formed vessels arising from mouse brain were first visible at 4 days and peaked at 12 days after VEGF administration (41). Cerebral microvascular plasma perfusion and vascular diame-

ters were examined in three dimensions by LSCM and analyzed using the MIRAGE program. Compared with the saline-treated rats (*n* = 6, Figure 1, a and g), treatment with rhVEGF₁₆₅ significantly (*P* < 0.05, *n* = 5) increased numbers of voxels of plasma-perfused vessels

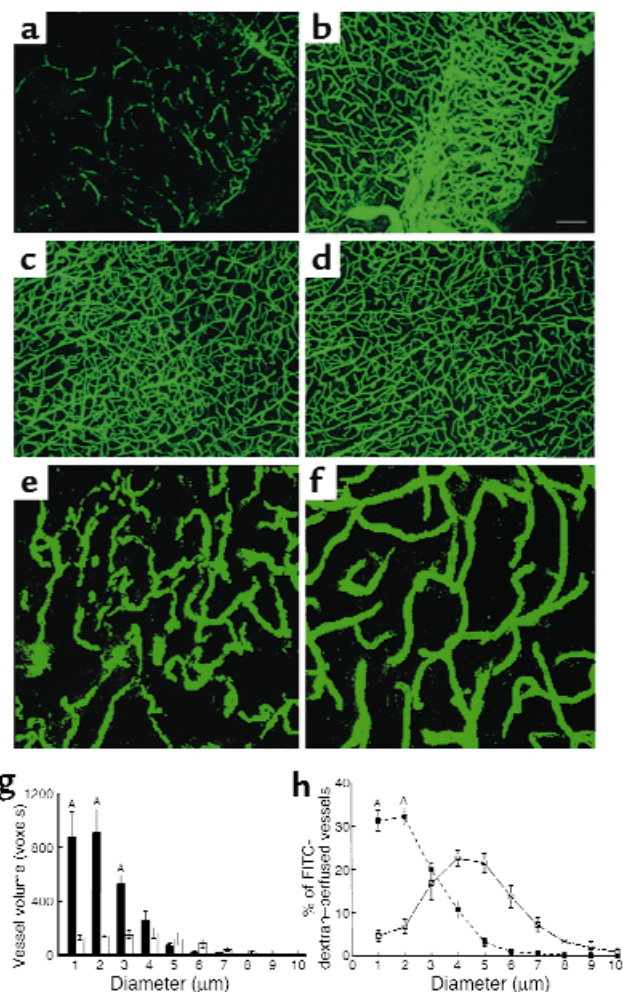


Figure 1

Composite images (through stacks of 14 optical sections of 5- μm thickness) of cerebral microvascular plasma perfusion obtained from the ischemic penumbra of the cortex 9 days after MCA occlusion. Late treatment with rhVEGF₁₆₅ enhances cerebral microvascular plasma perfusion in the penumbra (b) compared with the control rat (a). Treatment with rhVEGF₁₆₅ did not alter cerebral microvascular plasma perfusion in the contralateral hemisphere (d), compared with the contralateral hemisphere in the control rat (c). Bar, 100 μm . Quantitative data analyzed by the MIRAGE program (g) show a significant increase of plasma perfusion in vessels within 3 μm of diameters in the rhVEGF₁₆₅-treated group (filled bars; *n* = 5) compared with the saline-treated group (open bars; *n* = 6; ^A*P* < 0.05). Higher-magnification images show that cerebral microvessels in the ipsilateral hemisphere had an irregular pattern of tortuosity (e) compared with the vessels in the contralateral hemisphere (f). Dimension of the composite image is $260.6 \times 260.6 \times 20 \mu\text{m}^3$. (h) The percentage of vessels with different diameters in the entire vessels perfused by FITC-dextran. The number of vessels with diameters within 2 μm is significantly (*P* < 0.01) higher in the ipsilateral hemisphere (filled squares; *n* = 5) than in the contralateral hemisphere (open squares; *n* = 5).

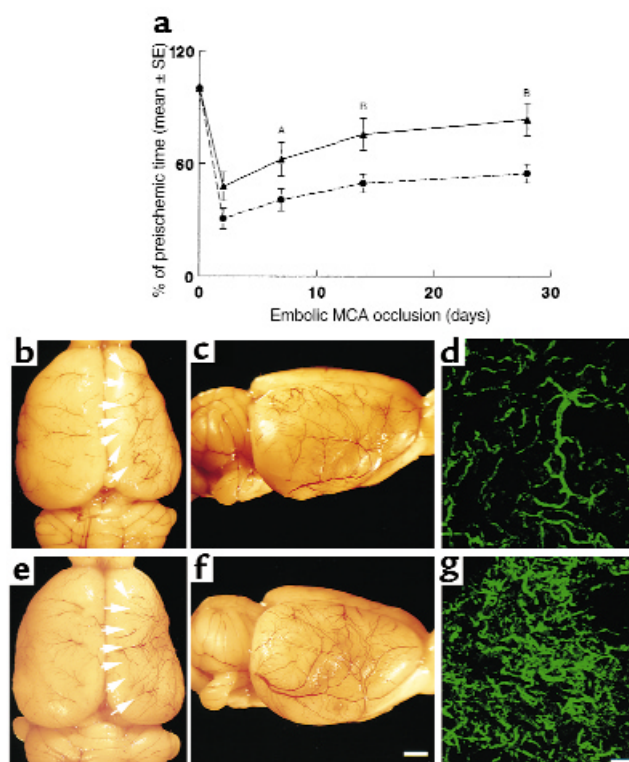


Figure 2

(a) The percentage of time that rats persisted on the rotarod after ischemia compared with the preischemic value as a function of time of stroke. Rotarod test was performed at 2, 7, 14, and 28 days after stroke on the ischemic rats treated with rhVEGF₁₆₅ or saline initiated at 48 hours after ischemia. Treatment with rhVEGF₁₆₅ (triangles; $n = 7$) significantly (^A $P < 0.05$ or ^B $P < 0.01$) increases time spent on the rotarod compared with the saline-treated group (circles; $n = 10$). Cerebral vessels and FITC-dextran-perfused cerebral microvessels from the saline-treated rat (b–d) and the rhVEGF₁₆₅-treated rat (e–g) 28 days after the right MCA occlusion. Dorsal view of the right and left hemisphere vessels (b and e), a lateral view of the right hemisphere vessels (c and f), and FITC-dextran-perfused cerebral microvessels in the penumbra of the cortex (d and g). More vessels in the ipsilateral ischemic border zone (e, arrows, and f) and more FITC-dextran-perfused vessels (g) were observed in the rhVEGF₁₆₅-treated rat compared with that in the saline-treated rat (b–d). Bar = 2 mm in f for b, c, e, and f; bar = 200 μ m in g for d and g. Rats were decapitated one minute after injection of FITC-dextran and their brains were rapidly removed from the severed heads and digitized at 8 \times magnification (for panels b, c, e, and f) using an MCID image analysis system (Imaging Research, St. Catharines, Canada). After digitization the brains were placed in 4% paraformaldehyde at 4°C for 48 hours (for panels d and g).

in the penumbra of the cortex (Figure 1b and 1g). However, treatment with rhVEGF₁₆₅ did not alter cerebral microvascular plasma perfusion in the contralateral hemisphere (Figure 1d) compared with the contralateral hemisphere from the saline-treated rats (Figure 1c). We measured in three dimensions the diameters of plasma-perfused cerebral microvessels in the penumbra (Figure 1e) and in the contralateral homologous tissue (Figure 1f). Eighty-five percent of plasma-perfused vessels in the penumbra had diameters within 3

μ m, whereas the majority of vessels in the contralateral tissue had diameters of 4 and 5 μ m (Figure 1h), suggesting that increased plasma-perfused vessels in the penumbra are newly grown vessels.

To determine whether late administration of rhVEGF₁₆₅ improves functional neurological recovery, embolic ischemic rats were treated with rhVEGF₁₆₅ at a dose of 1 mg/kg ($n = 7$) or saline ($n = 10$) at 48 hours after a stroke, and the rotarod and adhesive removal tests were performed on these rats at 2, 7, 14, and 28 days after stroke. To avert potential effects of rhVEGF₁₆₅ on the rotarod test due to the likelihood of hindlimb ischemia caused by cannulated femoral arteries and veins, rhVEGF₁₆₅ or saline was administered in a tail vein, and mean artery blood pressure was not measured during infusion of rhVEGF₁₆₅ or saline in these groups. Rats in the rhVEGF₁₆₅ or the saline-treated groups showed marked motor and somatosensory impairments 2 days after a stroke. Partial recovery was detected in the rhVEGF₁₆₅ and saline-treated groups at 7, 14, and 28 days after a stroke (Figure 2a). However, the ischemic rats treated with rhVEGF₁₆₅ exhibited significant ($P < 0.05$) improvements on the rotarod test at 7, 14, and 28 days after the stroke compared with the saline-treated rats (Figure 2a). The adhesive removal test did not reveal significant differences between the rhVEGF₁₆₅-treated and the saline-treated groups at 2 days (116 ± 3.6 vs. 115 ± 3.5 seconds), 7 days (66 ± 16.5 vs. 65.2 ± 11.1 seconds), and 14 days (40 ± 1.4 vs. 34.8 ± 3.2 seconds). However, a significant ($P < 0.05$) difference was detected at 28 days after stroke (11 ± 1.2 vs. 15 ± 1.1 seconds). In addition, brain tissue obtained at 28 days after ischemia exhibited more small vessels in the ipsilateral ischemic border zone (Figure 2, e and f) in rats treated with rhVEGF₁₆₅ when compared with the vehicle-treated rats (Figure 2, b and c). An increase in microvessels in the ischemic border zone was confirmed by detecting more FITC-dextran-perfused microvessels (Figure 2g) in the rats treated with rhVEGF₁₆₅ when compared with FITC-dextran-perfused microvessels in the penumbra from the saline-treated rats (Figure 2d).

The effects of rhVEGF₁₆₅ administration on BBB leakage. To examine early effects of rhVEGF₁₆₅ administration initiated 1 hour after MCA occlusion on BBB leakage, the MRI contrast agent Gd-DTPA, along with 1 ml 2% of Evans blue, were administered to the ischemic rats at the end of the 4-hour rhVEGF₁₆₅ ($n = 5$) or saline ($n = 4$) infusion, and FITC dextran (42) was administered intravenously at the end of the MRI measurements. These rats were sacrificed 1 minute after injection of FITC-dextran. Contrast agent Gd-DTPA within the tissue causes hyperintensity on T₂-weighted MRI and reflects a breakdown of the BBB (43, 44). Coadministration of Evans blue, which binds to albumin in plasma with Gd-DTPA contrast agent, permits comparison of changes in BBB leakage measured by MRI with disruption of BBB detected by LSCM; FITC-dextran, which remains dissolved and free in plasma, allows analysis of microvascular plasma perfusion (42). In the saline-treat-

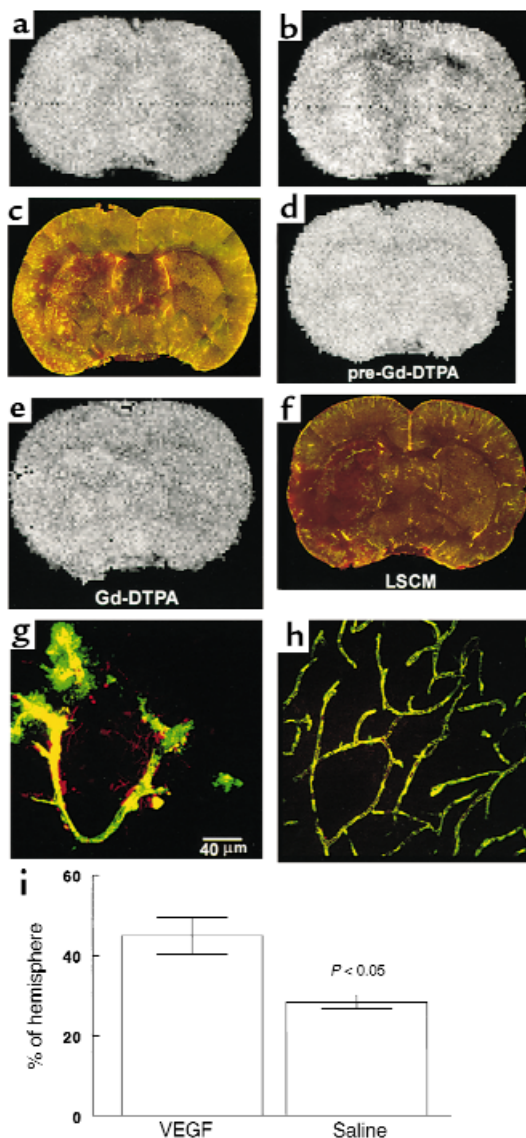


Figure 3

Gd-DTPA contrast-enhanced MRI images of ischemic brains treated with rhVEGF₁₆₅ at 1 hour after MCA occlusion (**a** and **b**) or 0.9% saline (**d** and **e**) and corresponding images of LSCM (**c** and **f**). Postcontrast images obtained at 30 minutes after injection of Gd-DTPA agent show the hyperintense areas in the ipsilateral MCA-supplied cortex and subcortex of a rat treated with rhVEGF₁₆₅ (**b**) and in the ipsilateral MCA-supplied subcortex of a rat treated with 0.9% saline (**e**), which are not seen on the precontrast images (**a** and **d**). Composite images of LSCM (80 μ m thick) from the same locations exhibit extravasation of Evans blue and FITC-dextran in the cortex and subcortex from the rat treated with rhVEGF₁₆₅ (**c**) and extravasation of Evans blue in the subcortex from the rat treated with saline (**f**) and sacrificed 6 hours after MCA occlusion. High-magnification images of LSCM from the rat treated with rhVEGF₁₆₅ show that the extravasation of Evans blue (**g**, red) and FITC-dextran (**g**, green) was present in capillaries with nonplasma and low-plasma perfusion in the ipsilateral hemisphere (**g**) and that cerebral microvessels in the contralateral hemisphere were well perfused with both dyes and did not exhibit any extravasation of fluorescent dyes (**h**). Bar, 40 μ m for **g** and **h**. Hyperintense areas on Gd-DTPA-enhanced MRI are significantly ($P < 0.05$) larger in rhVEGF₁₆₅-treated rats ($n = 5$) than the saline-treated rats ($n = 4$) (**i**). Hyperintense areas were measured on Gd-DTPA-enhanced images obtained at 30 minutes after injection of Gd-DTPA agent, which was injected after 4 hours of infusion of rhVEGF₁₆₅ or saline.

ed ischemic rats, the hyperintense area localized to the ipsilateral MCA supplied the subcortex (Figure 3e). Administration of rhVEGF₁₆₅ resulted in the hyperintense areas localized to the ipsilateral MCA-supplied cortex and subcortex (Figure 3b). The hyperintense areas on the MRI in VEGF-treated rats were significantly ($P < 0.05$) larger than the areas in the control animals (Figure 3i). LSCM images of coronal sections that matched MRI sections from the same animal revealed extensive Evans blue and FITC-dextran extravasation in rats treated with rhVEGF₁₆₅ (Figure 3c), compared with the control animals (Figure 3f). Areas with leakage of fluorescent dyes localized only to the ipsilateral area supplied by the MCA (Figure 3c) and corresponded closely to hyperintense areas detected on Gd-DTPA MRI (Figure 3b). High-magnification images of LSCM show that the extravasation of Evans blue and FITC-dextran occurred in capillaries and in the lesion boundary areas with nonplasma and low-plasma perfusion (Figure 3g), and extravasation of fluorescent dyes was not detected in cerebral vessels in the contralateral homologous tissue (Figure 3h). These data demonstrate that administration of rhVEGF₁₆₅ exacerbates BBB leakage. Disruption of the BBB was not detected in nonischemic rats treated with rhVEGF₁₆₅ ($n = 3$, data not shown).

To examine the effect of exogenous VEGF on BBB leakage and the incidence of hemorrhagic transformation, rats were sacrificed at 24 hours after a stroke, since in this embolic stroke model hemorrhagic transformation primarily appears during this time period (25, 45). Administration of rhVEGF₁₆₅ at the onset of 1 hour of MCA occlusion ($n = 6$) resulted in a significant ($P < 0.05$) increase in the incidence of gross hemorrhage (four out of six rats had gross hemorrhage,

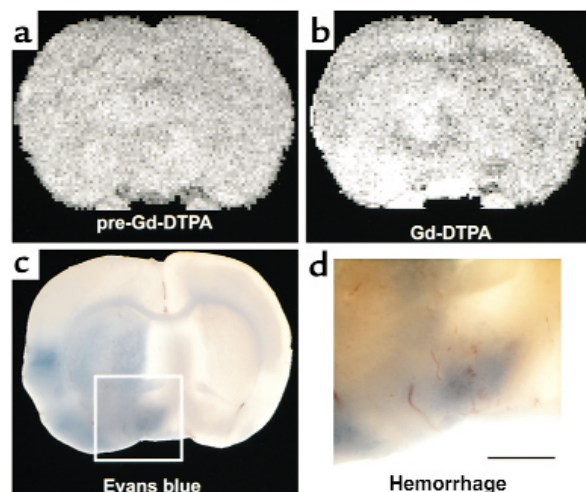


Figure 4

Increase in the BBB leakage and hemorrhagic transformation. Large hyperintense areas on Gd-DTPA-enhanced MRI were observed 30 minutes after injection of Gd-DTPA in a rat treated with rhVEGF₁₆₅ (**b**), compared with the precontrast image (**a**). A coronal section corresponding to the MRI section obtained at 24 hours after a stroke shows extensive Evans blue leakage and hemorrhage (**c** and **d**). Bar, 1 mm in **d**.

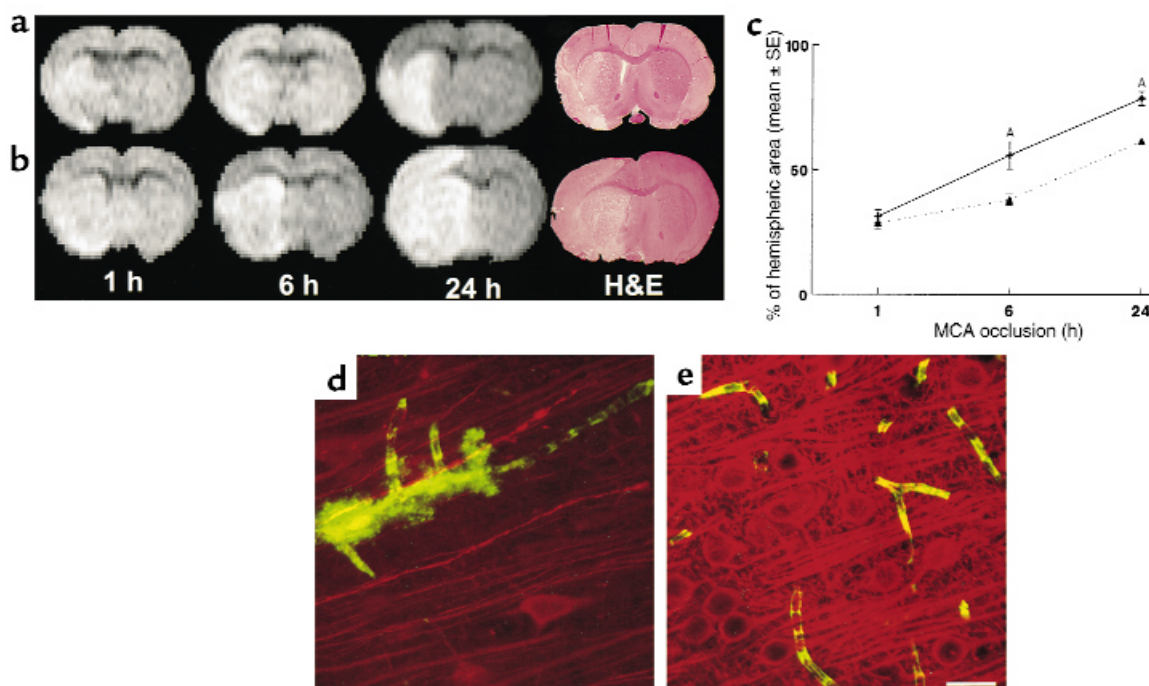


Figure 5

Evolution of the ischemic lesion from 1 hour to 24 hours after MCA occlusion in a rat treated with 0.9% saline (**a**) or rhVEGF₁₆₅ (**b**). Although areas of hyperintensity on the MRI were relatively similar at 1 hour of MCA occlusion in saline-treated (**a**) or rhVEGF₁₆₅-treated (**b**) rat, an area of hyperintensity on MRI was larger at 6 hours and 24 hours of ischemia in the rhVEGF-treated (**b**) rat compared with an area of hyperintensity at the same time points in the saline-treated rat (**a**). Differences of ischemic lesions observed on MRI between these two rats are confirmed on H&E-stained coronal sections obtained at 24 hours of ischemia (H&E) (**a** and **b**). Quantitative analysis of the ischemic area as a percentage of hemisphere measured on DWI at 6 hours and 24 hours of MCA occlusion (**c**) shows that ischemic areas are significantly ($P < 0.05$) larger in rats treated with rhVEGF₁₆₅ (solid line, $n = 6$) than in the control rats (dotted line, $n = 7$) at 6 hours and 24 hours of embolic MCA occlusion. Composite images ($135 \times 135 \times 20 \mu\text{m}^3$) of microvessels (FITC-dextran, green) and MAP2 immunoreactivity (Cy5, red) show much less MAP2 immunoreactivity (red) in low plasma-perfused areas with extravasation of FITC-dextran (green) in the ipsilateral hemisphere (**d**) compared with the homologous tissue in the contralateral hemisphere (**e**) in a rat treated with rhVEGF₁₆₅ (1 hour) and sacrificed at 6 hours after MCA occlusion. Bar, $20 \mu\text{m}$ for **d** and **e**.

80%) compared with non-rhVEGF₁₆₅-treated rats (two out of ten, 20%; $n = 10$). Areas with gross hemorrhage detected on the coronal sections at 24 hours after a stroke (Figure 4, c and d) match the anatomic hyperintense areas detected on Gd-DTPA-enhanced MRI measurements at 5 hours after a stroke (Figure 4b), suggesting that the early increase in the BBB leakage to large molecules may be related to the incidence of hemorrhage in this model.

To assess the effects of rhVEGF₁₆₅ given 1 hour after onset of ischemia on the evolution of the tissue damage and ultimate lesion volume, we performed diffusion-weighted MRIs to measure the evolution of the ischemic lesion in vivo, MAP2 immunohistochemistry to examine neuronal damage, and H&E staining to measure ischemic areas. The ischemic areas defined as the hyperintensity on the diffusion-weighted images (DWI) were relatively similar at 1 hour of ischemia for rats treated with saline (Figure 5a, 1 hour) or with rhVEGF₁₆₅ (Figure 5b, 1 hour), suggesting the same basal level of ischemic severity for rats in both groups. However, areas of hyperintensity in rhVEGF₁₆₅-treated rats increased substantially during and after infusion of

rhVEGF₁₆₅ (Figure 5b, 6 hours and 24 hours), compared with the vehicle-treated ischemic rats at the same time points (Figure 5a, 6 hours and 24 hours). Brain tissue obtained from the same animal immediately after the last MRI measurements at 24 hours after ischemia, shows a larger ischemic lesion on H&E-stained coronal section in the rhVEGF₁₆₅-treated rat (Figure 5b, H&E) than the ischemic lesion in the vehicle-treated rat (Figure 5a, H&E). Quantitative measurements of the ischemic areas defined as the hyperintensity on the DWI as a percentage of hemisphere reveal that the ischemic areas increased by 78% at 6 hours and 151% at 24 hours after ischemia compared with the ischemic areas at 1 hour after ischemia for rats treated with rhVEGF₁₆₅ ($n = 6$, Figure 5c). In contrast, the ischemic areas increased only by 32% at 6 hours and 114% at 24 hours of ischemia for rats treated with the vehicle ($n = 7$, Figure 5c). The differences of the increased ischemic areas between two groups at 6 hours and 24 hours of ischemia are statistically significant ($P < 0.05$; Figure 5c). Neurons in areas with extravasation of FITC-dextran (green, Figure 5d) exhibited much less MAP2 immunoreactivity (red, Figure 5d) at 6 hours of

ischemia compared with homologous tissue in the contralateral hemisphere (Figure 5e) for rats treated with rhVEGF₁₆₅, indicating acute ischemic neuronal damage. Taken together, these data suggest that exogenous VEGF accelerates ischemic damage.

To examine whether administration of rhVEGF₁₆₅ 48 hours after ischemia increased leakage of Gd-DTPA across the BBB, Gd-DTPA-enhanced MRI measurements were made on a subset of rats ($n = 6$) immediately after infusion of rhVEGF₁₆₅. These rats were sacrificed after the last MRI measurements, and cerebral microvascular leakage and plasma perfusion were measured by LSCM on these brains. Late administration of rhVEGF₁₆₅ did not result in a significant increase in the BBB leakage measured on Gd-DTPA-enhanced MRIs (Figure 6, a and b) and on LSCM (Figure 6c).

The effects of rhVEGF₁₆₅ administration on CBF. To examine the effects of early administration of rhVEGF₁₆₅ on CBF, we performed perfusion-weighted MRI measurements using an arterial spin-tagging technique (31). CBF increased in the ischemic hemisphere during the 1–3 hour infusion of rhVEGF₁₆₅ (Figure 7a, 2 hours) and returned to the preinfusion level at 4 hours of infusion of rhVEGF₁₆₅ (Figure 7a, 4 hours). Transient increases in CBF were not detected in the ischemic hemisphere in the control rats (Figure 7b), and the differences of changes in CBF between the VEGF ($n = 6$) and the control groups ($n = 6$) were statistically significant ($P < 0.05$; Figure 7d), suggesting that early administration of rhVEGF₁₆₅ induces a transient increase of CBF in the ischemic brain.

Hyperemia was detected by a perfusion-weighted MRI in the ischemic lesion at 48 hours after the stroke before infusion of rhVEGF₁₆₅ (Figure 7c). Administration of rhVEGF₁₆₅ at 48 hours after ischemia induced changes in the distribution of CBF within the ischemic lesion (Figure 7c). CBF was reduced in hyperemic areas during and after infusion of rhVEGF₁₆₅ (Figure 7c), suggesting that exogenous VEGF at 48 hours after ischemia causes a redistribution of CBF within the ischemic lesion.

Discussion

The present study demonstrates that rhVEGF₁₆₅ significantly enhances cerebral microvascular plasma perfusion and improves functional neurological recovery when rhVEGF₁₆₅ is administered to ischemic rats at 48 hours after a stroke. However, administration of rhVEGF₁₆₅ at 1 hour after stroke exacerbates BBB leakage, increases hemorrhagic transformation, and promotes ischemic cell damage. These findings provide the first evidence of a dual role of VEGF for enhancing cerebral microvascular perfusion and for increasing the BBB leakage in the ischemic brain.

Studies from human and experimental strokes indicate that angiogenesis is present in the focal ischemic brain (3, 6, 46). Development of angiogenesis in the ischemic brain is incompletely understood. The spatial and temporal profiles of VEGF and its receptor mRNA correspond closely with angiogenesis during embryonic development in the mouse brain, suggesting that VEGF and its receptors regulate brain angiogenesis (47, 48). Our data show that late treatment of ischemic rats with rhVEGF₁₆₅ resulted in a significant increase of cerebral microvascular plasma perfusion in the penumbra of the cortex. A significant increase in microvascular plasma perfusion may be attributed to newly formed capillaries, based on the morphology in which diameters ($\leq 3 \mu\text{m}$) of plasma-perfused vessels in the penumbra are significantly smaller than diameters ($5 \mu\text{m}$) in the contralateral homologous areas (49). In addition, microvessels in the ipsilateral border of ischemia in the VEGF-treated rats exceeded that observed in the saline-treated rats at 28 days after ischemia (Figure 2, b–g). Taken together, our data indicate that late treatment with rhVEGF₁₆₅ enhances angiogenesis in ischemic brain. Our observation is consistent with results obtained from a phase-1 trial, where administration of rhVEGF to patients with severe coronal artery disease significantly increased myocardial perfusion (10).

Our data also demonstrate that late administration of rhVEGF₁₆₅ to ischemic rats significantly improves neurological function. The rotarod test is a sensitive index

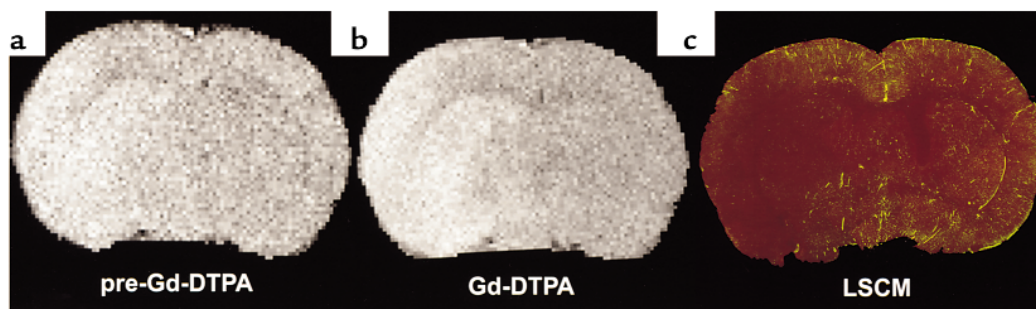


Figure 6

Late (48 hours) administration of rhVEGF₁₆₅ to ischemic rats did not result in increases in BBB leakage. Postcontrast images (b) obtained at 30 minutes after injection of the Gd-DTPA agent did not show an increase in areas of hyperintensity compared with the precontrast image (a) in a rat treated with rhVEGF₁₆₅ at 48 hours of MCA occlusion. Non- and low cerebral microvascular plasma perfusion was detected in the ipsilateral MCA-supplied territory on a composite image of LSCM (80 μm thickness) (c). However, extravasation of Evans blue and FITC-dextran was not detected (c). Images of LSCM were obtained immediately after the last Gd-DTPA-enhanced MRI measurements.

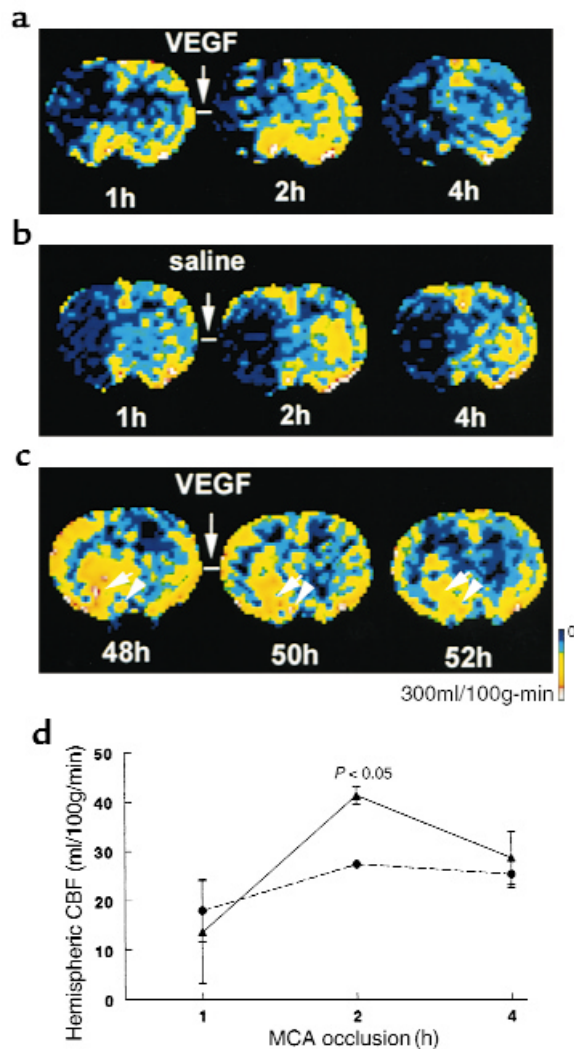


Figure 7

CBF maps of coronal sections measured by perfusion-weighted MRIs. Images of CBF were obtained before, during, and at the end of infusion of rhVEGF₁₆₅ (a) and saline (b) initiated at 1 hour after embolization. Increase of CBF was observed in the ischemic lesion during infusion of rhVEGF₁₆₅ (2 hours) (a). The increase in CBF in the rhVEGF₁₆₅-treated rats (triangles; $n = 6$) (d) was significant ($P < 0.05$) compared with that in the saline-treated animals (circles; $n = 6$). Images of CBF were obtained before, during, and at the end of infusion of rhVEGF₁₆₅ initiated at 48 hours after embolization (c). Hyperemia was evident in the ischemic lesion before infusion of rhVEGF₁₆₅ (48 hours) (c). Infusion of rhVEGF₁₆₅ decreases hyperemic areas in the ischemic lesion (50 hours and 52 hours) (c).

for assessing motor impairment after traumatic brain injury and focal cerebral ischemia (33, 34). In the present study, rats exhibited marked motor impairment at 2 days after a stroke, and partial recovery on the rotarod test was detected in the vehicle or the VEGF-treated groups at 7 and 28 days after a stroke, which is consistent with the fact that partial functional neurological recovery from a stroke occurs commonly (1, 2). However, VEGF-treated rats show a significant improvement on the rotarod test compared with vehicle-treated animals. An early significant improvement of the rotarod

test (which primarily detects motor impairment in the cortex), but late modest improvement on the adhesive removal test (which detects somatosensory impairment in the cortex and the striatum that is the ischemic core), is consistent with a significant increase in plasma perfusion in the penumbra of the cortex, suggesting that exogenous VEGF may promote therapeutic angiogenesis in the ischemic rats (49). Our data are consistent with findings that functional imaging of stroke patients shows increased cerebral blood flow and metabolism in tissue surrounding focal brain infarcts (4, 5) and that stroke patients with a higher blood vessel density in the ischemic penumbra appear to have better progress and longer survival (3), indicating that restoration of cerebral microvascular circulation is important in the ischemic brain for functional recovery after a stroke. Therefore, treatment with VEGF to potentiate new vessel formation may be an important therapy for functional neurological recovery during the repair process after a stroke.

Mechanisms by which exogenously administered VEGF enhances cerebral microvascular perfusion and improves functional neurological outcome cannot be deduced from the present study. Increases in microvascular perfusion may be attributed to angiogenesis. Regeneration of cerebral microvessels requires a vascular basement membrane upon which endothelial cells migrate and proliferate to reform vascular channels after brain injury (50). VEGF is a specific mitogen for endothelial cells (8). VEGF may bind to the vascular basement membrane and cause proliferation and migration of endothelial cells (51, 52). Angiogenesis may promote ischemic brain plasticity and thereby improve functional neurological outcome (53). In addition to its effect on angiogenesis, VEGF may have direct effects on neuronal plasticity by its neurotrophic activity and by stimulating axonal outgrowth (54–56).

Based on the acute appearance of disruption of the BBB and early upregulation of VEGF in the ischemic core (our unpublished observation), we tested the hypothesis that an early increase of endogenous VEGF increases BBB leakage. We administered rhVEGF₁₆₅ to the ischemic rats at 1 hour after onset of MCA occlusion and found a significant increase in BBB leakage in the ischemic lesion, but not in nonischemic areas, as measured by Gd-DTPA-enhanced MRI in the ischemic rats treated with rhVEGF₁₆₅ compared with the saline treated ischemic rats. An increase in BBB leakage was further confirmed by LSCM. Furthermore, intravenous administration of rhVEGF₁₆₅ did not increase BBB leakage in nonischemic rats. These findings are consistent with and extend previous studies (57–60). Intracarotid infusion of VEGF to nonischemic rats did not increase extravasation of FITC and ¹²⁵I-BSA from normal brain microvessels (59). However, topical application of VEGF to the parietal cortex in nonischemic rats resulted in increases of BBB leakage to FITC-labeled dextran of molecular mass 10,000 d (58). Studies in vitro show that when it was applied to the abluminal plasma membrane of the endothelial cells, but not to luminal side, VEGF

increased permeability of cultured brain microvessel endothelial cells (58, 60, 61). These data indicate that the receptors for VEGF are located on the abluminal side of vessels (58, 60, 61). However, our findings extend previous observations by demonstrating that intravenous administration of VEGF increases BBB leakage only in the ischemic lesion. An acute induction of VEGF receptors (Flt-1 and Flk-1) in the ischemic vessels and upregulation of neuropilin-1 (our unpublished observation) (62) may be responsible for the effect of exogenous VEGF on increasing BBB leakage only within the ischemic lesion. In contrast to early administration, late administration of VEGF does not increase the BBB leakage in the ischemic lesion, suggesting that there are some molecules that may inhibit VEGF-induced BBB leakage. Angiopoietin-1 has been demonstrated to counter peripheral vascular leakage induced by VEGF (63, 64). Expression of angiopoietin-1 is upregulated in this model (our unpublished observation). Future experiments to examine whether upregulation of angiopoietin-1 blocks ischemic cerebral vascular plasma leakage induced by VEGF are warranted.

Disruption of BBB was concomitant with increases in hemorrhagic transformation and ischemic lesions in rats treated with VEGF, suggesting that the early increase in the BBB leakage to large molecules causes subsequent hemorrhage and promotes ischemic cell damage. Overexpression of VEGF₁₆₅ causes tumor-associated intracranial hemorrhage in mice (65). Furthermore, astrocytes play an important role in BBB integrity (66). Early upregulation of endogenous VEGF on astrocytes in the ischemic core may interact with receptors for VEGF on the ischemic vessels and contribute to the disruption of BBB leakage, thereby promoting hemorrhagic transformation and ischemic cell damage. Indeed, a recent study shows that administration of soluble VEGF-receptor chimeric protein Flt-1(1-3)-IgG, inactivating VEGF to transient MCA-occluded mice, significantly reduced ischemic lesion volume measured by T₂-weighted MRI (67). In contrast, local application of VEGF to the ischemic rats subjected to 90 minutes of transient MCA occlusion resulted in a reduction of infarct volume and brain edema (68).

Mechanisms that contribute to increase of leakage of the BBB in response to VEGF administration are unknown. Some effects of exogenous VEGF on endothelial cells in vitro are partially attributed to the generation of nitric oxide (NO) from these cells (69, 70). In vivo, NO increases CBF (71). In the present study, a transient augmentation and redistribution of CBF were observed in the ischemic lesion after early and late administration of VEGF, respectively, suggesting that exogenous VEGF generates NO in ischemic brain. Therefore, a significant increase in BBB leakage after administration of rhVEGF₁₆₅ may result, in part, from action of NO, as others have demonstrated that exogenous VEGF administration increases permeability of the BBB through a NO synthase/cGMP-dependent pathway (57). In addition to generating NO, VEGF

acutely induces microvascular hyperpermeability through functional activation of vesicular-vacuolar organelles in the cytoplasm of endothelial cells (18, 72). In contrast to many vesicular-vacuolar organelles in endothelial cells of peripheral tissues, endothelial cells in brain capillaries contain few vesicular-vacuolar organelles under physiological conditions (73, 74). However, cerebral ischemia and brain trauma increase endothelial vesicles and vacuoles in cerebral capillaries (73, 74), which may contribute to increased leakage of the BBB in response to VEGF.

In summary, our data demonstrate that late administration of VEGF enhances angiogenesis in the ischemic brain, improving neurological recovery, and that early administration of VEGF exacerbates BBB leakage. This suggests a potentially effective therapeutic strategy aimed at administration of exogenous VEGF to promote therapeutic angiogenesis during the repair process after a stroke and inhibition of VEGF at the acute stage of stroke to reduce the BBB permeability and the risk of hemorrhagic transformation after cerebral ischemia.

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1. Kawamata, T.S.E., and Finklestein, S.P. 1997. The role of polypeptide growth factors in recovery from stroke. In *Brain plasticity*. H.J. Freund, B.A. Sabel, and O.W. Witte, editors. Lippincott-Raven. Philadelphia, Pennsylvania, USA. 377–382.
2. Pons, T.P. 1998. Reorganizing the brain. *Nat. Med.* 4:561–562.
3. Krupinski, J., Kaluza, J., Kumar, P., Kumar, S., and Wang, J.M. 1994. Role of angiogenesis in patients with cerebral ischemic stroke. *Stroke*. 25:1794–1798.
4. Weiller, C., Ramsay, S.C., Wise, R.J., Friston, K.J., and Frackowiak, R.S. 1993. Individual patterns of functional reorganization in the human cerebral cortex after capsular infarction. *Ann. Neurol.* 33:181–189.
5. Cramer, S.C., et al. 1997. A functional MRI study of subjects recovered from hemiparetic stroke. *Stroke*. 28:2518–2527.
6. Sbarbati, A., et al. 1996. The microvascular system in ischemic cortical lesions. *Acta Neuropathol. (Berl.)* 92:56–63.
7. Coyle, P., and Heistad, D.D. 1987. Blood flow through cerebral collateral vessels one month after middle cerebral artery occlusion. *Stroke*. 18:407–411.
8. Ferrara, N. 1995. The role of vascular endothelial growth factor in pathological angiogenesis. *Breast Cancer Res. Treat.* 36:127–137.
9. Carmeliet, P., and Collen, D. 1997. Molecular analysis of blood vessel formation and disease. *Am. J. Physiol.* 273:H2091–H2104.
10. Hendel, R.C., et al. 2000. Effect of intracoronary recombinant human vascular endothelial growth factor on myocardial perfusion: evidence for a dose-dependent effect. *Circulation*. 101:118–121.
11. Takeshita, S., et al. 1994. Therapeutic angiogenesis. A single intraarterial bolus of vascular endothelial growth factor augments revascularization in a rabbit ischemic hind limb model. *J. Clin. Invest.* 93:662–670.
12. Banai, S., et al. 1994. Angiogenic-induced enhancement of collateral blood flow to ischemic myocardium by vascular endothelial growth factor in dogs. *Circulation*. 89:2183–2189.
13. Aiello, L.P. 1997. Vascular endothelial growth factor. 20th-century mechanisms, 21st-century therapies. *Invest. Ophthalmol. Vis. Sci.* 38:1647–1652.
14. Bauters, C., et al. 1995. Site-specific therapeutic angiogenesis after systemic administration of vascular endothelial growth factor. *J. Vasc. Surg.* 21:314–324.
15. Kovacs, Z., Ikezaki, K., Samoto, K., Inamura, T., and Fukui, M. 1996. VEGF and flt. Expression time kinetics in rat brain infarct. *Stroke*. 27:1865–1872.
16. Hayashi, T., Abe, K., Suzuki, H., and Itoyama, Y. 1997. Rapid induction of vascular endothelial growth factor gene expression after transient middle cerebral artery occlusion in rats. *Stroke*. 28:2039–2044.
17. Lennmyr, F., Ata, K.A., Funai, K., Olsson, Y., and Terent, A. 1998. Expression of vascular endothelial growth factor (VEGF) and its receptors (Flt-1 and Flk-1) following permanent and transient occlusion of the middle cerebral artery

- in the rat. *J. Neuropathol. Exp. Neurol.* **57**:874–882.
18. Dvorak, H.F., Brown, L.F., Detmar, M., and Dvorak, A.M. 1995. Vascular permeability factor/vascular endothelial growth factor, microvascular hyperpermeability, and angiogenesis. *Am. J. Pathol.* **146**:1029–1039.
 19. Gotoh, O., Asano, T., Koide, T., and Takakura, K. 1985. Ischemic brain edema following occlusion of the middle cerebral artery in the rat. I. The time courses of the brain water, sodium and potassium contents and blood-brain barrier permeability to 125I-albumin. *Stroke* **16**:101–109.
 20. Belayev, L., Busto, R., Zhao, W., and Ginsberg, M.D. 1996. Quantitative evaluation of blood-brain barrier permeability following middle cerebral artery occlusion in rats. *Brain Res.* **739**:88–96.
 21. Hatashita, S., and Hoff, J.T. 1990. Role of blood-brain barrier permeability in focal ischemic brain edema. *Adv. Neurol.* **52**:327–333.
 22. Kuroiwa, T., Shibutani, M., and Okeda, R. 1988. Blood-brain barrier disruption and exacerbation of ischemic brain edema after restoration of blood flow in experimental focal cerebral ischemia. *Acta Neuropathol. (Berl)* **76**:62–70.
 23. Lyden, P.D., and Zivin, J.A. 1993. Hemorrhagic transformation after cerebral ischemia: mechanisms and incidence. *Cerebrovasc. Brain Metab. Rev.* **5**:1–16.
 24. Liu, H.M., and Chen, H.H. 1994. Correlation between fibroblast growth factor expression and cell proliferation in experimental brain infarct: studied with proliferating cell nuclear antigen immunohistochemistry. *J. Neuropathol. Exp. Neurol.* **53**:118–126.
 25. Zhang, R.L., Chopp, M., Zhang, Z.G., Jiang, Q., and Ewing, J.R. 1997. A rat model of embolic focal cerebral ischemia. *Brain Res.* **766**:83–92.
 26. Jiang, Q., et al. 1998. Diffusion-, T2-, and perfusion-weighted nuclear magnetic resonance imaging of middle cerebral artery embolic stroke and recombinant tissue plasminogen activator intervention in the rat. *J. Cereb. Blood Flow Metab.* **18**:758–767.
 27. Yang, R., et al. 1998. Substantially attenuated hemodynamic responses to Escherichia coli-derived vascular endothelial growth factor given by intravenous infusion compared with bolus injection. *J. Pharmacol. Exp. Ther.* **284**:103–110.
 28. Yang, R., et al. 1996. Effects of vascular endothelial growth factor on hemodynamics and cardiac performance. *J. Cardiovasc. Pharmacol.* **27**:838–844.
 29. LeBihan, D., et al. 1986. MR imaging of intravoxel incoherent motions: application to diffusion and perfusion in neurologic disorders. *Radiology.* **161**:401–407.
 30. Jiang, Q., et al. 1997. The temporal evolution of MRI tissue signatures after transient middle cerebral artery occlusion in rat. *J. Neurol. Sci.* **145**:15–23.
 31. Williams, D., Detre, J., Leigh, J., and Koretsky, A. 1992. Magnetic resonance imaging of perfusion using spin inversion of arterial water. *Proc. Natl. Acad. Sci. USA.* **89**:212–216.
 32. Peck, D., Spickler, E., Knight, R., Hearshen, D., and Windham, J. 1992. Analysis of the evolution of focal cerebral ischemia in the rat using the Eigenimage filter. *Magn. Reson. Med.* **26**:259–273.
 33. Zhang, L., Chen, J., Li, Y., Zhang, Z.G., and Chopp, M. 2000. Quantitative measurement of motor and somatosensory impairments after mild (30 min) and severe (2 h) transient middle cerebral artery occlusion in rats. *J. Neurol. Sci.* **174**:141–146.
 34. Hamm, R.J., Pike, B.R., O'Dell, D.M., Lyeth, B.G., and Jenkins, L.W. 1994. The rotarod test: an evaluation of its effectiveness in assessing motor deficits following traumatic brain injury. *J. Neurotrauma.* **11**:187–196.
 35. Schallert, T., et al. 1982. Tactile extinction: distinguishing between sensorimotor and motor asymmetries in rats with unilateral nigrostriatal damage. *Pharmacol. Biochem. Behav.* **16**:455–462.
 36. Matesic, D.F., and Lin, R.C. 1994. Microtubule-associated protein 2 as an early indicator of ischemia-induced neurodegeneration in the gerbil forebrain. *J. Neurochem.* **63**:1012–1020.
 37. Paxinos, G., and Watson, C. 1986. *The rat brain in stereotaxic coordinates*. 2nd edition. Academic Press Inc. New York, USA. 29–50.
 38. Gesztelyi, G., et al. 1993. Parenchymal microvascular systems and cerebral atrophy in spontaneously hypertensive rats. *Brain Res.* **611**:249–257.
 39. Hudetz, A.G., Greene, A.S., Feher, G., Knues, D.E., and Cowley, A.W., Jr. 1993. Imaging system for three-dimensional mapping of cerebrocortical capillary networks in vivo. *Microvasc. Res.* **46**:293–309.
 40. Miyoshi, Y., Date, I., and Ohmoto, T. 1995. Three-dimensional morphological study of microvascular regeneration in cavity wall of the rat cerebral cortex using the scanning electron microscope: implications for delayed neural grafting into brain cavities. *Exp. Neurol.* **131**:69–82.
 41. Dellian, M., Witwer, B.P., Salehi, H.A., Yuan, F., and Jain, R.K. 1996. Quantitation and physiological characterization of angiogenic vessels in mice: effect of basic fibroblast growth factor, vascular endothelial growth factor/vascular permeability factor, and host microenvironment. *Am. J. Pathol.* **149**:59–71.
 42. Zhang, Z.G., Davies, K., Probst, J., Fenstermacher, J., and Chopp, M. 1999. Quantitation of microvascular plasma perfusion and neuronal microtubule-associated protein in ischemic mouse brain by laser-scanning confocal microscopy. *J. Cereb. Blood Flow Metab.* **19**:68–78.
 43. Knight, R.A., et al. 1998. Prediction of impending hemorrhagic transformation in ischemic stroke using magnetic resonance imaging in rats. *Stroke.* **29**:144–151.
 44. Mathews, V., et al. 1995. Combined effects of magnetization transfer and gadolinium in cranial MR imaging and MR angiography. *Am. J. Roentgenol.* **164**:169–172.
 45. Zhang, R.L., Zhang, Z.G., and Chopp, M. 1999. Increased therapeutic efficacy with rt-PA and anti-CD18 antibody treatment of stroke in the rat. *Neurology.* **52**:273–279.
 46. Krupinski, J., Kaluza, J., Kumar, P., Wang, M., and Kumar, S. 1993. Prognostic value of blood vessel density in ischaemic stroke. *Lancet.* **342**:742.
 47. Breier, G., Albrecht, U., Sterrer, S., and Risau, W. 1992. Expression of vascular endothelial growth factor during embryonic angiogenesis and endothelial cell differentiation. *Development.* **114**:521–532.
 48. Risau, W. 1998. Development and differentiation of endothelium. *Kidney Int. Suppl.* **67**:S3–S6.
 49. Klagsbrun, M., and Folkman, J. 1990. Angiogenesis. In *Peptide growth factors and their receptors II*. M.B. Sporn and A.B. Roberts, editors. Springer-Verlag. New York, New York, USA. 459–586.
 50. Cancilla, P.A., Frommes, S.P., Kahn, L.E., and DeBault, L.E. 1979. Regeneration of cerebral microvessels: a morphologic and histochemical study after local freeze-injury. *Lab. Invest.* **40**:74–82.
 51. Rosenstein, J.M., Mani, N., Silverman, W.F., and Krum, J.M. 1998. Patterns of brain angiogenesis after vascular endothelial growth factor administration in vitro and in vivo. *Proc. Natl. Acad. Sci. USA.* **95**:7086–7091.
 52. Ingber, D.E., Madri, J.A., and Folkman, J. 1986. A possible mechanism for inhibition of angiogenesis by angiostatic steroids: induction of capillary basement membrane dissolution. *Endocrinology.* **119**:1768–1775.
 53. Stroemer, R.P., Kent, T.A., and Hulsebosch, C.E. 1998. Enhanced neocortical neural sprouting, synaptogenesis, and behavioral recovery with D-amphetamine therapy after neocortical infarction in rats. *Stroke.* **29**:2381–2393.
 54. Schratzberger, P., et al. 2000. Favorable effect of VEGF gene transfer on ischemic peripheral neuropathy. *Nat. Med.* **6**:405–413.
 55. Sondell, M., Lundborg, G., and Kanje, M. 1999. Vascular endothelial growth factor has neurotrophic activity and stimulates axonal outgrowth, enhancing cell survival and Schwann cell proliferation in the peripheral nervous system. *J. Neurosci.* **19**:5731–5740.
 56. Gale, N.W., and Yancopoulos, G.D. 1999. Growth factors acting via endothelial cell-specific receptor tyrosine kinases: VEGFs, angiopoietins, and ephrins in vascular development. *Genes Dev.* **13**:1055–1066.
 57. Mayhan, W.G. 1999. VEGF increases permeability of the blood-brain barrier via a nitric oxide synthase/cGMP-dependent pathway. *Am. J. Physiol.* **276**:C1148–C1153.
 58. Wang, W., Merrill, M.J., and Borchardt, R.T. 1996. Vascular endothelial growth factor affects permeability of brain microvessel endothelial cells in vitro. *Am. J. Physiol.* **271**:C1973–C1980.
 59. Criscuolo, G.R., Merrill, M.J., and Oldfield, E.H. 1990. Characterization of a protein product of human malignant glial tumors that induces microvascular permeability. *Adv. Neurol.* **52**:469–474.
 60. Zhao, L., Zhang, M.M., and Ng, K.Y. 1998. Effects of vascular permeability factor on the permeability of cultured endothelial cells from brain capillaries. *J. Cardiovasc. Pharmacol.* **32**:1–4.
 61. Qu, H., Nagy, J.A., Senger, D.R., Dvorak, H.F., and Dvorak, A.M. 1995. Ultrastructural localization of vascular permeability factor/vascular endothelial growth factor (VPF/VEGF) to the abluminal plasma membrane and vesiculovacuolar organelles of tumor microvascular endothelium. *J. Histochem. Cytochem.* **43**:381–389.
 62. Kawasaki, T., et al. 1999. A requirement for neuropilin-1 in embryonic vessel formation. *Development.* **126**:4895–4902.
 63. Thurston, G., et al. 1999. Leakage-resistant blood vessels in mice transgenically overexpressing angiopoietin-1. *Science.* **286**:2511–2514.
 64. Thurston, G., et al. 2000. Angiopoietin-1 protects the adult vasculature against plasma leakage. *Nat. Med.* **6**:460–463.
 65. Cheng, S.Y., Nagane, M., Huang, H.S., and Cavenee, W.K. 1997. Intracerebral tumor-associated hemorrhage caused by overexpression of the vascular endothelial growth factor isoforms VEGF121 and VEGF165 but not VEGF189. *Proc. Natl. Acad. Sci. USA.* **94**:12081–12087.
 66. Hatashita, S., and Hoff, J. 1990. Brain edema and cerebrovascular ischemia in rats. *Stroke.* **21**:582–588.
 67. van Bruggen, N., et al. 1999. VEGF antagonism reduces edema formation and tissue damage following ischemic reperfusion injury in the mouse brain. *J. Clin. Invest.* **104**:1613–1620.
 68. Hayashi, T., Abe, K., and Itoyama, Y. 1998. Reduction of ischemic damage by application of vascular endothelial growth factor in rat brain after transient ischemia. *J. Cereb. Blood Flow Metab.* **18**:887–895.
 69. Levin, E.R., et al. 1998. Role of vascular endothelial cell growth factor in Ovarian Hyperstimulation Syndrome. *J. Clin. Invest.* **102**:1978–1985.
 70. Wu, H.M., Huang, Q., Yuan, Y., and Granger, H.J. 1996. VEGF induces NO-dependent hyperpermeability in coronary venules. *Am. J. Physiol.* **271**:H2735–H2739.
 71. Iadecola, C. 1993. Regulation of the cerebral microcirculation during neural activity: is nitric oxide the missing link? *Trends Neurosci.* **16**:206–214.
 72. Feng, D., Nagy, J.A., Hipp, J., Dvorak, H.F., and Dvorak, A.M. 1996. Vesiculo-vacuolar organelles and the regulation of venule permeability to macromolecules by vascular permeability factor, histamine, and serotonin. *J. Exp. Med.* **183**:1981–1986.
 73. Reese, T.S., and Karnovsky, M.J. 1967. Fine structural localization of a blood-brain barrier to exogenous peroxidase. *J. Cell. Biol.* **34**:207–217.
 74. Simionescu, M., Simionescu, N., and Palade, G.E. 1974. Morphometric data on the endothelium of blood capillaries. *J. Cell. Biol.* **60**:128–152.