

## Congenital disorders of glycosylation caused by defects in mannose addition during N-linked oligosaccharide assembly

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### Commentary

Proteins that traverse the secretory pathway of eukaryotic cells can be covalently modified with carbohydrates, which are important for their stability and folding, and which mediate diverse recognition events in growth and development (1). Defects in the attachment of carbohydrate to protein give rise to mental and psychomotor retardation, dysmorphism, and blood coagulation defects (2, 3). These symptoms, referred to as CDG (for congenital disorders of glycosylation, or, until recently, for carbohydrate-deficient glycoprotein syndrome), are caused by mutations that affect the pathway for N-glycosylation. In this pathway (Figure 1), a branched saccharide of 14 sugars is built up on the polyisoprenoid carrier lipid dolichyl pyrophosphate (PP-Dol), then transferred to asparagines in the sequence Asn-X-Ser/Thr (where X is any amino acid except proline). Synthesis of the lipid-linked oligosaccharide (LLO) is carried out by enzymes in the membrane of the endoplasmic reticulum (ER) that act in a specific order, each transferring a specific sugar to the nascent glycolipid (4, 5). Two N-acetylglucosamines (GlcNAc's) and 5 mannoses (Man) are transferred from the sugar nucleotides UDP-GlcNAc and GDP-Man at the cytoplasmic face of the membrane, after which the Man<sub>5</sub>GlcNAc<sub>2</sub>-PP-Dol formed is flipped into the lumen of the ER, where 4 mannoses and 3 glucoses are added from dolichol-phosphate-mannose (Dol-P-Man) and Dol-P-glucose. The oligosaccharide is then attached to protein by oligosaccharyltransferase, a complex of at [...]

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Two reports in this issue (10, 11) and 1 elsewhere (12) describe new CDGs, types Id and Ie, in which LLO assembly is blocked at Man<sub>5</sub>GlcNAc<sub>2</sub>-PP-Dol. Fibroblasts from CDG-Ie patients (10, 11) have low activity of Dol-P-Man synthase, which transfers mannose from GDP-Man to Dol-P to generate the donor of the lumenally added mannoses. The human enzyme has 3 subunits (13, 14). Dpm1 is catalytic (15). Dpm2 and Dpm3 are small proteins; Dpm2 promotes binding of Dol-P to the enzyme. CDG-Ie patients have a point mutation in Dpm1, and the mutant enzyme's *K<sub>m</sub>* for GDP-Man is 6-fold higher than normal. In CDG-Id cells, Dol-P-Man synthase is normal, and transfer of mannose from Dol-P-Man to the LLO is affected by a missense mutation in the *hALG3* gene (12).

Of the new CDGs, type Id specifically affects N-glycosylation. However, the Dol-P-Man synthase deficiency that leads to CDG-Ie can have further biochemical consequences: Dol-P-Man donates the mannoses found in glycosylphosphatidylinositol (GPI) membrane anchors, the mannose attached through C-linkage to certain tryptophans, and the mannose in O-linked glycoproteins (16–18). A defect in GPI assembly leads to the disease paroxysmal nocturnal hemoglobinuria (PNH; 16), but it is not clear whether deficiencies in GPI synthesis or in O- or C-mannosylation contribute to the pathology of CDG-Ie; these patients apparently do not exhibit PNH symptoms, and the clinical features of CDG-Id and -Ie are similar. However, CDG-I cells always retain residual activity of an affected enzyme, which presumably permits them to glycosylate proteins and attach GPI anchors to them above a threshold level necessary for viability. It is possible that when GDP-Man and Dol-P-Man

are in limited supply, they are used more efficiently in other pathways, so that GPI anchoring occurs but N-glycosylation is incomplete (19).

Deficiencies in mannose-containing precursors in CDG-Ia, -Ib, and -Ie might be predicted to be bypassed if the cells are supplemented with mannose. Indeed, the LLO defect in PMI-defective (CDG-Ib) and PMM-defective (CDG-Ia) fibroblasts can be corrected by supplying mannose in their medium, but study results differ on whether this is possible in CDG-Ie cells. PMI deficiency can also be bypassed in the patient: oral mannose therapy has been used successfully to treat CDG-Ib (3).

Additional types of CDG-I could arise from defects at other steps in assembly of the LLO, in its translocation across the membrane, or in its transfer to asparagine. The challenge now is to determine the structures and biochemical activities of the enzymes involved, and the mechanism of Man<sub>5</sub>GlcNAc<sub>2</sub>-PP-Dol translocation. A further issue is whether CDG-I symptoms are due to hypoglycosylation of a few specific proteins. If so, the roles of these glycoproteins in, for example, the nervous system, are of great interest. Because the progress made in identifying the genes mutated in the CDG-I subtypes has relied on work done in yeast, these studies are an excellent example of how basic research with a model organism can help us understand human genetic disease.

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