COUP-TFII regulates satellite cell function and muscular dystrophy

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SUPPLEMENTAL INFORMATION:

SUPPLEMENTAL FIGURES.



Supplemental Figure 1. COUP-TFII expression in satellite cells and characterization of COUP-TFII-expressing mice. (A) COUP-TFII expression in DMD patients by querying public dataset. (B) COUP-TFII level in 1-month-old mdx and its control C57BL/10ScSnJ mice (n=6), as well as 2-month-old control and COUP-TFII transgenic animals (n=6). (C) Representative β -gal staining of TA muscles collected on day 0, 8, and 12 after injury. Images are representative of three different animals at each time point. (D) Freshly FACS-isolated SCs from control and COUP-TFII OE mice were examined for COUP-TFII levels one week after tamoxifen injection. (E) Body weight of control (n=5) and COUP-TFII OE mice (n=8) at 20 months of age. (F) Mean fiber CSA of soleus muscles in control (n=6) and COUP-TFII OE mice (n=5). (G) Percentage of eMHC⁺ cells in soleus muscles in control (n=6) and COUP-TFII OE mice (n=5). (H) Representative Myc photomicrograph of soleus muscles from 5 animals of each genotype. Arrowheads indicate the central-nucleated myc positive fibers. *P<0.05, **P<0.01, ***P<0.001 by Student's *t*-test. Data represent mean + SEM. Scare bars: 25 µm (C,H).



Supplemental Figure 2. Satellite cell specific COUP-TFII overexpression in COUP-TFII OE and COUP-TFII OE *mdx* animals. (A,B) Myc and PAX7 staining indicates myc-tagged COUP-TFII-expressing SCs in diaphragm muscles in COUP-TFII OE mice one week after tamoxifen treatment. (C,D) Representative images of Myc and PAX7 stained diaphragm in *mdx* and COUP-TFII OE *mdx* mice. Arrowhead indicates myc positive cells (A,C). Data are representative of three independent experiments. Scare bars: $25 \mu m$ (A,B,C,D).



Supplemental Figure 3. Characterization of *mdx* and COUP-TFII OE *mdx* mice. (A) Hematoxylin-stained diaphragm muscles indicate centrally positioned myonuclei. (B-D) The diaphragm muscles were stained with p-Histone H3 and BrdU antibodies. Quantification of the number of BrdU⁺ cell per 100 nuclei in *mdx* and COUP-TFII OE *mdx* mice (*n*=6). Arrowheads indicate the cycling SCs with p-Histone H3 signal. Scare bars: 25 μ m (A,B,C). ****P*<0.001 by Student's *t*-test. Results are the mean <u>+</u> SEM.



Supplemental Figure 4. Examination of cellular senescence and apoptosis in COUP-TFII OE *mdx* animals. (A,B) Representative photomicrograph (A) and percentage (B) of P16^{INK4a} positive cells in *mdx* (*n*=25 images from 5 animals) and COUP-TFII OE *mdx* (*n*=29 images from 6 animals) mice. (C-E) Diaphragm muscles from 4-month-old *mdx* and COUP-TFII OE *mdx* animals were assayed for p19^{ARF} (C), SA-β-gal activity (D) and cleaved caspase 3 (E). Images are representative of four animals for indicated genotypes. (F) Gross appearance of *mdx* and COUP-TFII OE *mdx* mice at 8 months of age. Arrowhead indicated immunoreactive signals in A,C,D,E. Scare bars: 25 µm (A,C,D,E). Statistical significance was determined by Student's *t*-test



Supplemental Figure 5. Defective regenerative response in COUP-TFII overexpression mice following transient injury. (A) Quantification of myofiber size as evaluated by cross-sectional area in control (n=6) and COUP-TFII OE mice (n=6) on day 12 post-injury. (B) H&E staining of TA muscles 12 days after BaCl₂ injection. Results are representative of 3 independent experiments. (C) Real-time PCR shows the expression of *COUP-TFII*, *ki67*, myogenin and *ckm* in regenerating skeletal myocytes (n=6). (D) Transcripts of denoted genes in TA muscles from control (n=6) and COUP-TFII OE (n=8) mice on day 2 after CTX injury. (E) p-Histone H3 stained proliferating SCs in control and COUP-TFII OE mice 2 days after damage. Scare bars: 25 µm (B,E). *P<0.05, **P<0.01 by Student's *t*-test. Results are the mean <u>+</u> SEM.



Supplemental Figure 6. COUP-TFII overexpression inhibits satellite cell proliferation. (A) Immunofluorescence staining of PAX7 in FACS-sorted satellite cells. (B) Differentiation of isolated satellite cells shown by MHC staining 5 days after induction. (C)The mRNA levels of cell cycle-associated genes in COUP-TFII OE myoblasts 3 days after COUP-TFII induction. Results are representative of 3 independent experiments. (D) COUP-TFII represses *Ccnd1* and *Myf5* transcription in primary myoblasts. (E) Quantification of CCND1⁺ cell evaluated as a fraction of CCND1⁺ cells in total nuclei in *mdx* (*n*=10) and COUP-TFII OE *mdx* (*n*=8) mice. (F) Measurement of MYF5⁺ cell in *mdx* (*n*=10) and COUP-TFII OE *mdx* (*n*=9) mice. Scare bars: 50 µm (A,B). **P*<0.05, ****P*<0.001 by Student's *t*-test. Data are presented as mean <u>+</u> SEM.



Supplemental Figure 7. IGF-1 and G-CSF expression in COUP-TFII OE *mdx* mice. (A,B) Representative images of IGF-1 (A) and G-CSF (B) staining of 4-month-old diaphragm from *mdx* and COUP-TFII OE *mdx* mice. (C,D) Expression of *IGF-1* and *G-CSF* genes in cultured COUP-TFII over- (C) and under-expression (D) myoblasts 3 days after virus infection. Results are representative of 3 independent experiments. Scare bars: 25 μ m (A,B). Statistical significance was determined by Student's *t*-test



Supplemental Figure 8. Fusion defects in COUP-TFII transgenic mice. (A) Expression profile of the indicated genes in control (n=6) and COUP-TFII OE (n=6) muscles from day 5 to day 12 after CTX injection. (B) Activation of *Cav3* and *Cxcr4* expression in COUP-TFII deficient myoblasts. Data are representative of 3 independent experiments. (C) Luciferase assay of wild type and mutant *Cxcr4* enhancer in the presence and absence of COUP-TFII in primary SCs. *Cxcr4* enhancer fragment was placed in front of the TATA basic promoter that drives the luciferase reporter. Data are representative of 3 independent experiments. **P*<0.05, ***P*<0.01, ****P*<0.001 by Student's *t*-test. Results are the mean <u>+</u> SEM.



Supplemental Figure 9. Characterization of COUP-TFII KO and COUP-TFII KO mdx mice. (A) COUP-TFII expression in FACS-sorted satellite cells from control and COUP-TFII KO mice. Results are representative of 3 independent experiments. (B) Histological analyses of diaphragm muscles in control and COUP-TFII KO mice. Interstitial fibrosis and mineralization were assessed by Masson's trichrome and Kossa staining, respectively. (C) von Immunofluorescence staining of PAX7 and COUP-TFII in diaphragm muscles showed the depletion of COUP-TFII protein in PAX7⁺ cells one week after tamoxifen injection. Images are representative of four different animals of indicated genotypes. Scare bars: 100 µm (B), 25 µm (C). ***P<0.001 by Student's t-test. Results are the mean + SEM.



Supplemental Figure 10. Histological examination of *mdx* and COUP-TFII KO *mdx* mice. (A) Representative photograph of trichrome stained diaphragm at 4 months of age. (B-D) 11-month-old diaphragm (B) and tibialis anterior muscles (C,D) were stained with H&E and trichrome. Images are representative of four animals for each genotype. Scare bars: 100 μ m (A,B), 50 μ m (C,D).



Supplemental Figure 11. EBD uptake in *mdx* and COUP-TFII KO *mdx* mice. (A) Body weight and lean mass of 4-month-old *mdx* (*n*=8) and COUP-TFII KO *mdx* mice (*n*=7). (B) Representative images for EBD infiltration in 4-month-old TA muscles. (C) Quantified mean data for EBD uptake in *mdx* and COUP-TFII KO *mdx* mice (*n*=42 images from 7 animals). For every mice, six images taken from three different muscle sections were counted for EBD absorption. TA, tibialis anterior, Gas, gastrocnemius. Scare bars: 200 µm (B). **P*<0.05 by Student's *t*-test. Results are shown as mean <u>+</u> SEM.



Supplemental Figure 12. Representative photomicrographs of H&E staining, macrophage and neutrophil infiltration. (A) Images of H&E-stained 4-month-old tibialis anterior muscles. (B) TA muscles were stained with neutrophil (MPO and Ly-6G) and macrophage (F4/80) markers. Data are representative of four independent experiments. Scare bars: 100 μ m (A), 50 μ m (B).