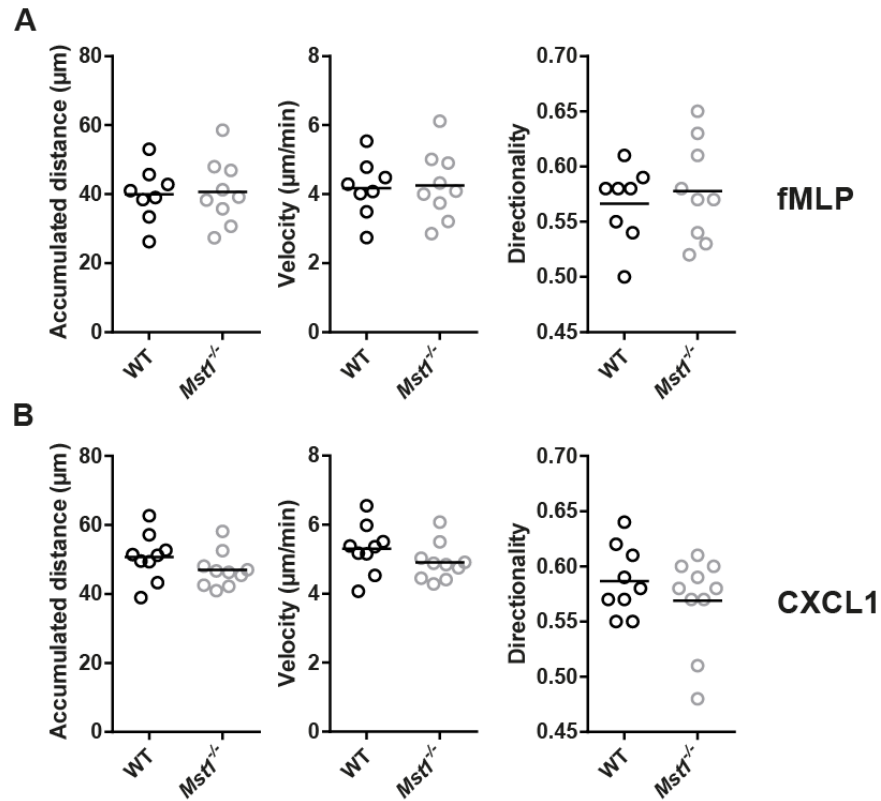
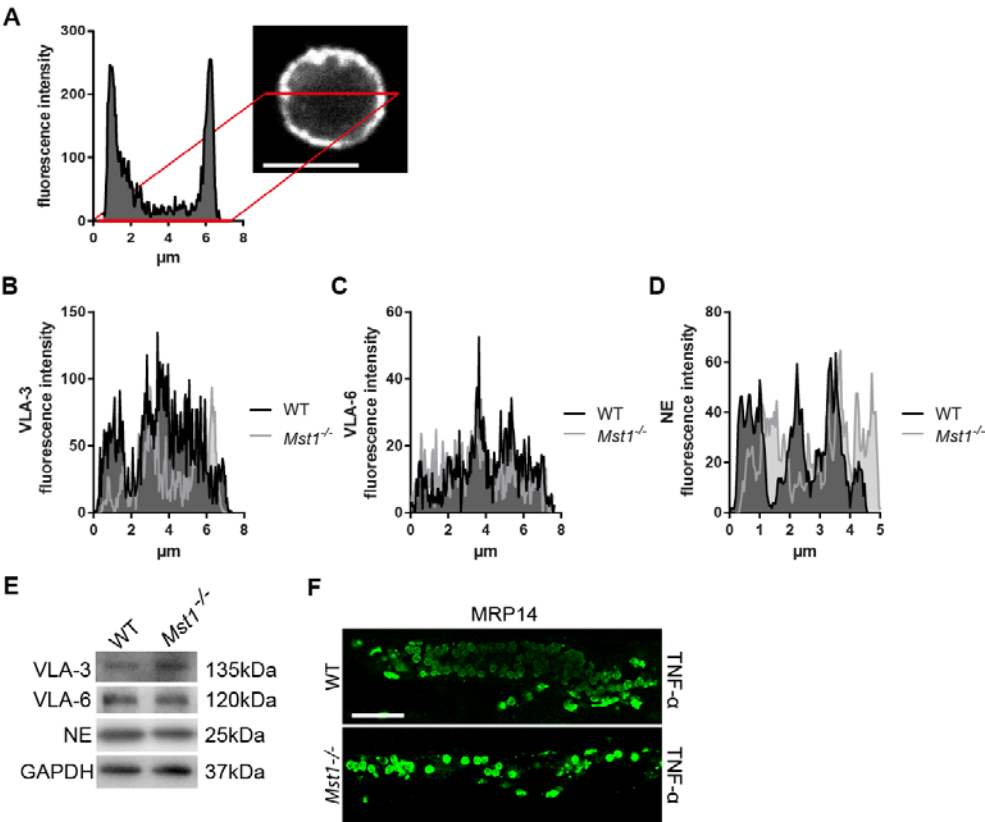


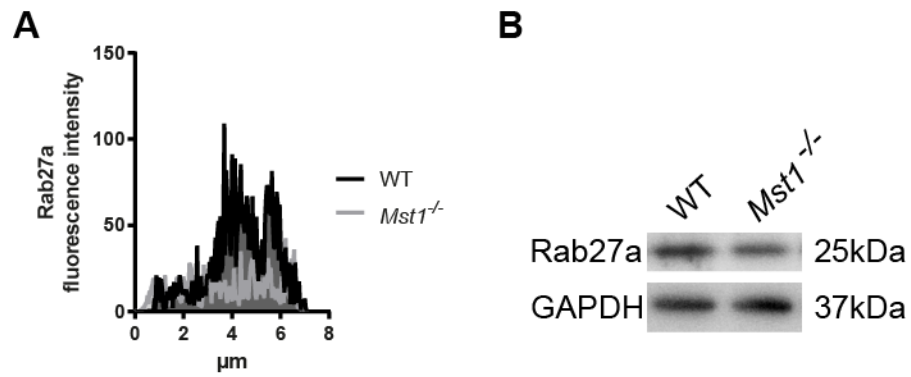
Supplemental Figure 1 Expression, viability and adhesion properties of WT and *Mst1*^{-/-} neutrophils. (A) MST2 protein levels of EGF-stimulated A431 cells, used as positive control, and of WT and *Mst1*^{-/-} neutrophils. GAPDH served as loading control (n=3). (B) FACS analysis of viable (TO-PRO3⁺ T-MRE⁺) and apoptotic (TO-PRO3⁺ T-MRE⁻) WT (back circles) and *Mst1*^{-/-} neutrophils (grey circles) after cultivation for indicated times (n=3 mice per group, mean ± SEM, n.s., 2way ANOVA, Sidak's multiple comparisons test) (C) Apoptosis rate (TO-PRO3⁺ T-MRE⁻) of neutrophils isolated from cremaster muscles of WT and *Mst1*^{-/-} mice 2 h after i.s. injection of TNF-α (n=3 mice per group, n.s. unpaired t-test). (D) Rolling flux fraction and (E) cumulative distribution of neutrophil rolling velocity in cremaster muscle venules of WT and *Mst1*^{-/-} mice 2 h after i.s. injection of TNF-α (n ≤ 5, mean ± SEM, n.s., unpaired t-test). (F,G) FACS analysis of ICAM-1 binding to (F) WT or (G) *Mst1*^{-/-} neutrophils with or without CXCL1 stimulation (n=5 * p < 0.05, ** p < 0.01 paired t-test). (H) FACS analysis of WT and *Mst1*^{-/-} neutrophils for surface expression of LFA-1, Mac-1, CXCR2, CD44, PSGL-1, CD62L and corresponding isotype controls. The mean fluorescent intensity (MFI) was evaluated relative to the isotype control on Ly6G⁺ cells (neutrophils) (n= 3 mice, mean ± SEM, n.s., unpaired t-test).



Supplemental Figure 2 Chemotactic migration of WT and *Mst1*^{-/-} neutrophils. (A,B)
 Chemotactic migration of WT and *Mst1*^{-/-} neutrophils seeded in 3D collagen gels towards a gradient of **(A)** fMLP or **(B)** CXCL1 for 30 min. Neutrophils were tracked using Fiji software (62). Accumulated distance, velocity and directionality of cells were evaluated (n= 3, scatter blots with mean, n.s., unpaired t-test).



Supplemental Figure 3 VLA-3, VLA-6 and NE – protein levels, distribution and NE activity. Fluorescence intensity profiles along a line cut through the center of the cell as exemplified in **(A)** for **(B)** VLA-3, **(C)** VLA-6 and **(D)** NE of a representative WT (black line) and *Mst1*^{-/-} neutrophil (grey line) seeded on BSA coated wells. Scale bar: 5 μm. **(E)** Total protein levels of VLA-3, VLA-6 and NE from neutrophils of WT and *Mst1*^{-/-} mice. GAPDH served as loading control (n=3 mice). **(F)** Immunostaining of MRP14 (green) in TNF-α stimulated cremaster muscle whole mounts from WT and *Mst1*^{-/-} mice (n=3 per group). Scale bar: 50 μm.



Supplemental Figure 4 Protein levels and distribution under unstimulated conditions of Rab27a. **(A)** Fluorescence intensity profile along a line cut through the center of the cell for Rab27a of a representative WT (black line) and *Mst1*^{-/-} neutrophil (grey line) seeded on BSA coated wells. **(B)** Total protein levels of Rab27a from neutrophils of WT and *Mst1*^{-/-} mice. GAPDH served as loading control (n=3 mice).

Supplemental Movie 1 LFA-1 clustering in WT and *Mst1*^{-/-} neutrophils. Whole blood from (A) WT and (B) *Mst1*^{-/-} mice was incubated with a non-blocking LFA-1 antibody and perfused through flow chambers coated with E-selectin, ICAM-1 and CXCL1. Using confocal microscopy, interacting cells were recorded under flow conditions. Images were recorded for 195 s (one stack every 13 s). The movies were generated and converted using Fiji software.

Supplemental Movie 2 Intravital multi photon laser scanning microscopy of transmigrating neutrophils. Extravasation of neutrophils (green) was recorded from postcapillary venules (red) in a TNF- α stimulated mouse cremaster muscle of (A) *Lyz2*^{GFP} and (B) *Mst1*^{-/-} x *Lyz2*^{GFP} mice. Z-stacks of postcapillary venules were obtained every 30 seconds over a time period of 240 min. Sequences of z-stacks were processed and converted using Imaris 7 and Fiji software.

Supplemental Movie 3 Intravital multi photon laser scanning microscopy of laser injury. Four hours after intrascrotal injection of TNF- α in (A) *Lyz2*^{GFP} and (B) *Mst1*^{-/-} x *Lyz2*^{GFP} mice, a laser injury was induced near postcapillary cremaster muscle venules (red). Swarming of neutrophils (green) was then recorded to the sites of laser injury. Z-stacks of postcapillary venules (red) were obtained every 30 seconds over a time period of 30 min. Sequences of z-stacks were processed and converted using Imaris 7 and Fiji software.