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Selective graft-versus-leukemia depends on magnitude and diversity of the alloreactive T cell response

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Supplemental Table 1: Patient characteristics

patient ID	age at alloSCT	don→pat gender ^A	disease	donor ^B	HLA-A	HLA-B	HLA-C	conditioning regimen ^C		stem cell source ^D	state pre DLI ^E	GvHD	GvHD treatment	follow up (years)
6268	42	f→f	AML-M4	sd	02:01 24:02	35:01 44:02	04:01 05:01	MA	cyclophosphamide TBI	PBSC	MC	no	no	8.4
7103	53	m→m	MDS	sd	02:01	07:02 44:02	05:01 07:02	NMA	fludarabin busulphan Alemtuzumab	PBSC	BM blasts cytoreduction	no	no	5.6
3356	45	m→f	CML-CP	sd	02:01 24:02	07:02 13:01	06:02 07:02	MA	cyclophosphamide TBI	PBSC	BCR-ABL pos	no	no	15.4
4461	23	f→m	CML-CP	sd	02:01	07:02 44:02	05:01 07:02	MA	cyclophosphamide TBI	PBSC	BCR-ABL pos	no	no	12.7
5835	50	m→m	CML-CP	sd	29:02	44:03 51:01	14:02 16:01	MA	cyclophosphamide TBI	PBSC	BCR-ABL pos	no	no	9.1
5866	55	f→f	CML-CP	sd	02:01 11:01	35:01 51:01	04:01 14:02	MA	cyclophosphamide TBI	PBSC	BCR-ABL pos	no	no	9.9
5596	66	m→m	AML	mud	01:01 02:01	08:01 51:01	07:01 15:02	NMA	fludarabin busulphan Alemtuzumab horse ATG	PBSC	bm blasts (smouldering)	grade II	dermovate, prednisone, cyclosporine	8.3
7995	63	m→m	AML	sd	01:01	35:02 52:01	04:01 12:02	NMA	fludarabin busulphan Alemtuzumab	PBSC	MC	grade II	prednisone, cyclosporin, mycophenolate mofetil	4.9
5852	56	m→f	MDS	mud	02:01	07:02 44:02	05:01 07:02	NMA	fludarabin busulphan horse ATG	PBSC	MC	grade II, skin+mouth	dermovate, prednisone, cyclosporin	9.7
6181	48	m→f	CML-CP	sd	11:01 24:02	44:02 51:01	05:01 15:02	MA	cyclophosphamide TBI	bone marrow	BCR-ABL pos	grade III, skin+mouth	dermovate, prednisone, cyclosporine	8.6
4716	36	f→m	AML-M1	sd	02:01 24:02	40:01 44:02	03:04 05:01	MA	cyclophosphamide TBI	PBSC	bm blasts (smouldering)	grade IV, deceased	prednisone, cyclosporin, mycophenolate mofetil	0.2
^A f=female m=male		^B sd=sibling donor mud=matched unrelated donor			^C MA=myeloablative NMA=non myeloablative TBI=total body irradiation			^D peripheral blood stem cells		^E MC (mixed chimerism) by STR-PCR BCR-ABL by real time PCR			median:	8.6

Supplemental Table 2: T cell receptor- β sequencing analysis

CD8 T cells were purified from patient samples obtained before and after DLI. Two control samples were prepared by defined mixtures of 23 cultures T cells clones each expressing a unique TCR- β sequence. These samples served to compare the input cellular fraction of every T cell clone with the observed fraction of clonotypic sequence reads. After mRNA isolation, ARTISAN PCR was performed for gene-specific cDNA synthesis and amplification (1). TCR- β libraries were barcoded, pooled and paired-end sequencing with a read length of 125 base pairs was performed on an Illumina HiSeq2500 device. Raw reads were processed by using MiXCR algorithms to obtain TCR- β V-gene and J-gene alignments followed by assembly into clonotypes (2). Sequences of oligonucleotides are given in 'Supplemental experimental procedures: Primers used for cDNA synthesis, amplification and sequencing of TCR- β '.

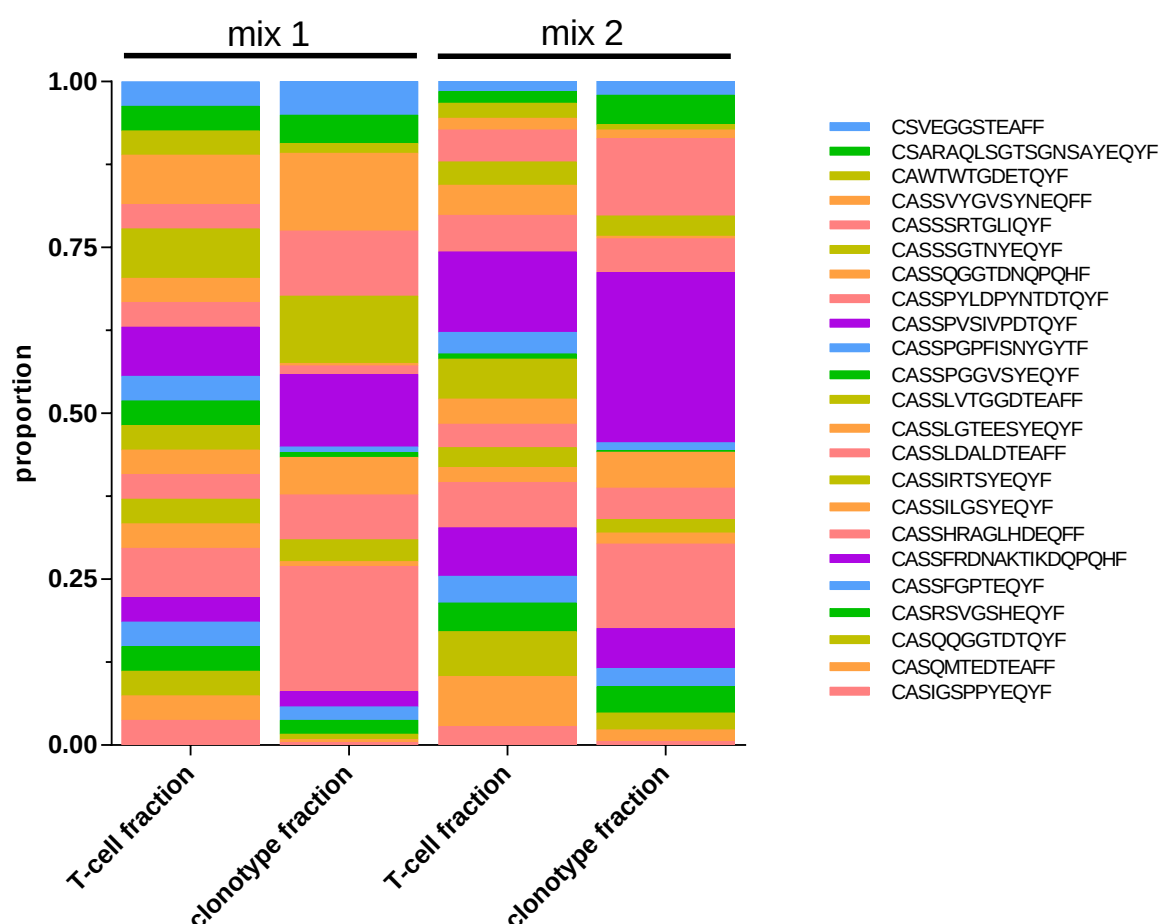
Table: Input of CD8 T cells, raw HiSeq output and MiXCR output.

sample ID	Patient ID	days after DLI	CD8 ^A T-cell input *10 ³	Raw read count *10 ⁶	% of $\geq$ Q30 Bases (PF)	Mean Quality Score (PF)	MiXCR TCR- β reads *10 ⁶	MiXCR clonotypes	Simpson's diversity index
1	7103	0	12	10.35	86.6	34.3	3.11	2,038	0.85
2		52	479	2.49	85.4	34.0	0.98	2,290	0.75
3	3356	0	382	4.00	86.3	34.2	0.94	6,212	0.92
4		56	518	8.49	87.6	34.5	1.03	5,957	0.93
5	4461	0	410	3.45	88.3	34.7	0.98	3,041	0.82
6		84	195	5.04	86.5	34.2	1.56	8,008	0.89
7	5835	0	339	3.09	83.0	33.4	0.64	4,581	0.94
8		72	187	6.06	87.9	34.6	0.64	4,202	0.96
9	5866	0	530	12.30	85.7	34.0	2.60	1,320	0.84
10		99	160	2.96	86.0	34.1	0.78	2,060	0.94
11	5596	0	6	4.86	86.2	34.2	1.11	3,337	0.79
12		49	158	7.63	87.2	34.3	1.07	6,763	0.98
13	7995	0	523	5.14	87.7	34.5	3.11	6,187	0.14
14		62	331	4.94	85.7	34.0	1.31	513	0.89
15	5852	0	409	3.80	82.8	33.3	1.15	1,753	0.88
16		49	239	6.92	87.9	34.6	0.72	3,892	0.95
17	6181	0	143	8.58	85.0	33.8	0.83	678	0.78
18		84	260	4.28	84.2	33.6	1.61	3,022	0.72
19	4716	0	506	4.44	84.6	33.8	1.47	5,563	0.92
20		23	89	9.00	87.0	34.3	1.56	9,209	0.98
median			296	5.0			1.09	3,615	0.89
min			6	2.5			0.64	513	0.14
max			530	12.3			3.11	9,209	0.98

^A CD8 T cells were purified by magnetic bead isolation from PBMCs.

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Expected and observed abundance of clonotypic TCR-β sequences in control samples of 2 defined mixtures of T cell clones



Cultured T cell clones with known TCR-β were mixed in predefined proportions and analyzed in parallel with primary purified CD8 T cells. Per mix, the fraction of input T-cells is plotted next to the clonotype fraction as calculated from HiSeq data by MiXCR algorithms. Significant correlations were observed between T-cell fraction and clonotype fraction: mix 1: $R^2=0.67$ $p<0.0001$, mix 2: $R^2=0.51$ $p=0.0001$.

References

1. Koning MT, Kielbasa SM, Boersma V, Buermans HP, van der Zeeuw SA, van Bergen CA, Cleven AH, Kluin PM, Griffioen M, Navarrete MA, et al. ARTISAN PCR: rapid identification of full-length immunoglobulin rearrangements without primer binding bias. *British journal of haematology*. 2016;bjh.14180.
2. Bolotin DA, Poslavsky S, Mitrophanov I, Shugay M, Mamedov IZ, Putintseva EV, and Chudakov DM. MiXCR: software for comprehensive adaptive immunity profiling. *Nature methods*. 2015;12(5):380-1.

Supplemental Table 3: Cloning and selection of HLA-DR^{pos} CD8 T cells and tested stimulator cells

patient	days after DLI	isolated cells	cloning format (well)	cloning efficiency % (growing clones)	tested target cells ^B						
					EBV-LCL		monoDC		CD40L-B		FB ^C
					pat	don	pat	don	pat	don	pat
6268	0	5760	384	3 (157)							
	56	14976		10 (1534)							
	105	11520		13 (1462)							
7103	0	3072	384	10 (75)							
	17	6604		10 (192)							
	24	6374		11 (192)							
	52	6604		31 (384)							
3356	0	480	96	10 (50)							
	56	2080		9 (191)							
	122	864		11 (96)							
4461	0	923	96	13 (123)							
	28	1212		7 (90)							
	84	960		4 (36)							
	182	1152		11 (131)							
5835	0 ^A	384	96	20 (78)							
	44 ^A	2016		6 (129)							
	72 ^A	480		16 (78)							
5866	0	864	96	15 (126)							
	43	3396		8 (272)							
	99	3168		3 (94)							
cloning efficiency, median (range)				8.6% (2.4%-20.3%)							
growing clones, median (range)				128 (36-1534)							
5596	0	480	384	8 (36)							
	42 ^A	4702		5 (237)							
	49 ^A	9310		2 (206)							
7995	62	16282	384	3 (518)							
	78	16282		6 (936)							
5852	45	2000	96	10 (132)							
6181	0	384	96	7 (25)							
	49	1440		14 (202)							
	84	960		12 (115)							
4716	0	480	384	11 (52)							
	23	16454		2 (287)							
cloning efficiency, median (range)				6.5% (1.7%-14.0%)							
growing clones, median (range)				202 (25-936)							

^AIndicated are the days after the second DLI.

^BShaded boxes indicate that cells in column header were tested.

^CFB were tested after 4 days incubation with 200 IU/ml IFN-γ

Supplemental Table 4: Response diversity

patient	specificity	clone	HLA restriction	MiHA frequency	clones	TCR-β V-gene ^A	TCR-β J-gene ^A	TCR-β CDR3 region	TCR-β % pre DLI ^B	TCR-β % post DLI ^B
7103	1	a	A*02	0.54	1	7-9	2-7	CASSLTRQGNEQYF	0.00	1.46
		b			1	7-9	2-5	CASSLVAGETQYF	0.00	2.68
3356	2		A*02	0.41	2	12-3	1-5	CASSFRDNAKIKDQPQHF	0.00	0.01
	3		A*02	0.04	1	11-2	2-1	CASSFRRTMNEQFF	0.00	0.00
	4		A*24	0.38	1	7-9	2-1	CASSSSTVSYEQYF	0.00	0.08
	5		B*07	0.69	2	6-2	2-1	CASSWTSGPGGNEQFF	0.00	0.09
	6		B*07	0.25	2	7-9	2-1	CASSLRGGTDEQFF	0.00	0.05
	7 ³⁾		B*07	0.71	1	30	1-2	CAWNVGQALYGYTF	0.00	0.03
	8		B*07	1.00	5	19	1-1	CASSTGQGVGTEAFF	0.00	0.01
	9		B*07	0.29	1	7-9	2-4	CASSRTGLIQYF	0.00	0.01
	10		B*13	0.31	2	7-9	2-7	CASSLVGRGHEQYF	0.00	0.47
	11		B*13	0.04	1	7-8	2-7	CASSPKQDFSSYEQYF	0.00	0.00
4461	12		A*02	0.75	1	9	2-1	CASSVSSSGDEQFF	0.00	0.20
	13		A*02	0.13	2	12-3	2-5	CASSFASGGGRGQYF	0.00	0.35
	14		B*07	0.88	1	7-2	2-1	CASSFREIGNEQFF	0.00	0.12
	15		B*44	0.29	1	24-1	2-1	CATSPTGGFDEQFF	0.00	0.00
5835	16		B*44	0.98	1	12-3	2-3	CASSLAGQGPDTQYF	0.00	0.00
	17	a	C*14	0.61	1	28	2-3	CASSLTGAPGGQPQHF	0.00	0.06
		b			1	6-5	1-5	CASSLVRFSGTDQYF	0.00	0.00
	18	a	C*14	0.04	1	7-8	2-5	CASSPMAGTSRTTDTQYF	0.00	0.02
		b			1	7-8	2-3	CASSSYRDREETQYF	0.00	0.02
5866	19		B*35	0.53	1	5-5	2-7	CASSLGTEESYEQYF	0.00	9.27
5596	20		A*01	0.67	2	5-1	2-3	CASSLAGLAGGRDTQYF	0.00	0.02
	21		A*01	0.67	1	6-2	1-2	CATLGQHYGYTF	0.00	0.03
	22		A*02	0.43	1	27	1-2	CASSPGPFISNYGYTF	0.00	0.14
	23		A*02	0.63	3	7-6	2-2	CASSLGQGAGELFF	0.02	6.58
	24		A*02	0.04	1	10-2	2-7	CATSPGHEQYF	0.00	0.55
	25 ^C		A*02	0.29	1	nd ¹⁾	nd	nd	nd	nd
	26 ^C		A*02	0.38	1	nd	nd	nd	nd	nd
	27	a	B*08	0.68	6	4-1	2-5	CASSLRQGGYEQYF	0.00	0.00
		b			1	6-1	2-7	CASSPRTSGGLQETQYF	0.00	0.46
	28		B*08	0.68	3	14	1-1	CASSLGLGAFF	0.00	0.14
	29	a	B*08	0.46	1	9	1-1	CASDLRPYTGELEFF	0.00	0.00
		b			1	9	2-2	CASSPEGNTEAFF	0.00	0.00
	30		B*51	0.96	1	28	2-1	CASSPRANSNEQFF	0.00	0.00
7995	32	a	A*01	0.96	5	5-6	2-7	CASSLWGAPSYEQYF	0.00	0.00
		b			1	20-1	2-5	CRASGGLWETQYF	0.00	0.00
	33		B*35	0.00	2	14	2-5	CASSQVRQALGQETQYF	0.00	0.00
	34	a	B*35	0.67	15	24-1	2-1	CASSPNPSTDQYF	0.00	2.33
		b			1	7-9	2-3	CATSDLPQQGGTYNEQFF	0.00	4.12
	35 ^C		C*04 or C*12	0.96	1	27	2-2	CASSLWASGRAGELFF	0.00	0.51

^ATCR-β V-gene and J-gene according to IMGT nomenclature

nd: Not determined due to loss of T cell clone.

^BNumbers represent percentages of MiHA specific TCR-β sequences within the total TCR-β repertoire of purified CD8 T cells.

^CT cell clone could not be tested in tissue specificity experiments due to insufficient in vitro expansion.

Supplemental Table 4: Response diversity (continued)

patient	specificity		HLA restriction	MiHA frequency	clones	TCR-β V-gene ^A	TCR-β J-gene ^A	TCR-β CDR3 region	TCR-β % pre DLI ^B	TCR-β % post DLI ^B
5852	36	a	A*02	0.50	3	19	2-7	CASSILGSYEQYF	0.02	0.02
		b			1	3-1	1-5	CASSQGGTDNQPQHF	0.00	0.21
	38		A*02	0.50	2	7-2	1-1	CASSLVTGGDTEAFF	0.00	0.03
	39	a	A*02	0.04	1	30	2-5	CASSPVSVIPDTQYF	0.30	0.27
		b			1	5-1	2-3	CAWTWTGDETQYF	0.01	0.04
	40	a	A*02	0.13	2	7-2	1-1	CASSHRAGLHDEQFF	0.14	0.52
		b			2	12-3	2-1	CASSLDALDTEAFF	0.05	0.06
	41		A*02	0.98	1	5-6	2-7	CASSFGPTEQYF	0.03	0.78
	42		A*02	0.08	1	nd	nd	nd	nd	nd
	43		B*07	0.67	1	9	2-1	CASSVYGVSYNEQFF	0.01	0.96
	44		B*07	0.60	1	6-1	2-7	CASIGSPPYEQYF	0.00	0.01
	45	a	B*07	0.37	2	7-9	2-7	CASSIRTSYEQYF	0.02	0.14
		b			1	7-9	2-7	CASSSGTNYEQYF	0.03	0.84
	46		B*07	0.53	1	6-1	2-7	CASSRGRGQRDEQYF	0.00	0.01
	47		B*07	0.13	3	20-1	2-7	CSARAQLSGTSGNSAYEQYF	0.05	0.00
	48		B*07	0.04	2	9	2-5	CASSAYNAGGYQETQYF	0.00	0.02
	50		B*44	0.71	1	nd	nd	nd	0.00	0.39
	51		B*44	0.04	1	7-2	2-3	CASSQVPLAGGRADTQYF	nd	nd
	52		C*05	0.96	2	13	2-7	CASSGTGDEQYF	0.01	5.55
	53		C*05 or C*07	0.04	3	3-1	2-7	CASSPGGVSYEQYF	0.00	0.50
	54		C*05 or C*07	0.08	1	19	1-1	CASQMTEDTEAFF	0.02	0.04
	55 ^C		nd	nd	2	nd	nd	nd	nd	nd
6181	56	a	A*11	0.63	3	10-3	1-5	CASSEGQGAYEQYF	0.00	0.00
		b			1	11-2	2-7	CAVGHSNQPQHF	0.01	1.58
	57		B*44	0.88	2	5-1	1-1	CASSLPLQGAREMNTEAFF	0.15	52.35
	58		B*51	0.51	13	7-9	1-2	CASSLVSGTDYGYTF	0.01	1.77
60		B*51	0.04	6	9	2-7	CASSVAGTGTDQYF	0.01	1.91	
4716	61	a	A*02	0.58	1	10-3	1-1	CAISEAAGLNTEAFF	0.00	0.02
		b			1	6-5	1-6	CASRTAAHSPLHF	0.00	0.11
	62		A*24	0.58	1	6-1	2-7	CASVPPYTGNYEQYF	0.00	0.00
	63		B*40	0.58	1	3-1	2-7	CASSQDLTAEQYF	0.00	0.06
	64	a	A*02	0.43	1	13	1-2	CASRRTLEGSYTF	0.00	0.11
		b			1	24-1	2-1	CASSPYLDPYNTDTQYF	0.00	0.04
		c			1	27	2-3	CATSDLTSPSLTF	0.00	0.02
	65		A*02	0.04	1	5-6	2-3	CASSLALAIATDTQYF	0.00	0.01
	66		B*40	0.30	1	11-3	2-3	CASSSRLAATDTQYF	0.00	0.00
	67	a	B*40	0.83	3	27	2-7	CASSGTGTEAFF	0.01	0.01
		b			1	27	1-1	CASSWTGDEQYF	0.93	0.31
	68		B*44	0.04	1	7-6	1-2	CASSQGQGYNYGYTF	0.00	0.07
	69	a	B*44	0.88	1	7-6	1-2	CASSPGQSNEQYF	0.01	1.55
		b			1	7-6	2-7	CASSQGQGITMATF	0.00	0.00
	70		B*44	0.96	2	7-6	2-1	CASSLESRRVSYNEQFF	0.00	0.64
	71		B or C	0.00	1	6-5	1-2	CASSYSIGQSNGYGYTF	0.14	0.06

^ATCR-β V-gene and J-gene according to IMGT nomenclature nd: Not determined due to loss of T cell clone.^BNumbers represent percentages of MiHA specific TCR-β sequences within the total TCR-β repertoire of purified CD8 T cells.^CT cell clone could not be tested in tissue specificity experiments due to insufficient in vitro expansion.

Supplemental Table 5: Expression of MiHA-encoding genes according to EMBL-EBI Expression Atlas

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tissue ^A		genes targeted in patient with selective GvL reactivity ^B									genes targeted in patients with GVHD ^B												
		HMHA1	CCL4	TMEM8A	MOB3A	PNP	NCAPD3	ZDHHC6	POLE	RWDD4	NDC80	APOBEC	GSTP1	GEMIN4	TTK	WNK1	ERAP1	ARHGDIB	BCAT2	PDCD11	C19orf48	KDM5D	ZNFx1
hematopoietic	bone marrow	46	84	17	24	190	10	11	17	9	13	19	249	6	10	20	16	760	4	3	33	6	12
	lymph node	45	25	23	50	37	9	26	8	9	14	5	207	8	6	25	30	605	10	9	17	0	12
	spleen	56	28	22	36	29	7	26	11	8	3	2	176	6	0.9	21	26	413	10	7	11	18	17
	mean	49	46	21	37	85	9	21	12	9	10	9	211	7	6	22	24	593	8	6	20	8	14
	sd	5	27	3	11	74	1	7	4	0	5	7	30	1	4	2	6	142	3	2	9	7	2
non-hematopoietic	adipose tissue	3	4	15	8	4	4	14	5	10	0.8	0.7	112	3	0.8	41	49	74	11	4	7	0	9
	adrenal gland	3	16	13	8	18	4	17	2	45	0	0	151	4	0	15	13	50	31	4	10	0	8
	animal ovary	0.6	2	8	15	2	4	19	5	33	0	1	232	11	0	20	18	17	21	8	22	0	9
	appendix ^C	34	30	28	32	51	8	25	6	9	7	5	191	7	4	26	34	421	9	9	15	0	15
	bladder	7	12	28	20	29	6	21	3	14	4	3	350	7	2	24	29	246	34	6	10	15	12
	cerebral cortex	2	18	11	7	10	4	13	1	6	0	0	69	3	0	32	12	30	7	4	4	9	9
	colon	8	5	36	8	22	5	20	6	12	3	7	155	4	4	15	28	53	18	6	13	10	10
	duodenum	12	6	33	9	97	5	34	5	9	4	5	249	5	3	8	46	54	12	4	13	22	8
	endometrium	3	8	13	14	11	5	23	4	11	2	1	271	8	0.7	18	19	53	13	8	15	0	9
	esophagus	0.8	5	28	13	62	5	15	3	13	3	2	1048	9	4	39	13	49	11	7	12	12	6
	gall bladder	5	38	29	15	60	5	23	3	14	1	1	368	7	0.7	21	22	137	10	4	6	16	14
	heart	0	1	14	5	5	2	8	2	10	0	0	156	4	0	29	9	47	21	2	3	0	7
	kidney	3	3	22	5	88	3	28	2	16	0	0	155	3	0	54	13	44	16	4	8	0	8
	liver	0	54	8	3	30	2	10	3	7	0.7	0.7	13	2	0	11	12	51	3	2	2	3	4
	lung	13	15	25	18	19	4	18	4	9	0.6	2	407	8	0	27	23	204	16	5	8	0	15
	pancreas	0	3	23	3	6	1	4	1	6	0	0.9	66	2	0	4	3	6	14	2	13	2	2
	placenta	7	16	60	13	96	7	20	5	9	7	2	307	5	4	20	39	91	14	5	18	5	9
	prostate	2	4	30	12	8	62	19	5	13	4	0	145	7	0.6	17	16	35	21	5	47	17	7
	salivary gland	5	3	20	6	4	2	10	3	13	0	0.7	92	4	0	7	9	45	14	4	9	0	5
	skin	0.9	0	4	13	12	8	20	11	9	3	2	334	19	3	34	26	40	8	11	18	0	9
	small intestine	12	9	31	10	43	5	36	6	9	4	3	210	4	3	10	43	72	12	4	13	35	8
	stomach	10	9	32	10	28	4	23	5	10	2	3	408	5	2	15	25	56	17	4	12	17	9
	testis	2	2	53	9	5	10	16	15	12	12	0.6	38	40	29	39	8	14	12	15	18	11	14
	thyroid	1	5	20	8	13	8	22	2	22	0.6	1	475	8	0	17	14	65	14	6	29	0	13
	mean	6	11	24	11	30	7	19	4	13	2	2	250	7	3	23	22	81	15	6	14	7	9
	sd	7	13	13	6	30	12	7	3	9	3	2	208	8	6	12	13	89	7	3	9	9	3
ratio hem/nonhem		9	4	1	3	3	1	1	3	1	4	5	1	1	2	1	1	7	1	1	2	1	1
		hemat	rel hem	broad	broad	broad	broad	broad	broad	broad	rel hem	rel hem	broad	broad	broad	broad	broad	hemat	broad	broad	broad	broad	broad

^ATissues were separated between hematopoietic (bone marrow, lymph node, spleen) and non-hematopoietic.^BNumbers represent fragments per kilobase of transcript per million mapped reads. The threshold for expression is 0.5.^CNo difference in ratios is observed when appendix is excluded from the analysis

Example: Value of HMHA1 in bone marrow: 46 fragments were mapped per 1000 basepairs of the HMHA1 transcript per 1 million mapped reads.

Mean expression level of bone marrow, lymph node and spleen is divided by the mean expression level of 24 other tissue types.

High ratios represent selective expression in hematopoietic tissues, low ratios represent tissue type independent expression

Classification of ratio hem/nonhem:

1 - 3 expressed in all tissues

4 - 6 more frequent in hematopoietic tissues

7 - 9 selective expression in hematopoietic tissues

Supplemental experimental procedures: Mixed models statistical analysis

Linear mixed-effects models are extensions of standard linear regression models including random effects to model correlation structures between dependent measurements. Parameters describing both fixed and random effects are estimated with a maximum likelihood approach. The estimated parameters can be used to calculate model-based predictions of outcome values for different values of the predictor variables, as shown in the formulas and graphs below. As stimulators were tested EBV-LCL and fibroblasts (FB). FB stimulation was tested after culturing of FB for 4 days in medium alone, in the presence of 200 IU/mL IFN- γ (stimFB-IFN γ) or in the presence of IFN- γ in combination with 10 ng/mL TNF- α and 100 IU/mL Interleukin-4 (stimFB-IT4). The data of the experiments with stimFB-IT4 are not presented in the main paper. The statistical model for FB however was built on experiments including this stimulator type and therefore the data are presented here. The effect of various ratios of stimulators was measured by production of IFN- γ by the T-cell clone and is expressed in fg per T cell per 20 h.

Predictors:

Stimulators:

EBV-LCL

steady-state fibroblasts (steady-state FB)

FB treated with IFN- γ (stimFB-IFN γ)

FB treated with IFN- γ , TNF- α and IL-4 (stimFB-IT4)

Ratios (as continuous variable):

0.11 0.33 1.0 3.0 9.0

Groups:

Selective GvL = 0

GvL+GvHD = 1

Outcome: IFN- γ production (fg/T cell/20h)

On the following pages parameter estimates, formulas and graphical representations for each of the stimulators are given.

In formulas: 'log' indicates $\log$

'I' indicates an indicator variable (1 if condition fulfilled, 0 otherwise).

Stimulator = EBV-LCL

Estimated mixed model parameters:

	Effect	Value	Std.Error	p-value
B0	Fixed (intercept)	1.56	0.27	<0.001
B1	Ratio	0.32	0.02	<0.001
B2	groupGVHD	0.13	0.32	0.69
B3	ratio*groupGVHD	0.04	0.03	0.20

*: interaction

formula to predict outcomes based on the mixed model:

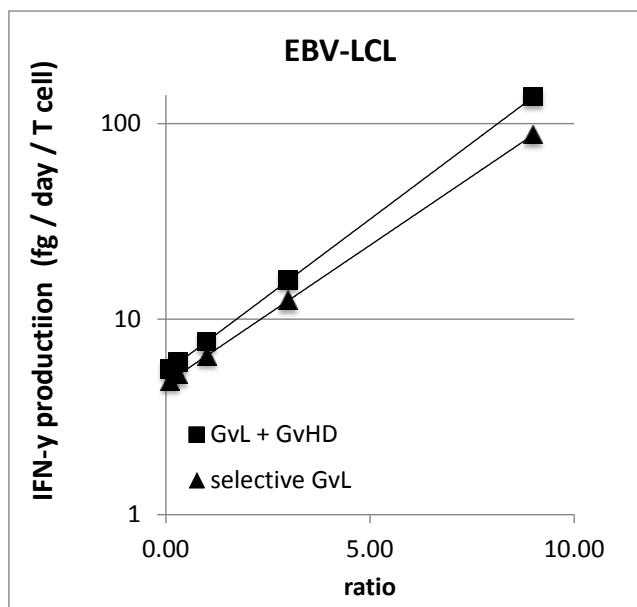
$$\log(\text{outcome} + 0.1) = B0 + (B1*\text{ratio}) + B2*(I_{\text{group=GVHD}}) + B3*\text{ratio}*(I_{\text{group=GVHD}})$$

with values for ratio= 0.11 – 9.0

$$\log(\text{outcome} + 0.1) = 1.56 + 0.32*\text{ratio} + 0.13*(I_{\text{group=GVHD}}) + 0.04*\text{ratio}*(I_{\text{group=GVHD}})$$

p-value=0.26 for the difference in outcome at ratio=9.0

Plotted:



Stimulator=FB (steady state / IFN- γ / IFN- γ +TNF- α +IL-4)

Estimated mixed model parameters:

	Effect	Value	Std.Error	p-value
B0	Fixed (Intercept)	-0.85	0.40	0.03
B1	log(ratio)	0.30	0.03	<0.001
B2	groupGVHD	0.87	0.46	0.097
B3	stimFB-IFN γ	0.53	0.09	<0.001
	stimFB-IT4	0.69	0.09	<0.001
B4	log(ratio)*groupGVHD	0.03	0.03	0.40
B5	groupGVHD*stimFB-IFN γ	0.32	0.11	0.003
	groupGVHD*stimFB-IT4	<0.001	0.11	1.00

*: interaction

Stimulator = steady-state FB

formula to predict outcomes based on the mixed model:

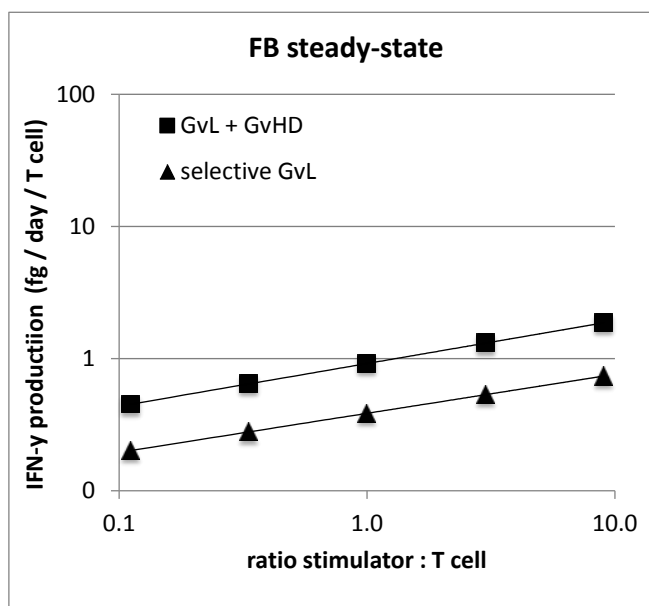
$$\log(\text{outcome} + 0.1) = B0 + B1 \cdot \log(\text{ratio}) + B2 \cdot (I_{\text{group}=\text{GVHD}}) + B3 \cdot (I_{\text{stim}=\text{FB-IFN}\gamma}) + B4 \cdot \log(\text{ratio}) \cdot (I_{\text{group}=\text{GVHD}}) + B5 \cdot (I_{\text{stim}=\text{FB-IFN}\gamma}) \cdot (I_{\text{group}=\text{GVHD}})$$

with values for ratio= 0.11 – 9.0:

$$\log(\text{outcome} + 0.1) = -0.85 + 0.30 \cdot \log(\text{ratio}) + 0.87 \cdot (I_{\text{group}=\text{GVHD}}) + 0 + 0.03 \cdot \log(\text{ratio}) \cdot (I_{\text{group}=\text{GVHD}}) + 0$$

p-value=0.10 for the difference in outcome at ratio=1.0

Plotted:



Stimulator = IFN-γ treated FB

formula to predict outcomes based on the mixed model:

$$\log(\text{outcome} + 0.1) = B_0 + B_1 \cdot \log(\text{ratio}) + B_2 \cdot (I_{\text{group}=\text{GVHD}}) + B_3 \cdot (I_{\text{stim}=\text{FB-IFN}\gamma}) + B_4 \cdot \log(\text{ratio}) \cdot (I_{\text{group}=\text{GVHD}}) + B_5 \cdot (I_{\text{stim}=\text{FB-IFN}\gamma}) \cdot (I_{\text{group}=\text{GVHD}})$$

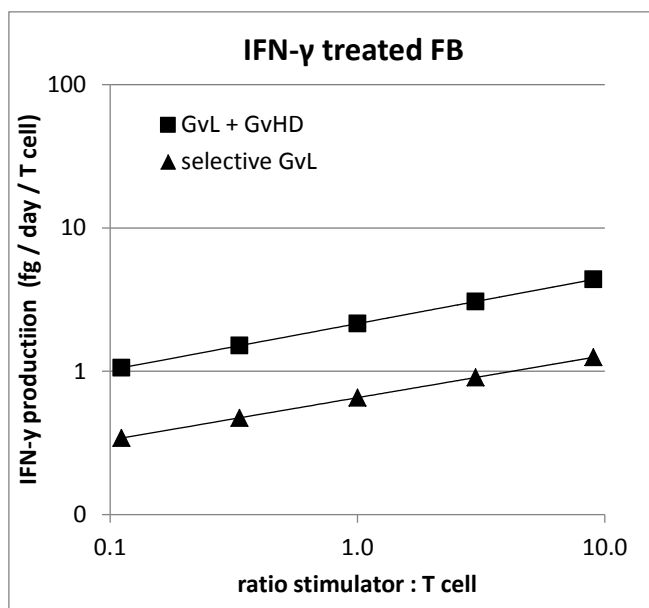
with values for ratio= 0.11 – 9.0

$$\log(\text{outcome} + 0.1) = -0.85 + 0.30 \cdot \log(\text{ratio}) + 0.87 \cdot (I_{\text{group}=\text{GVHD}}) + 0.53 + 0.03 \cdot \log(\text{ratio}) \cdot (I_{\text{group}=\text{GVHD}}) + 0.32 \cdot (I_{\text{group}=\text{GVHD}})$$

The p -value for difference in outcome between the groups noGVHD and GVHD for FB-IFN-γ was calculated using the FB model but changing the reference category to FB-IFN-γ.

p -value=0.03 for the difference in outcome at ratio=1.0

Plotted:



Stimulator = IFN- γ , TNF- α and IL-4 treated FB

formula to predict outcomes based on the mixed model:

$$\log(\text{outcome} + 0.1) = B_0 + B_1 \cdot \log(\text{ratio}) + B_2 \cdot (I_{\text{group}=\text{GVHD}}) + B_3 \cdot (I_{\text{stim}=\text{FB-IT4}}) + B_4 \cdot \log(\text{ratio}) \cdot (I_{\text{group}=\text{GVHD}}) + B_5 \cdot (I_{\text{stim}=\text{FB-IT4}}) \cdot (I_{\text{group}=\text{GVHD}})$$

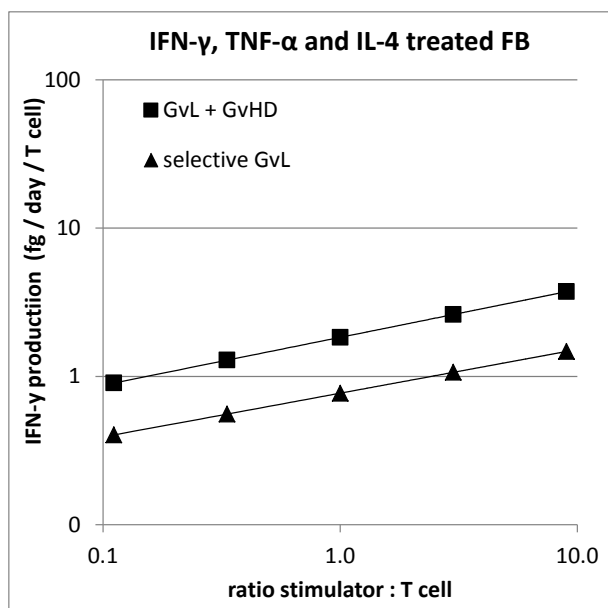
with values for ratio= 0.11 – 9.0

$$\log(\text{outcome} + 0.1) = -0.85 + 0.30 \cdot \log(\text{ratio}) + 0.87 \cdot (I_{\text{group}=\text{GVHD}}) + 0.69 + 0.03 \cdot \log(\text{ratio}) \cdot (I_{\text{group}=\text{GVHD}}) + 0.0004 \cdot (I_{\text{group}=\text{GVHD}})$$

The p -value for difference in outcome between the groups noGVHD and GVHD for FB-IT4 was calculated using the FB model but changing the reference category to FB-IT4.

p -value=0.10 for the difference in outcome at ratio=1.0

Plotted:



Supplemental experimental procedures: Primers used for amplification and sequencing of MiHA-encoding genes

WGAs results		NCBI Reference Sequence			PCR product ^B	Primer sequence 5' ► 3'	
SNP ID	gene name	Accession	Length (bp)	CDS ^A		Forward	Reverse
rs1719152	<i>CCL4</i>	NM_002984	667	80 ► 358	32 ► 583	TGAGTTCTGCAGCCTCACCTCTGA	ACAGTGACAGTGGACCATCCCCA
rs2071915	<i>TMEM8A</i>	NM_021259	3.656	130 ► 2,445	231 ► 1,018 737 ► 1,409	CGGCTACAGCGGGAAGAGCGA CCCTCCTCTCCCATCCAGCTAC	CAGCACTGAAAGCCACTGTCCC AAGGGCGAGGCAGCATTACAG
rs11541046	<i>MOB3A</i>	NM_130807	3.347	319 ► 972	361 ► 919	ACATTCCGCCCAAGCGCAA	CGATGAGGCCGAACCTCTTGACG
rs305087	<i>C16ORF99</i>	mRNA not known, PCR performed on genomic DNA			1 ► 368	ACGCACACAAACACCATCCTGC	ATCTTCCTGCCCTCCCTCACC
rs1049564	<i>PNP</i>	NM_000270	2.438	147 ► 1,016	132 ► 742	GCGTCTGCAGACCATGGAGAA	TGGGGCTGCCACCATCACATA
rs12292394	<i>NCAPD3</i>	NM_015261	5.661	607 ► 5,103	2,163 ► 2,753	TTCCGAACCCTCAGGGGAGATCA	GCCGAATGTTCCGTGCCAGT
rs4918752	<i>ZDHHC6</i>	NM_022494	2.187	425 ► 1,666	744 ► 1,400	ACAAGGCACCACGTTACATCACTG	ATTGAGAGGGCAGCAGGCACC
rs5745001	<i>POLE</i>	NM_006231	8.024	210 ► 7,070	437 ► 1,418 1,373 ► 2,322 2,312 ► 3,261 2,973 ► 3,954 3,934 ► 4,637 4,618 ► 5,539 5,326 ► 6,031 6,211 ► 7,014	GCGCTTAGGCAGTGCAGTGGA CTTCCAGAAGGACAGCCAGGGG GGGGCCAGCTCGGGCCTTTC GTCACCCGCTCAGAGAACAGCA CCCAGGACCTCACGCCGACT TGCCTCTCTGGCCAGTTCA TGGAGTTCGATGACCAAGCCACT CAGCGTACATCGTGGCCGTGT TCGTAGCCCACAGCCCACTGC	GATGCACTGGGGCGCCTTGT GAGCTGGCCCTCTGGGAACA ACAGCACGTCCAGCCAGTAGTCA CAGTCGGCGTGAGGTCCTGG GAACTGGGCCAGAGAGCGCAT CTGGCAGCTGACCACCCGT TGGGAGTCTTGCACTCCACAGTGA ACGACATGCCGTAGTGCTGGG CACAACATCAACCCAGTTCAGCC
rs4547833	<i>RWDD4A</i>	NM_152682	2.593	227 ► 793	26 ► 754	TCGTAGCCCACAGCCCACTGC	CACAACATCAACCCAGTTCAGCC
rs7233405	<i>NDC80</i>	NM_006101	2.209	183 ► 2,111	15 ► 745 702 ► 1483 1,400 ► 2,042	GCGTAATGACGTCAGCGCC GCTCCTCATACATGGCCTCAC AGCGATTGAAACACAATTAGCAGA	AGCCAACTAAGGTGCCAC CCTTTGGAATTCTCAGCACCT CGAGAGATCTTCTGACATGCATT
rs608962	<i>TTK</i>	NM_001166691	3.019	124 ► 2694	124 ► 1,515 1,343 ► 2,215 2,132 ► 2,963	ATGGAATCCGAGGATTTAAGTG GCAGATTCCGGAGTTAGCC ATGCAACCAGATACAACAAGTG	CAGCTCATGTAATCATCCAAG ACATATCTTTGATTGCTTCTGG CTTGCTATCCACCCACTATTC
rs26654	<i>ERAP1</i>	NM_001040458	5.085	348 ► 3173	140 ► 816 669 ► 1,290 1,268 ► 1,973 1,890 ► 2,583	GTTTACCCTTTCCCCAGCTC AGGAAGGGAGCTGGAGAGAG TCTTGCTGCTATTCCCGACT GTGGATGTGAAAACCATGATGA	CCGAAAGATTGCCAGCATAG GAAAGTCGGAATAGCAGCA TACATTCTCCCTCACTG TCTGTACGCACGGCTGATAG
rs4703	<i>ARHGDIB</i>	NM_001175	1.216	105 ► 710	69 ► 701 510 ► 1,216	CCGGACAGAGACGTGAAGC GTGGATAAAGCAACATTTATGG	CCACTCCTTCTTAATCGACAG CTCAGCCAGAACACACACAG
rs4801775	<i>BCAT2</i>	NM_001190	1.616	38 ► 1216	510 ► 972 918 ► 1,472	ATCGAAGTGACAAGGACTG CTGAATGGTGTATCCTGCCTG	GAGCCATGTCCAGTAGACTC GCACGACAAGGAGTAATGGG
rs4801853	<i>C19ORF48</i>	NM_199250	1.667	898 ► 1,251	776 ► 1,425	CAGCCCACCATGGACAGGGGTA	TCTGGCCAGTGACCCACAG
rs238199	<i>ZNFX1-AS1</i>	NR_003604	1.020	n.a.	9 ► 694 458 ► 980	CAGGGTGGAGAGCACGAG ATTGCTTCTTCGCGTCTGC	CCCGCATTATCTCTGGTTCA ATGCAGGTAGGCAGTTAGAAAT
rs238221	<i>ZNFX1</i>	NM_021035	7.371	248 ► 6,004	2,828 ► 3,300	GCTAAATGCTTCTGGCCATGAG	GCAATGGTATGGGCCTCAAG

^A first and last bp of the coding sequence

^B first to last bp of the PCR product

Supplemental experimental procedures: Primers used for cDNA synthesis, amplification and sequencing of TCR-β

Step	Orientation	Name	5'-to-3' DNA-sequence			
reverse transcription	TSO	SA.rt	AAGCAGTGGTATCAACGCAGAGTACGCrGrGrG			
	rev	TRB.rt	CACGTGGTCGGGGWAGAAGC			
amplification	for	SA.pcr	GAGTTCAGACGTGTGCTCTTCCGATCTAAGCAGTGGTATCAACGCAGAGTACGC			
	rev	TCRβ.1	CCTACACGACGCTCTTCCGATCTGTGGGAACACCTTGTTTCAGGTCCTC			
	rev	TCRβ.2	CCTACACGACGCTCTTCCGATCTGTGGGAACACGTTTTTCAGGTCCTC			
barcoding PacBio	for	SA.bc-xx	GGTAG [SAbc.xx] CTTAAGCAGTGGTATCAACGCAGAGTACG			
			SAbc.01	GCGCTCTGTGTGCAGC	SAbc.07	AGAGACACGATACTCA
			SAbc.02	TCATGAGTCGACACTA	SAbc.08	CTGCTAGAGTCTACAG
			SAbc.03	TATCTATCGTATACGC	SAbc.09	AGCACTCGCGTCAGTG
			SAbc.04	ATCACACTGCATCTGA	SAbc.10	TCATGCACGTCTCGCT
			SAbc.05	ACGTACGCTCGTCATA	SAbc.11	AGAGCATCTCTGTACT
			SAbc.06	TGTGAGTCAGTACGCG	SAbc.12	CGCATCGACTACGCTA
	rev	TR_D5xx	AATGATACGGCGACCACCGAGATCTACAC [D5xx] GACACTCTTTCCCTACACGACGCTCTTCCGATCT			
			D501	CGTGAT	D507	GATCTG
			D502	ACATCG	D508	TCAAGT
			D503	GCCTAA	D509	CTGATC
			D504	TGGTCA	D510	AAGCTA
			D505	CACTGT	D511	GTAGCC
			D506	ATTGGC	D512	TACAAG
barcoding Illumina HiSeq	for	TR_D7xx	CAAGCAGAAGACGGCATACGAGAT [D7xx] GTGACTGGAGTTCAGACGTGTGCTCTTCCGATC			
			D701	TTGACT		
			D702	GGAACT	D704	GGACGG
			D703	TGACAT	D705	CTCTAC
	rev	TR_D5xx	AATGATACGGCGACCACCGAGATCTACAC [D5xx] GACACTCTTTCCCTACACGACGCTCTTCCGATCT			

TSO=template switching oligo W=A/T rG=RNA