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SDF1 induction by acidosis from principal cells regulates intercalated cell subtype distribution

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The nephron cortical collecting duct (CCD) is composed of principal cells, which mediate Na, K, and water transport, and intercalated cells (ICs), which are specialized for acid-base transport. There are two canonical IC forms: acid-secreting α -ICs and HCO_3^- -secreting β -ICs. Chronic acidosis increases α -ICs at the expense of β -ICs, thereby increasing net acid secretion by the CCD. We found by growth factor quantitative PCR array that acidosis increases expression of mRNA encoding SDF1 (or CXCL12) in kidney cortex and isolated CCDs from mouse and rabbit kidney cortex. Exogenous SDF1 or pH 6.8 media increased H^+ secretion and decreased HCO_3^- secretion in isolated perfused rabbit CCDs. Acid-dependent changes in H^+ and HCO_3^- secretion were largely blunted by AMD3100, which selectively blocks the SDF1 receptor CXCR4. In mice, diet-induced chronic acidosis increased α -ICs and decreased β -ICs. Additionally, IC-specific *Cxcr4* deletion prevented IC subtype alterations and magnified metabolic acidosis. SDF1 was transcriptionally regulated and a target of the hypoxia-sensing transcription factor HIF1 α . IC-specific deletion of *Hif1a* produced no effect on mice fed an acid diet, as α -ICs increased and β -ICs decreased similarly to that observed in WT littermates. However, *Hif1a* deletion in all CCD cells prevented acidosis-induced IC subtype distribution, resulting in more severe acidosis. Cultured principal cells exhibited an HIF1 α -dependent increase of *Sdf1* transcription in response to media acidification. Thus, our results indicate that principal cells respond to acid by producing SDF1, which then acts on adjacent ICs.

Introduction

The kidney responds to an acid load by increasing the rate of net H^+ secretion in the proximal and collecting tubules as well as by increasing the capacity to excrete net acid through stimulation of NH_3 synthesis largely in the proximal tubule. Net H^+ secretion in the collecting tubule is mediated by intercalated cells (ICs), which are present in two forms: (a) α -ICs, which secrete H^+ by an apical H^+ -ATPase and a basolateral $\text{Cl}:\text{HCO}_3^-$ exchanger, AE1, and (b) β -ICs, which secrete HCO_3^- by an apical pendrin and a basolateral H^+ ATPase. Net acid secretion in this segment is thus the sum of these two processes. In the collecting tubule, as in the other acidifying segment (the proximal tubule), there are acute and chronic responses to an acid load. Acute responses might include the following: exocytosis of vesicles containing H^+ -ATPases, thereby increasing the number of proton pumps on the surface (1, 2); assembly and disassembly of various subunits; and kinetic effects on the ATPase mediated by cytoplasmic acidification, kinetic activation of soluble adenylyl cyclase (3), as well as by hormones such as angiotensin II (4) and aldosterone (5).

The chronic response in the collecting duct occurs by alteration in the numbers of the two subtypes of ICs (6), a change that

is complex and requires new RNA and protein synthesis (7). While this response is well documented and occurs in vivo, the mechanism by which these cells sense a change in blood pH, which causes cellular acidification, is obscure. It is clear that sensing the lowered blood pH and the response of the epithelial cell can be entirely local to the cortical collecting duct (CCD). This was demonstrated when isolated perfused rabbit cortical collecting tubules were incubated for 3 hours in acid media. A study of individually identified HCO_3^- -secreting cells showed that they lost their apical $\text{Cl}:\text{HCO}_3^-$ exchange activity, and some began to express basolateral $\text{Cl}:\text{HCO}_3^-$ exchange (7). The discovery that polymerization of the extracellular protein hensen (DMBT1) (8) was critical to this process again confirmed the role of local factors produced by the collecting tubule (7). The question remains: how does the cell receive the acid signal, and what is its immediate response? Further, given that the collecting tubule contains two cell types — principal cells and ICs — we examined whether the IC itself is autonomous in the control of acid-base homeostasis at the level of the collecting duct, i.e., whether it receives as well as responds to the acid signal.

Here, we show that the proximate response of the cell is to induce the expression of SDF1 (also known as CXCL12), likely by activating the hypoxia transcription pathway, which, by itself, is sufficient to induce chronic adaptation, whereby the proportion of acid- and bicarbonate-secreting cells is changed. The most surprising finding, however, is that it is the principal cell that is the sensor

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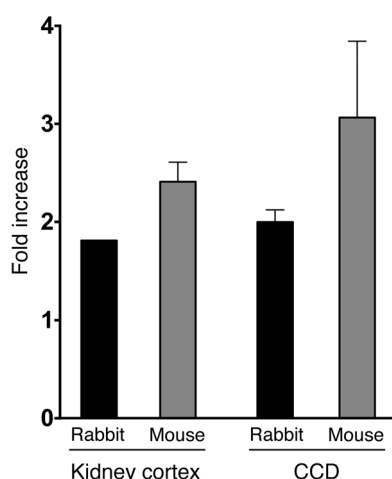


Figure 1. qPCR of *SDF1* (*CXCL12*) in kidney from normal and acidotic animals. qPCR was performed on kidney cortex mRNA from animals fed a normal diet and compared with kidney cortex mRNA from animals fed an acid diet for 2 days. Rabbit and mouse kidney cortex mRNA was normalized to *Atp5v1b1* (rabbit, $n = 4$) and *Gapdh* (mouse, $n = 4$). CCDs from transgenic mice expressing GFP in the ICs were isolated using the COPAS sorter and were normalized to *Gapdh*.

for the acid signal, a cell that heretofore had been thought to be dedicated to Na, K, and water transport but not to acid-base transport.

Results

Metabolic acidosis induces *SDF1* in CCDs. We induced metabolic acidosis in mice by feeding them an acid diet (0.1 g $\text{NH}_4\text{Cl/g}$ food), isolated renal cortex mRNA, and used a growth factor PCR array (SABiosciences, QIAGEN) to assess expression of 89 growth factors. Comparison of mRNA from mice fed a normal diet with that from mice fed an acid diet for 2 days showed that acidosis increased expression of the following factors: *Fgf2*, *Sdf1* (*Cxcl12*), *Bdnf*, *Gdnf*, *Lefty2*, and *Inhba* (Supplemental Table 1; supplemental material available online with this article; doi:10.1172/JCI80225DS1). However, confirmation by quantitative PCR (qPCR) showed that only *Sdf1* was increased consistently by acidosis (qPCR of *Sdf1/Gapdh* mRNA showed an ~2-fold

increase in acidosis over that seen in mice fed a normal diet, $n = 4$) (Figure 1). We isolated CCDs from mice expressing GFP in their ICs using a complex object parametric analyzer and sorter (COPAS) and found that feeding mice an acid diet for 3 days also resulted in a substantial induction of *Sdf1* mRNA (Figure 1). We extended these studies to another species and found that a similar induction of *SDF1* occurs in kidney cortex mRNA and in isolated CCDs from rabbits fed an acid diet for 3 days (Figure 1). We had previously found that SDF1 and its receptor CXCR4 are widely expressed in the kidney (9).

ISH showed that in mice fed an acid diet, there was increased expression of SDF1 in cortical and outer medullary collecting tubules (Figure 2). Remarkably, however, the S1 segment of the proximal tubule had the highest level of expression.

Although we made intensive efforts to provide evidence for an increase in protein expression using Western blotting, IHC, and ELISA, we were unable to detect a signal, despite the use of three independently produced Abs.

***SDF1* increases H^+ secretion and decreases HCO_3^- secretion in rabbit CCDs.** The CCD is capable of both HCO_3^- absorption (i.e., H^+ secretion) and HCO_3^- secretion, each of which is mediated by a different IC type. To evaluate the fluxes produced by each cell type, we performed H^+ and HCO_3^- transport studies in isolated perfused rabbit CCDs using methods described previously (6, 7).

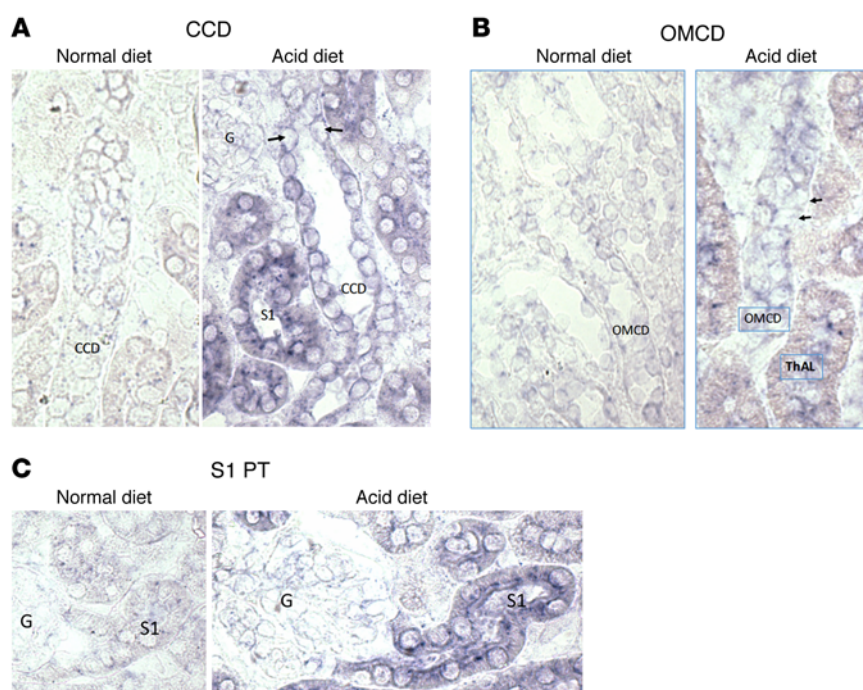


Figure 2. ISH of *SDF1* in mouse kidney cortex and medulla. (A) Images of the CCD and S1 segment of the proximal tubule. (B) Images of OMCDs. Note the thick ascending limb, which shows a high brown background but not much SDF1 staining. (C) Image of the S1 proximal tubule connecting with the glomerulus. Images were taken from mice fed a normal diet or a 2-day acid diet. Representative samples from 2 independent studies. Arrows in the CCD and OMCD images indicate heterogeneous, lower-intensity staining in different cells of these structures compared with more homogeneous staining in the S1 segment. Original magnification, $\times 600$. G, glomerulus; PT, proximal tubule; THAL, thick ascending limb.

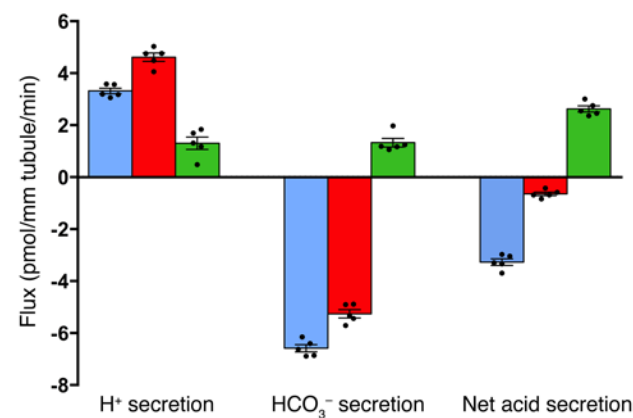


Figure 3. Effect of 10 nM SDF1 on acid-base transport in isolated perfused rabbit CCDs. Net acid secretion was measured as the change in HCO₃ concentration, after which the perfusate was changed to a Cl-free medium and the net acid-base transport measured again. Removal of Cl blocked HCO₃ secretion, thereby uncovering H⁺ secretion. After a period of control collections (blue bars), SDF1 was added to the bathing medium for 3 hours and the fluxes measured again (red bars). Green bars show the difference between SDF1 and control fluxes. Numerical values are provided in Table 1. Data represent the mean ± SEM; n = 5.

HCO₃ is secreted into the lumen by the β-IC, which has a Cl:HCO₃ exchanger (pendrin) in the apical membrane and a H⁺ ATPase in the basolateral membrane. HCO₃ is absorbed by the α-IC, which has an apical H⁺ ATPase and a basolateral Cl:HCO₃ exchanger. In the CCD, net HCO₃ transport is first measured as the change in HCO₃ concentration in the perfusate per unit time per unit length of CCD. Then, the perfusing medium is changed to a Cl-free solution, which inhibits the function of the β-IC, given that pendrin is a strict Cl:HCO₃ exchanger. The flux of HCO₃ remaining in the absence of luminal Cl is therefore that of HCO₃ absorption (i.e., H⁺ secretion) by the α-IC. Figure 3 and Table 1 provide the results of these studies (fluxes are given in pmol/min/mm tubule length). These fluxes were measured again (Figure 3, red bars) in the same tubules after a 3-hour incubation at pH 7.4 in the presence of 10 nM SDF1. SDF1 significantly (*P* < 0.01) increased HCO₃ absorption (H⁺ secretion) and reduced HCO₃ secretion compared with the fluxes observed during a 3-hour control incubation in the absence of SDF1.

Blockade of the receptor for SDF1 inhibits the acid-mediated change in acid-base transport. CXCR4 is the receptor for SDF1, and we used its selective inhibitor AMD3100 (plerixafor) to examine the role of the SDF1/CXCR4 axis in the response of the tubule to acid incubation (Figure 4 and Table 2). After an initial control period during which fluxes were measured at pH 7.4 (Figure 3, blue bars), tubules were exposed to an acid-bathing medium (pH 6.8) for 3 hours, and transport rates were measured again (red bars with [Figure 4B] or without [Figure 4A] 1 μM AMD3100; rates are in

pmol/min/mm tubule length). Note that acid media (Figure 4, red bars) changed net HCO₃ secretion to net HCO₃ absorption (compare the -3.8 blue bar, which changed to a +2.6 red bar in Figure 4A). The change in net flux was produced by a reduction in HCO₃ secretion and an increase in H⁺ secretion, but when the acid media contained AMD3100 (Figure 4B), the normal response to acidosis was prevented (compare the net acid secretion of +0.4 [red bar] in Figure 4B with that of +2.6 [red bar] in Figure 4A). Figure 4C shows that the 3-hour incubation at a constant pH of 7.4 did not alter the rates of acid-base transport. These results demonstrate in the aggregate that SDF1 mediates the effect of extracellular acidosis to increase H⁺ secretion and decrease HCO₃ secretion largely through its action on its receptor CXCR4.

Does the SDF1/CXCR4 pathway mediate change in the proportion of β- and α-ICs? The response of the CCD to acidosis might involve acute stimulation of H⁺ secretion and inhibition of HCO₃ secretion through processes such as a change in the abundance of various transporters on the cell membrane. Alternatively, it might mediate a change in the number of α-ICs and/or β-ICs, which occurs in states of chronic acidosis. To examine whether SDF1 affects the abundance of the different subtypes of ICs, we induced acidosis in mice and measured the number of α- and β-ICs in the cortex. We define α-ICs as those cells with apical H⁺ ATPase and a basolateral AE1 and β-ICs as those with apical pendrin and a basolateral H⁺ ATPase. In each study, the response of mutant mice and their non-mutant littermates to an acid diet was compared. The littermates are referred to herein as WT mice for simplicity.

Table 1. Effect of 10 nM SDF1 added to the bathing medium on H⁺ and HCO₃ fluxes in isolated perfused rabbit CCDs

	Before acid incubation			+3-hour incubation			Difference		
	JH ⁺	JHCO ₃	Jnet	JH ⁺	JHCO ₃	Jnet	ΔJH ⁺	ΔJHCO ₃	ΔJnet
Vehicle	Vehicle								
Average	3.23	-6.57	-3.34	3.5	-6.87	-3.37	0.27	-0.30	-0.03
SD	0.35	0.55	0.37	0.38	0.74	0.46	0.19	0.29	0.10
Vehicle	+10 nM SDF1								
Average	3.32	-6.58	-3.27	4.62 ^A	-5.26 ^A	-0.64 ^A	1.30 ^A	1.33 ^A	2.63 ^A
SD	0.29	0.66	0.48	0.33	0.43	0.21	0.39	0.46	0.33

After measurement of initial fluxes, the bathing medium was changed to contain SDF1 or its vehicle. After a 3-hour incubation, fluxes were measured again. All solutions were at pH 7.4. The reported studies are an average of 5 independent studies. ^A*P* < 0.01 using a 2-tailed paired *t* test. J, flux; Jnet, net flux.

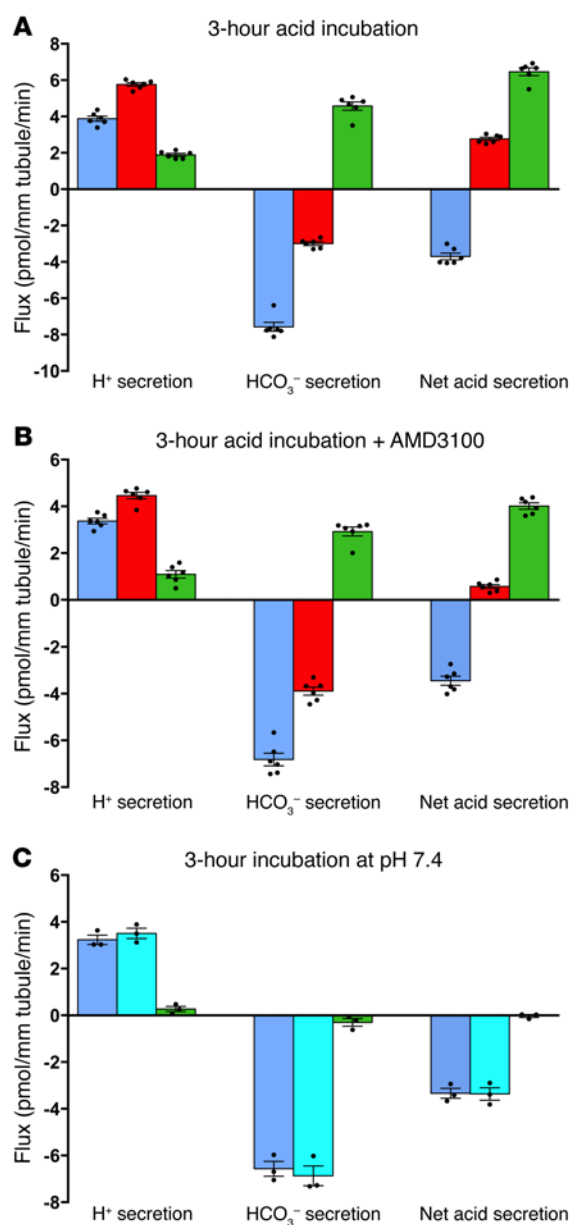


Figure 4. Acid-base transport in isolated perfused rabbit CCDs: effect of the CXCR4 antagonist AMD3100. Tubules were perfused and fluxes were measured at pH 7.4, following which the bathing medium was replaced with the same solution, except at pH 6.8. Three hours later, fluxes were measured again. **(A)** Initial fluxes at pH 7.4 (blue bars) and effect of a 3-hour incubation at pH 6.8 (red bars); $n = 6$. The difference between these 2 fluxes is indicated in the green bars. **(B)** Same as in **A**, except the medium contained 1 μ M AMD3100; $n = 6$. **(C)** Initial fluxes at pH 7.4 (blue bars) and the effect of a 3-hour incubation at pH 7.4 (cyan bars); $n = 3$. Data represent the average \pm SEM. Numerical values of the mean \pm SEM are provided in Table 2.

mice deleted for *Cxcr4* in the IC lineage, 39% were AE1⁺ (and 61% were pendrin⁺). However, acid feeding had no effect on the proportion of AE1⁺ or pendrin⁺ cells. These results represent studies of 4 mice in the *Cxcr4*-KO lineage and 12 WT mice. In each group of 4 mice, we counted 6,576–8,863 ICs (Supplemental Table 2). In other studies, in which we stained for aquaporin 2 and VB1 ATPase, the mutations (or acidosis) had no effect on the ratio of intercalated to principal cells (data not shown).

To confirm the role of this pathway in whole-body acid-base balance, we measured the plasma HCO₃ concentration and found that when WT littermates were fed an acid diet for 3 days, their plasma HCO₃ decreased by approximately 3.8 mEq/l. However, the same acid diet in the mutant mice caused a decrease of almost twice the level of that in their littermates (Table 3). These studies demonstrate that SDF1 and its receptor CXCR4 are critical for determining the response of ICs to acidosis.

These results show that the SDF1/CXCR4 pathway mediates the effect of acidosis on IC remodeling in the CCD. However, SDF1 binds and activates another G protein-coupled receptor, CXCR7 (13). We found that this alternative receptor was expressed in the collecting tubule at levels similar to those of CXCR4 (Supplemental Figure 2). As shown above, deletion of *Cxcr4* inhibited tubular adaptation by more than 80%, suggesting that it is the major receptor for SDF1 in these segments during acidosis. Whether CXCR7 signaling contributes to the effects of acidosis on ICs will require more definitive studies.

Which cell in the renal tubule produces SDF1 in response to acidosis? Since isolated collecting tubules by themselves are capable of producing SDF1 (Figure 1), we asked which of its two cell types, principal or intercalated, was responsible for SDF1 production. SDF1 and CXCR4 are targets of the hypoxia-sensing transcription factor HIF1 α (14). Regulation of SDF1 production is thought to be largely (or entirely) due to the transcriptional activity of HIF proteins rather than to the release of stored material in the cell. To identify the acid-sensing cell, we first crossed *Hif1 α* -floxed mice (*Hif1 α ^{f/f}*) with two Cre-expressing lines. In one line, we deleted *Hif1 α* from the intercalated lineage (using the VB1-Cre line; Figure 5B). Surprisingly, in response to an acid diet, the number of α -ICs increased and the number of β -ICs decreased in a quantitative manner nearly identical to that of their WT littermates. Similarly, the acid-base balance in the whole animal, as reflected by reductions in the plasma HCO₃ concentration, was similar to that of WT mice (Table 1). This result suggests that SDF1 continued to be active in the absence of its major regulator in the IC. Given that the effect of SDF1 was blocked by AMD3100 in isolated perfused tubules (Figure 4) and that it was produced by isolated CCDs

To investigate whether the SDF1/CXCR4 pathway is critical to the process of remodeling, we deleted *Cxcr4* in all ICs by crossing *Cxcr4*-floxed mice (*Cxcr4^{f/f}*) (a gift of Dan Littman, New York University [NYU], New York, New York, USA) with those carrying Cre under control of the promoter of the VB1 subunit of vacuolar ATPase [Tg(*ATP6V1B1*-Cre), herein referred to as VB1-Cre], a promoter whose renal activity is limited to the ICs (Supplemental Figure 1) (10). We stained kidney cortex sections for VB1 ATPase, AE1, or pendrin in mice with *Cxcr4* deletion and in their WT littermates. Figure 5A (see also Supplemental Figure 3 and Supplemental Table 2) shows that in the cortex of WT mice fed a normal diet (blue bars), 56% of the ICs stained for AE1 (i.e., they were α -ICs) and 44% were pendrin⁺ (i.e., they were β -ICs). When fed an acid diet for 2 days (Figure 5, red bars), the fraction of AE1⁺ cells increased to 78%, with a concomitant reduction of the pendrin⁺ cells to 22%. These results are similar to those of numerous other studies in different species in which acidosis increased α -ICs and reduced β -ICs (11, 12). We found that in

Table 2. Effect of 1 μ M AMD3100 on H^+ and HCO_3^- fluxes in isolated perfused rabbit CCDs

	Before acid incubation			+3-hour acid incubation			Difference		
	JH^+	$JHCO_3^-$	J_{net}	JH^+	$JHCO_3^-$	J_{net}	ΔJH^+	$\Delta JHCO_3^-$	ΔJ_{net}
Vehicle									
Average	3.88	-7.57	-3.70	5.76	-3.00	2.77	1.89	4.58	6.46
SD	0.34	0.60	0.45	0.24	0.25	0.22	0.21	0.56	0.51
Vehicle									
	+100 μM AMD3100								
Average	3.37	-6.82	-3.45	4.46 ^A	-3.89 ^A	0.57 ^A	1.09 ^A	2.93 ^A	4.02 ^A
SD	0.29	0.66	0.48	0.33	0.43	0.21	0.39	0.46	0.33

After measurement of initial fluxes, the bathing medium was changed to one with a pH of 6.8 as described in Methods. After a 3-hour incubation, fluxes were measured again. AMD3100 was added, and the reported studies are an average of 6 independent studies. ^A $P < 0.01$ using a 2-tailed unpaired t test. $n = 6$ independent studies for each group.

(Figure 1), it is likely that it was produced by the principal cell. SDF1 is poorly diffusible and tightly binds to a variety of extracellular matrix proteins (15).

To test the hypothesis that the adjacent principal cells might be producing SDF1, we deleted *Hif1a* from both principal cells and ICs (using the *Hoxb7-Cre* line; Figure 5C). When *Hif1a* was deleted from principal cells (and ICs), the response to acidosis was abolished. While the response in WT littermates to a 2-day acid challenge was an increase in α -IC and a decrease in β -IC numbers, these numbers remained constant in mice lacking HIF1 α in principal cells (and ICs), despite the acid challenge. We also observed a substantially defective response of the kidney to an acid challenge, with a decrease of 6 mEq/l in HCO_3^- concentration in the mutant mice compared with a 3 mEq/l decrease in their WT littermates (Table 3). These results confirm the physiological significance of changes in the proportion of α - and β -ICs in response to SDF1.

To confirm this conclusion, we performed immunolocalization of SDF1 in rabbit kidney cortex to demonstrate the generality of this finding. The results shown in Figure 6 demonstrate that in rabbit kidney cortex and medulla, SDF1 was expressed in the collecting tubule only in the principal cells. Similarly, in the medulla of the kidney, a region known to be hypoxic and hence to have higher levels of HIF1 α expression, higher expression levels of this transcription factor were found only in the principal cells. Western blot analysis of mouse kidneys had previously shown that SDF1 expression levels are higher in the medulla than in the cortex (9). ISH (Figure 2) showed that SDF1 was induced by acidosis in the S1 segment as well as in the CCDs and outer medullary collecting ducts (OMCDs). Examination of the OMCD images showed that, while the majority of the cells expressed SDF1, there were always a few cells that lacked any expression (Figure 2B, arrows). Similar results were obtained in the CCDs (Figure 2A, arrows); however, there were far fewer images of CCDs that we were able to rigorously identify as CCDs, since no counterstain was used. Given that the majority of cells in the collecting tubule are principal cells, we suggest that this staining pattern confirms the conclusions of the *Hif1a*-deletion studies. In summary, it appears that the ICs themselves do not produce SDF1 to provide paracrine signaling. Rather, when *Hif1a* (and thus, presumably, SDF1) was deleted from principal cells as well, the results were very different, demonstrating, surprisingly, that the acid-sensing cell in the CCD is actually the

principal cell, a cell heretofore not known to participate directly in acid-base balance.

While it is well known that *Sdf1* and *CXCR4* are target genes of the HIF system, we investigated whether the activation of *Sdf1* transcription was mediated entirely by HIF1 α and not some other acid-sensitive pathway such as the secretion of preformed SDF1. We used two CCD cell lines, M-1 (16) and mouse CCD (mCCD) (17) cells, which have many of the characteristics of principal cells of the collecting tubule. Using qPCR, we found that incubation of these cells at pH 6.8 for 18 hours increased SDF1 transcription by 1.8 ± 0.1 -fold in M-1 cells ($n = 3$) and by 1.8-fold in mCCD cells ($n = 2$), levels similar to those detected in mRNA of both kidney cortex and isolated CCDs (Figure 1). When these cells were exposed to acid media in the presence of 5 μ M acriflavine, a drug that interrupts the dimerization of HIF1 α and ARNT (18), qPCR showed that *Sdf1* transcription levels were drastically reduced to 0.24-fold of the levels detected in the control cell line incubated at pH 7.4. At this dose, acriflavine had no deleterious effect on cell survival in our studies or in studies in other cells (18). Acid incubation of M-1 cells also caused a 1.8-fold increase in transcription of carbonic anhydrase IX, a classic target gene of HIF1. Further, HIF1 transcription factors are hydroxylated on prolines by prolyl hydroxylases that utilize α -ketoglutarate and oxygen as substrates. Prolyl hydroxylation allows vHL to bind and ubiquitinate HIF proteins, thereby causing their degradation. Addition of a permeable α -ketoglutarate to acid-incubated M-1 and mCCD cells reduced the expression levels of *Sdf1* to 0.5-fold from 1.8-fold. These results demonstrate that the acid-sensing pathway causes an increase in SDF1 production through activation of HIF1 α .

Discussion

The renal response to chronic acidosis is mediated by ammoniogenesis, which occurs in the proximal tubule (19), and by an increase in proton secretion in the collecting tubule. Acidosis causes an increase in the number of acid-secreting cells at the expense of HCO_3^- -secreting cells (6). Proliferation of acid-secreting α -ICs was also reported in some studies (20, 21). In the outer medullary collecting tubule, there is also an increase in the amount of acid secreted by each cell in chronic acidosis (22). While, in principle, many mechanisms might be invoked to mediate this critical homeostatic response, it is clear that the machinery for regulation

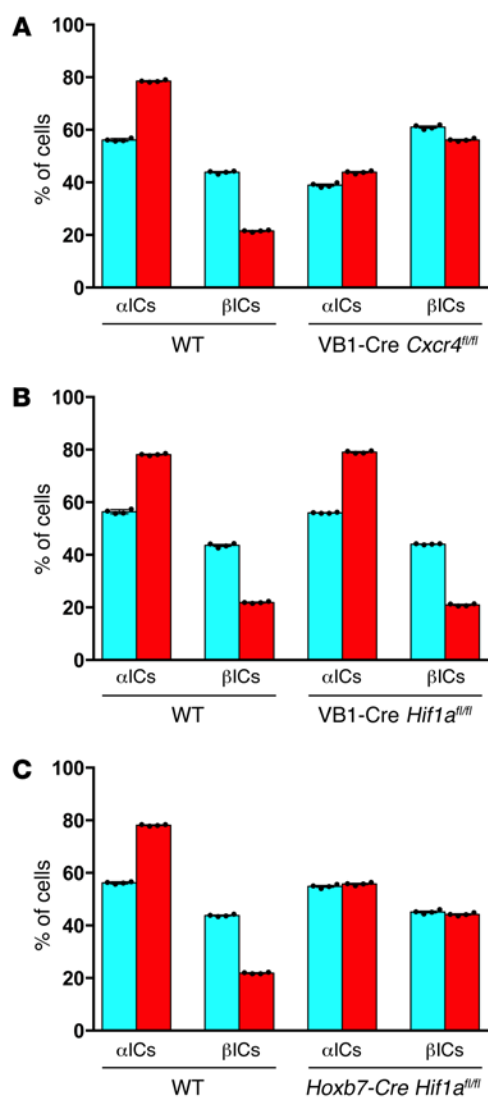


Figure 5. Distribution of IC subtypes in kidney cortex from control and KO mice. ICs were identified as those cells expressing the B1 subunit of the H⁺ATPase, α-ICs as those that also expressed KAE1, and β-ICs as those that expressed pendrin. (A–C) Each panel represents studies performed in 4 mice fed a normal diet (blue bars) and 4 littermates fed a pellet diet containing 2% NH₄Cl (red bars). Each bar represents the average (± SD) of the cell counts from 4 mice. For each study in A–C, we examined 8 mice expressing the active Cre recombinase and another 8 mice that were their WT littermates. Four mice of each genotype were fed a normal diet and the other 4 an acid-supplemented diet. In each group of 4 mice, we counted between 6,576 and 8,863 ICs (the raw data are provided in Supplemental Table 2).

of the chronic response is present in the CCD without any need to invoke freely circulating factors. Here, we show that activation of the hypoxia-signaling pathway occurs during acidosis and that one consequence of this activation is the production of the chemokine SDF1 (CXCL12). The most surprising finding is that it appears to be the principal cell, not the IC, that responds to acidosis by activating the HIF pathway.

SDF1 is a basic growth factor with limited diffusion capacity. It is well known to bind to extracellular matrix components, especially glycosaminoglycans, heparin sulfates, and other com-

ponents located at the cell surface and in the extracellular matrix (23). We previously found that all cells in the collecting tubule produce the extracellular matrix protein hensin (DMBT1), yet during acidosis, it is polymerized and deposited underneath the acid-secreting cell (24). SDF1 is perhaps bound to hensin and thus might be presented to its receptor on the basolateral surface of the IC. SDF1, when produced by a principal cell, needs to diffuse across only the intercellular space to the adjacent IC to exert its effect. Since principal cells outnumber ICs by a factor of approximately 2, each IC abuts at least 1 principal cell. Thus, acidosis has the capability of activating every IC.

Unfortunately, despite a large effort by the three laboratories involved in our collaborative work, we have failed to detect induction of SDF1A protein by acidosis while using three independently produced (commercial and private) Abs. Even though we were able to demonstrate such activities previously at one of our laboratories (9), it has been frustratingly difficult to reproduce these findings. Perhaps the reason for our previous success was that the kidneys became hypoxic or ischemic, either of which is a potent stimulus for HIF1 activation.

How might the principal cell respond to acidosis? In general, all cells contain acid-transporting proteins (e.g., Na:H or Cl:HCO₃ exchangers), and medium acidification will change the driving forces for proton/HCO₃ transport, leading to cellular acidification. Indeed, acidification of the media bathing the collecting tubules was found to acidify the cytoplasm of principal cells (25, 26). However, alternative (or additional) signaling mechanisms have been proposed, whereby proton-sensing G protein-coupled receptors might mediate the effect of acidosis; indeed, deletion of *Gpr4* led to renal tubular acidosis (RTA) (27).

How cell acidification might activate the hypoxia-signaling mechanism remains to be determined. Cellular acidosis might either reduce expression of vHL (or remove it from the cytoplasm), or inhibit the activity of prolyl hydroxylase(s). Acidosis was found to activate HIF1α in some cells in culture, as demonstrated in a recent study showing that exposure of cells in culture to acidosis resulted in activation of HIF1α due to the sequestration of vHL in the nucleolus (28). However, testing of this hypothesis in several cell lines revealed that this effect occurred in some lines, but did not appear to be a universal response (29). Prolyl hydroxylases require α-ketoglutarate as a substrate and produce succinate as a product. It is possible that acidosis alters cellular metabolism in such a way as to reduce the concentration of the substrate or increase that of the product. It will be necessary to perform studies in principal cells of the collecting tubule, where the role of these two possible mechanisms can be directly examined.

One of the important recent clinical discoveries has been the identification of acidosis as a mediator of worsening kidney function. Treatment of patients with chronic kidney disease (CKD) with oral sodium bicarbonate reduced the rate of progression of renal failure (30). Similarly, large-scale epidemiologic and clinical studies showed that the lower the serum HCO₃, the worse the outcome for kidney function (31). How could acidosis produce this progression? Acidosis induces the production of endothelin, a factor that might lead to progression (32). It is interesting to note that endothelin is also a target gene of HIF1α (33). There is also extensive evidence that endothelin-1 itself can activate HIF1α (reviewed in ref. 34), sug-

Table 3. HCO₃ concentration (in mEq/l) in WT and mutant mice fed a normal diet or an acid-supplemented diet for 3 days

	Normal	Acid	Δ		Normal	Acid	Δ	δΔ
WT	26.2 ± 0.3	22.4 ± 0.2	3.79 ± 0.7	VB1-Cre <i>Cxcr4</i> ^{fl/fl}	26.7 ± 0.9	20.1 ± 0.3	6.6 ± 1.3	2.8 ± 0.8 ^A
WT	26.3 ± 0.5	23.9 ± 0.6	2.35 ± 0.13	VB1-Cre <i>Hif1a</i> ^{fl/fl}	26.2 ± 0.8	24.4 ± 0.1	1.8 ± 0.7	0.7 ± 0.6
WT	23.9 ± 0.3	21 ± 0.7	3.1 ± 0.6	<i>Hoxb7</i> -Cre <i>Hif1a</i> ^{fl/fl}	24.2 ± 0.7	18.6 ± 0.7	6.03 ± 0.5	3.0 ± 0.9 ^A

Data represent ± SD. Each group consisted of 4 mice. WT mice were littermates of the mutants and hence were not, strictly speaking, WT. ^AP < 0.01.

gesting the possibility of a positive feedback system involving interaction of microvasculature and epithelial cells of the kidney.

SDF1 was discovered as a chemoattractant for a variety of inflammatory cells, and its s.c. injection led to skin invasion by lymphocytes (35). How might SDF1 injure the kidney? SDF1 attracts monocytes and lymphocytes, but not neutrophils, in vitro and in vivo (35). Since this discovery, a host of studies have continued to show the critical importance of SDF1 to the migration of BM-derived cells, including stem cells. Its receptor CXCR4 has been found to be highly expressed in all these cells as well as in macrophages and DCs. There is strong evidence for the involvement of SDF1/CXCR4 signaling in tissue inflammation in the kidney and other organs. In ischemic kidney injury, SDF1 is induced by epithelial cells of the nephron, leading to invasion by BM-derived cells (36). Wang et al. showed that all mouse models of lupus were associated with increased expression of SDF1 in the kidneys and invasion of CXCR4-expressing monocytes, B cell subsets, and plasma cells (37). Feeding animals a high-protein diet (which induces acidosis) was associated with invasion of the kidney by lymphocytes (38). Interestingly, studies of human renal biopsies from patients with CKD of various causes revealed local production of SDF1 in the collecting tubule and attraction of inflammatory cells to these discrete regions of the kidney (39). To our knowledge, no one has searched for invasion of immune cells into the kidney in states of “pure acidosis” such as RTA, probably because such patients rarely undergo biopsies and because tubulointerstitial disease is commonly a consequence and a cause of RTA. Further, in distal RTA, we often see nephrocalcinosis, which is likely due to Ca being leached out of bone, but which could also be due to acidosis-induced inflammation. Yet there are suggestions that the kidneys in acidotic patients are invaded by immune cells; half of the patients with methylmalonic aciduria (as pure an acidosis as one can get) develop significant renal failure (40) and most had tubulointerstitial infiltrates. Mice with dysfunctional NaHCO₃ cotransporters develop severe acidosis, and their kidneys show substantial amounts of mononuclear cell infiltrates (41). On the basis of these studies, we propose that SDF1 plays a pathogenic role in the progression of CKD by attracting immune cells to the kidney and priming them there.

Methods

Animals

VB1-Cre mice were generated by Miller et al. (10), and the *Hoxb7*-Cre line was generated by Bates et al. (42). All mouse lines used in this study were on a C57BL/6N background. The *Cxcr4*-floxed line was a gift of Dan Littman (NYU, New York, New York, USA) and have now been donated to and are available from The Jackson Laboratory (cat-

alog 008767 B6.129P2 *Cxcr4*<tm2Yzo>/J). The *Hif1a*-floxed line was purchased from The Jackson Laboratory (catalog 007561 B6.129-Hif1a<tm3Rsjo>/J). Breeding of the mice was performed under the supervision of the Columbia University Animal Care Facility. Mice were placed in metabolic cages and given water and food ad libitum. Blood was collected using intracardiac or retro-orbital puncture and injected into a blood gas apparatus for determination of pH, PCO₂, and HCO₃. All measurements were made in littermates of WT and mutant mice. When we tried to give the mice NH₄Cl (280 mM) in the drinking water (in a 2% sucrose solution), they did not drink it and became dehydrated. We estimated the amount of NH₄Cl needed to make the mice acidotic and their average food intake and asked a food supplier (Test-Diet) to make 2% NH₄Cl mouse food in pellet form. The mice tolerated this much better but were still given 2% sucrose in water to induce a high urine volume. Blood gases were measured using a clinical blood gas analyzer. Mice were placed in metabolic cages for 2 days before the diet was changed. The urine pH declined after 1 day of the acid diet, and urine for net acid excretion was collected daily after the beginning of the acid diet. Rabbits were given 75 mM NH₄Cl in the drinking water and restricted to 30 g per day of food to induce metabolic acidosis (7).

CCD sorting

Kidneys from VB1-GFP mice were prepared and collecting ducts sorted as previously described by Miller et al. (43). Acidosis in VB1-GFP mice was induced by adding 280 mM NH₄Cl to the drinking water.

Tissue fixation and preparation

Animals were anesthetized with i.p. ketamine (75–95 mg/kg BW) and perfused through the left ventricle first with PBS (0.09% NaCl in 10 mM phosphate buffer, pH 7.4), followed by pyridoxal-PLP fixative (4% paraformaldehyde [PFA], 75 mM lysine-HCl, 10 mM sodium periodate, and 0.15 M sucrose in 37.5 mM sodium phosphate). Both kidneys were removed and sliced and were further fixed by immersion in PLP

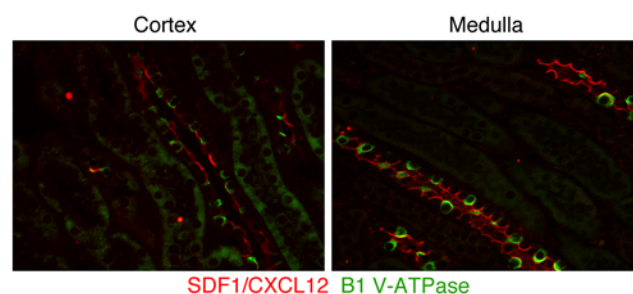


Figure 6. IHC of SDF1 in collecting ducts of rabbit kidney cortex and medulla. Red staining shows SDF1, and green staining shows the VB1 subunit of H⁺ ATPase (i.e., ICs). Original magnification, ×400.

for 4 hours at room temperature and overnight at 4°C, then extensively rinsed in PBS and stored at 4°C in PBS containing 0.02% sodium azide until use. To prepare frozen sections, tissues were cryoprotected by immersion in 0.9 M (30%) sucrose in PBS overnight at 4°C and mounted for cryosectioning in Tissue-Tek (Sakura) before freezing in liquid nitrogen and sectioning at 5-μm thickness with a Leica CM3050 cryostat (Belair Instrument Company). Sections were picked up on Fisherbrand Superfrost Plus charged glass slides (Fisher Scientific).

Immunofluorescence and confocal microscopy

Fixed sections were hydrated in PBS for 10 minutes and treated by immersion in sodium citrate buffer (10 mM sodium citrate, 0.05% Tween-20, pH 6.0) at 95°C for 10 minutes for retrieval of antigenic sites. Sections were washed in PBS 3 times for 5 minutes and blocked in a solution of 1% BSA/PBS/sodium azide for 15 minutes to prevent non-specific staining, followed by a 90-minute incubation in the primary Ab at room temperature. After three 5-minute PBS washes, the secondary Ab was applied for 1 hour at room temperature, and the slides were then rinsed again in PBS 3 times for 5 minutes. Slides were mounted in fluorescent mounting medium (Dako, Agilent Technologies). For dual staining with Abs raised in different species, the primary Abs were applied sequentially at the appropriate concentrations, as described below, with each primary Ab being followed by the corresponding secondary Ab. Digital images were acquired with a Microphot SA EPI-FL3 fluorescence microscope (Nikon Instruments) with a SPOT-RT 941 SE digital camera (SPOT Imaging Solutions, Diagnostic Instruments Inc.). Images were then analyzed using SPOT Advanced image-processing software and imported into and printed from Adobe Photoshop version 6.0 image-editing software (Adobe Systems). For confocal laser-scanning microscopy, tissue sections were prepared as described above. Confocal imaging was performed on an LSM 510 Meta confocal microscopy system (Carl Zeiss MicroImaging) using LSM Image Browser software, and images were edited as described above.

Abs

The following primary Abs were used: (a) an affinity-purified rabbit polyclonal Ab raised against rat AE1 anion exchanger at a dilution of 1:100 (Alpha Diagnostic; catalog AE11-A); (b) an affinity-purified rabbit polyclonal Ab raised against pendrin (Santa Cruz Biotechnology Inc.; catalog sc-50346); (c) a rabbit polyclonal Ab raised against AQP2 at a 1:100 dilution (Sigma-Aldrich; catalog A7310); (d) a rabbit polyclonal affinity-purified Ab raised against V-ATPase B1/2 at a dilution of 1:100 (Santa Cruz Biotechnology Inc.; catalog sc-20943); and (e) a mouse mAb raised against V-ATPase B1/2 at a dilution of 1:100 (Santa Cruz Biotechnology Inc.; catalog sc55544).

Abs against SDF1. For SDF1 immunofluorescence, we used anti-SDF1 goat polyclonal Ab (Santa Cruz Biotechnology Inc.; C-19, catalog sc-6193). We also used K15C mAb generated against the N-terminal peptide KPVLSYRSPSRFFC. For SDF1 ELISA, we used a kit (Abcam; ab 100741, specific to mouse SDF1A) containing Abs against aa 22–89, which encompass all of SDF1A. Secondary Abs labeled with Texas Red (TR), TRITC (rhodamine), and FITC (fluorescein) were from Jackson Immuno-Research and used at a dilution of 1:100 to 1:200 after reconstitution.

Microperfusion of isolated CCDs from rabbits

CCDs were microdissected from rabbit kidneys and perfused as described previously (7, 44). Three-hour incubations were performed

at pH 7.4 and pH 6.8 (7). Net bicarbonate transport was measured using a WPI NanoFlo microfluorometer (World Precision Instruments). Cl-free solutions were perfused to separate H⁺ and HCO₃ fluxes as recently described (7).

Microdissected mouse nephron segments

Microdissected mouse nephron segments were isolated for qPCR of CXCR4 and CXCR7 as previously described (45). qPCR was performed with a SYBR Green amplification kit (Roche).

Primers for qPCR and genotyping

RNA was isolated and followed by clean-up with the PureLink RNA Mini Kit (Invitrogen) according to the manufacturer's instructions. For qPCR analysis, RNA was reverse transcribed and subjected to PCR amplification using either SYBR Green PCR Master Mix or TaqMan Universal PCR Master Mix on an AB 7500 platform (all from Applied Biosystems). mRNA expression levels were quantified using the relative standard curve method according to the manufacturer's instructions (Applied Biosystems).

The following mouse primer pairs were used for amplification: *Sdfl*: forward, 5'-CCGCTGCCGCACTTCTACTCT-3', reverse, 5'-GCTTGACGTTGGCTCTGGCGA-3'; carbonic anhydrase IX: forward, 5'-GCCACGCAACCCTTGAATG-3', reverse, 5'-AGTTTCTGT-CATCTCTGCCGG-3'; *Cxcr4*: forward, AGCATGACGGACAAGTACC, reverse, GATGATATGGACAGCCTTACAC; *Rpl26*: forward, GCTAATGGCACAACCGTC, reverse, TCTCGATCGTTTCTTCCTTGAT. *Cxcr7* qPCR was performed using a TaqMan assay kit (Mm00432610_m1) from Life Technologies. The following rabbit primer pairs were used for amplification of *Sdfl*: forward, 5'-AAACCCGTCAGCCTGAGC-TACAGA-3', reverse, 5'-TCTTGAGATGCTTGACGTTGGCTC-3'. For *VB1*: forward, 5'-ATCAACGTGCTCCCTTCTCTGTCA, reverse, 5'-TGTGCAGGAATCCAGGTAGAGCA.

The following mouse primer pairs were used for genotyping: *VB1-Cre*: forward, 5'-CATTACCGGTCGATGCAACGAG-3', reverse, 5'-TGCCCCCTGTTTCTACTATCCAGG-3'; *Cxcr4* LoxP: forward: 5'-CCACCCAGGACAGTGTGACTCTAA-3', reverse, 5'-GATGGGAT-TTCTGTATGAGGATTAGC-3'; *Hif1a* LoxP: forward, 5'-TGCTCAT-CAGTTGCCACTT-3', reverse, 5'-GTTGGGGCAGTACTGGAAAG-3'; *HoxB7-Cre*: forward, 5'-GGTCACGTGGTCAGAAAGAGG-3', reverse, 5'-CTCATCACTCGTTGCATCGA-3'.

ISH of SDF1 in mouse kidneys

Probe synthesis for ISH. Mouse kidney mRNA was reverse transcribed into cDNA using SuperScript III First-Strand Synthesis SuperMix for qPCR (Invitrogen; catalog 11752-050), and a 222-bp fragment of the *Sdfl* gene was amplified using the following primers: forward, 5'-CCGCTGCCGCACTTCTACTCT-3' and T7-embedded reverse, 5'-GGATTACCTAA-TACGACTCACTATAGGGATATGCTATGGCGGAGTGCTT-3'. The PCR product was used as a template for in vitro transcription. Probe was synthesized by T7 RNA polymerase (Roche), and digoxigenin-labeled (DIG-labeled) RNA was subsequently purified by a PureLink RNA Mini Kit (Ambion, Life Technologies; catalog 12183018A).

ISH of frozen sections. Kidneys from mice fed a normal or acid diet (see above) were perfusion fixed, collected in ice-cold PBS, and fixed overnight at 4°C in 4% PFA in PBS and soaked in 30% sucrose in PBS for 24 hours, then processed directly for embedding in OCT. Frozen sections (8-μm) were air dried for 1 to 3 hours, then fixed in 4%

PFA for 10 minutes at room temperature. After washing 3 times with PBS, sections were treated with proteinase K (1 µg/ml for 5 minutes at room temperature), then washed 3 times with PBS, acetylated for 10 minutes at room temperature, and washed again with PBS. Prehybridization was performed for 2 hours or overnight at room temperature in a humidified chamber. Hybridizations were performed at 68°C to 72°C overnight in a humidified chamber. Prehybridization and hybridization solution was composed of 50% formamide, 5× SSC, 5× Denhardt's solution, 250 µg/ml baker's yeast RNA (Sigma-Aldrich), and 500 µg/ml herring sperm DNA (Sigma-Aldrich). Washes were performed at 72°C in 5× SSC for 5 to 10 minutes, then at 72°C in 0.2× SSC for 1 hour. Sections were stained overnight with anti-digoxigenin Ab (Boehringer Mannheim) at a 1:5,000 dilution in 0.1 M Tris-HCl, pH 7.5, containing 0.15 M NaCl with 1% heat-inactivated goat serum. After staining overnight at 4°C in a humidified chamber, slides were washed in 0.1 M Tris-HCl (pH 7.5) containing 0.15 M NaCl. Alkaline phosphatase activity was detected by developing slides in BCIP/NBT (Boehringer Mannheim) and 0.25 mg/ml levamisole in a humidified chamber for 6 to 24 hours in the dark. Sections were dehydrated and mounted in Permount (Fisher Scientific).

Statistics

The data in the flux studies shown in Figures 3 and 4 were all analyzed in paired samples, hence the paired *t* test was calculated using a 2-tailed test. For qPCR (Figure 1) and cell count results (Figure 5), SDs were calculated.

Study approval

All studies were conducted in accordance with protocols approved by the IACUCs of the Institutional Animal Care Facility of Columbia University and the University of Rochester School of Medicine. The studies performed in Paris were approved by the Comité d'éthique pour l'expérimentation animale of the Université René Descartes (Paris, France).

Author contributions

QA, GJS, and DE designed the studies; GJS, XBG, ST, JMP, HP, NP, and VD'A performed various parts of the study; GXB, GJS, DE, and QA analyzed the data; QA wrote the manuscript; QA, GJS, and DE edited the manuscript.

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