A precise equilibrium between cellular differentiation and proliferation is fundamental for tissue homeostasis. Maintaining this balance is particularly important for the liver, a highly differentiated organ with systemic metabolic functions that is endowed with unparalleled regenerative potential. Carcinogenesis in the liver develops as the result of hepatocellular de-differentiation and uncontrolled proliferation. Here, we identified SLU7, which encodes a pre-mRNA splicing regulator that is inhibited in hepatocarcinoma, as a pivotal gene for hepatocellular homeostasis. SLU7 knockdown in human liver cells and mouse liver resulted in profound changes in pre-mRNA splicing and gene expression, leading to impaired glucose and lipid metabolism, refractoriness to key metabolic hormones, and reversion to a fetal-like gene expression pattern. Additionally, loss of SLU7 also increased hepatocellular proliferation and induced a switch to a tumor-like glycolytic phenotype. Slu7 governed the splicing and/or expression of multiple genes essential for hepatocellular differentiation, including serine/arginine-rich splicing factor 3 (Srsf3) and hepatocyte nuclear factor 4α (Hnf4α), and was critical for cAMP-regulated gene transcription. Together, out data indicate that SLU7 is central regulator of hepatocyte identity and quiescence.

Introduction

The liver performs a variety of unique functions essential for the preservation of homeostasis, including glucose and lipid metabolism, xenobiotic detoxification, and serum protein synthesis. Most of these roles are performed by the hepatocyte, a quiescent and highly differentiated cell expressing a complement of enabling genes. The liver’s central position in systemic metabolism implies a prominent exposure to noxious stimuli derived from environmental toxicants, alcohol, viruses, and dietary habits, the principal causes of liver disease. To cope with these challenges, the liver has developed a remarkable and tightly controlled regenerative capacity based on the proliferation of hepatocytes in response to parenchymal loss (3). However, persisting injury may lead to the development of liver cirrhosis, a condition characterized by extracellular matrix accumulation and regenerative hepatocellular nodules (4). Another important hallmark of the chronically injured liver is progressive loss of function due to a gradual decrease in the expression of hepatospecific genes (5–8). These changes are often accompanied by unrestrained cell proliferation and reactivation of genes characteristic of the fetal hepatocyte (7, 9, 10).

Human and experimental studies indicate that hepatocellular dedifferentiation may be linked to the development of hepatocellular carcinoma (HCC), a most serious complication of chronic liver injury and cirrhosis (11). Clinical findings demonstrated that the degree of differentiation of HCC tissues positively correlated with patient survival (12). More recently, analyses of gene expression profiles in HCCs (13) and peritumoral cirrhotic liver tissues (14) provided molecular confirmation of these histological observations, linking expression patterns indicative of fetal and dedifferentiated status with poor patient prognosis. However, little is known about the mechanisms driving hepatocellular dedifferentiation during chronic liver disease and tumor development. Impaired expression of a limited set of genes generally known as liver-enriched transcription factors or hepatocyte nuclear factors (HNFs) (1) could play a fundamental role. The best characterized is the nuclear receptor HNF4α, a transcriptional regulator involved in embryonic liver development and in the maintenance of the differentiated phenotype of adult hepatocytes (15, 16). HNF4α expression is reduced in liver cirrhosis (7), contributing to experimental hepatocarcinogenesis (17). A recent study demonstrated that the pre-mRNA splicing regulator serine/arginine-rich splicing factor 3 (SRSF3) was also essential for hepatic maturation and metabolic function in mice (18), indicating that preservation of hepatocellular identity was not restricted solely to HNFs. Altered expression and activity of splicing factors has been lately reported in different tumor types, including HCC (19–21). These observations highlight the importance of accurate pre-mRNA splicing for liver homeostasis and the implication of deranged splicing in disease progression (22). Recently we reported that SLU7, a splicing factor critical for the correct selection of the 3’ splice site (23), was downregulated in human cirrhotic liver and HCC and that loss of SLU7 resulted in the aberrant production of a protumorigenic splice variant of the P73 tumor suppressor (24). Here we identified SLU7 as a fundamental gene for hepatocellular differentiation and quiescence. Slu7 knockdown in adult mouse liver gave raise to a striking phenotype, encompassing impaired glucose and lipid metabolism, loss of response to key metabolic hormones, and reversion to a fetal-like gene expression pattern. Moreover, Slu7 downregulation resulted in hepatocellular proliferation and a metabolic switch to a glycolytic metabolism.
phenotype reminiscent of that found in tumors (25–27). Mechanistically, SLU7 modulated the splicing and expression of genes essential for the preservation of the hepatocyte identity, including Srsf3 and Hnf4α, and we identified SLU7 as a key transcriptional regulator in the cAMP signaling pathway.

Results

Knockdown of SLU7 induces profound changes in pre-mRNA splicing and gene expression in human HCC cells. To investigate the role of SLU7 in human liver cell biology, we knocked down SLU7 expression in PLC/PRF/5 HCC cells and performed microarray analyses of exon utilization and gene expression. We detected 590 splice events (504 exons upregulated, 86 downregulated). Ingenuity Pathway Analysis of data according to cellular functions revealed that most of these altered exon use events occurred in genes involved in lipid and carbohydrate metabolism, cell cycle, proliferation, and survival pathways (Figure 1A and Supplemental Table 1; supplemental material available online with this article; doi:10.1172/JCI74382DS1). Interestingly, the most represented category was that of genes implicated in RNA posttranslational modification (Figure 1A). Gene expression microarray data were consistent with our observations in the exon utilization microarray: among the 243 genes with altered expression (85 upregulated, 158 downregulated), the majority of changes occurred in genes implicated in carbohydrate and lipid metabolism, cell proliferation and survival, and posttranscriptional RNA modification (Figure 1B and Supplemental Table 2). Most variations in exon utilization and gene expression affected metabolic genes typical of the mature hepatocyte phenotype, which suggests that SLU7 plays an important role in hepatocellular differentiation. This hypothesis was reinforced by our finding of altered SRSF3 pre-mRNA splicing upon SLU7 silencing (Supplemental Table 1). In view of the key role of SRSF3 in liver function (18), we further explored the influence of SLU7 on SRSF3 pre-mRNA splicing. SRSF3 pre-mRNA alternative splicing generates the full-length isoform lacking exon 4 (Iso1) and an alternative isoform including exon 4 (Iso2) (Figure 1C). As occurs for all SR genes, alternative isoforms like SRSF3 Iso2 contain a poison cassette exon introducing an in-frame premature termination codon promoting SR transcript degradation (28). SLU7 knockdown triggered an increase in the SRSF3 Iso2/Iso1 ratio in several human hepatoma cell lines (Figure 1, C and D). This response was validated in the nontransformed human liver cell line HepaRG (Figure 1E and ref. 29). Consistently, SLU7 overexpression in HepaRG cells had the opposite effect on the SRSF3 Iso2/Iso1 ratio (Figure 1F). SRSF3 expression in human liver tissues has not previously been examined. SRSF3 mRNA levels were reduced in HCC samples compared with control tissues, and a tendency toward reduced levels (without reaching statistical significance) was noted in cirrhotic liver tissues (Figure 1G). Interestingly, expression of SRSF3 and SLU7 directly correlated when all liver tissue samples were collectively analyzed (Figure 1H).

SLU7 defines a liver-specific gene expression pattern in human hepatocytes and is a developmentally regulated gene in the mouse liver. Next, we directly analyzed the influence of SLU7 on relevant genes characteristic of the hepatocellular identity. Upon SLU7 knockdown in HepaRG cells, the expression of albumin (ALB), HNF-4α, and methionine-adenosyltransferase 1A (MAT1A) (30) was significantly inhibited, while the oncofetal marker H19 (31) was induced (Figure 2A). Moreover, the ratio between the CYP4F3 splice variants CYP4F3A and CYP4F3B was markedly increased (Figure 1A), also indicative of hepatocellular dedifferentiation (32). Reduced CYP4F3B mRNA levels were also observed in our expression microarray analysis (Supplemental Table 2). Complementarily, SLU7 overexpression increased HNF4α and MAT1A expression (Figure 2B). These effects were reproduced in HepG2 cells (Supplemental Figure 1).

During mouse liver development, Slu7 mRNA followed a temporal expression pattern (Figure 2C), similar to that of Hnf4α and Mat1a (33), and opposite that displayed by Wilms’ tumor suppressor gene 1 (Wt1), which is expressed in fetal but not in adult hepatocytes and is reactivated in liver cirrhosis and HCC (7, 8). These findings were corroborated at the protein level (Figure 2D). Immunohistochemical analysis of SLU7 in adult mouse liver showed a predominantly hepatocellular and nuclear pattern (Figure 2E).

Knockdown of Slu7 in mouse liver disrupts the gene expression pattern characteristic of hepatocellular identity and markedly impairs physiological metabolic responses. We next assessed the role of SLU7 in vivo. Hepatocellular Slu7 expression was downregulated using an adeno-associated viral vector expressing a Slu7-specific shRNA regulated by a hepatocyte-specific promoter (referred to herein as AAV-shSLU7; Figure 3, A and B). Importantly, the effect of reduced SLU7 expression on SRSF3 alternative splicing that we had observed in vitro was reproduced in mouse livers (Figure 3C). Moreover, Slu7 knockdown markedly reduced Alb, Hnf1α, and Hnf4α gene expression and significantly altered the expression of genes involved in hepatic lipid, cholesterol, and glucose metabolism (Figure 3D). Importantly, these effects were accompanied by induction of the oncofetal markers alphafetoprotein (AfP) and Wt1 and of activating transcription factor 3 (Atf3) and P53, stress-responsive genes in the liver (Figure 3D and refs. 34, 35). The specificity of the effects of Slu7 on liver gene expression was better illustrated by reciprocal changes in specific isozymes transcripts. Slu7 knockdown resulted in downregulation of Mat1a, glutaminase 2 (Gl2), glucokinase (Gck), and L-pyruvate kinase (Lpk) and concomitant upregulation of Mat2a, Gl3, hexokinase 2 (Hk2), and pyruvate kinase m2 (Pkm2), together with a switch in the expression of Hnf4α isoforms derived from the P1 and P2 gene promoters and in the ratio of Insr A and B splice variants (Figure 3E and Supplemental Figure 2). The isoforms, Hnf4α P2-derived isoforms, and splice variants favored upon Slu7 downregulation were characteristic of the fetal and/or transformed hepatocyte, which exert profound effects on metabolism (26, 30, 36–40).

In association with these changes, Slu7 downregulation strongly influenced serum levels of glucose, cholesterol, and triglycerides and reduced intrahepatic glycogen stores (Figure 3F and Supplemental Figure 3). These effects were not related to liver damage, as only a mild elevation in serum alanine aminotransferase (ALT) was observed (98 ± 8 vs. 69 ± 5 UI/l, P = 0.008, n = 31), while aspartate aminotransferase (AST) remained unaltered (169 ± 14 vs. 163 ± 13 UI/l, P = 0.88, n = 31). No hepatic histopathological alterations were found in Slu7-depleted mice (data not shown).

The liver plays a central role in blood glucose homeostasis, breaking down glycogen stores in the postabsorptive phase and synthesizing glucose as fasting progresses (36). In addition to depleted glycogen stores, Slu7 knockdown induced fasting hypoglycemia (96 ± 3 vs. 112 ± 3 mg/dl blood glucose, P < 0.0025, n = 18). These findings prompted us to examine the expression of phosphoenolpyruvate carboxykinase (Pepck) and glucose-6-phosphatase (G6pc), which are the 2 rate-limiting enzymes in gluconeogenesis, and of glucose-6-phosphate transporter...
SLU7 strongly influences the gene expression profile of human liver cell lines and modulates SRSF3 splicing. (A and B) Top categories of genes undergoing changes in (A) splicing events and (B) expression in PLC/PRF/5 cells induced by SLU7 knockdown, classified in pathways according to Ingenuity Pathway Analysis. (C) SLU7 downregulation affected SRSF3 alternative splicing, promoting the generation of Iso2 targeted for nonsense-mediated decay (NMD). Bottom: Representative gel analyzing SRSF3 Iso1 and Iso2 PCR products in cells transfected with siSLU7 or siGL control. (D) qPCR analysis of SRSF3 Iso2/Iso1 ratio in PLC/PRF/5, HepG2, and Hep3B cells transfected with siGL or siSLU7. Representative Western blots of SLU7 and ACTIN protein levels are also shown. *P < 0.05 vs. siGL. (E and F) qPCR analysis of SRSF3 Iso2/Iso1 ratio (E) in HepaRG cells upon SLU7 knockdown and (F) in untransfected HepaRG cells (Control) or cells transfected with control vector (pEGFP) or SLU7 expression vector (pEGFP-SLU7). Bottom: Representative Western blot analysis of SLU7 and ACTIN protein levels. *P < 0.05 vs. siGL or pEGFP. (G) SRSF3 expression, analyzed by qPCR, in human liver tissues from control, cirrhotic, and HCC tissue samples. (H) Spearman correlation analysis of SLU7 and SRSF3 expression, analyzed by qPCR, in all groups of human liver tissue samples.
To gain mechanistic insights into the effects of SLU7 on gluconeogenesis, we directly examined its involvement in the regulation of PEPCK, which was identified as the second most downregulated gene in our microarray analysis (Supplemental Table 2). Basal levels of PEPCK gene expression were reduced upon SLU7 knockdown in human liver cells (Figure 4, D and E). This effect was markedly attenuated by the concomitant overexpression of HNF4α (Supplemental Figure 4C), which suggests that under resting conditions, SLU7-dependent PEPCK expression could be mediated to a great extent through HNF4α. We also found that cAMP-induced PEPCK protein and mRNA levels were strongly dependent on SLU7 expression in human liver cells and were also stimulated by forskolin (Figure 4, D and E, and Supplemental Figure 4D). This finding was extended to NR4A2 (also known as NURR1), another prototypical cAMP-regulated gene (41, 42), while ACTIN mRNA remained unaffected (Figure 4E). cAMP-induced transcription of PEPCK and NR4A2 genes is critically regulated by cAMP response elements (CREs) in their proximal promoters (42, 43). cAMP triggers phosphorylation of CRE-binding protein (CREB), facilitating its association with the coactivator CREB-binding protein (CBP). CBP is essential for RNA polymerase II (RNA-Pol II) recruitment to drive transcription of cAMP-responsive genes like PEPCK and NR4A2 (41, 42, 44, 45). Using ChIP assays, we demonstrated the association of SLU7 with PEPCK and NR4A2 proximal promoters (Figure 4F and Supplemental Figure 4E). Moreover, SLU7 was required for the association of CBP and RNA-Pol II with PEPCK and NR4A2 promoters, under both basal and cAMP-stimulated conditions (Figure 4G). Therefore, SLU7 seems essential for the assembly of the basic and cAMP-triggered transcriptional complex at the promoter of cAMP-responsive genes. A direct implication of SLU7 in this process is supported by our finding of communoprecipitation of phosphorylated CREB (P-CREB), CBP, and RNA-Pol II with SLU7 (Figure 4H). Given that glucagon stimulates the gluconeogenic program through the cAMP/CREB signaling pathway (45), these observations contribute to explain our in vivo findings.

Lipogenesis is also a fundamental metabolic liver function. We observed that Slu7 knockdown markedly reduced the basal expression of key lipogenic genes (Figure 3D). The lipogenic genetic program is tightly controlled by nutrients and hormones like insulin, being physiologically modulated in response to diet (46). Slu7 expression was also regulated in mouse liver upon fasting and refeeding. (G6pt), which is essential in the final steps of gluconeogenesis (36). Slu7-depleted mice showed impaired fasting-induced Pepck and G6pc upregulation and constitutive reduction in G6pt mRNA (Figure 4A). These observations were consistent with diminished de novo hepatic glucose production in a pyruvate tolerance test and a lower hyperglycemic response after glucagon challenge (Figure 4, B and C). Congruently, an enhanced hyperglycemic response was observed when a pyruvate tolerance test was performed in SLU7-overexpressing mice (infected with SLU7-expressing AAVs [AAV-SLU7]; Supplemental Figure 4, A and B). Together, these findings indicated that Slu7 is necessary for liver glucose production.

Figure 2
SLU7 modulates the expression of adult and fetal markers in human liver cells and is developmentally regulated in mice. (A) qPCR analysis of the expression of adult (ALB, HNF4α, and MAT1A) and fetal (H19 and CYP4F3A/B splice variant ratio) marker genes in HepaRG cells transfected with siGL or siSLU7. *P < 0.05, **P < 0.01 vs. siGL. (B) pEGFP-SLU7–mediated SLU7 overexpression in HepaRG cells promoted expression of the hepatocellular markers HNF4 and MAT1A. *P < 0.05 vs. pEGFP. Top: Representative Western blot analyses of SLU7 protein levels at the indicated time points after transfection. (C) qPCR analysis of Slu7, Mat1a, Hnf4α, and Wt1 expression in fetal and postnatal mouse liver. (D) Representative Western blot analyses of SLU7, MAT1A, and WT1 protein levels in fetal and postnatal mouse liver. ACTIN levels are shown as control. (E) Representative images of immunohistochemical detection of SLU7 in adult mouse liver. Original magnification, ×20; ×40 (insets).
**Figure 3**

SLU7 is required for the preservation of the genetic program characteristic of the differentiated, quiescent, and metabolically functional liver. (A) qPCR analysis of Slu7 expression in livers of mice infected with AAV-shSLU7 or control (AAV-Ren). ***P < 0.001. (B) Representative Western blot analysis of SLU7 protein in liver tissues. ACTIN levels are shown as control. Lanes were run on the same gel but were noncontiguous. (C) Effect of Slu7 knockdown in mouse liver on Srsf3 alternative splicing. Left: Representative gel analyzing Srsf3 Iso1 and Iso2 RT-PCR products. Right: qPCR analysis of Srsf3 Iso2/Iso1 ratio in livers. *P < 0.05 vs. AAV-Ren. (D) qPCR analysis of expression of selected liver-enriched genes along with the fetal and proliferative hepatocyte markers Afp, Wt1, and Atf3. *P < 0.05, **P < 0.01, ***P < 0.001 vs. AAV-Ren. (E) Liver-specific Slu7 knockdown induced a switch in the gene expression of metabolic enzymes, Hnf4α P1/P2 promoter usage, and Insr splicing isoforms toward a pattern characteristic of the fetal and transformed hepatocyte. *P < 0.05, **P < 0.01 vs. AAV-Ren. (F) Liver-specific Slu7 knockdown influenced serum levels of glucose, cholesterol, and triglycerides. *P < 0.05, **P < 0.01, ***P < 0.001 vs. AAV-Ren.
and tissue immunostaining revealed that predominantly pericentral expression became more apparent in the liver of fasted mice (Figure 5, A and B, and Supplemental Figure 5, A and B). Insulin may be a key factor in the nutritional control of \textit{Slu7} gene expression, as this hormone directly reduced \textit{Slu7} mRNA levels in primary mouse hepatocytes (Supplemental Figure 5C). Moreover, mice fed a high-fat diet (HFD), a condition associated with insulin resistance, had increased basal levels of \textit{Slu7} expression in the liver and showed impaired \textit{Slu7} downregulation upon feeding (Supplemental Figure 5, D and E). Importantly, \textit{Slu7} knockdown profoundly altered the expression of developmentally regulated and liver-specific genes (Figure 3, D and E). While the expression of the non-SREBP1 target gene \textit{Insig2a} decreased with feeding, as expected (47), in both mouse groups (Figure 5E).

Given the central role played by insulin in the dietary regulation of \textit{Srebp1} expression (47), we examined insulin sensitivity in \textit{AAV-shSLU7}–infected mice and found significant resistance to the action of the hormone (Figure 5F). The PI3K-AKT pathway is key in insulin-mediated \textit{Srebp1} expression (47). Consequently, liver AKT phosphorylation was clearly detected in control mice upon refeeding, but not in \textit{AAV-shSLU7}–infected mice (Figure 5G). Consistently with this was the impaired phosphorylation of the downstream AKT target GSK3\(\alpha\)/\(\beta\) (Figure 5G and ref. 48). Together, these data are indicative of abnormal hepatic insulin signaling upon reduced \textit{SLU7}. \textit{Slu7} depletion in mouse liver: \textit{SLU7} plays a central role in glucose production and is essential for cAMP-mediated gene expression. (A) qPCR analysis of \textit{Pepck}, \textit{G6pc}, and \textit{G6pt} expression in fed and fasted \textit{AAV-Ren}– and \textit{AAV-shSLU7}–infected mice. *\(P < 0.05\) vs. \textit{AAV-Ren}. (B and C) Hepatic glucose production after (B) pyruvate challenge and (C) glucagon challenge in \textit{AAV-Ren}– and \textit{AAV-shSLU7}–infected mice. *\(P < 0.05\) vs. \textit{AAV-shSLU7}. (D and E) PLC/PRF/5 and HepG2 cells were transfected with siGL or siSLU7 48 hours later, cells were treated with 10 \(\mu\)M forskolin (FK) for 4 hours, prior to analysis of \textit{PEPCK} and/or \textit{NR4A2} expression. (D) Representative Western blots of PEPCK, SLU7, and ACTIN control. (E) qPCR analysis of \textit{PEPCK}, \textit{NR4A2}, and \textACTIN control. (F) ChIP assays using control (IgG) or anti-SLU7 antibodies in siGL- or siSLU7-transfected PLC/PRF/5 cells analyzing \textit{PEPCK} and \textit{NR4A2} proximal promoters. (G) ChIP assays of \textit{PEPCK} and \textit{NR4A2} promoters in siGL- and siSLU7-transfected PLC/PRF/5 cells. 48 hours after transfection, cells were treated with 10 \(\mu\)M forskolin for 30 minutes, then subjected to ChIP with anti-CBP and anti–RNA-Pol II antibodies. (H) \textit{SLU7} coimmunoprecipitated with P-CREB (arrow), CBP, and RNA-Pol II in PLC/PRF/5 cells. Lysates were immunoprecipitated with anti \textit{SLU7} antibodies or preimmune IgG. Representative Western blots of 3 experiments are shown.

![Figure 4](https://doi.org/10.1172/JCI74382)
upregulated during liver development (Supplemental Figure 2 and ref. 49). We found that SLU7 knockdown had a strong influence on liver Insr expression (Figure 5H). Nevertheless, in addition to the regulation of the insulin signaling system, we cannot exclude a more direct effect of SLU7 on the regulation of SREBP1 expression. As we observed for gluconeogenic genes (Figure 4, A and D), basal expression of Srebp1 was significantly reduced upon Slu7 knockdown in vivo, as well as in human liver cells (Figure 5, C and D, and Supplemental Figure 5, F and G). Collectively, these findings indicate that SLU7 is required for the preservation of essential liver metabolic pathways and their hormonal regulation.

Reduced Slu7 expression promotes aerobic glycolysis and liver growth. One of our most intriguing observations was the switch in the hepatic expression of the glycolytic isoenzymes Lpk to Pkm2 and Gck to Hk2 upon Slu7 knockdown (Figure 3E). PKM2 and HK2 favor aerobic glycolysis, a hallmark of transformed cells (27). Upregulation of PKM2 and HK2 proteins was confirmed in livers from Slu7-depleted mice (Figure 6A). We demonstrated that Slu7 downregulation resulted in significantly increased hepatic glucose uptake and catabolism, as demonstrated by intrahepatic lactate concentrations (Figure 6, B–D). The forced expression of PKM2 and HK2 activates hepatocellular growth in normal liver (39). We found that the cell cycle–related genes Foxm1, Nor1, and early growth response 1 (Egr1) and the cyclins Ccnd1, Ccnb2, and Ccna2 were all upregulated in Slu7-depleted mouse livers and in SLU7-silenced HepG2 cells (Figure 7A and Supplemental Figure 6). Additionally, these mice showed increased expression of the HCC-promoting splicing protein heterogeneous nuclear ribonucleoprotein A1 (Hnrnpa1; Figure 7A and ref. 21), which was also identified in our microarray analysis (Supplemental Table 2). Overexpression

Figure 5

Slu7 depletion in mouse liver interferes with nutritional regulation of lipogenesis and blunts hepatic insulin responses. (A) Slu7 mRNA levels were analyzed by qPCR, and SLU7 protein levels by Western blot, in mice that were fed ad libitum, fasted for 14 hours, or fasted for 14 hours and subsequently refed for 4 hours. Lanes were run on the same gel but were noncontiguous. *P < 0.05 vs. refed. (B) Immunohistochemical detection of SLU7 in liver tissues of mice fasted for 14 hours. Original magnification, ×20. (C) qPCR analysis of Srebp1 expression in livers of AAV-Ren– and AAV-shSLU7–infected mice upon fasting and refeeding. (D) Western blot analysis of SREBP1 protein in livers upon fasting and refeeding. SLU7 and tubulin (TUBA) are also shown. (E) qPCR analysis of the mRNA levels of SREBP1 target genes and the regulatory genes Insig1 and Insig2a in livers upon fasting and refeeding. (F) Blood glucose concentrations during insulin tolerance test. **P < 0.01, ***P < 0.001 vs. AAV-Ren. (G) Intracellular signaling in livers upon fasting and refeeding. Representative Western blots show phosphorylated AKT and GSK3 as well as SLU7 and tubulin. Lanes were run on the same gel but were noncontiguous. (H) Expression of Insr mRNA determined by qPCR, and INSR protein (β chain) determined by Western blotting, in mouse livers. *P < 0.05 vs. AAV-Ren. A representative blot is shown.
of cell proliferation–related genes, including Myc (encoding c-Myc), and downregulation of Hnf4α were also validated at the protein level (Figure 7B). Interestingly, the fast-migrating anti-HNF4α–reactive bands we observed upon Slu7 knockdown may correspond to Hnf4α P2 promoter–derived transcripts (50), which we found to be favored upon Slu7 downregulation (Figure 3E). Quite remarkably, these genetic changes translated into hepatocellular proliferation and a 23% increase in liver mass (Figure 7, C and D). To further establish the influence of SLU7 on hepatocellular proliferation, we evaluated its expression during liver regeneration after two-thirds liver resection (Supplemental Figure 7, A and B). Most interestingly, we observed enhanced expression of EGR1 and CCND1 after PH in the face of Slu7 knockdown (mice infected with AAV-shSLU7) and reduced levels of CCND1 after PH in mice overexpressing SLU7 (infected with AAV-SLU7) (Supplemental Figure 7, C and D).

Discussion

We have identified a fundamental new role for SLU7 in the preservation of liver function and hepatocellular quiescence. SLU7 downregulation in human liver cells impaired the expression of multiple genes characteristic of the mature and differentiated hepatocyte, inducing that of immature and proliferative cells. These findings were corroborated in healthy adult mice with liver-specific Slu7 knockdown, which developed a strong metabolic and proliferative hepatic phenotype. SLU7 is known as a pre-mRNA splicing regulator (51). However, the physiological role of SLU7 in vivo has not been addressed, previous works being restricted to in vitro observations (24, 52). Our microarray analyses demonstrated SLU7 participation in the regulation of splicing and expression of genes implicated in RNA modification and liver metabolism. In particular, SLU7 knockdown modulated the splicing of SRSF3 not only in cultured cells, but also in normal mouse liver. SRSF3 is a RNA-binding protein and splice regulator, and its hepatocyte-specific deletion (SRSF3HKO mice) results in impaired hepatic maturation and disturbed lipid and glucose metabolism (18). We showed that SRSF3 expression was also reduced in human HCC, correlating significantly with that of SLU7 in healthy and diseased human liver tissues. This effect of SLU7 on SRSF3 expression may be of functional relevance, as the phenotype of the Slu7-deleted mice overlapped to a great extent with that of SRSF3HKO animals. Indeed, Slu7 downregulation also resulted in decreased expression of liver-specific genes, including those involved in glucose and lipid metabolism and liver-enriched transcription factors, and in the upregulation of oncogenic markers. These findings suggested that the phenotypic alterations found upon Slu7 knockdown could be mediated by the concomitant downregulation of Srf3. However, at variance with Slu7-depleted mice, SRSF3HKO animals did not show insulin resistance nor increased hepatocellular growth, and displayed extensive liver injury (18). This indicated that in addition to altering SRSF3 expression, SLU7 was also influencing other mechanisms essential for hepatocellular homeostasis. One key regulator of hepatocyte differentiation and function is HNF4α (1, 15). While SRSF3HKO mice showed no changes in HNF4α levels (18), we found the expression of this liver-enriched transcription factor to be consistently reduced upon Slu7 knockdown, both in cultured cells and in mouse livers. HNF4α gene expression can be driven from 2 promoters, P1 and P2. P2 activity gives rise to the isoforms characteristic of the adult differentiated hepatocyte, while P2-derived transcripts are found in fetal and HCC cells and code for HNF4α variants with less transactivation potential (40, 53, 54). Remarkably, we found that Slu7 was essential for preserving the P1 and P2 promoter–driven Hnf4α expression pattern of the adult liver. Hepatic HNF4α expression is necessary for lipogenesis, cholesterol synthesis, and gluconeogenesis (16, 55). The concomitant reduction in SRSF3 and HNF4α levels may explain, at least in part, the deranged expression of metabolic genes found in Slu7-depleted mice. Moreover, Hnf4α downregulation may also help to explain the robust liver proliferative response detected in these mice. Hepatocyte-specific deletion of Hnf4α results in hepatocellular dedifferentiation and proliferation and promotes chemically induced hepatocarcinogenesis. These responses were accompanied by upregulation of the promitogenic genes Egr1, Myc, and Ccn2 (17, 56–58), which were also detected in Slu7-depleted mice. Interestingly, Ccn2 can also interfere with Hnf4α transcriptional activity (57), while increased c-Myc levels modulate the switch in GLS1 and GLS2 expression (38) and, through the induction of hnRNP1, can promote the expression of HK2 and PKM2 favoring glycolytic metabolism (38, 59).
Figure 7

Slu7 knockdown triggers proliferation-related gene expression and promotes liver growth. (A) qPCR analysis of the expression of cell-cycle– and proliferation-related genes in the livers of AAV-Ren– and AAV-shSLU7–infected mice. *P < 0.05, **P < 0.01 vs. AAV-Ren. (B) Western blot analyses of selected proliferation-related proteins. SLU7 and tubulin are shown as controls. Asterisk indicates fast-migrating SLU7 isoforms. Lanes were run on the same gel but were noncontiguous. (C) Left: Immunohistochemical analysis of Ki67 in liver sections. Data were acquired from livers corresponding to 12 (AAV-Ren) and 10 (AAV-shSLU7) mice; 10^3 cells were counted per liver. Original magnification, ×10; ×20 (inset). Right: Quantification of Ki67-positive hepatocytes. ***P < 0.001. (D) Liver/body weight ratio.

The liver has an extraordinary regenerative capacity, with hepatocytes exhibiting almost unlimited proliferative potential (3). Characterization of the mechanisms governing the switch from quiescence to proliferation is a matter of great importance, not only for regenerative medicine, but also for its clear implications in hepatocarcinogenesis. Our current findings that HNF4α levels were influenced by Slu7 gene expression, and that Slu7 knockdown triggered proliferation in an otherwise normal liver and facilitated the expression of cell-cycle–driving genes after PH, attest to the fundamental role of SLU7 in liver biology. The mechanisms behind HNF4α downregulation and P1/P2 promoter usage switch upon SLU7 knockdown are not currently known, although this effect could be partly mediated by the transcription factor WT1. This is supported by our previous observations demonstrating a potent negative effect of WT1 on HNF4α gene transcription in hepatocytes (7) and our current findings showing WT1 upregulation upon Slu7 knockdown in mouse liver. Moreover, WT1 overexpression could also underlie the insulin resistance found in Slu7-deficient mice. Part passa with SLU7, INSR expression was progressively induced during hepatocellular maturation, concomitantly with the decay of WT1 mRNA levels. This correlation and the previous identification of WT1 as a potent repressor of insulin gene promoter (60) lent support to this hypothesis.

One remarkable observation was the profound dysregulation of gluconeogenesis in Slu7-depleted mice. We not only found reduced expression of gluconeogenic genes in fed conditions, which could be attributed in part to low HNF4α (61) and increased ATF3 expression (62), we also observed a marked impairment in the activation of the gluconeogenic transcriptional program by nutritional and hormonal stimuli. The cAMP signaling pathway plays a central role in this response, controlling the activity of CREB and the coactivators CBP and CRTCs (45, 61). The cAMP-mediated upregulation of PEPCK and NR4A2 gluconeogenic genes (41, 42, 63) was almost erased upon SLU7 knockdown in cultured cells. Importantly, SLU7 was essential for the recruitment of CBP and RNA-Pol II to cAMP-responsive gene promoters, in which SLU7 was also detected, and was found to interact with P-CREB, CBP, and RNA-Pol II. CAMP signaling activates gene transcription by promoting P-CREB and its association with coactivators like CBP, which in turn interact with components of the RNA-Pol II complex facilitating transcription (45). However, it has become apparent that the interaction of P-CREB with its coactivators would be too weak to drive gene activation per se, and that additional partners are required for stable and productive recruitment (41). In this context, we could be looking at a novel cellular function of SLU7 not related in principle to its role as splicing factor, but as a necessary component of the cAMP signaling pathway at the level of transcriptional complex assembly and target gene regulation. This functional diversity is not without precedent among splicing factors; for instance, hnRNPs have been identified as transcription regulators and coactivators for a number of genes (64), while NONO (also known as p54 nrb) has also been reported to be a necessary component of cAMP-CREB–mediated gene transcription (42). SLU7 may thus be regarded as a new coactivator in the cAMP-CREB pathway regulating gluconeogenic expression. Importantly, the interaction between SLU7 and cAMP signaling might have implications exceeding the regulation of gluconeogenesis, as the cAMP pathway has also been involved in the maturation of the hepatic lineage and the expression of liver-enriched genes (65). Our current observations of the strong effects of SLU7 on cAMP-CREB–regulated gene promoters, as well as its critical influence on HNF4α P1 and P2 promoter activity, suggest that transcriptional regulation would be a major biological role of this splicing factor. Future studies will need to delineate the mechanisms contributing to the recruitment of SLU7 to specific gene promoters and to identify the critical SLU7-interacting factors participating in the formation of transcriptional regulatory complexes.
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According to the significant roles played by SLU7 in the liver, regulation of its expression and availability must be tightly controlled processes. Previous studies demonstrated the nucleocytoplasmic shuttling of SLU7 protein in response to cellular stress (1, 52), and we also described how SLU7 transcription is repressed in hepatocytes by amphiregulin, a ligand of the epidermal growth factor receptor (2, 24). Here, we showed that hepatic SLU7 expression could be modulated by nutritional signals, being induced upon fasting and downregulated after feeding. Our in vitro findings demonstrated that CAMP stimulated expression of SLU7, whereas insulin exerted a negative effect on its transcription. These findings may suggest that, in vivo, the nutritional regulation of SLU7 expression could be mediated by glucagon and insulin signaling. The impaired postprandial downregulation of Slu7 transcription in mice fed a HFD would be consistent with the loss of insulin sensitivity that developed in this model. Nevertheless, further studies are warranted to fully understand the mechanisms underlying regulation of SLU7 gene expression in normal and diseased liver.

In summary, our current observations identified SLU7 as an essential mediator in the preservation of hepatocellular identity. Our finding that downregulation of SLU7 expression triggered a proliferative phenotype in healthy liver further emphasizes its pivotal role in the homeostasis of the organ. The loss of SLU7 expression occurring in the cirrhotic and preneoplastic liver (24) may be of high pathogenic relevance, contributing to the progressive deterioration of liver function up to HCC development. Further characterization of the mechanisms controlling SLU7 gene expression and its cellular targets may provide new avenues for therapeutic intervention in chronic liver disease.

Methods

Further information can be found in Supplemental Methods and Supplemental Tables 3 and 4.

RNAi. Human SLU7-specific siRNA (siSLU7) and the control siRNA (siCGL) were from Sigma-Aldrich (24). Transfections were performed using Lipofectamine RNAiMAX reagent (Invitrogen) following the manufacturer’s instructions. Silencing was confirmed by qPCR and Western blot.

9 siRNAs targeting mouse Slu7 were tested in vitro for their capacity to inhibit SLU7 expression. The 2 best performing sequences (siSLU7-767, TGGGCGAAGTTGACTCTTAA; siSLU7-723, GAGGATGAAGACGAGACAAA) and a siRNA targeting Renilla (CAGGAATTATAGCTCTATCT), used as negative control, were cloned as shRNAs in the context of miR-30 (66). The 3 miR-shRNAs cassettes were subcloned in adenoviral vectors (AAV8) flanked by AAV2 WT inverted terminal repeats, and under the regulation of a chimeric liver-specific promoter composed of the human c1-antitrypsin promoter (AAV) with regulatory sequences from the albumin enhancer (Eal) (67). The 2 AAV-shSLU7 vectors performed similarly in the in vivo experiments; the results presented correspond to siSLU7-767.

Production and purification of AAVs. To generate the AAV-SLU7 virus, mouse SLU7 cDNA was cloned in the AAV8 vector as described above. All rAAV viruses were produced by polyethylenimine-mediated (PEI-mediated) cotransfection of AAV8-miR-shRNAs or AAV8-SLU7 vectors with pDP8.ape plasmid (PlasmidFactory GmbH & Co.) in HEK-293 cells (68). Cells were harvested 72 hours after transfection, and virus was released from the cells by 3 rounds of freeze-thawing. Crude lysate from all batches was then treated with RNase and DNase for 1 hour at 37°C and then kept at −80°C until purification, which was performed by iodixanol gradients as described previously (69). The purified batches were concentrated and diafiltered by cross-flow filtration with a molecular mass cutoff of 400 kDa. Batches were then concentrated further by passage through Centricon tubes (YM-100; Millipore) to a final concentration of 1 × 1012 vg/ml, as determined by qPCR. Finally, viral batches were filtered (pore size, 0.22 mm) and stored at −80°C.

Animal experiments. C57BL/6 male mice (Harlan) were used. 16-week-old mice were injected i.v. with rAAVs (1 × 1011 pfu). Experiments were performed 2–3 weeks after AAV injection with at least 5 animals per group and were repeated at least twice.

Mice were maintained under fed conditions with regular chow or fasted for 4 hours or overnight. In some experiments, after overnight fasting, mice were refed with regular chow diet for 4 hours. At sacrifice, blood was collected, and livers were weighed, snap-frozen in liquid nitrogen, and paraffin embedded. In some experiments, mice were fed a HFD (D12451 OpenSource Diets) for 5 months before fasting overnight and refeeding for 4 hours.

For the insulin tolerance test, after a 4-hour fast, mice received an i.p. injection of 1 U/kg insulin (Novo Nordisk Pharma) in saline. For the pyruvate tolerance test, after an overnight fast, animals received an i.p. injection of 2 g/kg sodium pyruvate (Sigma-Aldrich) in saline. For glucagon challenge test, mice were fasted for 4 hours and i.p. injected with 100 μg/kg glucagon (Sigma-Aldrich). Blood glucose concentration was measured from the tail vein at different time points using an ACCU-CHEK Aviva glucometer (Roche).

For hepatic lactate determination, frozen liver tissues were ground to powder in liquid nitrogen using mortar and pestle. Liver (approximately 30 mg) was homogenized (1:3 w/v) with 1% perchloric acid on ice. The homogenate was centrifuged at 5,000 g for 10 minutes. The supernatant was collected, neutralized with KOH (1N), and centrifuged at 10,000 g for 10 minutes at 4°C. Pellets were discarded, and supernatants were used for the determination of total lactate concentration on Roche/Hitachi Cobas system.

For hepatic glycogen measurement, about 100 mg frozen liver tissue was ground to a powder, and 300 μl of 30% KOH was added. The homogenized tissue was heated to 100°C for 2 hours. To precipitate glycogen, 2 volumes of 95% ethanol were added, and samples were centrifuged at 1,500 × g for 10 minutes. Pellets were resuspended in a minimal amount of distilled water and acidified to pH 3 with HCl (5N). Glycogen was reprecipitated with ethanol, and pellets were dried and dissolved in hydrolysis buffer for analysis using Glycogen Assay Kit (BioVision).

18F-FDG PET. Liver glucose metabolism was assessed in vivo by PET with the glucose analog radiotracer 18 fluorodeoxyglucose (18F-FDG). To improve liver uptake of 18F-FDG, mice were fasted overnight and warmed with a heating pad at 30°C 30 minutes before the study and during the imaging period, and kept under continuous anesthesia with isoflurane (2% in 100% O2 gas) (70). PET scans were obtained in a small-animal dedicated imaging tomograph (Mosaic; Philips), with an axial and transaxial field of view of 11.9 and 12.8 cm, respectively, and a full width at half-maximum resolution of 2.1 mm. On the day of study, mice were placed prone on the PET scanner. Then, 18F-FDG (7.5 ± 0.5 MBq) was injected i.v. through the tail vein simultaneously to the beginning of a list mode study of 30 minutes. 15-frame dynamic sinograms were created (2 × 15, 7 × 30 s, 1 × 60 s, 1 × 120 s, 1 × 180 s, 2 × 300 s, and 1 × 600 s), and dynamic images were then reconstructed in a 128 × 128 matrix with a 1 × 1 × 1 mm3 voxel size using the 3D Ramla algorithm with 2 iterations, a relaxation parameter of 0.024, and applying of dead time, decay, random, and scattering corrections. For assessment of liver 18F-FDG uptake, all studies were acquired and analyzed using PMOD software (PMOD Technologies Ltd.). Images were expressed in standardized uptake value (SUV) units, calculated as [tissue activity concentration (Bq/cm3)/injected dose (Bq)] × body weight (g). Because of the limited
anatomical information of PET images, the volume of interest (VOI) of liver was drawn over an image of the first 2–4 minutes created from the dynamic study that reflects the vascular distribution of 18F-FDG. VOI was then transferred to an image of the last 10 minutes of the study (20–30 minutes after injection). From each VOI, mean SUV and maximum SUV were calculated.

PH. Two-thirds PH and sham operations were performed as described previously (71) in control mice and mice injected 2 weeks prior with AAV-shSLU7, AAV-SLU7, or control adenovirus (AAV-Ren). Animals were killed 3 or 34 hours after surgery, and liver samples were snap frozen.

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40. Torres-Padilla ME, Fougeré-Deschatrette C, Weiss MC. Expression of HNF4α isoforms in mouse liver development is regulated by sequential promoter usage and constitutive 3’ end splicing. Mech Dev.
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