**Introduction**

Humans are colonized with trillions of microbes, predominantly bacteria. Work by numerous investigators combined with the Human Microbiome Project (HMP), initiated in 2007, has provided remarkable data that have enhanced our understanding of the complexity, variability, and biology of human associations with our microbiota at diverse body sites (1). Microbiota science has been fostered and complemented by microbial whole genome sequencing, technical sequencing advances, and rapidly evolving bioinformatics. The first phase of the HMP, completed in 2012, focused on establishing the “normal” microbiota and now has evolved to address the key scientific challenge of translating findings from this phase to understand how the microbiota contributes to disease pathophysiology (2). As this translation occurs, there is optimism that new diagnostic and therapeutic approaches will be developed. This research frontier has already yielded exciting observations in several fields such as atherosclerosis, obesity, and colon cancer (3–5). Herein, we will describe the story of how *Bacteroides fragilis* subverts mucosal biology: from symbiont to colon carcinogenesis.

The human body comprises fewer host cells than bacterial cells, most of which are obligate anaerobes residing in the gut. The symbiont *Bacteroides fragilis* constitutes a relatively small proportion (up to 1%–2%) of cultured fecal bacteria, but colonizes most humans. There are 2 classes of *B. fragilis* distinguished by their ability to secrete a zinc-dependent metalloprotease toxin, *B. fragilis* toxin (BFT). Strains that do not secrete BFT are nontoxicogenic *B. fragilis* (NTBF), and those that do are called enterotoxigenic *B. fragilis* (ETBF). ETBF can induce clinical pathology, including inflammatory diarrhea, although asymptomatic colonization may be common. Intestinal inflammation is mediated by BFT, as yet the only known virulence factor of ETBF. Recent experimental evidence demonstrating that ETBF-driven colitis promotes colon tumorigenesis has generated interest in the potential contribution of ETBF to human colon carcinogenesis. Critical questions about the epidemiology of chronic, subclinical human colonization with ETBF and its impact on the biology of the colon need to be addressed.

Such observations may further parlay into new microbe-based approaches to prevention and detection of chronic diseases such as sporadic colon cancer.

**The evolution of our clinical perspective on *B. fragilis***

All *B. fragilis* are obligate anaerobes that inhabit and flourish along the entire length of the colon, where they are minority members of the normal colonic microbiota with a propensity for mucosal adherence. Interest in *B. fragilis* blossomed in the 1970s with the recognition that these organisms were the leading anaerobes in bloodstream infection and critical contributors to intra-abdominal abscess formation (7, 8). Our subsequent understanding that *B. fragilis* is relatively aerotolerant (i.e., able to grow in the presence of nanomolar oxygen concentrations) partially explains its success in mucosal colonization (where oxygen tension is higher), its survival following colon perforation when exposed to the peritoneal cavity before abscess formation, and its likelihood of inducing bacteremia (9). The capsule of *B. fragilis* emerged as a pivotal virulence factor that is key to the organism’s success in induction of abscess formation (10). Molecular characterization of the first 2 polysaccharides purified from the surface of *B. fragilis*—polysaccharide A (PSA) and polysaccharide B (PSB)—revealed that the charge and structure of PSA were, in fact, sufficient to induce abscess formation in rat or murine models (11). Namely, PSA contains a balanced positively charged amino group and negatively charged carboxyl group; modification of either charged group reduced by at least 2 orders of magnitude the biologic potency of experimental abscess induction by PSA administered intraperitoneally. PSB also contains oppositely charged groups but is an order of magnitude less potent in abscess induction. The concept that oppositely charged groups on bacterial polysaccharides are critical to abscess induction was confirmed by chemical modification of the Vi polysaccharide of *Salmonella typhi* to display both positive and negative charges.
charged groups (native Vi contains 1 negative charge). Unmodified Vi polysaccharide had no abscess-inducing activity whereas modified Vi polysaccharide was abscess-inducing (11). Conversely, subcutaneous treatment of rats with purified PSA protected against abscess formation (12). Thus, PSA alone can cause abscess formation and, conversely, can stimulate protective immunity against abscess formation. Investigations of the capsule of B. fragilis and the host immune response to this organism led to further transformative observations. First, B. fragilis possess the most diverse surface polysaccharide gene repertoire of any known bacterium with the ability to synthesize up to 8 distinct capsular polysaccharides (A–H) that likely populate the surface of B. fragilis one at a time (13). The benefit to the organism of its ability to “change its coat” remains unknown, given a single longitudinal human study suggesting that humans tend to be stably colonized with B. fragilis populations expressing a range of polysaccharide types (14). Second, immunologic studies support B. fragilis as a symbiont with the remarkable capacity to modulate homeostatic mucosal immunity as well as contribute to systemic immune development, effects mediated by PSA (15–17). Most intriguingly, colonization with B. fragilis displaying PSA (strain NCTC 9343 [ATCC 25285], a human strain from an appendix abscess) inhibits subsequent experimental chemically induced (e.g., by dextran sodium sulfate or 2,4,6-trinitrobenzenesulfonic acid) colitis, likely mediated, in part, by enhanced bacterial-mucosal contact (17, 18). Hence, this strain is now proposed as a potential beneficial microbe therapeutic to moderate inflammation in the colon (19); however, this concept is challenged by the fact that most individuals are already colonized by B. fragilis combined with recent data suggesting Bacteroides species-specific colonization resistance (20).

Nearly in parallel with this early B. fragilis interest, in the 1980s, Myers and colleagues determined that select strains of B. fragilis accounted for a portion of diarrheal disease in lambs (21) as well as other livestock including piglets, calves, and foals (6, 22–25). These initial studies used lamb and rabbit ligated ileal loops as well as experimental animal inoculations to provide evidence that these strains of B. fragilis induced intestinal secretion in association with histologic changes that all humans experience; whether human colonization or stems from one of the frequent, but undiagnosed, diarrheal illnesses that all humans experience; whether human colonization begins most often in childhood and then persists; or whether some hosts can eliminate ETBF colonization through as-yet unidentified immune mechanisms. The remainder of this Review will describe how a novel cause of diarrheal disease in animals and humans became a candidate instigator of human colon cancer.

BFT: a molecular link to colonic inflammation and oncogenesis

The chromosomal bft gene codes for a pre-protein metalloprotease holotoxin that is processed by ETBF and secreted as the
Figure 1. Molecular types of Bacteroides fragilis. B. fragilis commonly colonize humans and are considered as 2 molecular types based on expression of BFT protein. NTBF is a human symbiont not associated with diarrheal disease, but able to cause invasive human disease. In contrast, ETBF is associated with diarrheal disease in all age groups and expresses 1 of 3 subtypes of BFT (BFT-1–BFT-3). BFT is a pre-protoxin that is processed by ETBF to the secreted, mature 20-kDa protein toxin. Based on the HEXXH motif and other studies (69), the toxin is classified as a zinc-dependent metalloprotease toxin. BFT is related to other metalloprotease toxins important in human medicine, such as anthrax toxin, tetanus toxin, and botulinum toxin. Adapted with permission from Clinical Microbiology Reviews (6).

The range of CEC signal transduction activated by BFT is remarkably broad and incompletely understood. For example, BFT also activates NF-κB and MAPK signaling in CECs, resulting in their release of proinflammatory chemokines (such as IL-8 and TNF-α, among others) across the basolateral membrane of CEC monolayers (50–52). Colon epithelial cell release of chemokines and cytokines into the submucosa is predicted to foster recruitment of neutrophils and other immune cells to the colonic mucosa (53). BFT also induces CEC expression of COX-2 (but not COX-1), increasing mucosal prostaglandin E2 (54). These mechanisms likely contribute to ETBF-induced inflammatory diarrhea in animals and humans (28, 39).

Thus, cleavage of E-cadherin, activation of NF-κB (with antiapoptotic effects), increased polyamine metabolism, and induction of DNA damage within the colonic epithelium (Figure 2) are key BFT oncogenic mechanisms identified to date in studies in vitro. E-cadherin cleavage yields multiple potentially procarcinogenic triggers, including Wnt signaling, CEC proliferation, and epithelial barrier disruption that promote mucosal inflammation and colon tumor formation as is well demonstrated in murine models of colon carcinogenesis (38, 55). Additionally, colonization with ETBF in patients with colon cancer might contribute to cancer metastatic potential, as epithelial tumors with reduced E-cadherin exhibit increased metastases (56). Pro-oncogenic activities of NF-κB and SMO activation include induction of mucosal inflammation, enhanced epithelial cell survival or proliferation, DNA damage, and/or promotion of angiogenesis (reviewed in ref. 57). Mechanistic details of BFT-induced DNA damage, which is clearly important to oncogenic transformation, are needed.

ETBF, an etiologic candidate of bacterially induced colon cancer in humans

The clinical observation that ETBF causes human inflammatory diarrhea combined with the in vitro studies of BFT mechanisms of action led to the hypothesis that ETBF were carcinogenic bacteria. This hypothesis was tested in mice chronically colonized...
with ETBF. Notably, germ-free mice developed lethal colitis in 24 hours when colonized with ETBF (bft2) but not NTBF. In contrast, a single ETBF inoculation of conventional C57BL/6 mice resulted in acute, occasionally bloody, inflammatory diarrhea that gradually subsided over a week in nearly all mice (37). Despite resolution of symptoms, ETBF colonization persists in C57BL/6 mice, lasting up to 1 year, and is resistant to treatment with antiinflammatory antibiotics to which the bacterium was sensitive, likely because of small-intestinal antibiotic absorption and insufficient antibiotic delivery to the colon (C. Destefano-Shields and C.L. Sears, unpublished observation). Chronic ETBF colonization induces persistent, low-level, IL-17A–dominant colon inflammation with modest colon epithelial cell hyperplasia, foci of STAT3 activation, ROS production, and DNA damage (58). All BFT isotypes induce acute IL-17A–dominant colitis (S. Wu and C.L. Sears, unpublished observation); studies of ETBF (bft2) indicated that colonization with IL-17A–dominant colitis persists for up to 1 year in C57BL/6 mice (58). ETBF strains with an in-frame chromosomal deletion of bft do not induce colitis, and, conversely, transfection of NTBF with a plasmid bearing bft induces colitis similar to wild-type ETBF, demonstrating the central contribution of BFT (i.e., necessary and sufficient) to ETBF disease (37).

Multiple intestinal neoplasia (Min) mice are Apc heterozygous, most often expressing a truncated APC protein (59), which is known for its crucial role in colon cancer suppression. Loss of the second allele of Apc renders the mice susceptible to development of intestinal adenomas, predominantly in the small bowel. Despite the fact that the adenoma propensity of Min mice does not align with the characteristics of human disease, in which CECs are most likely to transform, the Min mouse is commonly used to understand potential mechanisms relevant to colon carcinogenesis. Thus, the Min mouse model was chosen to test the carcinogenic potential of ETBF. Notably, in Min mice, ETBF-induced inflammatory colitis progresses to gross colon tumorigenesis within 4 weeks (60). ETBF-colonized Min mice die within approximately 3 months because of colon adenoma burden, whereas parental Min mice survive at least 5 months. Time course studies revealed that ETBF promoted tumor initiation rather than tumor growth with excess histologic microadenomas identified within 1–2 weeks after ETBF colonization of Min mice. In contrast, microadenomas are rare in sham or NTBF-colonized mice at this early time point. One striking feature of this model is that ETBF-induced carcinogenesis is unevenly distributed along the colon axis with marked excess tumorigenesis in the distal colon, similar to the predominant location of human colon cancer (Figure 3A). This asymmetric disease distribution occurs despite relatively uniform ETBF colonization throughout the murine colon. Mechanistically, ETBF induces widespread, immediate (within 2 days) then prolonged, sporadic activation of STAT3 in CECs and a subset of associated infiltrating immune cells (58, 60). In the microenvironment of ETBF colon tumors, STAT3 activation is accentuated, compared with parental Min mouse colon tumors, where more modest STAT3 activation is detected (60). Predictably, since activation of the STAT3 transcription factor is integral to adaptive CD4+ Th17 cell differentiation, IL-17–secreting Th17 cells dominate the early ETBF-associated mucosal inflammatory immune response (60). IL-17 blockade as well as depletion of CD4+ cells inhibits ETBF colon tumorigenesis, confirming that ETBF triggers IL-17–dependent carcinogenesis (60). IL-17 is a potent chemoattractant for neutrophils, and neutrophils are prominent in ETBF colitis, likely releasing mutagenic ROS (61). ETBF also induces SMO expression in vivo, and treatment of Min mice with an SMO inhibitor reduced ETBF-induced colonic inflammation, epithelial cell proliferation, and colon tumorigenesis (48).

Thus, these ETBF murine models confirmed the in vitro activities of BFT to induce NF-κB, SMO, and Wnt signaling, ROS production, and DNA damage. Most importantly, the oncogenic potential of ETBF was confirmed, and, for the first time, endogenous adaptive immune responses, specifically Th17 adaptive immunity, were identified as carcinogenic. Since these observations, Th17 immunity has been demonstrated to contribute to carcinogenesis in numerous models and in human disease. In fact, activation of the STAT3/IL-17 pathway correlates with a worse prognosis in human colon cancer (62). The ETBF Min model of colon carcinogenesis offers the opportunity, for example, to identify why the distal colon is more sensitive to bacterially mediated oncogenesis; to examine somatic and
epigenetic interactions in bacterial oncogenesis; and to establish mechanisms of bacterially induced DNA damage or gene mutations contributing to colon cancer initiation, all in a time frame vastly accelerated compared with that of human disease (Figure 3B).

Concluding remarks: back to the clinic

Although the link between pathogenic inflammation and cancer has become remarkably clearer in the past few years, colon cancer has not yet been epidemiologically linked to a single microbe. This may suggest that rather than a single microbe, a community of microbes and their associated genome (microbiome) may promote colon carcinogenesis (5, 63). The distinction between colon tumorigenesis due to the carcinogenic activity of intestinal microbiota (causal implication) and establishment of an altered microbiota resulting from the tumor environment (a consequence) will remain a challenge requiring complex longitudinal human studies. Murine experiments have established the carcinogenic potential of ETBF along with the critical contributions of BFT and the Th17/IL-17 axis in ETBF carcinogenesis. Human data support the inflammatory potential of ETBF and suggest that exposure often occurs early in life, may persist, and is likely common even in countries with higher socioeconomic status. These observations are consistent with the long lead time (up to 40 years) observed in human colon cancer. Preliminary but insufficient human data support a link between ETBF, human colon cancer, and inflammatory bowel disease, known to predispose to colon cancer (29, 30, 64). One clinical study in Turkey reported significantly higher ETBF detection in stools of hospitalized colon cancer patients (21/56, 38%) than outpatient controls (5/40, 12%; P < 0.009) (64). Critical questions include whether the highly related BFT isotypes exhibit similar carcinogenic potential and whether long-standing, asymptomatic ETBF colonization in humans results in focal colon inflammation with activation of oncogenic mediators as seen in mice. Similar to the contribution of other zinc-dependent metalloprotease toxins to human disease (e.g., tetanus toxin, anthrax toxin), experimental studies support the potency of BFT with in vitro biologic activity detectable at femtomolar concentrations (65). Since B. fragilis prefers the mucosal environment (66–68), it is possible that BFT delivery to CECs by low levels of mucosal ETBF colonization, not necessarily readily detected in stool, is biologically relevant in the colon. We do not know the relationship of human colon ETBF mucosal colonization to ETBF detection in the stool, nor are we certain how to accurately detect persistent ETBF colonization in humans. Interactions with putative bacteria carcinogenic to the colon (e.g., *Fusobacterium*, *E. coli* possessing the *pks* island) are unknown, but current knowledge supports the idea that, collectively, these bacteria may trigger additive or even synergistic carcinogenic mechanisms in the colon mucosa (5). For example, both ETBF and *E. coli* possessing the *pks* island induce DNA damage; the activated complex of the FadA adhesin of *Fusobacterium* as well as BFT secreted by ETBF triggers CEC Wnt signaling that enhances epithelial cell proliferation; and *Fusobacterium nucleatum* recruits colon mucosal myeloid populations, while ETBF recruits polymorphonuclear leukocytes and IL-17–secreting inflammatory cells (reviewed in ref. 5). Thus, co-colonization of the susceptible host with more than 1 of these bacteria yields the potential for critical interactions in DNA mutations, cell signaling, and procarcinogenic inflammation known to be highly relevant to the promotion of human colon cancer. There is much to do, but we have every expectation that insights from ETBF and bacterial pathogenesis will inform our understanding of the initiation and progression of human colon cancer and, more importantly, yield tools to address the global threat of human colon cancer.

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