Emerging therapies for food allergy

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Food allergy is a common condition for which there are currently no approved treatments except avoidance of the allergenic food and treatment of accidental reactions. There are several potential treatments that are under active investigation in animal and human studies, but it is not yet clear what the best approach may be. Here, we review approaches that are currently in clinical trials, including oral, sublingual, and epicutaneous immunotherapy, immunotherapy combined with anti-IgE, and Chinese herbal medicine as well as approaches that are in preclinical or early clinical investigation, including modified protein immunotherapy, adjuvants, DNA vaccines, and helminth administration. We discuss the importance of fully exploring the risks and benefits of any treatment before it is taken to general clinical practice and the need for clarity about the goals of treatment.

Pathophysiology of food allergy

IgE-mediated food allergy is characterized by Th2-dominant immunologic responses, with allergen-specific IgE present in circulating forms and bound to mast cells and basophils. Allergen-specific, Th2-deviated CD4⁺ T cells predominantly produce cytokines, such as IL-4, IL-5, IL-9, and IL-13, which promote IgE production, eosinophil proliferation, and trafficking of inflammatory cells to tissues. The predominant mechanism by which food allergy develops remains controversial, with some studies suggesting that primary sensitization through skin contact may be even more important than exposure via the gut (9). Active gastrointestinal tolerogenic mechanisms appear to be important in preventing food allergy in general, as children with genetic defects in generating regulatory T cells frequently have severe allergic disease (10, 11). APCs in the gut, particularly DCs, clearly direct these T cell responses and are themselves responsive to the context in which they receive antigen. Abnormal function of both DCs and T cells has been linked to food allergy (12–20). Contextual clues that influence DC responses include costimulatory signals through a variety of receptors, including the TLRs. These signals come from multiple sources, including those associated with tissue damage, commensal bacteria, and the allergen itself (20, 21). Responses to these signals may vary considerably due to genetic predisposition (22).

Although basic science has been helpful in understanding the mechanisms by which potential treatments for food allergy might work, to date, the most promising therapies have not come as a result of these discoveries but instead from clinical observation and the modification of therapies previously developed for other allergic diseases. For example, immunotherapy, discussed at length below, was first described as a treatment for IgE-mediated allergic disease in 1911 (23), more than 50 years before the discovery of IgE (24). Some therapies, such as the recombinant peanut vaccine described below, emerged from basic science and appeared effective in animal models, but failed when tested in humans. Still other approaches, such as DNA vaccines, which appear promising in animal models, may be difficult or impossible to safely translate to humans. In the following sections, we will first review approaches that are under active clinical investigation, after which we will review potential approaches that are in preclinical or early clinical investigation.

Current clinical investigations

Current clinical investigations are summarized in Table 1. For venom and Aero allergies, immunotherapy using intact, and often
rather crude, allergens remains the only disease-modifying therapy available. Although sublingual delivery is emerging as a treatment option for inhalant allergens, especially in Europe, most immunotherapy has traditionally been provided by s.c. injection of gradually increasing allergen doses over months, followed by several years of maintenance dosing. Limited study of s.c. injection for food allergy, however, resulted in unacceptably high rates of systemic side effects (25) such that subsequent approaches under investigation have attempted to improve this risk/benefit ratio through a variety of methods, including different routes of delivery, modification of the allergens, cotreatment with medications to reduce adverse reactions, and the use of adjuvants to achieve maximum benefit at the lowest possible allergen dose.

**Oral and sublingual immunotherapy.** Published case reports of sublingual immunotherapy or oral immunotherapy for the treatment of food allergy date back to at least 1908 through the 1940s (26–29), but the first randomized, placebo-controlled study of either method was not published until 2005 (30). In recent years, there has been a proliferation of small and medium-sized studies of these methods (28, 30–74), but small sample sizes and variable study designs have made interpretation of the evidence difficult. A Cochrane review of milk oral immunotherapy (75) up to October 2012 found five trials that met their quality criteria, with a total of 196 patients. Of these trials, only three were blinded with a placebo arm, and each study used a different protocol. In total, 62% of the treated group was able to consume a full serving of milk at the end of the treatment, compared with 8% of the control subjects. An additional 25% of treated subjects could consume a partial serving of milk. The rate of adverse events was difficult to summarize because of the variability between studies, but overall, 9% of treated subjects required epinephrine.

These findings are similar to other controlled studies of oral immunotherapy. A multi-center, randomized, placebo-controlled trial of egg oral immunotherapy conducted by the Consortium for Food Allergy Research (CoFAR) in 55 children found that 55% were able to consume a 5-gram serving of egg after 10 months of therapy compared with 0% of placebo-treated patients. After an additional 12 months of therapy, 75% of those on active treatment were able to consume a full 10-gram serving (53). For peanut, a randomized, placebo-controlled trial of oral immunotherapy in 28 children found that 84% were able to tolerate a 5-gram challenge at one year compared with none of the placebo subjects (44). Most of the studies have been done in school-aged children, but a randomized trial of 60 toddlers found that milk oral immunotherapy was also effective in young children, with 90% of treated subjects tolerating the full challenge compared with 23% of control subjects (72). Generally, these early studies with motivated volunteers have had a 10%–20% dropout rate for adverse events, typically gastrointestinal side effects or acute reactions. Eosinophilic esophagitis has been reported in some studies, and it is not clear how frequently undiagnosed disease may complicate immunotherapy. Oral symptoms are very common with dosing, and moderate side effects, while generally occurring with less than 2% of doses (53, 76), affect a high percentage of participants because doses are administered daily. For example, in the study of young children, 47% of subjects developed moderate reactions during treatment (72).

Fewer studies have been done with sublingual immunotherapy, but thus far it appears that efficacy is much less than with oral immunotherapy. A CoFAR placebo-controlled study of peanut sublingual immunotherapy in 40 adolescents and adults found that 70% of treated subjects increased their food challenge thresh-

### Table 1

Selected recent clinical studies

<table>
<thead>
<tr>
<th>Selected recent clinical studies</th>
<th>Drug name (if any)</th>
<th>Foods studied/ description</th>
<th>Types of studies done</th>
<th>Efficacy</th>
<th>Safety</th>
<th>Selected references</th>
</tr>
</thead>
<tbody>
<tr>
<td>Allergen-specific immunotherapies</td>
<td>Oral immunotherapy</td>
<td>N/A Milk, egg, peanut, others</td>
<td>Small to medium phase I/II</td>
<td>&gt;60% desensitized in most studies</td>
<td>10%–20% rate of withdrawal for AEs</td>
<td>28, 30–74</td>
</tr>
<tr>
<td></td>
<td>Sublingual immunotherapy</td>
<td>NA Milk, peanut, hazelnut, others</td>
<td>Small phase I/II</td>
<td>High rate of partial desensitization, full desensitization rare</td>
<td>Appears safer than oral immunotherapy</td>
<td>47, 76</td>
</tr>
<tr>
<td></td>
<td>Oral immunotherapy plus omalizumab</td>
<td>Omalizumab Milk, peanut</td>
<td>Small phase I</td>
<td>Desensitization in 80%–90%</td>
<td>Up to 10% rate of withdrawal for AEs in rush protocols</td>
<td>88, 89</td>
</tr>
<tr>
<td></td>
<td>Recombinant protein</td>
<td>EMP-123 Rectally administered, recombinant peanut within &lt;i&gt;E. coli&lt;/i&gt;</td>
<td>Small phase I</td>
<td>NA</td>
<td>50% withdrawal for AEs, 20% anaphylaxis</td>
<td>113</td>
</tr>
<tr>
<td></td>
<td>Epicutaneous patch</td>
<td>ViaSkin, others Epicutaneous patch for milk</td>
<td>Phase I/II</td>
<td>Unknown</td>
<td>Well tolerated</td>
<td>96</td>
</tr>
<tr>
<td>Nonspecific</td>
<td>Chinese herbal medicine</td>
<td>FAHF-2 Mixture of traditional Chinese medicine herbs</td>
<td>Phase I trials</td>
<td>Unknown</td>
<td>Well tolerated</td>
<td>104, 106</td>
</tr>
<tr>
<td></td>
<td>Anti-IgE</td>
<td>Omalizumab, TNX-901 Peanut</td>
<td>Phase I trials</td>
<td>At least partial desensitization in most subjects</td>
<td>Well tolerated</td>
<td>81, 82</td>
</tr>
</tbody>
</table>

AE, adverse event.
old by at least 10-fold at 44 weeks, compared with 15% of placebo-treated subjects, but none of the treated subjects were able to pass a full food challenge at that time (47). Our group compared milk oral immunotherapy to sublingual immunotherapy in 30 children and found that after approximately 18 months, 70% of oral immunotherapy subjects tolerated a full serving of milk compared with 10% of sublingual immunotherapy subjects (76). However, sublingual immunotherapy appears to be safer than oral immunotherapy, with significantly fewer multi-system, gastrointestinal, and lower respiratory reactions (76).

It is likely that the differences between oral immunotherapy and sublingual immunotherapy, with regard to both efficacy and safety, primarily represent differences in the antigen doses that are used, given that typical sublingual immunotherapy maintenance doses are under 10 mg compared with 1 to 4 grams for oral immunotherapy. Improved sublingual immunotherapy efficacy might therefore be possible if higher doses were used, and in theory, an ideal sublingual immunotherapy dose might achieve efficacy similar to oral immunotherapy at far lower — and therefore safer — doses, given the high density of tolerogenic APCs in the sublingual space (77). However, significantly higher sublingual immunotherapy doses will not be possible unless more concentrated extracts or alternative delivery systems, such as the tablets that have been formulated for grass pollen sublingual immunotherapy (78), are developed. Until then, maximum sublingual immunotherapy dosing will remain limited by the concentration of the available aqueous extracts and the volume of liquid that can be safely administered sublingually.

The mechanisms of action underlying oral immunotherapy and sublingual immunotherapy are not entirely clear, although it is likely that a combination of suppressive mechanisms, anergy, and deletion of reactive T cells is important (refs. 21, 35, and Figure 1). With treatment, allergen-specific IgE tends to rise initially and then fall marginally by the completion of treatment. Specific IgG4 levels increase, and markers of basophil activation and mast cell reactivity (as evidenced by skin test responses) typically decrease (47, 53, 76). Although these changes occur in most subjects, especially with oral immunotherapy, and may be associated with more positive outcomes, the data thus far do not allow for any of these measures to be used as reliable biomarkers of clinical response.

Although short-term desensitization appears to be common with oral immunotherapy, the prospect for complete resolution of allergy — or even a sustained period of unresponsiveness (also sometimes called “clinical tolerance”) — appears to be limited with current methods. Although the persistence of desensitization after avoidance of the allergen has only been assessed in a few studies, the results are sobering overall. In our study of milk allergy, 40% of subjects desensitized with oral immunotherapy remained unresponsive at six weeks, and some regained reactivity within a week (76). Similarly, in the egg oral immunotherapy study, only 36% of desensitized subjects had sustained unresponsiveness four to six weeks after the discontinuation of therapy (39). This concept is very important because if sustained protection is not the norm, the long-term safety of food immunotherapy after treatment may be far more problematic than the known short-term risks during treatment. Although data on long-term outcomes are limited, we recently reported in a follow-up of two milk oral immunotherapy studies three to five years after study completion that only 25% of subjects were consuming normal servings of milk without any symptoms, that almost 30% were having regular or predictable symptoms, and that, most concerning, almost 20% had anaphylaxis during the follow-up period, including some who appeared to have had an excellent response to treatment (79). While ongoing consumption of the problem food may afford protection for most, this does not appear to always be the case and, even if it were, this is something that can be difficult to achieve, even in the research setting (80).

**Figure 1**

Potential mechanisms by which specific immunotherapy to food may act. Multiple cellular responses to immunotherapy may contribute to reduced immune activation, including deletion of effector Th2 cells, desensitization of mast cells and basophils, and induction of tolerogenic DCs. In addition, immunotherapy may promote allergen-specific Tregs, which in turn suppress effector T cells, reduce activation of mast cells and basophils, and trigger B cells to first increase IgG4 production and then decrease IgE production, leading to decreased activation of mast cells and basophils. Mechanisms of other therapeutic approaches (not shown) may include (a) inhibition of IgE binding (anti-IgE), (b) reduced basophil activation (FAHF-2), (c) reduced Th2 responses (FAHF-2, helminths), and (d) induction of tolerogenic DCs (DNA vaccines).
specific and thus may be effective for those who are allergic to multiple foods. However, their effects are dependent on continued injections of the antibody (81, 82), and this treatment is very expensive.

Theoretically, addition of an anti-IgE antibody alongside oral immunotherapy might reduce the rate of adverse events and/or improve long-term outcomes. In conjunction with s.c. injection using aeroallergens, omalizumab improved the tolerability of dose escalation while retaining and perhaps improving the long-lasting benefits of immunotherapy (83–87). Two small, uncontrolled studies of omalizumab combined with rush food oral immunotherapy, where dose escalation was performed at a more rapid pace than usual using either peanut or milk, have recently been published.

In the first, nearly all subjects were able to complete dose escalation, but 40% of subjects experienced a grade 2 or 3 adverse event, defined as the presence of moderate to severe symptoms (88). In the second, 30% of subjects still had reactions requiring epinephrine (89). Thus, although omalizumab may improve the tolerability of oral immunotherapy, it may only benefit a subset of patients, and determination of its specific role awaits the results of controlled studies that are currently underway (ClinicalTrials.gov identifier: NCT01157117).

**Epicutaneous immunotherapy and other routes of immunotherapy.**

Recently, there has been increasing interest in the potential to deliver immunotherapy via the skin and other routes. It has now been shown that with epicutaneous (e.c.) immunotherapy, antigen presented to intact skin is captured by DCs and brought to deliver immunotherapy via the skin and other routes. It has now been shown that with epicutaneous (e.c.) immunotherapy, antigen presented to intact skin is captured by DCs and brought to deliver immunotherapy via the skin and other routes.

Epicutaneous immunotherapy is safe and well tolerated and showed immunologic effects, including decreased IL-5 levels and basophil activation, as measured by CD63 expression (104, 106). A controlled efficacy study is currently being conducted in adolescents and adults (ClinicalTrials.gov identifier: NCT00602160), and studies using FAHF-2 along with oral immunotherapy will hopefully begin in the near future.

**Novel therapeutic strategies in preclinical or early clinical investigation**

**Modified protein immunotherapy.** Immunotherapy with proteins that are modified so that IgE-binding epitopes are removed or significantly altered, while maintaining relevant T cell binding, could provide efficacy similar to that of the unmodified protein, with an improved safety profile. This approach could theoretically make it possible to induce tolerance with far shorter courses of therapy by safely providing higher doses of the tolerogenic epitopes with little or no need for gradual dose escalation. Two general approaches are currently underway, one relying on the modification of the IgE-binding sites to reduce reactivity and the second based upon the identification of specific tolerogenic epitopes that are spliced from the larger molecule and provided as “peptide immunotherapy.” The latter approach is now in phase 3 trials for cat allergy (107, 108) and is being actively pursued in preclinical trials for other allergens, including egg and fish (109, 110).

The former approach has been applied to peanut allergy by developing a recombinant vaccine in which the IgE-binding epitopes on the three major peanut allergens, Ara h1, h2, and h3, were modified by single amino acid substitutions, then encapsulated in inactivated E. coli (EMP-123). In mouse models, the modified allergens did not bind IgE or induce basophil reactivity (111); however, they reduced anaphylactic symptoms on rechallenge with allergen in sensitized animals (112). In spite of the encouraging results in the animal model, a recent phase I trial of EMP-123 administered per rectum was disappointing (113). In fact, acute allergic reactions were so common that five of ten peanut-allergic subjects were unable to complete dosing, indicating that IgE binding was not adequately reduced by this modification.

**Adjuvants.** Another promising approach is to combine the immunotherapeutic protein — either intact or modified — with other components to enhance tolerogenic responses. For example, the TLR9 agonist CpG oligodeoxynucleotide leads to a Th1-skewed DC response. Further, linking CpG with allergen has reduced anaphylactic reactions to allergen when used either as a preventative therapy or as treatment in the form of s.c. injection or e.c. immunotherapy (114–118) in murine models. In humans, a similar TLR9 agonist, phosphorothioate oligodeoxynucleotide DNA, was coupled to ragweed antigen and showed some efficacy in a pilot s.c. injection study (119). Another potential adjuvant is chitosan, a polymer commonly found in the cell walls of fungi and many invertebrates, which activates macrophages and modulates Th2 inflammation (119–121). Mice fed chitosan did not exhibit orally induced peanut sensitization and had reduced IL-4, IL-5, and IL-10 production and increased IFN-γ (119–121). A more direct approach is to fuse inhibitory human molecules, in this case the human IgG FcγI, to allergen. Linking FcγI directly to Ara h2 causes aggregation of the inhibitory receptor FcγRIIB and the high-affinity IgE receptor FcεRI and thus inhibits degranulation of mast cells and basophils. In a murine model, s.c. treatment with FcεI-Ara h2 inhibited anaphylaxis to peanut (122). However, the likelihood of efficacy in humans and the long-term disease-modifying potential of these therapies are unclear. Although we are not aware of any current human trials using these or other adjuvant strategies for the treatment of food allergy, it is critical that...
these be pursued in future trials, as they may be the best means of optimizing outcome with therapies that might permit the use of lower allergen doses over shorter treatment periods.

DNA vaccines. A distinct immunotherapeutic approach is to eschew protein administration altogether and instead provide exposure to allergen in the form of DNA. Using a variety of vectors, DNA encoding allergen is administered and then incorporated into APCs, where it is ultimately translated into protein, potentially leading to Th1-biased responses (123). In a mouse model, oral gene delivery with chitosan-DNA nanoparticles protected against the development of peanut allergy (123). In humans, DNA vaccines for infectious diseases exhibit only modest immunological effects and have not yet shown efficacy when used alone (124). For allergy, a DNA plasmid vaccine for treatment of Japanese red cedar allergy is currently in phase I trials (ClinicalTrials.gov identifiers: NCT01707069 and NCT01966224). Although the limited human trials conducted thus far have shown good tolerability, the possibility that DNA may be integrated into the host genome remains a concern and may limit application to food allergy. Further, this approach shows more promise for prevention than it does for treatment, and while preventative strategies are certainly desirable, more general approaches would be preferable to food-specific approaches such as these.

Helminth administration. In an extension of the hygiene hypothesis, which states that the relative lack of infectious exposures of modern life has contributed to the epidemic of allergic diseases, some researchers have tried administration of helminths for control of allergic diseases, including allergic rhinitis and atopic eczema. Helminths secrete a variety of factors known to inhibit B cell and mast cell activity (125). In small studies of allergic and autoimmune diseases done thus far, administration of Trichuris suis (pig whipworm) was relatively well tolerated, with some gastrointestinal side effects and eosinophilia, but efficacy for allergic diseases has not yet been shown (125).

Effects on clinical practice

Although the FDA has not licensed any of these treatments for food allergy and most experts strongly oppose their use in current clinical practice, both oral immunotherapy and sublingual immunotherapy are being used increasingly in practice settings (126). Is this a justifiable practice? On the one hand, there is accumulating evidence that desensitization can be widely achieved with current protocols. On the other, the safety data we do have indicate that oral immunotherapy is a more risky treatment than we normally tolerate for diseases of this severity, with a relatively high per-patient rate of systemic reactions. Reactions can be unpredictable, with factors such as illness and menses that can lead to reactions with previously tolerated doses (34). Unlike s.c. injection, oral immunotherapy and sublingual immunotherapy are administered at home, without medical supervision. Most concerning, however, is our lack of understanding of the long-term trajectory of patients treated with oral immunotherapy or sublingual immunotherapy and whether they are durably protected from serious reactions. Patients and their families who choose these therapies may not fully understand the risk/benefit ratio, especially if they are being treated in clinical settings without informed consent. Although some patients and providers may believe that these therapies, especially oral immunotherapy, will limit the risk of serious or fatal reactions from food, there are currently no data that show a decreased rate of serious life-threatening reactions with oral immunotherapy, much less cost-effectiveness. In fact, all placebo-controlled trials show a higher rate of serious reactions overall in treated subjects, and it is our clear impression that the risk of significant adverse reactions is far higher in those being treated than in those practicing strict avoidance (47, 53). These risks may be justified depending on the long-term treatment outcome, but long-term results are currently unavailable, and studies to date are certainly not reassuring. More research is clearly needed to understand both sides of the risk/benefit equation and to optimize therapy to reduce risks. In addition, research should address patient preferences and quality of life with these therapies.

Future directions

Future research will likely maintain some form of immunotherapy at its core and focus on increasing efficacy and/or safety. It is possible that some therapies may provide a level of desensitization that protects from accidental reactions but does not eliminate all reactivity. It may also be possible that long-term, even lifetime, treatment with all of these therapies may be needed to sustain protection. Adjuvants, recombinant or pepsinized proteins, DNA vaccines, and/or coadministration with anti-IgE or Chinese herbs could all allow for safer and more effective therapy, but additional research is clearly needed. In the next decade, despite concerns about safety and efficacy, we do anticipate the wide use of food immunotherapy in general practice, but our hope for the next two to three decades is that therapies can be developed that are both safer and more effective and include the induction of sustained protection, or even true immunologic tolerance, for the vast majority of patients with persistent, severe food allergy.

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