Cilia are critical mediators of paracrine signaling; however, it is unknown whether proteins that contribute to ciliopathies converge on multiple paracrine pathways through a common mechanism. Here, we show that loss of ciliopathy-associated proteins Bardet-Biedl syndrome 4 (BBS4) or oral-facial-digital syndrome 1 (OFD1) results in the accumulation of signaling mediators normally targeted for proteasomal degradation. In WT cells, several BBS proteins and OFD1 interacted with proteasomal subunits, and loss of either BBS4 or OFD1 led to depletion of multiple subunits from the centrosomal proteasome. Furthermore, overexpression of proteosomal regulatory components or treatment with proteasomal activators sulforaphane (SFN) and mevalonolactone (MVA) ameliorated signaling defects in cells lacking BBS1, BBS4, and OFD1, in mormon zebrafish embryos, and in induced neurons from Ofd1-deficient mice. Finally, we tested the hypothesis that other proteasome-dependent pathways not known to be associated with ciliopathies are defective in the absence of ciliopathy proteins. We found that loss of BBS1, BBS4, or OFD1 led to decreased NF-κB activity and concomitant IκBα accumulation and that these defects were ameliorated with SFN treatment. Taken together, our data indicate that basal body proteasomal regulation governs paracrine signaling pathways and suggest that augmenting proteasomal function might benefit ciliopathy patients.

Introduction
In vertebrates, the cilium and the basal body are key components of paracrine signaling transduction. This has, in turn, suggested that phenotypes of ciliopathy patients might be attributed to defective paracrine signaling (1), including polyactyly due to defective sonic hedgehog (Shh) signaling (2) and renal cysts attributed to unbalanced Wnt signaling (3). Some data have raised the possibility that a fraction of these defects, especially Wnt, are driven by nonciliary functions of ciliary and basal body proteins (2, 4); other findings have indicated that the cilium/basal body Wnt roles are likely specific to discrete spatiotemporal contexts (5–9).

Although basal body and ciliary proteins are not signaling molecules per se, these structures are thought to operate as a hub for coordinating networks of signaling cascades. Components of various signaling pathways localize to basal body and cilium (10–13). Moreover, mutations in a single basal body or ciliary gene can lead to defects in more than one signaling pathway (11, 14), while loss-of-function mutations in signaling molecules such as the Shh regulator kinesin family member 7 (KIF7) (15, 16) and the Wnt/planar cell polarity (PCP) effector Fritz (17) cause ciliopathies in some families.

The increasing association of ciliary and basal body proteins with signaling defects led us to ask whether a common mechanism might account for the convergence of multiple pathways. One candidate, the ubiquitin-proteasome system (UPS), is attractive for three reasons. First, the basal body is known to be a proteolytic center (18–21). Second, we reported previously that disruption of some basal body proteins results in loss of proteasome-dependent degradation of β-catenin (5), a phenotype reproduced subsequently (22). Finally, we noted that, in addition to Wnt, proteasome degradation is implicated in most paracrine signaling cascades known to be defective in basal body mutants. For example, in Notch-mediated lateral inhibition, a transmembrane ligand of the Delta-Serrate-LAG2 (DSL) family binds a transmembrane Notch receptor in the adjacent target cells (23). To reduce Notch signaling, DSL ligands are ubiquitinated, internalized, and degraded by the proteasome (24); similarly, the intracellular domain of the Notch receptor is degraded by the proteasome to reduce Notch signaling (25). Likewise, when Shh ligand is present, glioma-associated oncopogenes 2 and 3 (GLI2/3) exist in their full-length activator forms, whereas they are truncated by proteasome-mediated proteolysis to their repressor forms when Shh is removed (26); both the activator and repressor forms of GLI2/3 are also degraded by the proteasome (27). Besides GLI2/3, suppressor of fused homolog (SUFU), a negative regulator of Shh signaling that physically localizes at the cilium, is also degraded in a proteasome-dependent manner (28).

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Ciliopathy proteins regulate paracrine signaling by modulating proteasomal degradation of mediators

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Given these observations, we wondered whether basal body and ciliary proteins might regulate multiple signaling pathways by controlling proteasome-mediated degradation of signaling mediators. We show that suppression of basal body–localized ciliopathy proteins led to defective proteasomal degradation of such mediators, which in turn caused dysfunction in three major cilia-associated signaling pathways (Shh, Wnt, and notch) in vitro and in vivo. These observations are unlikely to reflect non-specific cellular malaise; not only could the ciliopathy proteins tested here interact with specific regulatory subunits of the proteasome holoenzyme, but also depletion of each of Bardet-Biedl syndrome 4 (BBS4) and oral-facial-digital syndrome 1 (OFD1) selectively perturbed the subunit composition of the centrosomal proteasome. Further, signaling phenotypes due to depletion of BBS1, BBS4, and OFD1 could be rescued by activating proteasomal function. Finally, because the model predicts that other proteasome-dependent paracrine signaling pathways should also be defective in the absence of basal body proteins, we examined NF-κB signaling, a pathway that has not been linked previously to basal body (dys)function. Consistent with our prediction, loss of each of BBS1, BBS4, and OFD1 repressed NF-κB activity, which could be restored by activating the proteasome. Taken together, these findings suggest that proteasome-mediated basal body regulation might be a common mechanism for a multitude of signaling cascades and that proteasomal activation might be a potential treatment paradigm for ciliopathies.

Results

Accumulation of signaling mediators upon depletion of ciliopathy proteins. We and others have reported previously that perturbation of some ciliopathy proteins alters the stability of Wnt signaling mediators such as β-catenin and Dishevelled (5, 8, 21, 29, 30). To study the physiological relevance of these findings in vivo, we mated Bbs4–/– mice with a transgenic proteasome reporter mouse line expressing unstable ubiquitin–tagged GFP (31) to generate UbG76V-Gfp Bbs4–/– mice. As a hallmark of potential proteasomal dysfunction, we examined GFP degradation defects in the photoreceptors of mice. As a hallmark of potential proteasomal dysfunction, we examined NF-κB signaling, a pathway that has not been linked previously to basal body (dys)function. Consistent with our prediction, loss of each of BBS1, BBS4, and OFD1 repressed NF-κB activity, which could be restored by activating the proteasome. Taken together, these findings suggest that proteasome-mediated basal body regulation might be a common mechanism for a multitude of signaling cascades and that proteasomal activation might be a potential treatment paradigm for ciliopathies.

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Selective disruption of UPS caused by loss of ciliopathy proteins. Given our data on Wnt, Shh, and Notch and the fact that the processing of components from these paracrine signaling pathways is known to be mediated by the proteasome (11, 35), we asked whether proteasomal agonists could ameliorate the signaling phenotypes of basal body mutants.

The expression levels of three catalytic subunits of the proteasome, proteasome subunit β type 5, 6, and 7 (PSMB5, PSMB6, and PSMB7), as well as the corresponding peptidase activities, increased upon treatment with sulforaphane (36) [SFN; 1-isothiocyanato-4(R)-methylsulfinylbutane], an isothiocyanate extracted from cruciferous vegetables, rendering this compound an attractive initial substrate for our studies. We found that treatment with SFN rescued the accumulation of GLI2FL,
GLI3FL, and SUFU in T8-derived Ofd1KO neurons (Figure 3A).

Consistent with the participation of the proteasome in both the processing and degradation of GLI2/3, treatment with SFN revealed a reduction in total NICD levels (Figure 3C and Supplemental Figure 3E) and an approximately 50% ± 16% reduction in total JAG1 levels (Supplemental Figure 3F), suggesting that defects in proteasome-dependent processing and degradation of GLI2/3, SUFU, and β-catenin can be ameliorated.

Taken together, our data argue for a role of basal body proteins in regulating proteasome-mediated degradation and that we should observe a reciprocal signaling/proteasomal phenotype upon overexpressing our proteins of interest. We therefore overexpressed BBS4 in cells and observed a depletion of both NICD (Figure 3C and Supplemental Figure 3E) and JAG1 (Supplemental Figure 3F); further, we treated BBS4-overexpressing cells with two proteasome inhibitors, N-carbobenzoxyl-l-leucinyl-l-leucinyl-l-norleucinal (MG132) and lactacystin as well as DMSO vehicle as a control. While cells overexpressing BBS4 showed a 21%–50% reduction in total NICD levels (Figure 3C and Supplemental Figure 3E) and an approximately 50% ± 16% reduction in total JAG1 levels (Supplemental Figure 3F), treatment with MG132 and lactacystin restored NICD protein levels to 93% (Figure 3C) and 138% (Supplemental Figure 3E) of basal levels, respectively, and restored JAG1 levels to 99% ± 38% of basal levels (Supplemental Figure 3F), suggesting that BBS4 can facilitate the degradation of Notch signaling mediators in a proteasome-dependent manner.

Figure 1
Accumulation of GFP in Bbs4−/− mice. (A and B) Immunoblotting with anti-GFP to examine the kidney (P80), liver (P144), several brain components (P80), and retina (P12–P126) of UbG76V-Gfp Bbs4−/− mice, with UbG76V-Gfp WT littermates used as controls. Samples in each panel for the brain were run on the same gel but were noncontiguous. (C) Immunohistochemistry of retinal sections of P23 and P126 UbG76V-Gfp transgenic mice. Progressive retinal degeneration and GFP accumulation in photoreceptors (OS and IS, and ONL) were observed in Bbs4−/− mice, but not in WT littermates. OS was immunolabeled with anti-α-opsin; ONL and INL were labeled with DAPI staining. RPE, retinal pigment epithelium; OS, outer segment of the photoreceptors; IS, inner segment; ONL, outer nuclear layer; OPL, outer plexiform layer; INL, inner nuclear layer; IPL, inner plexiform layer; GCL, ganglion cell layer. White boxes delimit the enlarged images, showing OS and IS. Scale bar: 25 μm in images and inserts. Bar graphs showing standard error of the mean are plotted adjacent to each blot. *P < 0.05; **P < 0.01.
10 [RPN10], regulatory proteasome non-ATPase subunit 13 [RPN13], RPT6, and non-19S regulatory subunit [PA28γ], selected based on antibodies available for subsequent experiments). Under stringent detergent conditions (1% Triton X-100), we confirmed the previously reported interaction between BBS4 and RPN10 (5) by semiendogenous coimmunoprecipitation (Figure 4A) and also observed a biochemical interaction of some BBS proteins (BBS1, BBS2, BBS4, BBS6, BBS7, and BBS8) with proteasomal components, while BBS5 and BBS10 did not interact with any tested proteasomal subunits (Supplemental Figure 4A). To test the physiological relevance of a BBS-proteasome interactome, we determined that BBS1 and RPN10 interacted at endogenous levels in protein lysates isolated from C57BL/6 mouse testes (Figure 4A).

To probe the relevance of these interactions, we asked whether loss of OFD1 or BBS4 alters the composition of the proteasome with respect to its subunits. Using a stable cell line expressing tagged regulatory proteasome non-ATPase subunit 11 (RPN11) (37), we suppressed OFD1, purified 26S proteasome complexes, and observed a robust reduction (60% ± 8%) of 26S-bound regulatory proteasome ATPase subunit 2 (RPT2) in OFD1-depleted cells in comparison with that in control, while total RPT2 protein abundance was not affected by OFD1 depletion (Figure 4B). We also assayed the fraction of 26S-bound RPN10 in BBS4-depleted cells and observed a modest but significant reduction (20% ± 3%) in RPN10 (Supplemental Figure 4B). Furthermore, we also tested the localization of RPN10 around the centrosome in HEK-293 cells depleted of BBS4 and OFD1 by quantifying the RPN10 signal that localized around centrosomal γ-tubulin; we did not observe appreciable changes in BBS4-depleted cells (data not shown), possibly because of a lack of spatial resolution. By contrast, we observed a 43% ± 2% reduction of pericentriolar RPN10 levels (normalized to cytoplasmic RPN10 levels) in OFD1-depleted cells (Figure 4C). Given these data, we tested whether loss of OFD1 or BBS4 altered the composition of RPN10, RPT2, and other proteasomal components by sedimentation. Based on the commercial availability of reliable antibodies against proteasomal subunits, we found that RPN10, RPT6, and RPT2 were enriched in γ-tubulin-enriched fractions in control cells (Figure 4D and Supplemental Figure 4C). Depletion of OFD1 or BBS4, however, resulted in a shift in the peak expression of the proteasomal subunits (Figure 4D and Supplemental Figure 4C). Taken together, these data suggest that BBS proteins and OFD1 regulate the composition of the centrosomal proteasome, likely through direct biochemical interactions.

Activation of proteasomal components ameliorates signaling defects caused by loss of ciliopathy proteins that reside at the basal body. Our observations next led us to speculate that (a) if reduction of proteasomal

Figure 2
Accumulation of Shh and Notch signaling mediators upon depletion of ciliopathy proteins. (A) Accumulation of GLI2FL, GLI3FL, and SUFU as well as reduction of GLI3R in T8-derived Old1KO neurons. (B) At E10.5, accumulation of GLI2FL, GLI3FL, and SUFU as well as reduction of GLI3R were detected in protein lysates from Old1KO mice, with Old1CO mice used as controls. (C) Suppression of BBS4 increases Flag-tagged NICD levels compared with those in pSuper controls. (D) Suppression of BBS4 led to a 2-fold increase in GFP-tagged JAG1. The samples in each panel in B and C were run on the same gel but were noncontiguous. Bar graphs showing the SEM are plotted adjacent to each blot. "P < 0.05; "**P < 0.01.
subunits in the centrosomal fraction of BBS4- or OFD1-depleted cells results in hampered degradation of signaling mediators, then increasing expression of the corresponding proteasomal subunits may ameliorate defective signaling phenotypes; and (b) activation of the proteasome might compensate for signaling defects.

We and others have reported previously that loss of ciliopathy proteins produces defects in convergent extension (CE) movements during gastrulation in zebrafish embryos (5, 29, 30). In addition to and independent of these phenotypes, we observed defects in the definition of somite boundaries in zebrafish embryos injected with morpholinos against \textit{bbs} genes (\textit{bbs} morphants [\textit{bbs} MO]) (Supplemental Figure 5A and ref. 38). In vertebrates, somites develop as epithelial blocks at a temporal and spatial periodicity that is controlled, in part, by Notch signaling (39). Given the phenotypic overlap between \textit{bbs} morphants and segmentation mutants (\textit{mib}, \textit{bea}, \textit{des}, and \textit{aei}) (39), as well as our biochemical Notch observations in BBS4-deficient cultured mammalian cells, we wondered whether dysfunction of these proteins might influence Notch signaling. Upon injection of the established \textit{bbs4} morpholino (5, 38) into 1- to 8-cell-stage embryos, and scoring at the 9 ± 1 somite stage (9 ± 1 ss), 92% of embryos exhibited an expansion of the anteroposterior midline expression domain of \textit{her4} (Supplemental Figure 5A), whose transcription is targeted directly by \textit{notch1} activation (40). To differentiate between ectopic expression at developmental stages, when Notch signaling is already active, and the possibility that signaling is persistent throughout stages when it should be diminished, we performed an incremental developmental series of \textit{her4} in situ hybridization over the first 5 days of development. Scoring WT embryos (staged by number of somites and, later, by the presence of anatomical features such as the swim bladder to ensure that embryos of the same age were compared across experiments), we observed that the expression of \textit{her4} in neural structures of the head, including the developing forebrain, midbrain, hindbrain, and eye, was robust through 2.5 days post fertilization (dpf) and then began to wane. In \textit{bbs4} morphants, we saw persistent \textit{her4} expression through 5 dpf, especially in the eye (Supplemental Figure 5C).

Given the availability of experimentally tractable signaling phenotypes, we asked whether overexpression of proteasomal subunits that were seen to mislocalize in BBS4- and OFD1-depleted cells might rescue the \textit{bbs4} and \textit{ofd1} morphant phenotype. Upon blind scoring for the effects of coinjecting human \textit{RPN10}, \textit{RPN13}, or \textit{RPT6} mRNA with the \textit{bbs4} or \textit{ofd1} morpholino, we measured substantial rescue of defective CE, somitic defects, and persistent \textit{her4} expression (Figure 5A).

Next, as an independent test, we examined \textit{bbs4} morphants coinjected with SFN, a known transcriptional activator of proteasomal subunits that were seen to mislocalize in BBS4- and OFD1-depleted cells might rescue the \textit{bbs4} and \textit{ofd1} morphant phenotype. Upon blind scoring for the effects of coinjecting human \textit{RPN10}, \textit{RPN13}, or \textit{RPT6} mRNA with the \textit{bbs4} or \textit{ofd1} morpholino, we measured substantial rescue of defective CE, somitic defects, and persistent \textit{her4} expression (Figure 5A).

Figure 3 Disruption of proteasomal degradation caused by loss of ciliopathy proteins. (A) Treatment of proteasomal agonist SFN ameliorated the accumulation of GLI2FL, GLI3FL, and SUFU in T8-derived \textit{Ofd1KO} neurons. (B) Suppression of \textit{BBS4} in HEK-293-FT cells led to a 1.57-fold increase in β-catenin protein levels that could be rescued by SFN. (C) Overexpression of \textit{BBS4} reduced NICD levels. MG132 treatment restored Flag-NICD levels. Samples in each panel in C were run on the same gel but were noncontiguous. Bar graphs showing SEM are plotted adjacent to each blot. *P < 0.05; **P < 0.01; ***P < 0.001.
To substantiate the rescue effects of SFN, we used another proteasomal agonist, mevalonolactone [known as mevalonic acid lactone, mevalonate, and (±)-β-hydroxy-β-methyl-δ-valerolactone and abbreviated hereafter as MVA], to ask whether broad activation of the proteasome can alleviate basal body–dependent phenotypes. We coinjected MVA with the bbs4 morpholino into zebrafish embryos, and upon blind scoring at 9 ± 1 ss, we found a reduction in bbs4 morphant zebrafish with CE defects from 47.8% to 11.4% with MVA (Supplemental Figure 6). To investigate the persistence of Notch signaling in bbs4 morphant zebrafish coinjected with MVA, we performed in situ hybridization for her4. Coinjection of MVA with a bbs4 morpholino reduced her4...
expression levels in neural structures, especially the retina, to those in WT zebrafish by 4.5 dpf (Supplemental Figure 6).

NF-κB signaling defects in basal body ciliopathies can be rescued by activation of the proteasome. Our findings indicate that Wnt, Notch, and Shh phenotypes generated upon loss of three basal body proteins might converge at the point of proteasomal degradation. We therefore considered a model in which the basal body region serves as a broad regulator of paracrine signaling through proteasomal degradation. If this model is true, then (a) other paracrine pathways not implicated previously in ciliary/basal body biology, but known to be regulated by the proteasome, should exhibit signaling defects upon depletion of BBS proteins or OFD1; and (b) the model should also be able to predict the direction of the phenotype (suppressed or excessive signaling). To test this, we turned to NF-κB signaling, a pathway involved in inflammatory responses and lymphoid organogenesis with no known link to basal body (dys)function. In response to stimuli, IκBs are degraded by the proteasome, releasing NF-κB to translocate to the nucleus and activate the transcription of target genes (41).

We transfected HEK-293-FT cells with an NF-κB luciferase reporter plasmid containing three copies of the κB response element of the murine MHC class I promoter (3XκB-L). Cells stimulated by TNF-α and cotransfected with the pSuperBBS4, pSuperBBS1, and pSuperOFD1 plasmids displayed a 55%, 53%, and 72% reduction in NF-κB activity, respectively, compared with that of control cells; incubation with SFN for 6 hours restored NF-κB activity (Figure 6, A–C). In cells with depleted basal body proteins, the direction of the NF-κB activity change (suppressive signaling) is opposite that of Wnt and Notch activity change (excessive signaling) (5, 22).

This inverse relationship is consistent with our model, since the predicted basal body–regulated proteasomal degradation substrates are IκBs, which are negative regulators, while β-catenin and NICD are positive regulators. Also consistent with the luciferase assays, we observed accumulation of GFP-tagged IκBβ in BBS4-, BBS1-, or OFD1-suppressing cells, which can also be ameliorated by SFN treatment (Figure 6, D and E).

Finally, to probe the potential physiological relevance of the observed NF-κB data in vivo, we examined the expression and protein levels of the proteasomal substrate IκBβ in mice lacking Ofd1. Immunoblot analysis showed no differences in IκBβ protein levels on protein lysates from E10.5 Ofd1<sup>−/−</sup> mutant embryos and controls (data not shown). Since loss of Ofd1 in mice results in prenatal lethality, we generated and crossed Ofd1-floxed mice (Ofd1<sup>fl/fl</sup>) with a CAG-Cre-ER<sup>TM</sup>–inducible general deleter line to examine IκBβ protein levels in postnatal Ofd1 knockout mice. In Ofd1<sup>fl/fl</sup> CAG-Cre-ER<sup>TM</sup> (Ofd1<sup>fl/fl</sup> CKO) mice, Ofd1 inactivation was achieved at E18.5 by tamoxifen injection, and renal cysts were not observed at P8 (precystic stage). By P20, we observed a replacement of the renal parenchyma by cysts (cystic stage). Immunoblot analysis revealed an accumulation of IκBβ in protein lysates from kidney of Ofd1<sup>fl/fl</sup> CKO male mice compared with controls; crucially, this phenotype was evident in both pre- and cystic stages (Figure 6F), suggesting that the accumulation of IκBβ is not a by-product of tissue disorganization and cystogenesis. At the precystic stage, mRNA levels of the IκB gene (Nfκb1b) were lower in Ofd1<sup>fl/fl</sup> CKO mice compared with mRNA levels in WT animals (Figure 6G), confirming that the accumulation of IκBβ is not due to increased mRNA transcription levels. Nfκb1b mRNA levels were comparable between Ofd1<sup>fl/fl</sup> CKO and controls at the cystic stage (Figure 6G).

Discussion

The study of ciliary and basal body proteins has highlighted a complex role for this cellular region in the regulation of signaling pathways. These observations have raised critical questions, including whether dedicated signaling transduction machinery aggregate around the cilium and basal body. Several transduction components have been localized to the basal body and/or the ciliary axoneme, including Smoothened, GLI1 protein, SUFU, β-catenin, adenomatosis polyposis coli (APC), and Notch3 (8, 10, 11, 13, 29). At the same time, a model in which the cilium facilitates each pathway independently is difficult to reconcile with the fact that (a) the possible exception of the PCP transducer Fritz (17) and the Shh motor KIF7 (15, 16), none of the other 60-plus genes and proteins mutated in human ciliopathies are components of a specific paracrine signaling pathway; (b) the phenotype of ciliopathy patients is a mixture of defects more consistent with a context-specific pathway dysfunction; and (c) animal models ablated for specific ciliary or basal body proteins exhibit multiple signaling defects (1).

Our data suggest a simpler model, in which at least some basal body proteins play a role in signal transduction regulation by exerting their primary effect not on a given pathway per se, but by regulating context-dependent proteolytic degradation. The alternative would be that the observed phenotypes are the non-specific consequence of generalized cellular malaise and that the observed rescue effects were reflective of broad improvement in the ability of the cell to eliminate proteins targeted for degradation. Taken together, our experiments favor the former model. Cells and embryos suppressed or ablated for each of BBS1, BBS4, and OFD1 had defects in proteasomal clearance of both reporter proteins and specific signaling components that included β-catenin, NICD, JAG1, GLI2, GLI3, SUFU, and IκBβ. It is also notable that the pronounced defects in proteasomal activity were concomitant with overt anatomical pathology in our murine models of disease, such as in the retina of Bbr mice and the cystic kidneys of conditional Ofd1 animals, whereas no GFP accumulation was observed in Bbr mutant kidneys that, in our colony, never exhibited cyst formation.

We tested the robustness of the model in four ways: (a) by predicting that NF-κB signaling, which requires proteasomal degradation (41), but has no known ciliary link, would be defective in the absence of our proteins of interest; (b) by predicting the direction of the defect in NF-κB signaling; (c) by ameliorating the signaling defects for each tested pathway in vivo through the chemical upregulation of proteasomal components; and (d) by demonstrating that in the absence of some basal body proteins, the centromosomal proteasome is partially depleted of the various regulatory subunits whose overexpression also has an ameliorating defect in vivo. Notably, we observed sucrose fraction sedimentation changes in multiple proteasomal subunits in the absence of OFD1 or BBS4, arguing that ciliopathy phenotypes are unlikely to be driven by specific defects in only one subunit, consistent with the observations that the mice haploinsufficient for the subunit RPN10 are phenotypically normal, at least by gross pathology, while homozygous Rpn10<sup>−/−</sup> mutants are embryonic lethal (42). An attractive mechanism is one in which basal body proteins regulate the composition of multiple subunits in the proteasome holoenzyme in a context-dependent manner. This is known to occur during cellular stress (43), and it is plausible that ciliary signaling can have a similar effect.
Moreover, it is likely that proteasomal dysfunction is only one contributor to the pathology of ciliopathies. The BBS proteins have been proposed to perforate vesicles and regulate vesicular transport, possibly into the ciliary axoneme, in a GTPase-dependent fashion (44), while BBS4 has also been reported to bind to EXOC7 (45). These data intimate additional layers of complexity, in which these proteins can either have both promoting and inhibitory functions with regard to proteasomal targeting or other subcellular surveying mechanisms, or might be themselves inhibited or activated by other ciliary and axonemal proteins in a spatiotemporal context.

Emergent data suggest that the composition and function of primary cilia vary significantly across cell types (46). Such complexity might account for conflicting observations, in which each of the major signaling pathways mapped to date on this subcellular region have been shown to be activated, suppressed, or irrelevant in different tissues (4–6, 46, 47).

Finally, our findings indicate potential therapeutic applications. SFN has shown no toxic effects in humans and is currently being tested in several anticancer trials (48, 49). Should this compound show therapeutic benefit in rodent ciliopathy mod-

Figure 5
Activation of proteasome ameliorates signaling defects in bbs and ofd1 morphant zebrafish embryos. (A) Coinjection of human RPN10, RPN13, and RPT6 mRNA into bbs4 and ofd1 morphant zebrafish embryos rescued somitic and CE defects at the 9 ± 1 ss and ectopic expression of her4 in the eye (arrowheads in lower row) at 4.5 dpf. CE defects were scored based on the body gap angle (arrowheads in upper row). Expression of her4 was detected by whole-mount RNA in situ hybridization. (B, C, and D) Coinjection of SFN rescued somitic and CE defects as well as ectopic her4 expression in bbs4 (B), bbs1 (C), and ofd1 (D) morphants and were shortened in morphant zebrafish embryos coinjected with SFN. In the second row (lateral view), the body gap angle (arrowheads) was greater in morphants and reduced in the presence of SFN. Dashed boxes delimit the enlarged images in the third row, showing the effects of SFN treatment on somite boundary definition defects. Percentage of embryos with somite boundary definition and CE defects and sample size (n) are noted below the images of each condition. Scale bars: 100 μm.
els, it would be a candidate for clinical trials in some ciliopathy patients. More broadly, other SFN derivatives as well as unrelated proteasome agonists might also represent natural candidates. However, SFN is likely to confer a partial benefit that is possibly both genotype and subphenotype dependent. First, the observed rescue of embryos injected with the compound, although significant, was not complete. Second, SFN is unlikely to be of benefit to all pathway defects. The proteasome is known to degrade the negative regulator SUFU and both the activator and the repressor forms of GLI2/3; as such, it is likely that downstream Shh signaling defects will not be rescued. Likewise, long-term exposure to SFN repressed NF-κB activity (data not shown), suggesting that delivery, dosage, and duration of exposure might be critical. Finally, if basal body ciliopathy proteins participate in other cellular processes that regulate signaling, proteasomal modification will only be partially effective. Given our findings, however, a compound approach that includes targeting the proteasome may be of benefit.

Methods

Immunoblotting. Transfected cells and mouse tissues were lysed in modified RIPA buffer [150 mM sodium chloride, 50 mM Tris-HCl, pH 7.4, 1% nonidet P-40, 0.1% sodium deoxycholate, 1 mM EDTA] with 1× proteasome inhibitor (Roche) and centrifuged at 4°C for 15 minutes. Protein concentration was measured by Lowry assay using the DC Protein Assay Kit (Bio-Rad) on a DU 530 Life Science UV/Vis Spectrophotometer (Beckman Coulter). Total protein in each sample was separated by SDS-PAGE on 4% to 15% Mini-PROTEAN TGX Precast Gel (Bio-Rad) with a Spectra Multicolor Broad Range Protein Ladder (Fermentas) and transferred to an Immun-Blot PVDF Membrane (Bio-Rad). The membrane was blocked with 5% nonfat milk or 5% BSA (Sigma-Aldrich) and probed with the following commercial primary antibodies: anti-GAPDH (ab9484 from Abcam or sc-32233 from Santa Cruz Biotechnology Inc.); anti-GFP (sc-8334; Santa Cruz Biotechnology Inc. or ab13970; Abcam); anti-HSP90 (sc-7947; Santa Cruz Biotechnology Inc.); anti-GLI2 (AF3635; R&D Systems); anti-GLI3 (AF3690; R&D Systems); anti-SUFU (sc-10934; Santa Cruz Biotechnology Inc.); anti-Flag (F7425; Sigma-Aldrich); anti-catenin (sc-7199; Santa Cruz Biotechnology Inc.).

Figure 6
NF-κB signaling defects in BBS4-, BBS1-, and OFD1-depleted cells, and Ofd1 conditional knockout mice. (A–C) Suppression of BBS4 (A), BBS1 (B), or OFD1 (C) in HEK-293-FT cells led to decreased 3X-κB-L luciferase reporter responsiveness to a 12-hour TNF-α treatment. Incubation with SFN for 6 hours partially restored sensitivity to TNF-α stimulation (n = 3). (D and E) GFP-tagged IkBα protein levels were higher in HEK-293-FT cells depleted of BBS4, BBS1 (D), or OFD1 (E) compared with those in control. SFN incubation rescued IkBα-GFP accumulation. (F) Protein levels of IkBα in kidney tissues isolated from Ofd1fl/y CAG-Cre-ERTM (Ofd1fl/y CKO) mice at P8 (precystic stage) and P20 (cystic stage) were higher than the levels in kidney tissues from WT littermates. (G) mRNA levels of IkBα (Nfkbib) in kidney tissues were lower in Ofd1fl/y CKO compared with mRNA levels in WT animals at P8 and were comparable between Ofd1fl/y CAG-Cre-ERTM and controls at P20 (n = 3). Bar graphs showing SEM are plotted adjacent to each blot. *P < 0.05; **P < 0.01; ***P < 0.001.
Biotechnology Inc.); anti-α-tubulin (T6199; Sigma-Aldrich); anti-NICD (ab8925; Abcam); anti-IsOFD1 (rabbit polyclonal antisera against human full-length OFD1 NM_003611); anti-mmOFD1 (rabbit polyclonal antisera against a portion of murine OFD1 NM_177429 aa 461-884); anti-BBS4 (AB15009; Millipore); anti-BBS1 (a16663; Abcam); anti-γ-tubulin (T7451; Sigma-Aldrich); anti-20S (NB600-1016; Novus); anti-P28 (NB1P-54587; Novus); anti-RPN10 (ab20239; Abcam); anti-RPN13 (H0001037-M01; Novus); anti-RPT2 (AP-107; Boston Biochem); anti-RPT6 (BML-PW9265; Enzo Life Sciences); anti-IkBβ (sc-9248; Santa Cruz Biotechnology Inc.); anti-Myc (m4439; Sigma-Aldrich); and anti-HA (ab16918; Abcam). Densitometric analysis was performed with Image Lab (Bio-Rad), Quantity One (Bio-Rad), or ImageJ 1.44p (NIH) software.

Immunohistochemistry. Mouse eyes were fixed in 4% PFA, followed by immersion in sucrose (10%, 20%, and 30%) in PBS. With the lens removed, eyecups were embedded in Optimal Cutting Temperature Compound (Sakura) and flash frozen. Cryosections were blocked with 10% FBS in PBS and probed with primary antibodies anti-GFP (ab13970; Abcam) and anti-β-opsin (a gift from Jeremy Nathans, Johns Hopkins University, Baltimore, Maryland, USA), followed by secondary antibodies Alexa Fluor 488 IgG (Invitrogen) and Alexa Fluor 594 IgG (Invitrogen). Nuclei were stained with DAPI (Roche). Images were captured with a Nikon Eclipse 90i microscope.

Bioinformatic analyses. We selected a subset of 271 transcripts that included: (a) transcripts mutated in human ciliopathies; (b) transcripts that, when mutated in animal models, give rise to ciliary dysfunction; (c) a group of transcripts found in at least three of the available datasets of ciliary proteins (50); and (d) a subset of transcripts recently shown to be modulators of ciliogenesis and ciliatum length (51). The publicly available online tool http://netview.tigem.it was used to analyze the regulatory interactions among genes from genome-scale measurements of gene expression profiles (microarrays) (32).

E9 cell derived in vitro neural differentiation. The Ofd1-deficient E14Tg2A.4 KOES line was obtained from BayGenomics. Both WT and Ofd1 KOESs were maintained in an undifferentiated state by culture on a monolayer of mitomycin C–inactivated fibroblasts in the presence of leukemia-inhibiting factor (LIF). To induce neural differentiation, we followed previously described protocols (33). Briefly, 48 hours after ES cells were seeded on gelatin-coated plates, they were dissociated and plated on gelatin-coated plates at 1,000 cells/cm2 on day 0 (T0). The culture medium for neuronal differentiation (serum-free KnockOut Serum Replacement–supplemented medium; Invitrogen) contained knockout DMEM supplemented with 15% KSR (Invitrogen), 2 mM L-glutamine, 100 U/ml penicillin-streptomycin, and 0.1 mM β-mercaptoethanol and was replaced daily during the differentiation process.

Cell culture, DNA transfection, and drug treatment. HEK-293 or HEK-293-FT cells and human dermal fibroblasts were grown in DMEM (Invitrogen) containing 10% FBS (Invitrogen) and 2 mM L-glutamine (Invitrogen), iTERT-RPE1 cells in DMEM and Ham’s F-12 Nutrient 1:1 mixture (DMEM/F-12; Invitrogen) with 10% FBS and 2 mM L-glutamine. FuGene6 Transfection Reagent (Roche) was used for transfection of expression constructs (including the Notch1-NICD expression construct, a gift from Nicholas Gaiano, Johns Hopkins University, Baltimore, Maryland, USA), shRNA-expressing plasmids, and luciferase reporter plasmids; then cells were cultured for 72 hours. Drug treatment was carried out at a final concentration of 10 μM SFN (Sigma-Aldrich) for 6 to 24 hours, 30 μM N-carbobenzoxyl-l-leucinyl-l-leucinyl-l-norleucinal (MG132; Calbiochem) for 5 hours, 20 μM lactacystin (EMD Bioscience) for 5 hours, and 50 ng/ml TNF-α (Sigma-Aldrich) for 12 hours.

IP. For IP, approximately 1 mg of whole-cell, embryo, or tissue lysate was incubated with anti-OFD1, anti-Flag, or anti-GFP at 4°C overnight, followed by incubation with protein G–coupled agarose beads (Santa Cruz Biotechnology Inc.) at 4°C for 1 hour, or directly with anti-Flag M2 beads (A2220 Sigma-Aldrich). The beads were collected and washed with IP buffer (10% glycerol, 50 mM Tris-HCl [pH 7.5], 2.5 mM MgCl2, 1% NP40, and 200 mM NaCl). Proteins conjugated with the beads were then denatured and separated from the beads by boiling at 95°C to 100°C for 5 minutes before processing for immunoblotting.

Affinity purification of proteasome complex. The 26S proteasome complex was purified following a previously described protocol (52) with modifications. Briefly, HEK-293-FT cells expressing stable HTBH-tagged hRPN11 (a gift from Lan Huang, University of California, Irvine, California, USA) were transfected with either pSuper control plasmid or pSuperBBS4 to knock down BBS4 expression. Seventy-two hours after transfection, cells were lysed in buffer A (100 mM NaCl, 50 mM Tris-HCl [pH 7.5], 10% glycerol, 2 mM ATP, 1 mM DTT, and 5 mM MgCl2) with 1× protease inhibitor (Roche). Lysates were centrifuged at 4°C for 15 minutes to remove cell debris. To purify proteasomes, an aliquot of the supernatant was incubated with streptavidin beads at 4°C overnight to precipitate HTBH-RPN11. The beads were then washed with buffer A three times, followed by one washing with TEB buffer (50 mM Tris-HCl, pH 7.5 and 10% glycerol). Finally, the beads were incubated in TEB buffer containing 1% TEV protease at 30°C for 1 hour, before SDS-PAGE and immunoblotting with anti-RPN10 (ab20239; Abcam).

Immunocytochemistry. HEK-293 cells cultured on coverslips were fixed in methanol, blocked in normal goat serum (1:10 in PBS containing 5% BSA), and then probed with anti-RPN10 and anti-γ-tubulin, followed by secondary antibodies Alexa Fluor 488 IgG and Alexa Fluor 568 IgG. Finally, nuclei were visualized with Hoechst 33258 (Sigma-Aldrich). Images were captured with a Zeiss LSM 710 confocal microscope and analyzed with ImageJ 1.44p software.

Sucrose gradient sedimentation. HEK-293-FT cells were transfected and treated with nocodazole (10 μg/ml) and cytochalasin B (5 μg/ml) for 1 hour at 72 hours after transfection. To collect cytoplasmic lysates, cells were harvested in lysis buffer (1 mM HEPES [pH 7.3], 0.5% NP-40, 0.5 mM MgCl2, 0.1% β-ME, and 1× protease inhibitor), followed by centrifugation at 2,500 g for 10 minutes. After 10 mM HEPES and 5 U/ml DNase treatment for 30 minutes on ice, 1 ml of cytoplasmic lysates was layered on a discontinuous sucrose gradient (70%, 50%, and 40% sucrose in the buffer containing 10 mM PIPES [pH 7.2], 0.1% NP-40, and 0.1% β-ME) and centrifuged for 1 hour at 195,000 g; 2% of lysates were kept before ultracentrifugation and served as an input. After ultracentrifugation, 13 fractions were collected and analyzed by immunoblotting.

Microinjection of morpholinos, mRNA, and sulfopharane. Morpholinos against bb1 (5′-CACACGTCATCATCACTAACAATAGC-3′), bb4 (5′-CGTTTCTCAGGTTCCGCGCAT-3′), and ofd1 (5′-ATCTTCTC-TACTGCAAACACATAC-3′) were purchased from Gene Tools, LLC. Human RPN10, RPN13, and RPT6 mRNA were in vitro transcribed with an mMESSAGE mMACHINE SP6 Kit (Ambion). SFN was dissolved in DMSO (Sigma-Aldrich) at a stock concentration of 1 mM and further diluted in water to 10 mM. The morpholino and mRNA or SFN were mixed, and a volume of 0.5 nl was microinjected.

Whole-mount RNA in situ hybridization. Zebrafish embryos were fixed overnight in 4% PFA at 4°C. Residual pigment was removed by bleaching with 3% H2O2/0.5% KOH. Whole-mount RNA in situ hybridization was performed with a digoxigenin-labeled anti-her4 RNA probe (a gift from Tohru Ishitan, Kyush University, Fukuoka, Kyushu, Japan) synthesized by in vitro transcription (Roche), followed by immunological detection with Anti-Digoxigenin-AP, Fab Fragments (Roche) and nitro blue tetrazolium/5-bromo-4-chloro-3-indolyl-phosphate staining (Roche).
Luciferase reporter system assays. HEK-293-FT cells were seeded in 24-well plates at a density of 10^4 cells/well. After 24 hours, cells were transfected with expression constructs, short-hairpin plasmids, and a 3x-Kl-B reporter (gift from Tom Gilmore, Boston University, Boston, Massachusetts, USA) for NF-kB signaling. A pRL-SV40 plasmid expressing Remilla luciferase was used as an internal control. Seventy-two hours after transfection, cells were lysed with Passive Lysis Buffer (Promega). The luciferase activity of lysates was measured using the Dual Luciferase Reporter Assay System (Promega) on a FLUostar Omega microplate reader (BMG LABTECH) and analyzed with MARS Data Analysis Software (BMG LABTECH).

Generation of Odd1^flp/CAG-CreERT2^/ mice by tamoxifen injections. Odd1^flp/ mice were crossed with CAG-Cre-ER^T2/ mice, a general doletor Cre line in which Cre-ER is ubiquitously expressed after tamoxifen injection. We treated pregnant mothers with a single i.p. injection of 100 μg tamoxifen/g of weight at E18.5. Tamoxifen (Sigma-Aldrich) was diluted in 10% ethanol and 90% sesame oil at a final concentration of 10 ml/g. Quantification of Odd1 inactivation was assessed by quantitative RT-PCR (qRT-PCR) (data not shown).

Real-time qRT-PCR analyses. Total RNA was isolated with TRIzol (Invitrogen). The cDNA was synthesized from 5 μg of total RNA using SuperScript III (Applied Biosystems). The primers Nfkbib forward (5′-TGGCAGACCACTTACAAAGATG-3′) and Nfkbib reverse (5′-GACTGTGCTGATGAATCACGC-3′) were used to test mRNA levels of Nfkbib, while the primers Odd1 forward (5′-TGGCGACACCTTACAAAGATG-3′) and Odd1 reverse (AGACTGGATGAGGGTTAATC-3′) were used to examine the conditional knockdown efficiency, and the primers gapd1 forward (5′-CTCTTGGGTTGCAGGTAGT-3′) and gapd1 reverse (5′-TGCAACCAACTGCTTAGC-3′) were used as internal controls. Real-time controls were real-time collected and analyzed with the Sequence Detection System software package, version 2.3 (Applied Biosystems).

Statistics. A one-tailed student’s t-test was performed to compare the means of two populations. In the bar graphs, data represent the mean ± SEM of multiple repeats (n ≥3). A χ2 test was performed to compare two populations with several subgroups of different proportions. Statistical significance of differences between samples is indicated by *P < 0.05, **P < 0.01, and ***P < 0.001. A P-value less than 0.05 was considered significant.

Study approval. Zebrafish and mouse studies were approved by the IACUC of Duke University (protocol A229-12-08 and A251-12-09) and by the IACUC of Cardarelli Hospital (Naples, Italy), to which the Telethon Institute of Genetics and Medicine (TIGEM) refers, and were authorized by the Italian Ministry of Health. The Cardarelli Hospital Ethics Committee (Naples, Italy) approved this study.

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