SPARC promotes leukemic cell growth and predicts acute myeloid leukemia outcome

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Aberrant expression of the secreted protein, acidic, cysteine-rich (osteonectin) (SPARC) gene, which encodes a matricellular protein that participates in normal tissue remodeling, is associated with a variety of diseases including cancer, but the contribution of SPARC to malignant growth remains controversial. We previously reported that SPARC was among the most upregulated genes in cytogenetically normal acute myeloid leukemia (CN-AML) patients with gene-expression profiles predictive of unfavorable outcome, such as mutations in isocitrate dehydrogenase 2 (IDH2-R172) and overexpression of the oncoproteins brain and acute leukemia, cytoplasmic (BAALC) and v-ets erythroblastosis virus E26 oncoprotein homolog (ERG). In contrast, SPARC was downregulated in CN-AML patients harboring mutations in nucleophosmin (NPM1) that are associated with favorable prognosis. Based on these observations, we hypothesized that SPARC expression is clinically relevant in AML. Here, we found that SPARC overexpression is associated with adverse outcome in CN-AML patients and promotes aggressive leukemia growth in murine models of AML. In leukemia cells, SPARC expression was mediated by the SP1/NF-κB transactivation complex. Furthermore, secreted SPARC activated the integrin-linked kinase/AKT (ILK/AKT) pathway, likely via integrin interaction, and subsequent β-catenin signaling, which is involved in leukemia cell self-renewal. Pharmacologic inhibition of the SP1/NF-κB complex resulted in SPARC downregulation and leukemia growth inhibition. Together, our data indicate that evaluation of SPARC expression has prognosticative value and SPARC is a potential therapeutic target for AML.

Introduction

The secreted protein, acidic, cysteine-rich (SPARC) gene, located at chromosome bands 5q31–q33 (1), is highly conserved in vertebrates and encodes a 32-kDa matricellular protein also known as osteonectin or BM-40 (2, 3) that regulates cell-matrix interactions and tissue remodeling through the binding of collagen and other extracellular matrix proteins and activation of matrix metalloproteinases (2). SPARC also interacts with and regulates several growth factors, including TGF-β, FGF, VEGF, and PDGF (4–7). This pleiotropic activity suggests that the SPARC protein is likely to be expressed when tissue undergoes normal or pathologic remodeling.

SPARC expression has been shown to be altered in a variety of human conditions (e.g., diabetes, obesity) and in several types of cancer. However, the biologic and clinical significance of this gene is controversial, and conflicting reports have classified SPARC as either a tumor suppressor gene or an oncogene. The difficulty in assigning a specific function to the SPARC protein is related to the diverse roles that it can play both intracellularly in malignant cells and extracellularly in the surrounding microenvironment (8). Low expression levels of SPARC were found in ovarian (9), colorectal (10, 11), and pancreatic cancer (12), whereas high SPARC expression was reported in breast cancer (13, 14), melanoma (15, 16), and glioblastoma (17). Stromal SPARC expression was associated with poor prognosis in non–small cell lung cancer (18) and with disease recurrence in breast ductal carcinoma in situ (19), whereas low stromal expression of SPARC predicted poor prognosis in colon cancer (20).

In hematologic malignancies, the role of SPARC is equally controversial. It was found to be downregulated at diagnosis in patients with del(5q) myelodysplastic syndromes (MDS) and upregulated following treatment with lenalidomide (21–23). SPARC was also found to be downregulated in acute myeloid leukemia (AML) with MLL rearrangements, usually associated with unfavorable prognosis, and upregulated in AML with t(8;21) or inv(16), which is usually associated with favorable prognosis, although no correlation of SPARC expression with outcome was reported (24). In chronic myelogenous leukemia, the accumulation of intracellular SPARC mediated by the Fyn/ERK signaling pathway seemingly contributed to imatinib resistance (25).

Recently, we observed that SPARC was upregulated in gene expression profiles (GEPs) associated with prognostically unfa-
SPARC overexpression is associated with adverse clinical outcome in CN-AML. Outcome of CN-AML patients (n = 153) according to SPARC expression levels. Median value of the measured SPARC expression levels was used to dichotomize patients into high and low expressers. (A) CR rates and DFS. (B) OS.

Patients were dichotomized into higher and lower SPARC expressers using the median value cut-off. With a median follow-up of 8.7 years, higher SPARC expressers had lower odds of achieving a complete remission (CR) (P = 0.03) and shorter disease-free survival (DFS) (P = 0.009; 5-year DFS 28% vs. 55%) and overall survival (OS) (P = 0.001; 5-year OS 29% vs. 56%) than lower expressers (Figure 1, A and B). In multivariable analyses, higher SPARC expression was independently associated with lower odds of CR (P = 0.007), once adjusting for white blood count (WBC) (P = 0.003), and shorter OS (P = 0.03), once adjusting for FLT3 internal tandem duplication (FLT3-ITD) (P < 0.001), WT1 (P = 0.003), and RUNX1 (P = 0.006) mutations and WBC (P < 0.001). There was also a trend for shorter DFS (P = 0.08) once adjusting for FLT3-ITD (P < 0.001) and WT1 mutation (P = 0.004). These data support the notion that SPARC is differentially expressed across AML patients and that the difference in the SPARC expression in this patient population has potentially biologic and clinical relevance.

SPARC promotes leukemia growth in vitro and an aggressive disease in vivo. Having shown that higher SPARC expression was associated with adverse outcome, next we interrogated the SPARC-dependent mechanisms mediating the aggressive phenotype. THP-1 cells express a relatively lower endogenous level of SPARC mRNA (Supplemental Figure 1) and therefore represent a suitable in vitro model for gain-of-function experiments. THP-1 cells were stably infected with pLenti-SPARC (THP-1/SPARC) or empty vector as a control (THP-1/EV). Ectopic expression of SPARC was confirmed by Western blotting (Figure 2A). A significant increase in the ability of colony formation was observed in THP-1/SPARC cells compared with the THP-1/EV cells (mean, 209 vs. 72; P = 0.007; Figure 2A). Likewise, when we knocked down SPARC (Figure 2B) in Kasumi-1 cells that express a relatively higher endogenous levels of the gene (Supplemental Figure 1), we observed a decrease in the colony-forming ability in SPARC siRNA–transfected cells compared with the control siRNA–transfected cells (mean, 106 vs. 50; P = 0.002; Figure 2B). Similar experiments were also conducted in primary blasts from 5 AML patients (numbers 1–5) expressing a relatively lower level of endogenous SPARC and 2 patients (numbers 6, 7) expressing a relatively higher level of endogenous SPARC (see patients’ cytogenetic and molecular features in Supplemental Table 2). For patients 1–5; blasts were infected with either the pLenti-SPARC or the pLenti-EV vector. A significant increase in colony forming ability was observed in SPARC-infected blasts compared with EV controls for patients 1–4 (means, 16 vs. 7; P = 0.005; 109 vs. 73; P = 0.02; 23 vs. 16; P = 0.006 and 357 vs. 170; P = 0.06) (Figure 2C). In blasts from a patient 5, ectopic SPARC expression did not significantly increase the colony-forming ability after first plating, but a significant increase in the number of colonies was observed after the secondary plating (mean, 80 vs. 3; P = 0.001; Figure 2D). For patients 6 and 7, we observed an increase in spontaneous apoptosis in AML blasts when SPARC endogenous expression was knocked down by siRNAs (Figure 2E), suggesting that SPARC promotes both growth and survival of leukemia cells.

vorable gene mutations (i.e., those in isocitrate dehydrogenase 2 [IDH2-R172]; ref. 26) or overexpressed genes (i.e., brain and acute leukemia, cytoplasmic [BAALC] and v-ets erythroblastosis virus E26 oncogene homolog [ERG]; ref. 27), and downregulated in GEP associated with the prognostically favorable nucleophosmin (NPM1) mutation (28) in cytogenetically normal AML (CN-AML). This led us to hypothesize that SPARC overexpression contributes to a more aggressive phenotype in AML. Thus, we dissected the clinical significance of SPARC overexpression in AML, the mechanisms by which this gene is deregulated, and the downstream effects of this aberrantly expressed gene. We show that SPARC overexpression independently predicts adverse outcome in CN-AML patients, thus representing what we believe to be a novel prognostic marker in AML. Consistent with these findings, we also demonstrate that SPARC contributes to leukemia growth in vitro and aggressive disease in vivo. SPARC overexpression activates integrin-linked kinase/AKT (ILK/AKT) and in turn β-catenin and could be targeted by modulating the SP1/NF-κB/miR-29b network, thereby also representing a potential therapeutic target in AML.

Results
SPARC overexpression is associated with adverse clinical outcome in CN-AML. SPARC expression was analyzed by nCounter assays (NanoString Technologies) (29) in 153 younger (age range, 18–59 years) adults with primary CN-AML treated with cytarabine-daunorubicin–based regimens; clinical and molecular characteristics are shown in Supplemental Table 1 (supplemental material available online with this article; doi:10.1172/JCI70921DS1).
To examine the contribution of SPARC to leukemia growth in vivo, NOD/SCID/gamma (NGS) mice were injected with THP-1/SPARC cells or THP-1/EV cells via tail vein. Eight weeks later, 3 mice in each group were sacrificed. Bone marrow engraftment of the leukemia cells was confirmed by CD45 antibody staining. The THP-1/SPARC mice had significantly larger livers ($P = 0.04$; Figure 3A) and spleens ($P = 0.008$; Figure 3B) than THP-1/EV controls. THP-1/SPARC cells formed coalescing masses obliterating large expanses of the liver parenchyma (Supplemental Figure 2), while THP-1/EV cells were observed either alone or in small clusters within hepatic sinusoids without effacing the hepatic cords.

The THP-1/SPARC cells had a monomorphic phenotype and a seemingly higher proliferative capacity compared with the more pleomorphic character and increased apoptotic tendency of the THP-1/EV cells (see immunohistochemistry (IHC) staining for SPARC, CD45, and Ki67 in Figure 3C and quantification of Ki67 in Figure 3D). Histopathology of spleen sections showed increased infiltration of leukemic cells and destruction of splenic architecture in tissues obtained from THP-1/SPARC mice compared with tissues obtained from THP-1/EV mice (Supplemental Figure 2). The THP-1/SPARC mice ($n = 9$) also had a significantly shorter survival than the THP-1/EV mice ($n = 9$) ($2 \times 10^6$ cells were injected

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**Figure 2**

SPARC promotes growth advantage in AML. (A) SPARC protein levels (right panel) and quantification of colony formation (left panel) in THP-1 cells ectopically expressing SPARC. (B) SPARC knockdown (right panel) and quantification of colony formation (left panel) in Kasumi-1 cells. (C) SPARC ectopic expression in primary blasts from 4 AML patients (numbers 1–4; bottom panel) and quantification of colony formation assays in these cells (top panel). (D) Western blotting showing SPARC protein levels (right panel) and quantification of colony formation in primary blasts from AML patient 5 ectopically expressing SPARC scored at 14 days following first and second plating (left panel). (E) SPARC protein levels measured in primary blasts from 2 AML patients (numbers 6–7) transfected with SPARC siRNA (right panel) and quantification of apoptosis determined by annexin V staining in SPARC-siRNAs versus control siRNA–transfected blasts (left panel). Data represent mean ± SEM.
per mouse; median survival, 49 vs. 66 days; \( P = 0.001 \); Figure 3E). Similar results were obtained in a repeated experiment where a higher number of cells \( (5 \times 10^6) \) were injected into each mouse (Supplemental Figure 3). Consistent with these results, mice injected with the SPARC-shRNA–infected CG-SH cells, which otherwise express high levels of endogenous SPARC, showed a trend for smaller spleens than mice injected with control shRNA–infected cells (Supplemental Figure 4).

**Figure 3**

SPARC promotes aggressive disease in vivo. NSG mice were engrafted with THP-1 cells infected with pLenti-EV or pLenti-SPARC vectors; mice were sacrificed and tissues were collected 8 weeks following engraftment. (A) Liver sizes (mean ± SEM). (B) Spleen sizes (mean ± SEM). (C) Immunohistochemical staining for SPARC, CD45, and Ki67 expression in liver tissues obtained from THP-1/EV and THP-1/SPARC mice. Original magnification, \( \times 400 \). (D) Quantitative analysis for Ki67 (proliferation marker) expression in liver tissues obtained from THP-1/EV and THP-1/SPARC mice (using ImmunoRatio Software). (E) Survival analysis of THP-1/SPARC mice \( (n = 9) \) compared with THP-1/EV controls \( (n = 9) \).
Figure 4
SPARC activates ILK/AKT/β-catenin signaling pathways in THP-1 cells and enhances β-catenin nuclear translocation and transcriptional activity. THP-1 cells were transfected with pLenti-EV or pLenti-SPARC vectors. (A) Western blot measuring secreted SPARC protein levels in media collected from cell culture of THP-1 cells infected with pLenti-SPARC vector compared with cells infected with pLenti-EV. (B) SPARC and integrin coimmunoprecipitation assay. (C) ILK kinase activity assay. Lysates from THP-1/SPARC and THP-1/EV cells were immunoprecipitated with anti-ILK antibody and assayed utilizing GSK3 fusion protein (ILK substrate). Numbers represent ratio of band intensity to control band. (D) Western blot analysis of P-ILK(Ser473) and P-GSK3β(Ser9) levels in THP-1/SPARC cells compared with THP-1/EV in the presence of endogenous ILK or following ILK knockdown or (E) following blocking integrin receptors with integrin antibodies. Lanes separated by a line were run on the same gel but were noncontiguous. (F) Western blot analysis of P-β-catenin (Ser552) in THP-1/SPARC cells compared with THP-1/EV cells. (G) Immunofluorescence confocal microscopy images showing β-catenin (green) and cell nuclei (blue) in EV- or SPARC-transfected THP-1 cells stained with β-catenin antibody and Draq5 antibody (nuclear staining); arrows indicate cells exhibiting β-catenin nuclear translocation. Scale bars: 10 μm. (H) Luciferase activity measured in 293T cells following cotransfection with pBART (TCF/LEF reporter vector) or pμBART (TCF/LEF reporter vector with mutated site) and pLenti-EV or pLenti-SPARC and (I) in the presence of control siRNAs or ILK siRNAs. (J) MYC mRNA expression (left panel) and protein expression (right panel) in THP-1/SPARC cells compared with THP-1/EV. Data represent mean ± SEM for 3 different experiments.

It has been reported that β-catenin is expressed and activated in LSCs and that it is required for self-renewal (37, 38). Thus, we postulated that SPARC expression may be higher in less differentiated hematopoietic cell subpopulations. To prove this, we showed that SPARC expression was significantly higher in the CD34+ compared with CD34− cells in normal bone marrow cells (NBM) (n = 4; P < 0.001; Figure 6A), and in CD34+/CD38−HSC/multipotent progenitors (HSC/MPP) compared with CD34+/CD38− cells (common-myeloid progenitors [CMP] and granulocyte-monocyte progenitors [GMP]) both in NBM (n = 3) and AML blasts (n = 3; patients n. 1, 5 and 9) (Figure 6, B and C). Furthermore, we showed that SPARC-transduced primary AML blasts (patient 9) indeed exhibited a higher percentage of engraftment in NSG mice than those transduced with GFP/EV control (3.1% vs 0.4% circulating blasts 12 weeks after blast i.v. injection; Figure 6D).

SP1, NF-kB, and miR-29b modulate SPARC expression in AML. Next, we asked which mechanisms lead to aberrant SPARC expression in AML. We identified binding sites for SP1, NF-kB, and RUNX1 in the (−651/1) promoter region (Transcriptional Regulatory Element Database [TRED] website; http://rulai.cshl.edu/cgi-bin/TRED/tred.cgi?process=home) and binding sites for miR-29 in the 3′ UTR region of the human SPARC gene. Indeed, SPARC has been reported to be a miR-29 target (44). Notably, we have recently shown that activation and overexpression of oncoproteins involved in AML (i.e., KIT, FLT3, and DNMT3) require interaction and transactivation of the transcription factors SP1 and NF-kB and inhibition of miR-29b (45). Therefore, we postulated that a similar mechanism could be responsible for SPARC overexpression.

To test this hypothesis, we first performed luciferase gene reporter assays by cloning the SPARC promoter region spanning the SP1-, NF-kB-, and RUNX1-binding sites into the pGL4.11 luciferase vector. Then, 293T cells were cotransfected with the reporter gene and vectors expressing SP1 and p65 (NF-kB subunit). A signifi-
cant increase in luciferase activity was found in cells transfected with SP1- or p65-expressing vectors compared with EV controls ($P < 0.001$ for both; Figure 7A). Ectopic expression of SP1 and p65 also resulted in significant increases in endogenous SPARC expression in THP-1 cells (Figure 7B and C), whereas SP1 and p65 siRNA knockdown resulted in significant decrease of SPARC expression in Kasumi-1 cells (Figure 7D and E).

A significant decrease of SPARC promoter luciferase activity was found in 293T cells transfected with RUNX1 expression vector ($P < 0.001$; Figure 7F). In contrast, RUNX1-WT knockdown resulted in SPARC upregulation in MV4-11 cells (Figure 7G), while knockdown of RUNX1-RUNX1T1 translocation by siRNAs resulted in a significant SPARC downregulation in RUNX1-RUNX1T1-positive Kasumi-1 cells (Figure 7H).

Notably, ectopic expression of p65 and SP1 increased their respective enrichment on the SPARC promoter (Figure 7I and J), and forced RUNX1-WT expression decreased SP1, but not p65, enrichment as measured by ChIP in THP-1 cells (Figure 7K).

These results are also consistent with the finding that RUNXI-mutated CN-AML patients exhibit higher SPARC expression when compared with the RUNXI-WT patients (Supplemental Figure 12). Together, these results indicate that SP1, p65 and RUNXI loss-of-function promoted SPARC expression, while RUNXI-WT inhibited SPARC expression.

As SPARC was found to be a target for miR-29a and miR-29c[44] and the SPARC 3′ UTR region was predicted to have binding sites for miR-29b[46, 47], we cloned the SPARC 3′ UTR region in luciferase reporter vector (pGL3) and cotransfected this construct with the synthetic microRNA (miR-29b and miR-9*) or scramble oligos in 293T cells. We found a 60% decrease of SPARC 3′ UTR–luciferase activity in cells cotransfected with miR-29b compared with the activity in cells cotransfected with scramble ($P < 0.001$) or an unrelated microRNA: miR-9* (Figure 8A). Kasumi-1 cells transfected with miR-29b showed significant decreases in endogenous SPARC (Figure 8B). AML blasts transfected with miR-29b also showed significant decreases in SPARC levels at 24 hours (Figure 8C).
We have previously shown that miR-29b is downregulated in AML via a SP1/NF-κB/HDAC inhibitory complex and that pharmacologic disruption of this complex results in miR-29b upregulation and downregulation of direct (i.e., SP1, DNMT3A, DNMT3B) and indirect (i.e., DNMT1, FLT3, KIT) miR-29b targets (45, 48, 49). Consistent with these reports, we showed that bortezomib at concentrations achievable in vivo (45, 50) significantly increased miR-29b expression (Figure 8D) and in turn decreased SP1 and SPARC mRNA and protein levels in Kasumi-1 cells (Figure 8, E and F) and in blasts from 3 different patients with primary AML (numbers 6, 10, and 7) (Figure 8, G and H). Consistent with the in vitro results, we found SPARC expression was significantly reduced in blood and spleen from CG-SH cell–engrafted NSG mice treated with bortezomib (1 mg/kg) twice a week for 4 weeks compared with vehicle-treated mice (Supplemental Figure 13).

Discussion

Although altered SPARC expression has been observed in cancer, the clinical significance of both upregulation and downregulation of the gene and the encoded protein remain to be fully elucidated (15, 17, 51–58). In contrast with solid tumors, the number of reports investigating this gene in hematologic malignancies is relatively small. In MDS, 5q deletion results in allelic loss and significant downregulation of SPARC expression in the progenitor compartment (22, 23), and a dramatic upregulation of SPARC occurs in response to lenalidomide (23). Although this suggests that SPARC expression is a treatment-response predictor and perhaps plays a role in normal hematopoiesis, to date, no biological evidence supporting this notion has been reported. Indeed, SPARC was found to be dispensable for murine hematopoiesis (59). Nevertheless, the complexity of the hematopoietic role of SPARC is illustrated by a recent report (60). These authors showed in a myeloproliferative murine model that increased SPARC expression in the bone marrow stroma favored fibrotic changes, while loss of SPARC expression resulted in a defective stromal niche (60).

Here, we sought to dissect the role of SPARC in AML, starting from the observation that this gene was found overexpressed in GEPs associated with distinct molecular subsets of patients harboring prognostically unfavorable mutations or overexpressed genes. Supporting the hypothesis that SPARC overexpression likely contributes to aggressive myeloid leukemogenesis, we showed that high SPARC expression was associated with adverse outcomes in CN-AML patients. Although SPARC overexpression was associated with other unfavorable molecular markers at diagnosis, in multivariable models for outcome, the impact of SPARC remained significant even after adjusting for other molecular prognosticators, supporting an independent role of this gene in determining an aggressive clinical phenotype. To our knowledge, this is the first study reporting the clinical relevance of SPARC expression in AML. We validated these findings through mechanistic experiments using AML preclinical models and primary blasts. We showed that SPARC upregulation promoted an aggressive phenotype in AML cells. Mice engrafted with AML cells forced to express SPARC had increased leukemia burden and shorter survival compared with controls.

The mechanisms leading to SPARC upregulation in AML blasts are multifactorial and likely converge to NF-κB–dependent pathways, which have been previously shown to be constitutively activated in AML LSCs (61). We found that SPARC may function through an autocrine mechanism; once secreted by AML cells,
A finding in our study was that SPARC enhanced β-catenin activity in AML cells, known to be required for leukemia growth and stemness. The increase in β-catenin activity was seemingly enhanced via the SPARC-integrin-ILK axis. ILK is a kinase that links the cell adhesion receptors, integrins, and growth factors with the downstream signaling pathways involving AKT and β-catenin.
oncogenes, constitutive SPARC expression was dependent on a previously reported regulatory molecular network that involves SP1/NF-κB and miR-29b (45). The leukemogenic role of this network that can be targeted pharmacologically has been extensively studied by our group not only in AML (45) but also in other types of leukemia (45, 62). Our data support the view that SPARC overexpression is a potentially actionable therapeutic target via pharmacological disruption of the SP1/NF-κB/miR-29b network by, for example, bortezomib, used here as a proof of concept. Other compounds, including inhibitors of HDAC (e.g., vorinostat, AR-42) or SP1 (e.g., mithramycin A) or synthetic miR-29b mimics may also interfere with the SP1/NF-κB/miR-29b network and downregulate SPARC (45, 50, 65, 66).

Figure 8
Targeting SPARC expression in AML. (A) SPARC 3′ UTR–luciferase activity in the presence of miR-29b was measured by cotransfecting 293T cells with SPARC–3′ UTR reporter and a synthetic microRNA; values were normalized to those of cells transfected with scramble. (B) Ectopic expression of synthetic miR-29b significantly decreases SPARC mRNA expression in Kasumi-1 cells. (C) SPARC protein in AML blast samples (patients 6 and 10) at 24 hours following transfection with miR-29b. (D) miR-29b levels, (E) mRNA levels of SPARC and SP1, and (F) Western blot analysis showing SPARC, SP1, and MYC protein levels, measured in Kasumi-1 cells treated with 20, 60, and 100 nM of bortezomib. (G) SPARC mRNA and (H) protein levels in blasts from 3 primary AML samples (patients 6, 10, and 7) treated with 100 nM bortezomib and assessed 24 hours later.

GSK3β (43). This ultimately results in stabilization and nuclear localization of β-catenin and activation of genes promoting cell-cycle progression and proliferation (63). Consistent with these results, we observed that SPARC levels were higher in both normal and immature AML cell subpopulations that indeed displayed a higher engraftment potential in immunodeficient mice. While SPARC/ILK/AKT signaling was found involved in glioma cell invasion and survival (64), to our knowledge, the relevance of this mechanism in AML has not been previously reported.

Having observed that SPARC overexpression contributes to more aggressive phenotype in AML patients, we asked whether it is possible to target SPARC expression and demonstrate the clinical relevance of our findings. We showed that, similarly to other
While our findings emphasize a role of SPARC in the homeostasis of myeloblasts, several other reports have underscored the interplay of SPARC protein with the microenvironment to create a niche favorable for cancer growth (8). Thus, future studies of SPARC in AML need to address the dual intracellular and extracellular role of this protein.

In conclusion, SPARC deregulation is clinically relevant in AML because overexpression of this gene independently predicts adverse outcome in subsets of AML patients, contributes to aggressive AML growth likely via β-catenin activation, and is an actionable therapeutic target. This suggests that future clinical studies targeting the activity of SPARC protein are warranted in AML.

Methods

Plasmids and reagents. SPARC clone was purchased from Invitrogen and inserted in pcDNA3.1/V5-DEST (Invitrogen) and MIGR (Addgene). pCMV-p65, pSUPER-SP1, and pCMV-RUNX1 expression vectors were also used (45). OFF-TARGET control and ON-TARGET plus siRNA-SMARTpool reagents against SPARC, SP1, p65, and RUNX1 (each contains a mix of at least 4 different sequences) were purchased from Dharmacon. siRNA against RUNX1-RUNX1T1 was custom designed (Dharmacon). SPARC shRNA (a set of 6 different sequences) was purchased from Thermo Fisher Scientific.

Cell lines and primary blasts. THP-1, MV4-11, Kasumi-1, and CG-SH cells were cultured in RPMI medium supplemented with 10%–20% FBS. Blasts from AML patients were maintained in RPMI medium supplemented with 20% FBS and 1x StemSpan CC100 (StemCell Technologies). AML blasts used in the experiments were obtained from apheresis blood samples collected from patients treated at OSU and stored in the OSU Leukemia Tissue Bank.

Transient transfection, RNA interference, and viral induction. Transient transfection of cells was performed utilizing 1 to 2 μg of plasmid or 1 nmol of siRNA per reaction and Nucleofector Kit (Amaza) according to the manufacturer’s instructions (67). Retroviral and lentiviral infection were performed with an MOI of 3 and greater than 10, respectively, to obtain efficiencies that approximated 40%, as previously reported (68, 69). Cells were selected by antibiotic selection with blasticidin or sorting for GFP-positive cells.

RNA extraction and RNA expression quantification. Total RNA was extracted using Trizol reagent (Invitrogen). SPARC expression in CN-AML patients was measured by NanoString nCounter system according to the manufacturer’s instructions (Nano-String). Gene cDNA was synthesized using SuperScript III reagents (Invitrogen) and the TaqMan MicroRNA Reverse Transcription Kit (Applied Biosystems) according to the manufacturer’s instructions. Quantitative real-time RT-PCR (qRT-PCR) was performed using commercially available TaqMan Gene Expression Assay primers and probes and the 7900HT Fast Real-Time PCR System (Applied Biosystems). The expression levels were normalized to 18S for gene or U44 for microRNA expression.

Immunoprecipitation, Western blot analysis, CHIP, and antibodies. Immunoprecipitation and Western blot were performed as previously described (70). ChIP assays were performed using the EZ ChIP Kit (Millipore) according to the manufacturer’s protocol. DNA was quantified using qRT-PCR with SYBR green incorporation (Applied Biosystems) and primers specific to the promoter and normalized to their input. Antibodies and primers are listed in Supplemental Experimental Procedures.

Clonogenic and viability analyses. Methocellulose clonogenic assays were carried out by plating 1 × 10⁴ cells of different cell lines or 2 × 10⁴ primary blasts in 0.9% MethoCult (StemCell Technologies) (68). Colony numbers were scored 10 days later.

For viability and apoptosis analysis, cells were washed with PBS and resuspended in 50 μl binding buffer containing 2 μl of annexin V (eBioscience) and 5 μl propidium iodide (PI) (eBioscience). After 20 minutes incubation, fluorescence was quantified by flow cytometry on a FACSCalibur instrument.

Immunofluorescent staining, cell sorting, flow cytometry and confocal microscopy. Cells were washed and stained with CD34, CD38, CD45RA, and CD123 antibodies (eBioscience) and sorted into hematopoietic stem cells and HSC/MP/MPP (CD34+/CD38⁻), CMP (CD34+/CD45RA⁻/CD123⁻) and GMP (CD34+/CD45RA⁺/CD123⁺) using BD FACSAriaII cell sorter (BD Biosciences).

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59. Siva K, Jaako P, Miharada K. SPARC is dispensable among its ILK.


