

***Smap1* deficiency perturbs receptor trafficking and predisposes mice to myelodysplasia**

Shunsuke Kon, Naoko Minegishi, Kenji Tanabe, Toshio Watanabe, Tomo Funaki, Won Fen Wong, Daisuke Sakamoto, Yudai Higuchi, Hiroshi Kiyonari, Katsutoshi Asano, Yoichiro Iwakura, Manabu Fukumoto, Motomi Osato, Masashi Sanada, Seishi Ogawa, Takuro Nakamura, Masanobu Satake

J Clin Invest. 2013;123(3):1123-1137. <https://doi.org/10.1172/JCI63711>.

Research Article **Hematology**

The formation of clathrin-coated vesicles is essential for intracellular membrane trafficking between subcellular compartments and is triggered by the ARF family of small GTPases. We previously identified SMAP1 as an ARF6 GTPase-activating protein that functions in clathrin-dependent endocytosis. Because abnormalities in clathrin-dependent trafficking are often associated with oncogenesis, we targeted *Smap1* in mice to examine its physiological and pathological significance. *Smap1*-deficient mice exhibited healthy growth, but their erythroblasts showed enhanced transferrin endocytosis. In mast cells cultured in SCF, *Smap1* deficiency did not affect the internalization of c-KIT but impaired the sorting of internalized c-KIT from multivesicular bodies to lysosomes, resulting in intracellular accumulation of undegraded c-KIT that was accompanied by enhanced activation of ERK and increased cell growth. Interestingly, approximately 50% of aged *Smap1*-deficient mice developed anemia associated with morphologically dysplastic cells of erythroid-myeloid lineage, which are hematological abnormalities similar to myelodysplastic syndrome (MDS) in humans. Furthermore, some *Smap1*-deficient mice developed acute myeloid leukemia (AML) of various subtypes. Collectively, to our knowledge these results provide the first evidence in a mouse model that the deregulation of clathrin-dependent membrane trafficking may be involved in the development of MDS and subsequent AML.

Find the latest version:

<https://jci.me/63711/pdf>





Smap1 deficiency perturbs receptor trafficking and predisposes mice to myelodysplasia

Shunsuke Kon,¹ Naoko Minegishi,² Kenji Tanabe,³ Toshio Watanabe,¹ Tomo Funaki,¹ Won Fen Wong,¹ Daisuke Sakamoto,¹ Yudai Higuchi,¹ Hiroshi Kiyonari,⁴ Katsutoshi Asano,⁵ Yoichiro Iwakura,⁶ Manabu Fukumoto,⁷ Motomi Osato,⁸ Masashi Sanada,⁹ Seishi Ogawa,⁹ Takuro Nakamura,¹⁰ and Masanobu Satake¹

¹Department of Molecular Immunology, Institute of Development, Aging and Cancer, and ²Tohoku Medical Megabank Organization, Tohoku University, Sendai, Japan. ³Medical Research Institute, Tokyo Women's Medical University, Tokyo, Japan. ⁴Laboratory for Animal Resources and Genetic Engineering, RIKEN Center for Developmental Biology, Kobe, Japan. ⁵Nihon Gene Research Laboratories, Sendai, Japan. ⁶Institute of Medical Science, The University of Tokyo, Tokyo, Japan. ⁷Department of Pathology, Institute of Development, Aging and Cancer, Tohoku University, Sendai, Japan. ⁸Cancer Science Institute, National University of Singapore, Singapore. ⁹Cancer Genomics Project, Faculty of Medicine, The University of Tokyo, Tokyo, Japan. ¹⁰Division of Carcinogenesis, The Cancer Institute, Japanese Foundation for Cancer Research, Tokyo, Japan.

The formation of clathrin-coated vesicles is essential for intracellular membrane trafficking between subcellular compartments and is triggered by the ARF family of small GTPases. We previously identified SMAP1 as an ARF6 GTPase-activating protein that functions in clathrin-dependent endocytosis. Because abnormalities in clathrin-dependent trafficking are often associated with oncogenesis, we targeted *Smap1* in mice to examine its physiological and pathological significance. *Smap1*-deficient mice exhibited healthy growth, but their erythroblasts showed enhanced transferrin endocytosis. In mast cells cultured in SCF, *Smap1* deficiency did not affect the internalization of c-KIT but impaired the sorting of internalized c-KIT from multivesicular bodies to lysosomes, resulting in intracellular accumulation of undegraded c-KIT that was accompanied by enhanced activation of ERK and increased cell growth. Interestingly, approximately 50% of aged *Smap1*-deficient mice developed anemia associated with morphologically dysplastic cells of erythroid-myeloid lineage, which are hematological abnormalities similar to myelodysplastic syndrome (MDS) in humans. Furthermore, some *Smap1*-deficient mice developed acute myeloid leukemia (AML) of various subtypes. Collectively, to our knowledge these results provide the first evidence in a mouse model that the deregulation of clathrin-dependent membrane trafficking may be involved in the development of MDS and subsequent AML.

Introduction

Intracellular and extracellular homeostasis is maintained by a vesicle transport system that mediates the trafficking of membrane proteins to appropriate organelles. Clathrin-coated vesicles are formed at donor membrane sites in a highly ordered manner, and a number of molecules are involved in this process. Among them, small GTPases of the ARF family play a central role in vesicle formation. An ARF molecule cycles between two conformations, an active GTP-bound form and an inactive GDP-bound form. This cycling is mediated by a guanine nucleotide exchange factor that replaces GDP with GTP and a GTPase-activating protein (GAP) that hydrolyzes GTP to GDP and converts ARF into its inactive form. There are 6 ARFs (ARF1–ARF6) and several ARF-related proteins in mammals (1, 2). ARF6 is an isoform that localizes mainly to the plasma membrane and functions in the endocytosis and recycling of vesicles as well as in actin rearrangement and lipid metabolism (3, 4).

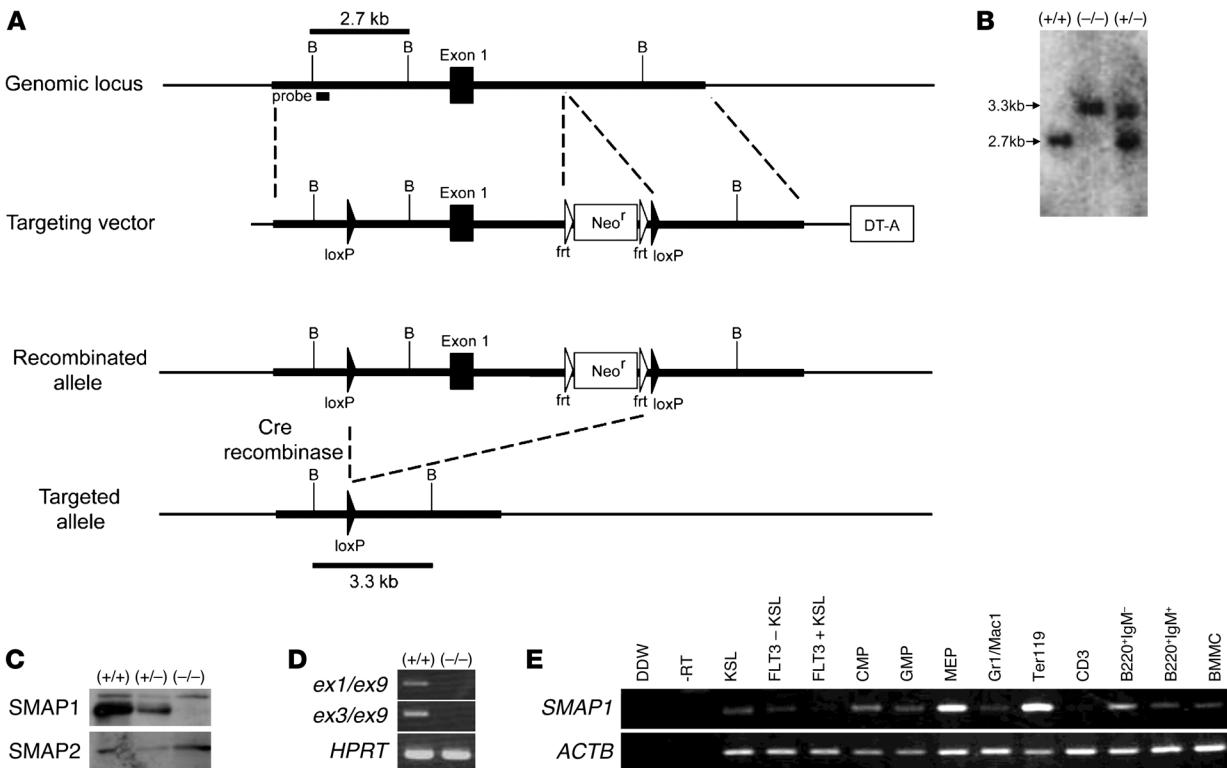
We previously demonstrated that small ARF GAP1 (referred to as SMAP1) is a regulator of clathrin-dependent endocytosis, based on a series of observations (5, 6). First, SMAP1 exhibits GAP activity against ARF6, as assessed by an in vitro GAP assay. Second, SMAP1 localizes to juxta-plasma membrane regions in which ARF6 also exists. Third, SMAP1 binds to the clathrin heavy chain directly via its clathrin-binding box. Fourth, overexpression of SMAP1 abrogates clathrin-dependent internalization of the transferrin receptor and E-cadherin.

Conflict of interest: The authors have declared that no conflict of interest exists.

Citation for this article: *J Clin Invest.* 2013;123(3):1123–1137. doi:10.1172/JCI63711.

Recently, mutations and chromosomal translocations associated with various human cancers and leukemia have been identified in the genes that encode endocytosis-related proteins (7–10). However, the precise molecular mechanisms that underlie the effect of these genetic alterations on membrane trafficking and lead to disorders in cell growth and/or differentiation remain poorly understood. Therefore, the significance of these mutations needs to be clarified. One process that could link membrane traffic to alterations in cell growth/differentiation is the deregulation of receptor tyrosine kinase (RTK) downregulation. Alterations in the endocytosis and/or lysosomal degradation of RTKs result in the persistence of these molecules on the membrane, which leads to the activation of growth and differentiation pathways (11–13).

Several studies have reported the involvement of *SMAP1* in oncogenesis in humans. For example, the *MLL* gene is a frequent target for recurrent chromosomal translocations in acute myeloid leukemia (AML), and more than 50 *MLL* fusion partners have been identified, including endocytosis-related genes, such as *EPS15*, *CALM*, and *EEN* (10). Interestingly, *SMAP1* was previously identified as one of the fusion partners of *MLL* (14). In colorectal cancers displaying microsatellite instability, mutations causing the truncation of the polypeptide chain have been detected in *SMAP1* (11% homozygous and 73% heterozygous) (15). This finding suggests that *SMAP1* may be acting as a tumor suppressor gene in intestinal cells. Based on these findings, we generated *Smap1*-targeted mice to examine the function of SMAP1 in clathrin-dependent vesicle trafficking and to determine the potential role of SMAP1 in cell growth and differentiation *in vivo*.

**Figure 1**

Establishment of *Smap1*-targeted mice and SMAP1 expression. (A) Physical maps of the *SMAP1* gene locus and its targeting vector. Features of the recombined and targeted *SMAP1* alleles are also depicted. Horizontal lines indicate the genomic sequences. The thick lines indicate the sequence incorporated into the targeting vector. Exon 1, neomycin resistance gene, and diphtheria toxin subunit A gene are indicated. Black and white arrowheads indicate the loxP and frt sequences, respectively. The small rectangle under the line corresponds to the probe that was used for Southern blot hybridization. B indicates a *Bam*H I restriction site. (B) Southern blot analysis of genomic DNA prepared from *Smap1*^{+/+} (+/), *Smap1*^{+/-} (+/-), and *Smap1*^{-/-} (-/-) mice. DNA was digested by *Bam*H I and processed for Southern blotting using the hybridization probe shown in A. The wild-type and targeted alleles give rise to 2.7-kb and 3.3-kb bands, respectively. (C) Immunoblot analysis of protein lysates prepared from bone marrow cells of *Smap1*^{+/+}, *Smap1*^{+/-}, and *Smap1*^{-/-} genotypes. The 50-kDa band represents SMAP1, whereas SMAP2 served as a control. Three independent experiments were performed, and one representative reproducible result is shown. (D) RT-PCR analyses of *Smap1* transcripts in bone marrow cells from *Smap1*^{+/+} and *Smap1*^{-/-} mice. Primers were set between exons 1 or 3 and exon 9. *HPRT* served as a control. (E) *SMAP1* expression in hematopoietic cells. Fractions of various hematopoietic lineages were sorted from bone marrow cells of wild-type mice by flow cytometry, and RNA was prepared from each and processed for semiquantitative RT-PCR analyses. DDW, distilled deionized water; -RT, without reverse transcription.

Results

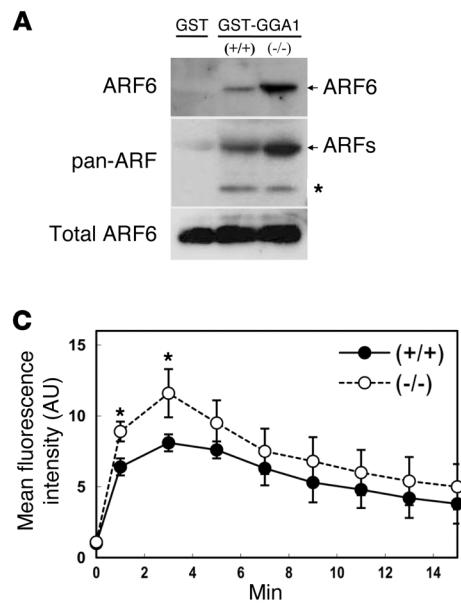
Establishment of *Smap1*-targeted mice and SMAP1 expression. The functions of SMAP1 in vivo were analyzed using a gene targeting approach. Figure 1A illustrates the genomic structure of *SMAP1* around exon 1 and the configuration of the targeting vector. Exon 1 was chosen as the targeting site because it harbors the SMAP1-initiating methionine codon. Two independent *Smap1*^{-/-} mouse lines (44 and 64) were established and crossed to each other to generate *Smap1*^{-/-} mice. Genomic DNA was processed for Southern blot analysis (Figure 1B). Based on the size of the detected bands, mouse genotypes were determined as wild-type, heterozygous, or homozygous targeting.

To confirm the expression of SMAP1, protein extracts from bone marrow cells were analyzed by immunoblotting (Figure 1C). SMAP1 was detected in wild-type cells but was substantially reduced in heterozygous cells and not detected in homozygous targeted cells. SMAP2, a homolog of SMAP1 (16), was detected in equal amounts in the 3 cell types. RT-PCR analyses did not detect *SMAP1* transcripts spanning exons 1 or 3 through to exon 9 in the *Smap1*^{-/-} cells (Figure 1D).

Thus, homozygous targeting was confirmed to correspond to a *Smap1*-deficient status. *Smap1*^{-/-} mice exhibited no particular abnormality, and *Smap1*^{-/-} mice also grew to adulthood and were apparently healthy. Both male and female *Smap1*^{-/-} mice were fertile, and pups were born following the Mendelian ratio of inheritance.

SMAP1 expression was examined in various hematopoietic lineages isolated from the bone marrow of wild-type mice, as this information is relevant to the phenotypes of targeted mice, as described below. RT-PCR analyses (Figure 1E) showed that a substantial amount of SMAP1 transcript was detected in the MEP (megakaryo/erythroid progenitor) and Ter119⁺ fractions, whereas a lower amount was detected in the remaining fractions. This indicates that SMAP1 is expressed abundantly in the erythroid lineage but is also distributed broadly in the other hematopoietic lineages, including progenitors.

Endocytosis of transferrin is enhanced in *Smap1*-targeted cells. The identification of SMAP1 as an ARF6 GAP was based on the effects of SMAP1 overexpression on the endocytosis of the transferrin receptor using tissue culture cells (5, 6). Here, we examined whether


Figure 2

ARF6 activation and transferrin endocytosis in bone marrow cells. **(A)** Protein lysates were prepared from *Smap1*^{+/+} and *Smap1*^{−/−} bone marrow cells and incubated with GST or GST-GGA1 coupled to glutathione-Sepharose. The bound fraction was processed for immunoblot detection by anti-ARF6-specific and anti-panARF antibodies, as indicated. An asterisk represents nonspecific bands. The amounts of ARF6 in each lysate prior to incubation with GST or GST-GGA1 were also evaluated by immunoblotting (see “Total ARF6”). **(B and C)** Bone marrow cells were prepared from *Smap1*^{+/+} and *Smap1*^{−/−} mice and labeled with fluorescein-transferrin at 4°C. Excessive transferrin in the medium was washed away (initially bound transferrin at this time is shown as “4°C” as indicated in the top left of **B**), and, after incubation of cells at 37°C for the indicated time, surface-remaining transferrin was stripped off. Cells were labeled with PE-anti-Ter119 and processed for flow cytometry. The Ter119⁺ fraction was gated, and the transferrin-derived fluorescence intensities are displayed. Relative amounts of internalized fluorescein were measured by comparing fluorescence intensities at 0 minutes and each given time. Cells were prepared from 3 independent pairs of *Smap1*^{+/+} and *Smap1*^{−/−} mice and processed for assays. Averages \pm SD of internalized transferrin were calculated for each incubation time at 37°C ($n = 3$). * $P < 0.05$.

SMAP1 functions as an ARF6 GAP in mouse tissues. Figure 2A shows the incubation of protein lysates from bone marrow cells with the GST-GGA1 fusion protein. GGA1 is a clathrin-adaptor protein that binds GTP-bound forms but not GDP-bound forms of ARF (17). The GST-GGA1-bound fraction was processed for immunoblot analysis using anti-ARF6 or anti-panARF antibodies, which showed that the amount of GGA1-bound ARF6 was increased by 3.1 fold in *Smap1*^{−/−} cells as compared with that in wild-type cells. Expression of ARF6 itself was not affected by SMAP1 targeting, as shown by the immunoblotting of lysates prior to the application of GST-GGA1. These results indicate that SMAP1 indeed functions as an ARF6 GAP in vivo.

Erythroblasts, which show highly active transferrin endocytosis, were used to examine the effect of SMAP1 targeting on ARF6-regulated endocytosis. Bone marrow cells were incubated with transferrin at 4°C, excessive transferrin was washed away, and the cells were incubated at 37°C for various time periods. Then, the remaining surface-bound transferrin was stripped off, leaving only the intracellularly incorporated molecules intact. Figure 2B depicts a time course of transferrin fluorescence intensity that was obtained by gating the Ter119⁺ erythroid cell fraction. The fluorescent intensities were quantified and are shown in Figure 2C. Although

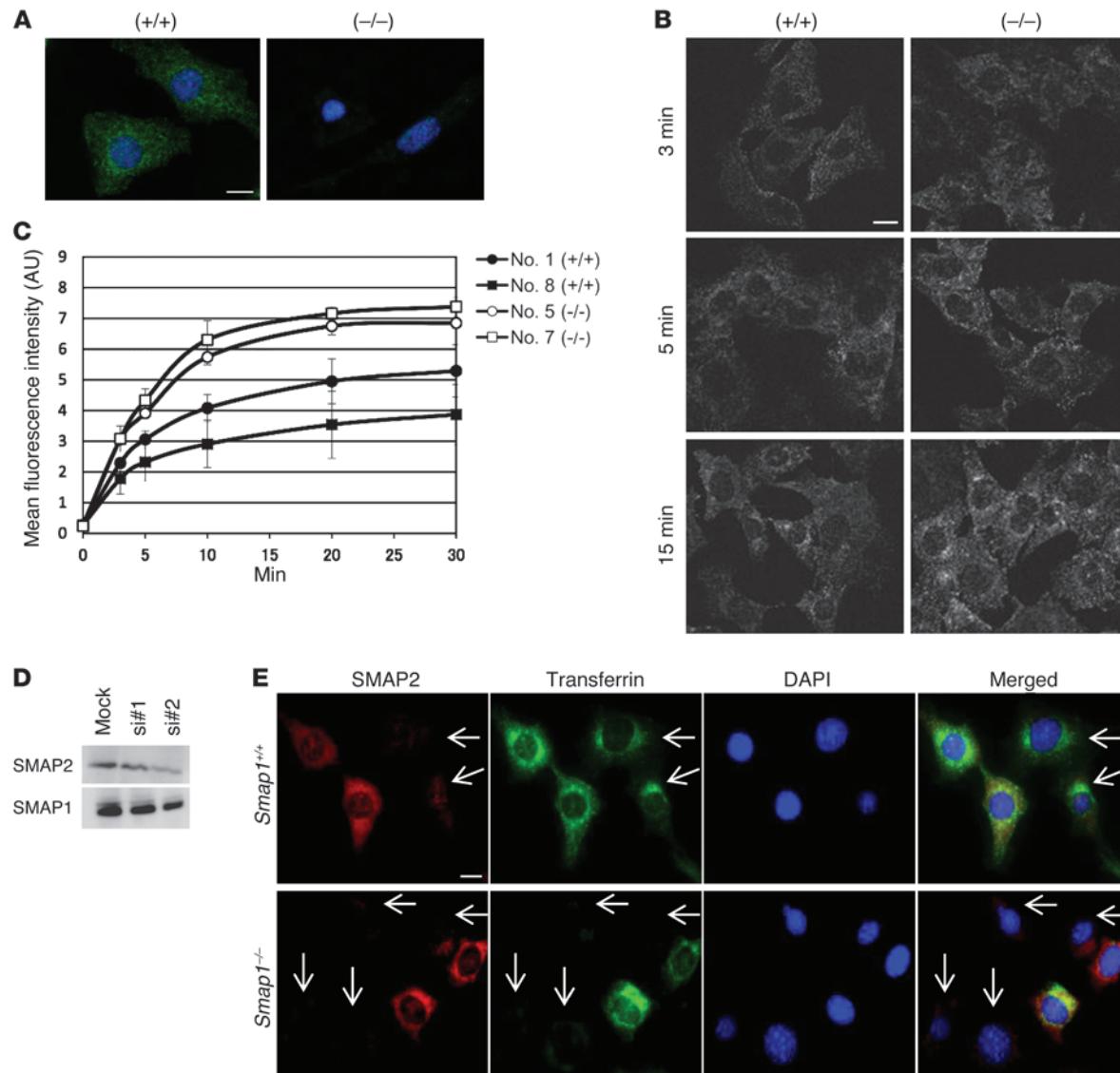
no differences were detected during the recycling phase (after 5 minutes), a significant increase in the amount of transferrin was incorporated into *Smap1*^{−/−} cells compared with wild-type cells during the initial uptake at 1 and 3 minutes. Notably, prior to the incubation at 37°C, amounts of the initially cell surface-bound transferrin at 4°C were similar between the 2 genotypes of Ter119⁺ bone marrow cells (Figure 2B, top left).

Transferrin endocytosis in Smap1-targeted cells is mediated by SMAP2. The effect of SMAP1 deficiency on transferrin endocytosis was investigated in cells of different lineages. Two independent wild-type and *Smap1*^{−/−} mouse embryo fibroblast (MEF) cultures were established. Figure 3A shows the immunofluorescence of endogenous SMAP1 on the cell surface and, as multiple dots in the cytoplasm, of wild-type cells but not targeted cells.

MEFs were incubated with transferrin for various time periods and then washed and fixed. Figure 3B depicts the fluorescence signals derived from internalized transferrin and shows that the intensity of fluorescence is stronger in *Smap1*^{−/−} cells than in wild-type cells. MEFs were recovered as a suspension and processed for flow cytometry. Figure 3C shows the gradual accumulation of transferrin in the cytoplasm. Transferrin accumulation was 1.3- to 1.9-fold more effective in the targeted cells as compared with that in the wild-type cells. When endocytosis and recycling were separately assayed using biotinylated transferrin, internalization was enhanced but

recycling was not affected in *Smap1*^{−/−} cells as compared with that in wild-type cells (Supplemental Figure 1; supplemental material available online with this article; doi:10.1172/JCI63711DS1), indicating that the enhanced accumulation of transferrin in *Smap1*^{−/−} MEFs (Figure 3C) is likely due to the enhanced incorporation of molecules (Supplemental Figure 1). Note that the fluorescence intensity of transferrin initially bound to the cell surface was similar in wild-type and *Smap1*^{−/−} MEFs (Supplemental Figure 1A).

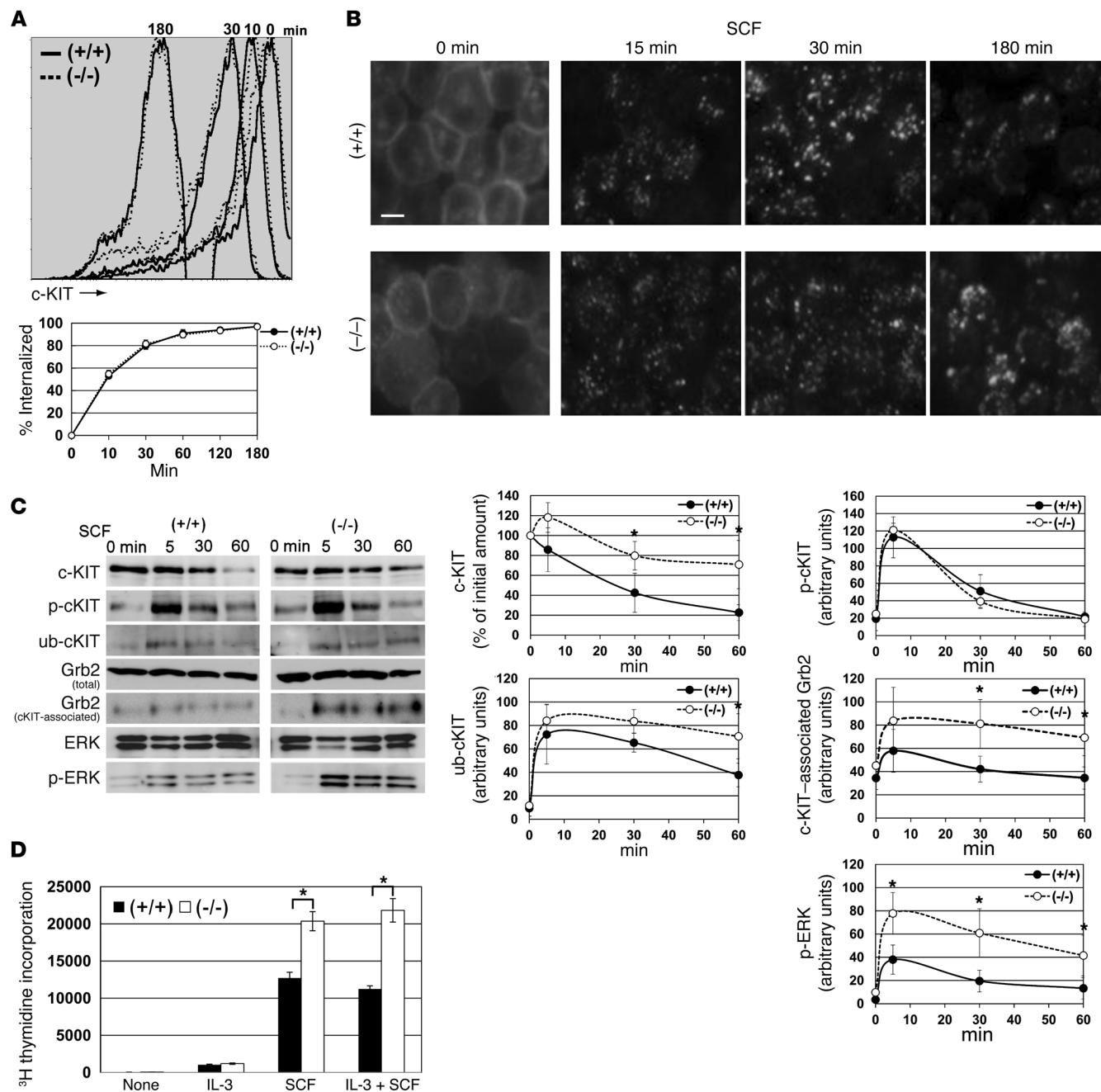
We then examined why transferrin endocytosis was not abrogated in *Smap1*^{−/−} cells. The effectiveness of siRNAs against SMAP2 was tested using wild-type MEFs (Figure 3D), and immunoblot analysis showed that siRNA2 worked more efficiently. Figure 3E shows the internalization of transferrin and SMAP2 levels in wild-type and *Smap1*^{−/−} MEFs after siRNA2-mediated silencing of SMAP2. Interestingly, the effects of siRNA2 appeared random and differed among cells, because endogenous SMAP2 remained intact in some cells, whereas it was almost abolished in other cells. Under these conditions, and in the case of *Smap1*^{+/+} MEFs, transferrin was equally incorporated regardless of the levels of SMAP2. In contrast, in *Smap1*^{−/−} MEFs, transferrin was not incorporated in SMAP2-silenced cells. These results suggest that SMAP2 likely compensates for the lack of SMAP1 in *Smap1*^{−/−} MEFs.

**Figure 3**

Transferrin transport in MEFs. **(A)** Immunofluorescence detection of endogenous SMAP1 in wild-type and *Smap1*^{-/-} MEFs using an anti-SMAP1 antibody (green). Blue indicates DAPI staining. **(B and C)** Internalization of transferrin in wild-type and *Smap1*^{-/-} MEFs. Cells were incubated with fluorescein-transferrin for the indicated times, and then surface-remaining transferrin was stripped off. The cells were then processed for analyses by **(B)** fluorescence microscopy or **(C)** flow cytometry. In **(C)**, the intensities of intracytoplasmic fluorescence were measured and expressed in relative arbitrary units. Independent cultures were prepared in triplicate from the indicated MEF clones, and averages \pm SD are shown ($n = 3$). “No. 1,” “No. 8,” “No. 5,” and “No. 7” refer to the MEF cell line numbers. **(D)** Effects of siRNA against SMAP2. Wild-type MEFs were treated with or without siRNA against SMAP2 (2 differentially designed siRNAs, siRNA1 and siRNA2, were used). Protein lysates were prepared and processed for immunoblot analyses using anti-SMAP2 and anti-SMAP1 antibodies. **(E)** Effects of SMAP2 knockdown on transferrin incorporation. The *Smap1*^{+/+} and *Smap1*^{-/-} MEFs were incubated with siRNA2 against SMAP2 and then with fluorescent transferrin and processed for immunofluorescence detection using an anti-SMAP2 antibody. The arrows indicate the reduction in fluorescence intensity from SMAP2, whereas DAPI staining indicates the location of cell nuclei. Scale bar: 10 μ m.

*Accumulation and enhanced signaling of c-KIT in *Smap1*^{-/-} cells.* c-KIT is highly expressed in hematopoietic progenitors and mast cells and is internalized through clathrin-coated vesicles. Because SMAP1 was detected in both types of cells (Figure 1E), the effects of SMAP1 targeting on c-KIT internalization were examined. Bone marrow-derived mast cells (BMMCs) were prepared and incubated with stem cell factor (SCF), and cell surface-located

c-KIT was measured by flow cytometry (Figure 4A). Cycloheximide was added to prevent the de novo synthesis of c-KIT, thereby preventing its expression on the cell surface. The top panel of Figure 4A shows the fluorescence intensity of cell surface c-KIT, as detected by anti-c-KIT, and the bottom panel of Figure 4A shows the percentage of internalized c-KIT. No difference was detected between the 2 genotypes, indicating that the SCF-induced endo-


Figure 4

Transport kinetics and c-KIT signaling in BMMCs. **(A)** Endocytosis of c-KIT. *Smap1*^{+/+} and *Smap1*^{-/-} BMMCs were cultured, starved in the presence of cycloheximide, incubated with SCF at 37°C for the indicated times, and processed for flow cytometry analyses. The top panel displays the fluorescence intensity of c-KIT and cell numbers, whereas the bottom panel plots the percentages of internalized c-KIT calculated by considering the initial surface fluorescence to be 100%. BMMCs were prepared from 3 independent pairs of *Smap1*^{+/+} and *Smap1*^{-/-} mice and processed for assays. Averages \pm SD of internalized c-KIT were calculated for each incubation time ($n = 3$). **(B)** Immunofluorescence detection of c-KIT in BMMCs. The *Smap1*^{+/+} and *Smap1*^{-/-} cells were incubated in the presence of SCF for the indicated times and stained for c-KIT. Scale bar: 10 μ m. **(C)** Activation status of c-KIT signaling molecules. Wild-type and *Smap1*^{-/-} BMMCs were incubated with SCF for the indicated times, and protein lysates were prepared and processed for immunoprecipitation/immunoblot analyses. Band densities were quantified, and averages \pm SD are shown ($n = 3$). p-cKIT, phosphorylated form of c-KIT; p-ERK1/2, phosphorylated form of ERK1/2; ub-cKIT, ubiquitinylated c-KIT; c-KIT-associated Grb2, Grb2 recruited into anti-c-KIT immunoprecipitates. **(D)** DNA synthesis in BMMCs. Triplicate cultures of cells were prepared from each of the wild-type and *Smap1*^{-/-} mice, incubated in the presence of IL-3 and/or SCF for 16 hours, and then treated with ³H-thymidine for 8 hours. The incorporation of ³H-thymidine into acid-insoluble fractions was measured, and averages \pm SD are shown ($n = 3$). * $P < 0.05$.

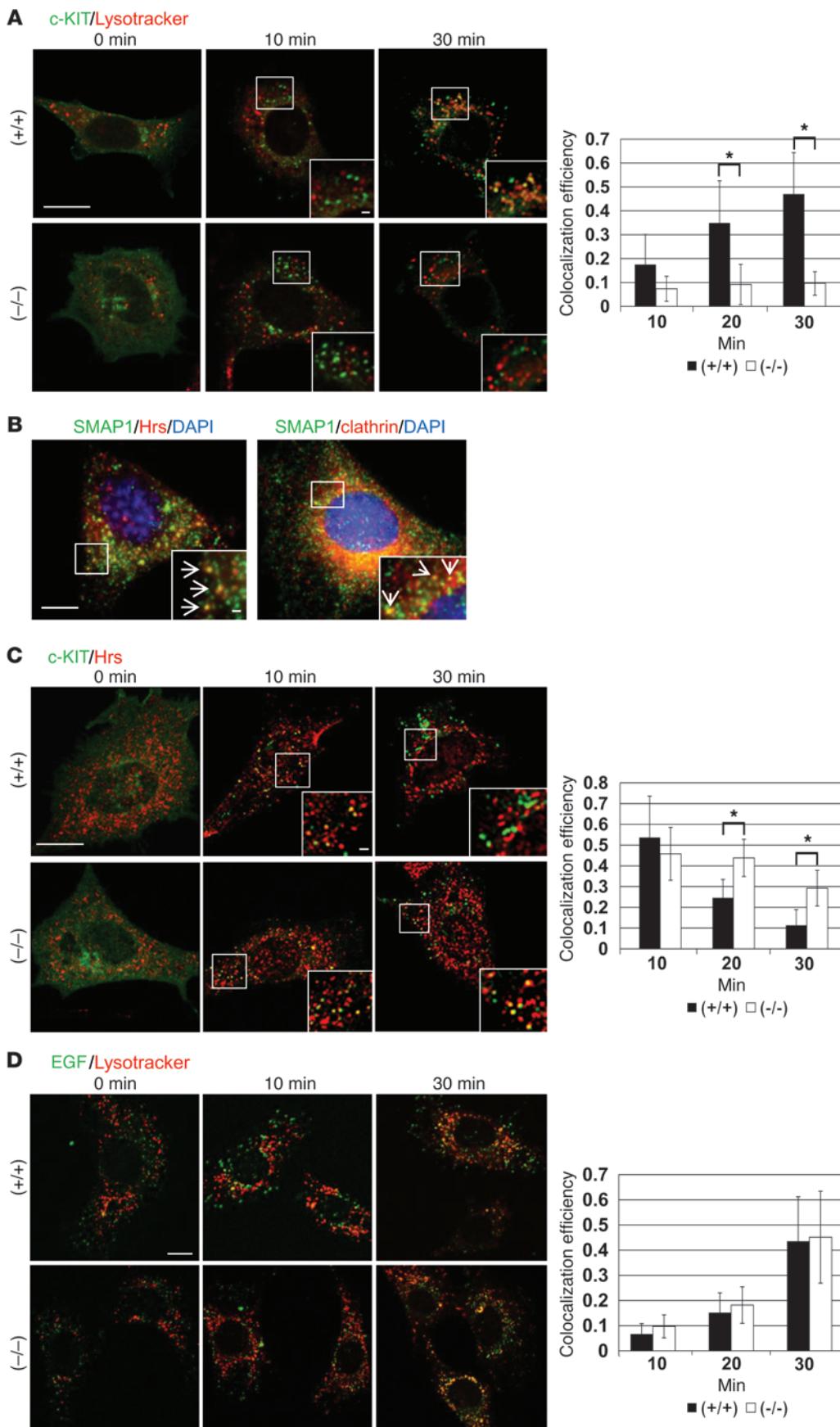


Figure 5

Transport of c-KIT and EGFR in MEFs. **(A and C)** Wild-type and *Smap1*^{-/-} MEFs were transfected by EYFP-c-KIT, incubated with SCF for the indicated times, and processed for double-fluorescence detection of **(A)** c-KIT and lysotracker or **(C)** c-KIT and Hrs. **(D)** MEFs were incubated with dye-conjugated EGF for the indicated times and processed for double-fluorescence detection of EGF and lysotracker. In **A**, **C**, and **D**, the colocalization of the 2 molecules was analyzed and plotted as histograms for the indicated incubation period. Data are shown as averages \pm SD ($n = 50$ –70). Reproducible results were obtained for 2 independent *Smap1*^{-/-} MEF cultures. $^*P < 0.05$. **(B)** Double-immunofluorescence detection of endogenous SMAP1 and the indicated organelle marker in wild-type MEFs. The nuclei were stained by DAPI. The arrows in insets indicate colocalization of SMAP1 with Hrs or clathrin. Scale bars: 10 μ m; 1 μ m (insets).

cytosis of cell surface c-KIT was not affected by *SMAP1* targeting, contrary to the effect on transferrin endocytosis.

Immunofluorescence analysis using anti-c-KIT antibodies (Figure 4B) showed that, prior to the addition of SCF, c-KIT was similarly detected on the cell surface in both types of cells. Fifteen and thirty minutes after addition of SCF, internalized c-KIT was detected in a punctate pattern in the cytoplasm. Prolonged incubations for up to 180 minutes resulted in the disappearance of the c-KIT signal in wild-type cells, whereas it was still clearly visible in the targeted cells, suggesting that the downregulation of internalized c-KIT might be delayed in the *Smap1*-targeted BMMCs.

Immunoblot analyses (Figure 4C) confirmed this effect by showing the c-KIT protein at comparable levels in both genotypes before addition of SCF and a significant downregulation of the protein 60 minutes after addition of ligand in the wild-type cells but not the targeted cells. Immunoblotting and immunofluorescence results collectively indicate that although c-KIT endocytosis was not affected by *SMAP1* targeting, the downregulation of internalized c-KIT was delayed in the targeted BMMCs.

Then, we examined whether c-KIT remaining in targeted cells was capable of transmitting growth signals to downstream molecules. SCF binding triggers tyrosine phosphorylation of c-KIT, which is followed by monoubiquitination and Grb2 association. Ubiquitination and Grb2 association are the necessary events leading to endocytosis

of c-KIT and signal transmission to ERK1/2, respectively. As seen in Figure 4C, the induction and downregulation of c-KIT phosphorylation and the levels of Grb2 and ERK1/2 did not differ between the 2 genotypes. On the other hand, c-KIT ubiquitination, Grb2 association with c-KIT, and ERK1/2 phosphorylation increased 2 fold in *Smap1*^{-/-} BMMCs as compared with that in the wild-type cells (see the quantification of band densities in Figure 4C, right panels). Figure 4D shows the incorporation of ³H-thymidine into an acid-insoluble fraction and demonstrates that SCF treatment induced DNA synthesis and a 1.6-fold increase in ³H-thymidine incorporation in the targeted cells as compared with that in the wild-type cells. Collectively, the above results indicate that, in the presence of SCF, *Smap1*-targeted BMMCs tend to accumulate c-KIT in the cytoplasm, resulting in enhanced signaling and cell growth activity.

*Sorting of c-KIT to lysosomes is delayed in *Smap1*^{-/-} MEFs.* Ligand-engaged and internalized c-KIT is transported first to early endosomes and then transits through multivesicular bodies (MVBs) and finally to lysosomes, in which the protein is degraded by digestive enzymes (18, 19). Because the persistent accumulation of c-KIT in the cytoplasm of *Smap1*^{-/-} BMMCs suggests an alteration in the transport pathway, the intracellular trafficking of c-KIT was examined in MEFs.

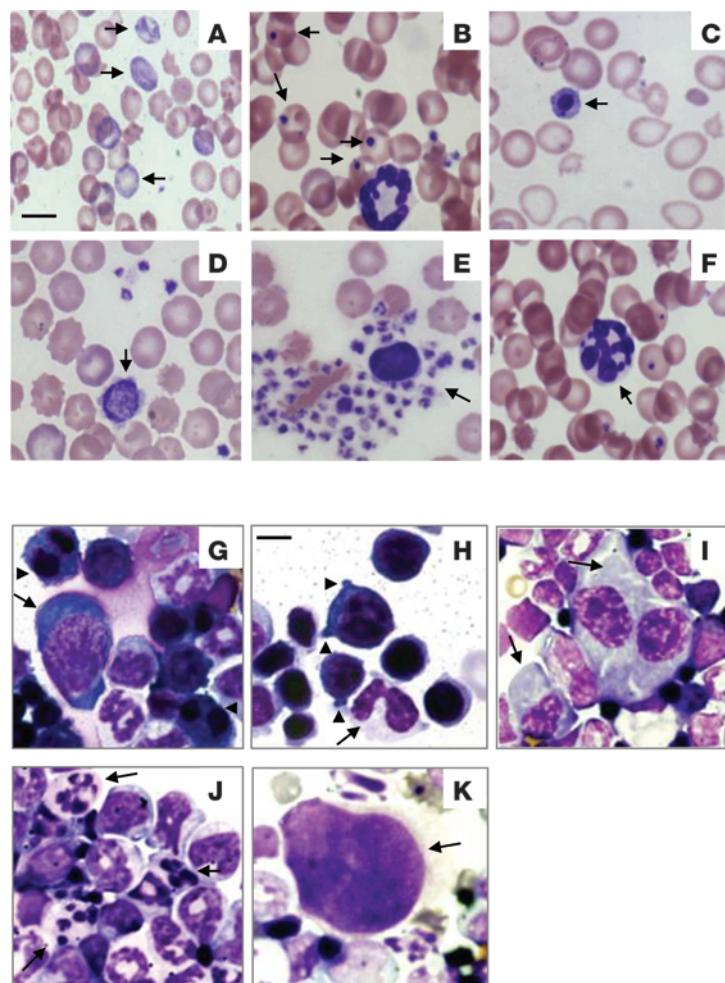
As shown in Figure 5A, c-KIT was detected on the cell surface prior to SCF stimulation (see 0 minutes) and then internalized into the cytoplasm after 10 minutes of SCF treatment in both *SMAP1* genotypes. However, after 30 minutes, a substantial fraction of c-KIT colocalized with lysotracker in the wild-type MEFs but not in targeted MEFs. The colocalization efficiency of the 2 molecules was quantified, and the result is shown as a histogram (Figure 5A). In *Smap1*^{-/-} cells, although c-KIT was incorporated into the cytoplasm upon SCF addition, its transport to lysosomes appeared impaired.

To identify the specific step in the transport of c-KIT that was affected by *SMAP1* targeting, wild-type MEFs were costained for endogenous SMAP1 and various organelle markers. SMAP1 fluorescence did not overlap with that of EEA1, Rab5, Rab11, and LBPA, and no colocalization with lysotracker was observed (data not shown). However, SMAP1 showed partial colocalization with Hrs, an MVB marker (Figure 5B). Substantial colocalization of SMAP1 and clathrin was as previously reported (5, 6). These obser-

Table 1Peripheral blood counts in *SMAP1*^{-/-} mice

Genotype	No. of mice	rbc (10^4 / μ l)	Hematocrit (%)	Hemoglobin (g/dl)	MCV (fl)	MCH (pg)	Reticulocytes (%)	PLT (10^4 / μ l)	wbc (10^2 / μ l)
<i>Smap1</i> ^{+/+}	24	1,002 \pm 49	45.2 \pm 1.8	14.8 \pm 0.6	44.8 \pm 1.0	14.7 \pm 0.3	4.7 \pm 0.9	145.2 \pm 21.9	108 \pm 27
<i>Smap1</i> ^{-/-} (nonanemic)	16	1,012 \pm 81	45.8 \pm 2.0	15.1 \pm 0.7	45.5 \pm 2.8	14.9 \pm 0.9	4.1 \pm 1.2	158.6 \pm 47.6	123 \pm 59
<i>Smap1</i> ^{-/-} (anemic MDS)	10	704 \pm 120 ^A	35.8 \pm 5.6 ^A	11.0 \pm 2.1 ^A	51.4 \pm 3.6 ^A	15.3 \pm 1.1	16.8 \pm 9.0 ^B	90.5 \pm 46.9 ^B	101 \pm 39
<i>Smap1</i> ^{-/-} (MPD/MDS)	2	777 \pm 69 ^A	42.1 \pm 4.3	14.0 \pm 1.6	54.2 \pm 0.8 ^A	18.0 \pm 0.5 ^A	nd	45.4 \pm 43.1 ^A	201 \pm 11 ^A
<i>Smap1</i> ^{-/-} (AML)	5 ^C	752 \pm 93 ^A	36.5 \pm 4.0 ^B	12.0 \pm 1.1 ^A	48.7 \pm 2.9	15.6 \pm 1.1	8.0 \pm 3.5	150.0 \pm 97.5	164 \pm 13 ^B

Statistically significant differences were detected between *Smap1*^{+/+} and *Smap1*^{-/-} mice by Student's *t* test (^A $P < 0.001$, ^B $P < 0.01$). ^CNote that, out of 5 AML-suffering mice, 3 mice were examined for their peripheral blood counts (see Supplemental Table 1 as well). MCV, mean corpuscular volume; MCH, mean corpuscular hemoglobin; PLT, platelets; nd, not determined.

**Figure 6**

Morphology of peripheral blood and bone marrow cells from *Smap1*^{-/-} mice. Smear samples were stained by May-Grunwald Giemsa. (A–F) In peripheral blood (PB), the cells are (A) polychromatic erythrocytes (arrows), (B) Howell-Jolly bodies (arrows), (C) erythroblasts (arrows), (D) giant platelets (arrow), (E) micromegakaryocytes (arrow), and (F) hypersegmented neutrophils (arrow). (G–K) In bone marrow cells (BM), the abnormal cells include (G) megaloblasts (arrow) and binucleated basophilic erythroblasts (arrowheads), (H) basophilic erythroblasts with cytoplasmic blebs (arrowheads) and pseudo-Pelger-Huet neutrophils (arrow), (I) multinucleated polychromatic megaloblasts, (J) orthochromatic erythroblasts with fragmented nuclei (arrows), and (K) megakaryocytes with hypolobulated nuclei. Scale bar: 10 μ m.

appears to be functioning in the MVB-to-lysosome transport of c-KIT but not that of EGFR.

Smap1^{-/-} aged mice develop phenotypes similar to those of myelodysplastic syndrome in humans. The results described above (Figures 2–5) were based on MEFs from embryos and bone marrow cells from 2- to 4-month-old mice, and despite the alterations in membrane traffic, *Smap1*^{-/-} mice were healthy up to the age of 12 months old. When Kaplan-Meier curves were plotted from the 35-month observation period, no statistically significant differences were detected between the survival percentages of *Smap1*-targeted and wild-type mice, although homozygously targeted mice showed a tendency to die at a somewhat younger age (Supplemental Figure 4A). Notably, a substantially higher percentage of homozygously targeted mice showed ill-health conditions after 12 months (Supplemental Figure 4B), suggesting the development of age-related diseases.

In fact, certain mice older than 1 year developed hematological disorders, and 33 of these *Smap1*^{-/-} mice older than 1 year were analyzed by measuring the number of peripheral blood cells in each individual mouse (Table 1 and Supplemental Table 1). Based on the number of rbc, *Smap1*^{-/-} mice were categorized into nonanemic and anemic/myelodysplastic syndrome (anemic/MDS) groups (100% and 70% rbc count as compared with wild type, respectively).

The average hematocrit and hemoglobin values in the anemic *Smap1*^{-/-} group were lower (70%–75%) than those in the nonanemic *Smap1*^{-/-} group and wild-type mice. MCV values were significantly increased in the anemic group, whereas MCH values did not differ significantly between the 2 groups, indicating the presence of macrocytic and normochromic anemia in approximately half of *Smap1*^{-/-} mice. A remarkable increase in the number of reticul-

vations suggest a possible involvement of SMAP1 in the MVB-to-lysosome transport pathway.

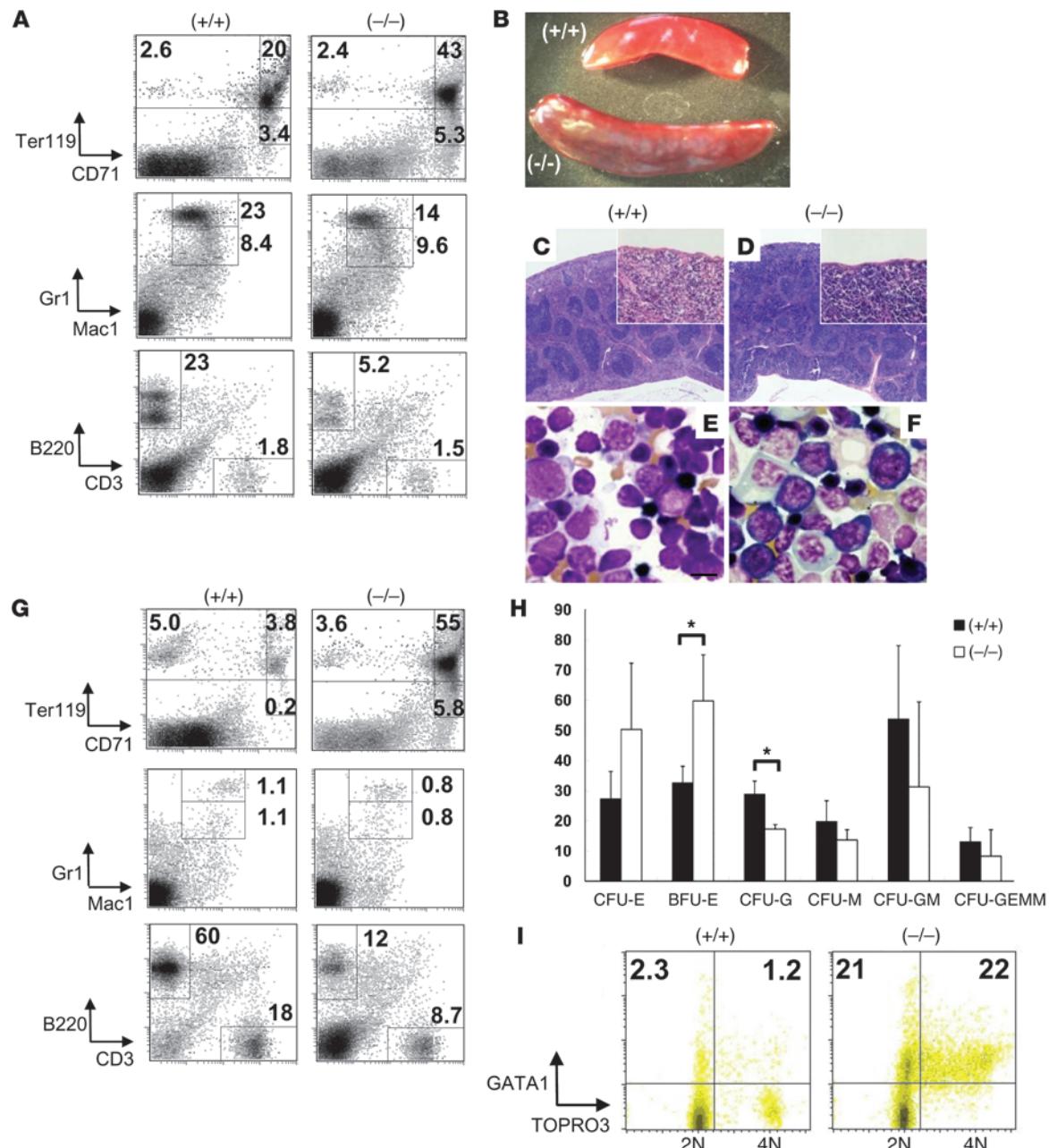
To verify this, the fate of internalized c-KIT was monitored in relation to Hrs (Figure 5C). After 10 minutes of SCF stimulation, a substantial proportion of internalized c-KIT was colocalized with Hrs, indicating the localization of c-KIT in MVBs. After 20 and 30 minutes, c-KIT colocalization with Hrs was not detected in wild-type MEFs but was present in *Smap1*^{-/-} MEFs, indicating that the transport of c-KIT from the MVB to the lysosome was impaired in *Smap1*-targeted MEFs and possibly explaining the accumulation of internalized c-KIT in *Smap1*^{-/-} BMMCs. The delay in the exit of c-KIT from late endosomes/MVBs in *Smap1*^{-/-} MEFs was confirmed by another marker, Rab7 (Supplemental Figure 2A). However, the exit of c-KIT from early endosomes was not affected by SMAP1 targeting, as shown by its colocalization with the early endosome marker Rab5 (Supplemental Figure 2B).

The possible involvement of ARF GTPase itself in the intracellular transport of c-KIT was examined in COS7 cells. Overexpression of the active form of ARF, which can mimic SMAP1 deficiency, induced the substantial colocalization of c-KIT, Hrs, and ARF (Supplemental Figure 3). This implies the delay or block of c-KIT exit from MVBs. On the other hand, the EGF-induced transport of EGFR, another RTK, to the lysosome was not affected in *Smap1*^{-/-} MEFs (Figure 5D). Therefore, an ARF/SMAP1 system

Table 2

Summary of the hematological diagnosis seen in mice

Genotype of mice	No. of mice	Age (mo.)	Diagnosis (no.)	Spleen weight (g)
<i>Smap1</i> ^{+/+}	24	12–25	Nonanemic (24)	0.11 \pm 0.07
<i>Smap1</i> ^{-/-}	33	13–25	Nonanemic (16)	0.10 \pm 0.05
			MDS (10)	0.23 \pm 0.16
			MPD/MDS (2)	0.68 \pm 0.37
			AML (5)	0.99 \pm 0.59


Figure 7

Enhanced erythropoiesis in *Smap1*^{-/-} mice with MDS (ID no. 47; see Table 3). (A and G) Flow cytometry analyses of (A) bone marrow cells and (G) splenocytes prepared from *Smap1*^{+/+} and *Smap1*^{-/-} mice. The cells were stained for indicated hematopoietic lineage markers. The numbers represent percentages of cells in each gated box. Flow cytometry analyses were performed for all *Smap1*^{-/-} mice with MDS, and reproducible results were obtained. (B–F) Histology of spleens. (B) A macroscopic view of the spleen. Note the enlargement of the *Smap1*^{-/-} spleen. (C and D) Sections were stained by hematoxylin and eosin. Note the enrichment of cells with densely stained nuclei in the red pulp of the targeted spleen. (E and F) Smears of Giemsa-stained splenocytes. Erythroblasts with densely stained nuclei are evident in the *Smap1*^{-/-} smear. Scale bar: 10 μ m. (H) CFU-C assay of bone marrow cells. Cells from wild-type and *Smap1*^{-/-} mice were assayed in vitro for their CFU-C activity. The numbers of colonies were counted under a microscope, and the morphology was classified as shown. Triplicate cultures were prepared from each mouse, and 4 independent pairs of older than 1 year *Smap1*^{+/+} and *Smap1*^{-/-} mice were used. The panel shows the averages \pm SD of CFU-C values obtained from 12 cultures from each genotype. * P < 0.0. (I) Estimation of replicating cells in the spleen. Splenocytes from *Smap1*^{+/+} and *Smap1*^{-/-} mice were processed for flow cytometry analysis of GATA1 and TOPRO3. 2N and 4N represent the diploid and tetraploid status of chromatin.

**Table 3**Details of the hematological disorder seen in *Smap1*^{-/-} mice

Mouse ID no.	Founder	Sex	Age (mo.)	Bone marrow	% Blasts in bone marrow cells ^A	Diagnosis
47	44	F	13	Erythroid hyperplasia	2.9	MDS
675	64	F	15	Erythroid hyperplasia	0.4	MDS
834	44	F	18	Erythroid hyperplasia	nd	MDS
826	64	F	18	Erythroid hyperplasia	7.4	MDS
524	64	F	19	Erythroid hyperplasia	4.6	MDS
231	44	F	23	Erythroid hyperplasia	6.3	MDS
576	64	M	18	Erythroid hyperplasia	0.9	MDS
69	64	F	24	ns	3.4	MDS
72	64	M	17	ns	nd	MDS
199	64	M	25	ns	5.0	MDS
448	44	F	23	Myeloproliferative	12	MPD/MDS
518	64	F	17	Myeloproliferative	16	MPD/MDS
192	44	F	14	Erythroleukemia	28	AML
987	64	F	20	Erythroleukemia	24	AML
831	44	F	18	Monocytic leukemia	20	AML
34	64	M	19	Monocytic leukemia	22	AML
138	64	F	25	Monocytic leukemia	36	AML

^ANote that the average percentage of blasts in bone marrow cells was 0.4 ± 0.3 , as counted for 6 *Smap1*^{+/+} mice. Numbers in the "Founder" column indicate the mouse line (the ES number). ns, no significant hyperplasia.

locytes was observed in the *Smap1*^{-/-} anemic group, which likely reflects a mechanism of compensatory erythropoiesis. The number of platelets in *Smap1*^{-/-} mice also decreased to 62%.

Peripheral blood and bone marrow cells from *Smap1*^{-/-} mice were visualized using May-Grunwald-Giemsa staining (Figure 6), which revealed several morphological abnormalities mainly in erythrocytes and erythroid cells but also in megakaryocytic and even myeloid lineages (see the legend Figure 6 for details). Overall, the hematological disorders of *Smap1*^{-/-} mice included (a) macrocytic and normochromic anemia, (b) thrombocytopenia, and (c) abnormal cellular morphologies observed mainly in the erythroid lineage. These hematological disorders were detected exclusively in aged mice. Based on the Bethesda guidelines (20), these phenotypes correspond to features seen in MDS in aged humans.

Based on the above results, 10 *Smap1*^{-/-} mice were diagnosed with MDS (Tables 1 and 2). Mice showing moribund conditions were sacrificed, and their bone marrow cells were examined by flow cytometry (Figure 7A). Erythroid lineage differentiation stages include Ter119^{med}CD71^{hi} (proerythroblasts), Ter119^{hi}CD71^{hi} (basophilic erythroblasts), and Ter119^{hi}CD71^{med/lo} (late erythroblasts, including polychromatic/orthochromatic erythroblasts) (21). The Ter119^{hi}CD71^{hi} fraction increased substantially in the *Smap1*^{-/-} marrow as compared with the wild-type marrow (43% vs. 20%). In addition, the bone marrow from *Smap1*^{-/-} mice showed hypercellularity, suggesting erythroid hyperplasia. Based on smears and flow cytometry, erythroid hyperplasia was detected in 7 out of 10 MDS-diagnosed *Smap1*^{-/-} bone marrow samples (Table 3).

MDS-bearing *Smap1*^{-/-} mice showed splenomegaly (Table 2), as illustrated in Figure 7B. Stained sections of *Smap1*^{-/-} spleens (Figure 7D) revealed structurally intact red and white pulps, but the red pulp was extensively replaced by cells with densely stained nuclei. Splenocyte smears (Figure 7F) showed that the majority of these densely stained cells were erythroblasts. This was confirmed by flow cytometry analysis (Figure 7G), which revealed an increase

in the Ter119^{hi}CD71^{hi} fraction from 3.8% in the wild-type splenocytes to 55% in *Smap1*^{-/-} splenocytes. Thus, splenomegaly seen in *Smap1*^{-/-} mice most likely reflects erythroid hyperplasia.

To examine whether erythroid hyperplasia was accompanied by an enhancement of cell growth activity, a CFU assay was performed by in vitro culture of bone marrow cells (Figure 7H). BFU-E activity in *Smap1*^{-/-} marrow cells was 1.8-fold higher than that in wild-type cells. This increase of BFU-E was apparent only in cells from mice older than 1 year of age, as shown in Figure 7H (data from younger mice is shown in Supplemental Figure 5). GATA1 is a master transcription factor of erythroid lineage. Splenocytes were stained for GATA1 expression and TOPRO3 (Figure 7I). In the *Smap1*-targeted spleen, the number of GATA1⁺ cells with a DNA content above the diploid value (>2N), which represent those undergoing DNA replication, increased to 22%.

The BFU-E assay and GATA1/TOPRO3 staining demonstrated that erythroid cell growth was enhanced in *Smap1*^{-/-} mice. On the other hand, as shown above, these mice were characterized by anemia. Therefore, although erythropoiesis was enhanced in *Smap1*-targeted hematopoietic organs, the presence of morphological abnormalities in erythroid cells was indicative of a dysregulation of this process, resulting in overall ineffective erythropoiesis and eventually anemia.

AML and myeloproliferative disease in *Smap1*-targeted mice. In humans, patients with MDS often develop AML. We therefore assessed the incidence of AML in *Smap1*-targeted mice and found that 5 out of 33 *Smap1*^{-/-} mice developed AML (Tables 1–3). Hematological subtypes of leukemia included erythroid (2 mice) and monocytic (3 mice). Figure 8A shows an example of a monocytic *Smap1*^{-/-} AML mouse in which the liver and spleen were enlarged. Tissue sections of *Smap1*^{-/-} mice (Figure 8B) show the infiltration of leukemic cells in the liver and spleen. Peripheral blood and bone marrow cell smears (Figure 8C) revealed the presence of immature monoblasts with large nuclei in the targeted mice. Figure 8, D and E, shows the results of flow cytometry analyses of bone marrow

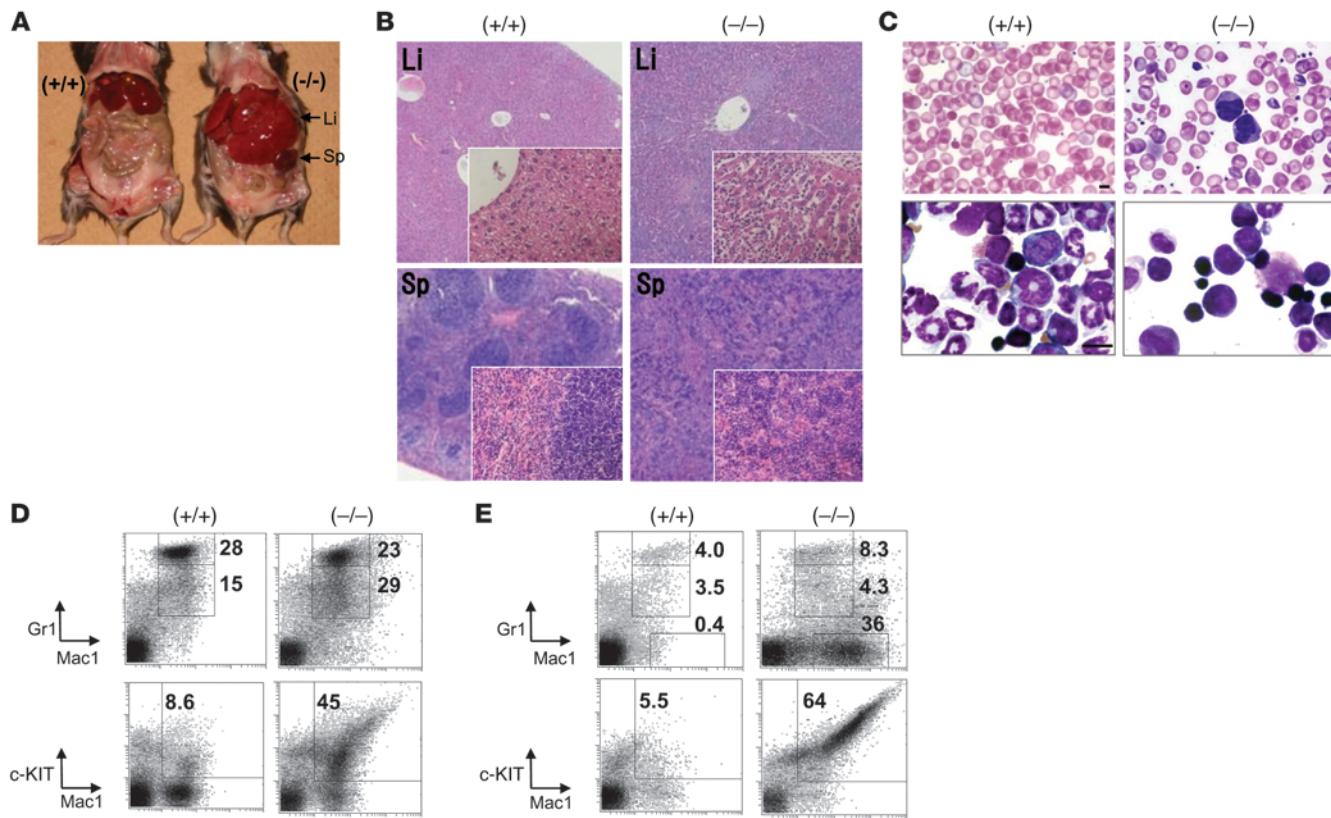


Figure 8

Monocytic AML in a *Smap1*^{-/-} mouse (ID no. 831; see Table 3). **(A and B)** Macroscopic and histological views of the liver and spleen. Note the remarkably enlarged liver and spleen in the *Smap1*^{-/-} mouse. Sections of liver and spleen were stained by hematoxylin and eosin. Note the massive infiltration of leukemic cells in the *Smap1*^{-/-} tissues. The leukemic cells were relatively small in size and possessed densely stained nuclei. Original magnification, $\times 40$; $\times 100$ (insets). Li, liver; Sp, spleen. **(C)** Peripheral blood (top) and bone marrow cell (bottom) smears. Scale bar: 10 μ m. **(D and E)** Flow cytometry analyses of **(D)** bone marrow cells and **(E)** splenocytes prepared from *Smap1*^{+/+} and *Smap1*^{-/-} mice. The cells were stained for hematopoietic lineage markers. The numbers indicate the percentage of cells in each gated box.

cells and splenocytes, respectively. *Smap1*^{-/-} cells showed a substantial increase in the c-KIT⁺Mac1⁺ fraction (45% and 64% in bone marrow cells and splenocytes, respectively, as compared with 8.6% and 5.5% in the wild type) and an increase in the Gr1⁺Mac1⁺ fraction in the spleen (36% vs. 0.4% in the wild type). In addition, the immature Ter119^{hi}CD71^{hi} fraction increased to 27% in the targeted spleen compared with 4.4% in the wild-type spleen (data not shown). This is likely the effect of enhanced compensatory erythropoiesis in response to anemia, suggesting the occurrence of AML in an MDS background. In fact, dysplastic cells were found in the peripheral blood smear (see *Smap1*^{-/-} in Figure 8C).

In addition to monocytic leukemia, *Smap1*^{-/-} mice developed erythroleukemia, as shown in Supplemental Figure 6. Overall, in the 5 mice that developed AML, the percentage of bone marrow blasts was above 20%, compared with 0.4%–7.4% in MDS-only mice (Table 2).

Two *Smap1*^{-/-} mice developed a mixture of myeloproliferative disease (MPD) and MDS (Tables 1–3). The diagnosis of MPD was based on the increase in myeloid lineage cells, according to the Bethesda guidelines. Leukocytosis (as well as anemia and the presence of dysplastic cells) was observed in the peripheral blood, whereas in the bone marrow, the percentage of myeloid blasts was 12%–16%. Supplemental Figure 7 shows the results of analyses performed in a MPD *Smap1*^{-/-} mouse.

Role of c-KIT signaling in the growth of a *Smap1*^{-/-} cell line derived from AML. Finally, we examined the possible involvement of c-KIT signaling in the growth of *Smap1*^{-/-} cells (Supplemental Figure 8). A cell line was established from the bone marrow of a monocytic AML mouse (ID no. 831; see Table 3). This cell line displayed a macrophage-like morphology and expressed c-KIT^{lo}, Mac1^{hi}, CD71^{lo}, and SCF (Supplemental Figure 8, A and B). Interestingly, as seen in Supplemental Figure 8C, which shows cells cultured without exogenous SCF, c-KIT was detected not only on the cell surface but also in the cytoplasm, and a fraction of cytoplasmic c-KIT colocalized with Hrs. In addition, treatment of cells with imatinib (a tyrosine kinase inhibitor) (22) or ISCK03 (a c-KIT inhibitor) (23) reduced cell viability and ERK1/2 phosphorylation (Supplemental Figure 8, D–F). Therefore, c-KIT signaling appears to have a positive effect on the viability of this AML-derived cell line.

Discussion

We previously reported that the overexpression of SMAP1 impairs the endocytosis of the transferrin receptor in cultured cells (refs. 5, 6, and see Supplemental Figure 9 for the effects of SMAP1 overexpression on c-KIT transport). In this study, we confirmed that SMAP1 is similarly involved in transferrin endocytosis in mouse tissues but unexpectedly found that *SMAP1*-deleted cells (both erythroblasts



and MEFs) incorporated transferrin more efficiently than wild-type cells. One possible explanation for this enhanced endocytosis of transferrin is the upregulation of the active form of ARF6 in the absence of SMAP1. In this case, SMAP1 could be conferred a negative role in endocytosis. However, a number of reports provided evidence that, for vesicles to be formed properly, ARF has to exert its GTPase activity and itself be converted to an inactive form (24–28). Therefore, as an alternative mechanism of transferrin endocytosis, we hypothesized that endogenous SMAP1 could play a positive role by converting ARF6 to an inactive form and speculated the possible involvement of SMAP2, a SMAP1 homolog. Namely, SMAP1 deficiency might be accompanied by the mobilization of SMAP2 as a compensatory mechanism in *Smap1*-targeted cells (29). Since SMAP2 can function as an ARF1 GAP and exhibit higher GAP activity than SMAP1 (16), the recruitment of SMAP2 might lead to enhancement of transferrin endocytosis. This mechanism was supported by the abolishment of transferrin endocytosis in SMAP2-silenced *Smap1*^{−/−} MEFs. A positive role for SMAP1 is compatible with the function of other ARF GAP proteins whose knockdown impairs vesicle transport. For example, targeting of ARF GAP1 impaired transferrin endocytosis (30), suggesting an ARF GAP1-dependent but SMAP1/2-independent route of transferrin endocytosis. In any case, our results support a positive role of SMAP1 in vesicle formation and may contribute to the discussion on the putative terminator versus effector functions of ARF GAPs (31, 32).

In this study, the loss of SMAP1 from BMMCs had no impact on SCF-mediated endocytosis of c-KIT, indicating that SMAP1 may play a role in the constitutive endocytosis of transferrin but not in the ligand-induced internalization of c-KIT. This result supports the idea that ARF GAP proteins function in distinctive, cargo-dependent pathways of endocytosis (33, 34). Alternatively, the present results may reveal a novel and possibly important function of SMAP1 in the degradation of c-KIT. SCF induces the phosphorylation and monoubiquitination of c-KIT, which are necessary for Grb2 association and endocytosis, respectively. The dephosphorylation of internalized RTKs occurs on early endosomes (7, 35). We observed that the kinetics of c-KIT in the early phases, such as phosphorylation, dephosphorylation, endocytosis from the cell surface, and exit from early endosomes, were not affected by the loss of SMAP1. In the late phases, c-KIT is transported to MVBs and eventually to the lysosomes (18, 19, 36). The endocytic adaptor protein Hrs functions at the MVB together with clathrin to sort ubiquitinated cargo to the lysosome (37, 38). Our results show that endogenous SMAP1 colocalized at least partially with clathrin and Hrs, suggesting that SMAP1 may be involved in the sorting of cargo at the MVB. In line with this notion, transport of c-KIT from MVB to lysosomes was substantially delayed in *Smap1*^{−/−} cells. Importantly, undegraded c-KIT, ubiquitinated c-KIT, and Grb2-associated c-KIT increased in *Smap1*-targeted cells. Collectively, our results suggest that, in *Smap1*^{−/−} cells, c-KIT persisting on MVBs may be in the dephosphorylated form but still capable of associating with Grb2, thereby leading to elevated levels of phosphorylated ERK. This suggests an enhancement of SCF-triggered c-KIT signaling, which was confirmed by an increase in thymidine uptake in these cells as an indicator of enhanced cell growth activity.

The alterations in transferrin and c-KIT transport discussed above were based on the analysis of *Smap1*-targeted cells derived from embryos and/or young mice exhibiting no obvious pathologies. We also revealed that aged *Smap1*-targeted mice exhibited phenotypes that resembled those of hematological disorders in

human patients with MDS. Importantly, the signs of MDS were observed in 50% of *Smap1*-targeted aged mice, suggesting that disturbances in membrane transport may act as a predisposing but not a deterministic factor for MDS development. For MDS to occur, a genetic alteration may need to be present in addition to *SMAP1* deficiency.

The MDS-like features observed in *Smap1*-targeted mice included anemia, thrombocytopenia, and the presence of dysplastic blood cells. The majority of these mice showed signs of accelerated erythropoiesis in the bone marrow and spleen, which were confirmed by an increase in the number of erythroid-committed progenitors. Erythroid cells may have acquired enhanced proliferation abilities associated with differentiation defects, resulting in the occurrence of anemia. Enhanced expression of *SMAP1* in the MEPs and Ter119⁺ fractions is in good accordance with the erythroid lineage-specific effects of *SMAP1* targeting.

Patients with MDS are particularly prone to developing AML. In this study, *Smap1*-targeted mice developed a range of AML subtypes, such as erythroid and monocytic leukemia. This is consistent with the notion that, in humans, MDS clones arise in CD34⁺ progenitor cells (39) and with the observation that *SMAP1* expression is detected broadly in various hematopoietic lineages. Several *Smap1*^{−/−} animals that had been diagnosed with MDS in our study subsequently developed AML (data not shown). It must be noted that not all MDS mice progressed to AML. This suggests that an additional genetic alteration(s) might be necessary to fully confer AML phenotypes. Screening and identification of such secondary mutations might provide information on the possible cooperation between *SMAP1* and another gene, resulting in the progression from MDS to AML.

Several clathrin-related molecules have been reported to enhance both transferrin endocytosis and cell growth. Huntingtin-interacting protein 1 (HIP1) is a clathrin-associated protein, and its expression level is frequently elevated in primary human cancers. In addition, overexpression of HIP1 alters the distribution patterns of clathrin and the AP-2 adaptor protein and promotes endocytosis of the transferrin receptor (40). Hrb/AGFG, another ARF GAP, functions positively in transferrin endocytosis in leukemic cells overexpressing Notch (41). It is widely accepted that transferrin receptor expression is increased in malignant tumors, including hematological malignancies, and may promote cell growth (42–44). Therefore, the enhanced endocytosis of transferrin in *Smap1*-targeted mice might facilitate the active iron metabolism and c-KIT-induced growth of erythroblasts and MDS/AML cells.

Furthermore, certain molecules that regulate the endocytosis of RTKs, such as c-KIT, have been associated with several human cancers. *c-Cbl* encodes an E3 ligase that ubiquitinates RTKs and is mutated in some cases of human MDS/AML (45, 46). *Tsg101* is a component of the ESCRT-1 complex that functions downstream of Hrs to sort ubiquitinated RTKs to MVBs. In various human cancers, *TSG101* is deleted or its splicing pattern is altered (7, 47). Furthermore, *SMAP1* is frequently mutated in human colon cancer associated with microsatellite instability (15). Base deletion or addition in the (A)₁₀ repeat causes a frameshift in the ARF GAP domain, resulting in a loss-of-function type mutation of *SMAP1* (occurrence of colon cancer has not been observed in our *Smap1*^{−/−} mice so far). Although one case of a *MLL-SMAP1* chimeric gene was reported in monocytic AML, the possible involvement of *SMAP1* in human MDS/AML remains to be investigated in the future.



In summary, this study revealed the predisposing role of alterations in clathrin-dependent protein trafficking in the development of MDS (and subsequent AML). To the best of our knowledge, this is the first report describing this mechanism using a mouse model.

Methods

Mice. The 4.8-kb and 6.4-kb fragments corresponding to the 5' and 3' sequences, respectively, of exon 1 of *SMAP1* were obtained from the corresponding BAC clones by appropriate restriction digestion. The genomic fragments, as well as the loxP site, *frt*-flanked neomycin resistance cassette, and diphtheria toxin subunit A gene, were inserted into the targeting vector. The resulting plasmid DNA was linearized and electrophoretically transfected into TT2 ES cells, which were derived from an F1 mouse of a C57BL/6 and CBA mating (48). Positive and negative selection and PCR genotyping yielded 14 colonies. Recombinant alleles were detected by PCR screening using the forward and reverse primers (5'-CTGACCGCTTCCTC-GTGTCTTACG-3' and 5'-AATACACATGGCCTAGATATTAACCTATAG-3') derived from the neomycin resistance cassette and 3' external region. Recombination was verified by Southern blot analysis, and 3 independent clones were each injected into 8-cell stage embryos of CD-1 mice. Two clones, 44 and 64, were successfully transmitted through the germ line, and *Smap1*-heterozygous mice were backcrossed to C57BL/6 mice for more than 10 generations. During these matings, heterozygous mice were crossed with *E2A-Cre* transgenic mice, causing the deletion of exon 1 in all tissues. *Smap1*^{+/−} mice (acc. no. CDB0427K; <http://www.cdb.riken.jp/arg/mutant%20mice%20list.html>) were thus established. For PCR genotyping of mice, the reverse primer was 5'-CCTCTGCTAACTCTACTCAG-3', and the forward primers were 5'-GTCATCCTGGTTAGCCTCAGTCTTG-3' for the wild-type alleles, 5'-CGCCTTCTATCGCCTCTTGACG-3' for the floxed alleles, and 5'-CCTGCCCTTACCCAGACTGTCTTAG-3' for the targeted alleles. The expected sizes of PCR products were 330 bp, 554 bp, and 480 bp for wild-type, floxed, and targeted alleles, respectively. Mice were maintained in the Animal Facility of the Institute of Development, Aging, and Cancer, Tohoku University, an environmentally controlled and specific pathogen-free facility.

Cultures of BMMCs, the AML-derived cell line, and MEFs. Femoral bone marrow cells were cultured in RPMI1640 supplemented with 10% (v/v) FBS, 2 mM L-glutamine, 50 μM 2-mercaptoethanol, 10 mM HEPES (pH 7.4), 0.2 mM nonessential amino acids, 1 mM sodium pyruvate, 100 U/ml penicillin, and 100 μg/ml streptomycin. The cytokines added were 20 ng/ml IL-3 and 10 ng/ml SCF for the first 2 weeks followed by 20 ng/ml IL-3 only for the next 2 weeks. The purity of BMMCs reached over 95%, as assessed by the surface expression of c-KIT and FcεRI. A cell line was established from the bone marrow of an AML-bearing *Smap1*^{+/−} mouse (ID no. 831; see Table 3) and cultured (deposited as TKG 0661 at the Cell Resource Center for Biomedical Research, Institute of Development, Aging and Cancer, Tohoku University). The medium was the same as that used for BMMCs but without any cytokine added. Cell viability was measured using a Cell Counting Kit-8 (Dojindo). In the indicated cases, cells were treated with imatinib mesylate (Santa Cruz Biotechnology Inc.) or ISCK03 (Sigma-Aldrich).

MEFs were prepared according to a previously published procedure (49). Briefly, 16.5-day-old embryos were isolated from pregnant mice. Single cell suspensions were prepared by trypsin digestion, cultured in a monolayer in DMEM supplemented with serum for 3 days, and immortalized by transfection with the SV40 large T antigen. Two independent cultures were established for wild-type and *Smap1*^{+/−} genotypes. Two siRNAs against SMAP2, siRNA1 and siRNA2, were purchased from Japan BioServices Co. LTD. and used to downregulate endogenous SMAP2 in MEFs. The sequences of siRNA1 and siRNA2 were 5'-GGAUUUAUUCGA-GAUAAAUTT-3' and 5'-CCUGUUGUUUUUGAGAAAGTT-3', respectively.

Hematological and histological examination. Peripheral blood was collected from the tail veins of mice, and hematological parameters were measured using an automated cell counter (XT-4000i, Sysmex). Smears of peripheral blood and bone marrow cells were prepared on glass slides and stained with May-Grunwald-Giemsa (Wako). Tissues, such as those derived from the spleen and liver, were fixed in 4% (w/v) paraformaldehyde in PBS for 18 hours at 4°C and embedded in paraffin. Microsections of each specimen were fixed on glass slides, deparaffinized, and stained with hematoxylin and eosin (Wako).

Flow cytometry analyses. Single cell suspensions were prepared from the bone marrow and spleen and incubated with CD16/32 mAb (BD Pharmingen) for 15 minutes, followed by incubation with an appropriately diluted, fluorescein-conjugated mAb on ice for 30 minutes. The mAbs used were PE-anti-CD71 (eBioscience) and FITC-anti-c-KIT, PE-anti-Ter119, APC-anti-Ter119, PE-anti-Gr1, APC-anti-Mac1, APC-anti-B220, and FITC-anti-CD3ε (all from BD Pharmingen). For DNA labeling, cells were fixed in 4% (w/v) paraformaldehyde in PBS and permeabilized with 0.25% (v/v) Triton X-100 and 5% (w/v) DMSO in PBS. After blocking with 10% (v/v) goat serum in PBS, cells were incubated with anti-GATA1 mAb (Santa Cruz Biotechnology Inc.) and TOPRO3 (Molecular Probes), followed by a secondary antibody reaction. The labeled cells were separated with an analytical flow cytometer (Beckman Coulter), and the data were analyzed with EXPO32 software. Various hematopoietic progenitor fractions were identified using the appropriate antibodies. These populations were designated as follows: KSL, Lin[−]/c-KIT⁺/Sca-1⁺; FLT3-KSL, Lin[−]/c-KIT⁺/Sca-1⁺/FLT3[−]; FLT3+KSL, Lin[−]/c-KIT⁺/Sca-1⁺/FLT3⁺; CMP, Lin[−]/c-KIT⁺/Sca-1[−]/CD34^{hi}/FcγR^{lo}; GMP, Lin[−]/c-KIT⁺/Sca-1[−]/CD34^{hi}/FcγR^{hi}; and MEP, Lin[−]/c-KIT⁺/Sca-1[−]/CD34^{lo}/FcγR^{lo}. Cell sorting was performed using a FACSaria (Becton Dickinson).

Colony formation and ³H-thymidine incorporation assays. Single cell suspensions were prepared from the femoral bone marrow and plated at a density of 1×10^4 cells per ml of methylcellulose (M3234, Stem Cell Technologies) in a 3.5-cm-diameter dish. The culture medium contained 50 ng/ml rmSCF (Kirin Brewery Company Ltd.), 10 ng/ml rmIL-3 (Wako), 10 ng/ml rmIL-6 (Wako), and 3 U/ml rhEPO (Peprotech). Colonies formed were observed through a phase-contrast microscope, and their numbers were counted on the third day for CFU-E and twelfth day for BFU-E, CFU-G, CFU-M, CFU-GM, and CFU-GEMM. In certain cases, 20 kBq of ³H-thymidine (GE Healthcare) was added to the culture of 2×10^4 BMMCs for 8 hours, and its incorporation into an acid-insoluble fraction was measured by a beta-counter, Matrix 9600 (Packard), according to the described method (50).

Transport assay. Intracellular uptake and recycling of transferrin were evaluated using an erythroblast-containing fraction. Bone marrow cells were incubated in serum-free medium at 37°C for 2 hours and then in RPMI1640 containing 50 μg/ml Alexa Fluor 488-conjugated transferrin (Molecular Probes), 20 mM HEPES (pH 7.4), and 1% (w/v) BSA on ice for 30 minutes. After washing 3 times, transferrin internalization was induced by incubating cells in RPMI1640 containing 10% (v/v) FBS at 37°C for the indicated times. Transferrin remaining on the plasma membrane was removed by incubating cells in a prechilled buffer consisting of 20 mM MES (pH 5), 130 mM NaCl, 50 μM deferoxamine, 2 mM CaCl₂, and 0.1% (w/v) BSA on ice for 20 minutes. After washing 3 times, cells were labeled with PE-anti-Ter119, and the fluorescence intensity of internalized transferrin in each Ter119⁺ fraction was quantified by flow cytometry.

For measuring the internalization of c-KIT, BMMCs were first serum starved and then incubated in RPMI1640 supplemented with 100 ng/ml SCF and 0.1% (w/v) BSA in the presence of 100 μg/ml cycloheximide (Sigma-Aldrich), and the c-KIT remaining on the cell surface was measured by flow cytometry.



Transferrin internalization in MEFs was measured by adjusting the procedure to a monolayer culture. MEFs were incubated in serum-free medium at 37°C for 2 hours and then in the presence of dye-conjugated transferrin at 37°C for various time periods. After treatment of cells with an acid buffer to remove cell surface transferrin, cells were harvested by trypsin-EDTA treatment and processed for flow cytometry. In the indicated cases, endocytosis and recycling were measured separately using biotinylated transferrin that was prepared by coupling Holo-Transferrin (R&D Systems) and NHS-SS-biotin (Thermo SCIENTIFIC). In the internalization assay, MEFs grown on gelatin-coated dishes were incubated in serum-free medium at 37°C for 2 hours and then in the presence of 20 µg/ml biotinylated transferrin for 30 minutes at 4°C, washed, and further incubated at 37°C for various times. Then, the surface-bound biotinylated transferrin was stripped by treating cells with 50 mM sodium 2-mercaptoethane sulfonate (MESNA) in TNB buffer (50 mM Tris-HCl [pH 8.6], 100 mM NaCl, 0.2% [w/v] BSA) for 30 minutes at 4°C, and MESNA was quenched by 75 mM iodoacetamide in TNB buffer for 30 minutes at 4°C. Lysates were prepared in lysis buffer (10 mM Tris-HCl [pH 7.4], 50 mM NaCl, 1 mM EDTA, 0.2% [w/v] BSA, 0.1% [w/v] SDS, 1% [v/v] Triton X-100) and applied to ELISA plates coated with anti-transferrin antibodies. After washing, antibody-trapped biotinylated transferrin was detected using HRP-conjugated streptavidin (DAKO). In the recycling assay, MEFs were incubated with 20 µg/ml biotinylated transferrin for 40 minutes at 37°C, followed by reduction with MESNA and incubation with 200 µg/ml unlabeled transferrin at 37°C for various times. Preparation of cell lysates and ELISA were the same as that above.

To monitor endocytosis and intracellular transport of c-KIT, MEFs were transfected by signal sequence-tagged EYFP-c-KIT (51) (provided by J. Duyster, Technical University of Munich, Munich, Germany) using a 4D-Nucleofector System (Lonza), incubated with 100 ng/ml SCF for different times, fixed with 4% (w/v) paraformaldehyde in PBS, permeabilized with 0.1 (v/v) Triton X-100 in PBS, and then processed for immunofluorescence detection. To examine the possible involvement of ARF GTPases in the intracellular transport of internalized c-KIT, COS7 cells were transfected with dominant-active forms of ARF1(Q71A), ARF5(Q71A), or ARF6(Q67A) (provided by K. Nakayama, Kyoto University, Kyoto, Japan) together with EYFP-c-KIT. To monitor the internalization of the EGFR, MEFs were incubated with 100 µg/ml Alexa Fluor 488-conjugated EGF (Molecular Probes).

Immunoblotting, immunofluorescence, and RT-PCR. The immunoblotting and immunofluorescence protocols used were as previously described (6). Anti-SMAP1 and anti-SMAP2 antibodies were purchased from Sigma-Aldrich. Anti-SMAP1 antibody was raised against amino acids 210–306 of the SMAP1 protein (440 amino acids), as specified by the manufacturer. The mAbs used were as follows: anti-panARF (Affinity Bioreagents); anti-clathrin heavy chain (Thermo Fisher Scientific); anti-c-KIT and anti-ARF6 (both from Santa Cruz Biotechnology Inc.); anti-ubiquitinated proteins (BIOMOL); anti-Rab11, anti-EEA1, anti-Grb2, and anti-phospho-tyrosine (all from BD Transduction Labs); anti-Rab5, anti-Rab7, anti-phosphorylated ERK1/2, and anti-ERK1/2 (all from Cell Signaling Tech-

nology); anti-Hrs (provided by N. Tanaka, Miyagi Cancer Research Center, Natori, Japan); and anti-LBPA (provided by T. Kobayashi, Riken Institute, Wako, Japan). Lysotracker was purchased from Molecular Probes. For immunoprecipitation, BMMCs were lysed with a buffer consisting of 25 mM HEPES-KOH (pH 7.4), 150 mM NaCl, 5 mM EDTA, 1% (v/v) Triton X-100, 2 mM DTT, 5 mM NaF, 5 mM Na₂VO₄, and a mixture of protease inhibitors (Roche Diagnostics). The cellular fluorescence intensities and extent of fluorescence colocalization were measured using a confocal microscope, LSM-510, and the LSM5 Image Examiner tool (Zeiss). Colocalization coefficient was calculated as the ratio of pixels^{colocalized c-KIT}/pixels^{total c-KIT}. For RT-PCR, RNA was extracted from isolated cells using the TRIZOL reagent and reverse transcribed using SSRT II (Invitrogen). The primers used were as follows: for SMAP1, 5'-CTGAGGGAGGAGGACAA-CAAGTAC-3' (forward on exon 1), 5'-GAAGCCAATCTTCCAGAGAAC-3' (forward on exon 3), and 5'-GTAACGGTAGACAGGGTAGCAGGT-3' (reverse on exon 9); for SCF, 5'-GAAGAAAACGCACCGAAGAA-3' (forward) and 5'-TAAGGCTCCAAAAGCAAAGC-3' (reverse).

Pull-down assay using GST-GGA1. A DH5 α strain of *E. coli* was transformed by a GST-GGA1 fusion cDNA (52) (provided by K. Nakayama). Bacteria were grown in LB media and treated with 0.4 mM IPTG for 60 minutes to induce protein expression. Cells were lysed in B-PER buffer (Pierce), and the lysate was centrifuged to obtain a supernatant containing GST-GGA1. Bone marrow cells were lysed with a buffer consisting of 150 mM KCl, 2 mM MgCl₂, 10% (v/v) glycerol, 1 mM DTT, 1 mM EGTA, 1 mM EDTA, 1% (v/v) Triton X-100, 50 mM Tris-HCl (pH 8.0), and 1× protease inhibitors and incubated with purified GST-GGA1 that had been coupled with glutathione-Sepharose beads (Pierce). The bound protein was recovered by eluting with SDS-loading buffer and processed for immunoblot analyses.

Statistics. Statistical significance was evaluated using 2-tailed Student's *t* test, and differences of *P* < 0.05 were considered statistically significant.

Study approval. Animal protocols were reviewed and approved by the Animal Studies Committee of the Tohoku University.

Acknowledgments

This work was supported by research grants from the Ministry of Education, Science, Sports, Culture and Technology of Japan; Gonryo Medical Foundation; and the Creative Interdisciplinary Research Program of the Center for Interdisciplinary Research, Tohoku University, Japan. M. Satake is supported by a GCOE program, "Network Medicine," at Tohoku University.

Received for publication March 5, 2012, and accepted in revised form January 3, 2013.

Address correspondence to: Masanobu Satake, Department of Molecular Immunology, Institute of Development, Aging and Cancer, Tohoku University, Seiryo-machi 4-1, Aoba-ku, Sendai 980-8575, Japan. Phone: 81.22.717.8477; Fax: 81.22.717.8482; E-mail: satake@idac.tohoku.ac.jp.

1. D'Souza-Schorey C, Chavrier P. ARF proteins: roles in membrane traffic and beyond. *Nat Rev Mol Cell Biol.* 2006;7(5):347–358.
2. Donaldson JG, Jackson CL. ARF family G proteins and their regulators: roles in membrane transport, development and disease. *Nat Rev Mol Cell Biol.* 2011;12(6):362–375.
3. Donaldson JG. Multiple roles for Arf6: sorting, structuring, and signaling at the plasma membrane. *J Biol Chem.* 2003;278(43):41573–41576.
4. Gillingham AK, Munro S. The small G proteins of the Arf family and their regulators. *Annu Rev Cell Dev Biol.* 2007;23:579–611.
5. Tanabe K, Torii T, Natsume W, Braesch-Andersen S, Watanabe T, Satake M. A novel GTPase-activating protein for ARF6 directly interacts with clathrin and regulates clathrin-dependent endocytosis. *Mol Biol Cell.* 2005;16(4):1617–1628.
6. Kori S, Tanabe K, Watanabe T, Sabe H, Satake M. Clathrin dependent endocytosis of E-cadherin is regulated by the Arf6GAP isoform SMAP1. *Exp Cell Res.* 2008;314(7):1415–1428.
7. Bache KG, Slagsvold T, Stenmark H. Defective downregulation of receptor tyrosine kinases in cancer. *EMBO J.* 2004;23(14):2707–2712.
8. Crosetto N, Tikkkanen R, Dikic I. Oncogenic break-downs in endocytic adaptor proteins. *FEBS Lett.* 2005;579(15):3231–3238.
9. Lanzetti L, Di Fiore PP. Endocytosis and cancer: an 'insider' network with dangerous liaisons. *Traffic.* 2008;9(12):2011–2021.
10. McMahon HT, Boucrot E. Molecular mechanism and physiological functions of clathrin-mediated endocytosis. *Nat Rev Mol Cell Biol.* 2011;12(8):517–533.
11. Kalesnikoff J, et al. RabGEF1 regulates stem cell factor/c-Kit-mediated signaling events and biological responses in mast cells. *Proc Natl Acad Sci U S A.* 2006;103(8):2659–2664.
12. Sigismund S, Argenzio D, Tosoni D, Cavallaro E, Polo



S, Di Fiore PP. Clathrin-mediated internalization is essential for sustained EGFR signaling but dispensable for degradation. *Dev Cell*. 2008;15(2):209–219.

13. Joffre C, Barrow R, Ménard L, Calleja V, Hart IR, Kermorgant S. A direct role for Met endocytosis in tumorigenesis. *Nat Cell Biol*. 2011;13(7):827–837.

14. Meyer C, et al. Diagnostic tool for the identification of MLL rearrangements including unknown partner genes. *Proc Natl Acad Sci USA*. 2005;102(2):449–454.

15. El-Bchiri J, et al. Nonsense-mediated mRNA decay impacts MSI-driven carcinogenesis and anti-tumor immunity in colorectal cancers. *PLoS One*. 2008;3(7):e2583.

16. Natsume W, et al. SMAP2, a novel ARF GTPase-activating protein, interacts with clathrin and clathrin assembly protein and functions on the AP-1-positive early endosome/trans-Golgi network. *Mol Biol Cell*. 2006;17(6):2592–2603.

17. Collins BM, Watson PJ, Owen DJ. The structure of the GGA1-GAT domain reveals the molecular basis for ARF binding and membrane association of GGAs. *Dev Cell*. 2003;4(3):321–332.

18. Sorkin A, Von Zastrow M. Signal transduction and endocytosis: close encounters of many kinds. *Nat Rev Mol Cell Biol*. 2002;3(8):600–614.

19. Masson K, Heiss E, Band H, Ronnstrand L. Direct binding of Cbl to Tyr568 and Tyr936 of the stem cell factor receptor/c-Kit is required for ligand-induced ubiquitination, internalization and degradation. *Biochem J*. 2006;399(1):59–67.

20. Kogan SC. Bethesda proposals for classification of nonlymphoid hematopoietic neoplasms in mice. *Blood*. 2002;100(1):238–245.

21. Socolovsky M. Ineffective erythropoiesis in $\text{St}5\alpha^{-/-}\text{Sb}^{-/-}$ mice due to decreased survival of early erythroblasts. *Blood*. 2001;98(12):3261–3273.

22. Heinrich MC, Griffith DJ, Druker BJ, Wait CL, Ott KA, Zigler AJ. Inhibition of c-kit receptor tyrosine kinase activity by ST1 571, a selective tyrosine kinase inhibitor. *Blood*. 2000;96(3):925–932.

23. Na YJ, Baek HS, Ahn SM, Shin HJ, Chang IS, Hwang JS. [4-t-butylphenyl]-N-(4-imidazol-1-yl phenyl)sulfonamide (ISCK03) inhibits SCF/c-kit signaling in 501mel human melanoma cells and abolishes melanin production in mice and brownish guinea pigs. *Biochem Pharmacol*. 2007;74(5):780–786.

24. Lanoix J, et al. Sorting of Golgi resident proteins into different subpopulations of COPI vesicles: a role for ArfGAP1. *J Cell Biol*. 2001;155(7):1199–1212.

25. Yang JS, et al. ARFGAP1 promotes the formation of COPI vesicles, suggesting function as a component of the coat. *J Cell Biol*. 2002;159(1):69–78.

26. Lee SY, Yang JS, Hong W, Premont RT, Hsu VW. ARFGAP1 plays a central role in coupling COPI cargo sorting with vesicle formation. *J Cell Biol*. 2005;168(2):281–290.

27. Kahn RA. Toward a model for Arf GTPases as regulators of traffic at the Golgi. *FEBS Lett*. 2009;583(23):3872–3879.

28. Spang A, Shiba Y, Randazzo PA. Arf GAPs: gatekeepers of vesicle generation. *FEBS Lett*. 2010;584(12):2646–2651.

29. Sakakura I, Tanabe K, Nouki N, Suzuki M, Satake M, Watanabe T. The carboxy-terminal region of SMAP2 directs subcellular localization as well as Arf protein specificity. *Biochem Biophys Res Commun*. 2011;404(2):661–666.

30. Bai M, et al. ARFGAP1 promotes AP-2-dependent endocytosis. *Nat Cell Biol*. 2011;13(5):559–567.

31. Kahn RA. GAPs: Terminator versus effector functions and the role(s) of ArfGAP1 in vesicle biogenesis. *Cell Logist*. 2011;1(2):49–51.

32. Kon S, Funaki T, Satake M. Putative terminator and/or effector functions of Arf GAPs in the trafficking of clathrin-coated vesicles. *Cell Logist*. 2011;1(3):86–89.

33. Randazzo PA, Hirsch DS. Arf GAPs: multifunctional proteins that regulate membrane traffic and actin remodelling. *Cell Sig*. 2004;16(4):401–413.

34. Tanabe K, Kon S, Natsume W, Torii T, Watanabe T, Satake M. Involvement of a novel ADP-ribosylation factor GTPase-activating protein, SMAP, in membrane trafficking: implications in cancer cell biology. *Cancer Sci*. 2006;97(9):801–806.

35. Authier F, Chauvet G. In vitro endosome-lysosome transfer of dephosphorylated EGF receptor and Shc in rat liver. *FEBS Lett*. 1999;461(1–2):25–31.

36. Zeng S, Xu Z, Lipkowitz S, Longley JB. Regulation of stem cell factor receptor signaling by Cbl family proteins (Cbl-b/c-Cbl). *Blood*. 2005;105(1):226–232.

37. Raiborg C, Bache KG, Mehlum A, Stang E, Stenmark H. Hrs recruits clathrin to early endosomes. *EMBO J*. 2001;20(17):5008–5021.

38. Raiborg C, Bache KG, Gillooly DJ, Madshus IH, Stang E, Stenmark H. Hrs sorts ubiquitinated proteins into clathrin-coated microdomains of early endosomes. *Nat Cell Biol*. 2002;4(5):394–398.

39. Corey SJ, Minden MD, Barber DL, Kantarjian H, Wang JC, Schimmer AD. Myelodysplastic syndromes: the complexity of stem-cell diseases. *Nat Rev Cancer*. 2007;7(2):118–129.

40. Rao DS, et al. Altered receptor trafficking in Huntington interacting protein 1-transformed cells. *Cancer Cell*. 2003;3(5):471–482.

41. Khwaja SS, et al. HIV-1 Rev-binding protein accelerates cellular uptake of iron to drive Notch-induced T cell leukemogenesis in mice. *J Clin Invest*. 2010;120(7):2537–2548.

42. Smilevska T, et al. Transferrin receptor-1 and 2 expression in chronic lymphocytic leukemia. *Leuk Res*. 2006;30(2):183–189.

43. O'Donnell KA, et al. Activation of transferrin receptor 1 by c-Myc enhances cellular proliferation and tumorigenesis. *Mol Cell Biol*. 2006;26(6):2373–2386.

44. Habashy HO, et al. Transferrin receptor (CD71) is a marker of poor prognosis in breast cancer and can predict response to tamoxifen. *Breast Cancer Res Treat*. 2010;119(2):283–293.

45. Caligiuri MA, et al. Novel c-CBL and CBL-b ubiquitin ligase mutations in human acute myeloid leukemia. *Blood*. 2007;110(3):1022–1024.

46. Sanada M, et al. Gain-of-function of mutated C-CBL tumour suppressor in myeloid neoplasms. *Nature*. 2009;460(7257):904–908.

47. Tanaka N, Kyuuma M, Sugamura K. Endosomal sorting complex required for transport proteins in cancer pathogenesis, vesicular transport, and non-endosomal functions. *Cancer Sci*. 2008;99(7):1293–1303.

48. Yagi T, et al. A novel ES cell line, TT2, with high germline-differentiating potency. *Anal Biochem*. 1993;214(1):70–76.

49. Funaki T, et al. Localization of SMAP2 to the TGN and its function in the regulation of TGN protein transport. *Cell Struct Funct*. 2011;36(1):83–95.

50. Endo S, Sakamoto Y, Kobayashi E, Nakamura A, Takai T. Regulation of cytotoxic T lymphocyte triggering by PIR-B on dendritic cells. *Proc Natl Acad Sci USA*. 2008;105(38):14515–14520.

51. Jahn T, et al. Analysing c-kit internalization using a functional c-kit-EGFP chimera containing the fluorochrome within the extracellular domain. *Oncogene*. 2002;21(29):4508–4520.

52. Takatsu H, Yoshino K, Toda K, Nakayama K. GGA proteins associate with Golgi membranes through interaction between their GGAH domains and ADP-ribosylation factors. *Biochem J*. 2002;365(pt 2):369–378.