Type 1 angiotensin receptors on macrophages ameliorate IL-1 receptor–mediated kidney fibrosis

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In a wide array of kidney diseases, type 1 angiotensin (AT1) receptors are present on the immune cells that infiltrate the renal interstitium. Here, we examined the actions of AT1 receptors on macrophages in progressive renal fibrosis and found that macrophage-specific AT1 receptor deficiency exacerbates kidney fibrosis induced by unilateral ureteral obstruction (UUO). Macrophages isolated from obstructed kidneys of mice lacking AT1 receptors solely on macrophages had heightened expression of proinflammatory M1 cytokines, including IL-1. Evaluation of isolated AT1 receptor–deficient macrophages confirmed the propensity of these cells to produce exaggerated levels of M1 cytokines, which led to more severe renal epithelial cell damage via IL-1 receptor activation in coculture compared with WT macrophages. A murine kidney crosstransplantation concomitant with UUO model revealed that augmentation of renal fibrosis instigated by AT1 receptor–deficient macrophages is mediated by IL-1 receptor stimulation in the kidney. This study indicates that a key role of AT1 receptors on macrophages is to protect the kidney from fibrosis by limiting activation of IL-1 receptors in the kidney.

Introduction

Chronic kidney disease (CKD) constitutes a major public health concern. Hematopoietic cells infiltrate the injured kidney and modulate the progression of immune-mediated glomerulonephritis and even “nonimmune” forms of kidney damage related to hypertension and ischemia-reperfusion (1–3). Among BM-derived cell lineages, macrophages play a prominent role in regulating kidney fibrosis (4), and the extent of fibrosis predicts organ failure in chronic diseases of the kidney and other tissues (5).

After infiltrating the kidney, macrophages can promote inflammatory injury or direct tissue repair depending on their phenotype, summarized in a dichotomous “M1 versus M2” paradigm (6). In this vernacular, proinflammatory M1 macrophages induce damage by secreting cytokines such as TNF-α and IL-1β, whereas reparative M2 macrophages secrete the decoy IL-1 receptor (IL-1R2) and other immunosuppressive mediators. However, the mechanisms through which key drivers of kidney fibrosis contribute to macrophage polarization require elucidation.

As the principal effector molecule of the renin-angiotensin system (RAS), Ang II is a fundamental driver of renal fibrosis (7). Accordingly, treatment with type 1 angiotensin (AT1) receptor blockers has achieved considerable success in managing CKD patients (8). Ang II is a fundamental driver of renal fibrosis (7). Accord-

Results and Discussion

AT1A receptor deficiency on macrophages exacerbates kidney fibrosis. To examine infiltration of LysM-expressing macrophages in the kidney during renal fibrosis, we subjected LysM-Cre+ Agtr1afl/fl mice to UUO. By day 7 of UUO, robust accumulation of GFP+ macrophages was detected in the interstitial areas of obstructed kidneys, but not the contralateral, unobstructed kidneys (Figure 1A), confirming that LysM-Cre–mediated gene excision affects macrophages infiltrating the diseased kidney. To evaluate the actions of AT1 receptors on macrophages that infiltrate the kidney undergoing fibrosis, we harvested kidneys from mice with macrophage-specific deletion of Agtr1a (LysM-Cre+ Agtr1afl/fl, herein referred to as Macro KO) and WT mice 7 days following UUO. Obstructed kidneys from Macro KO mice contained 64%
more interstitial fibrosis (Figure 1B and Supplemental Figure 5), 80% more collagen-producing myofibroblasts (Supplemental Figure 6), and 80% more collagen I (Col I) protein (Figure 1C) than in WT mice. Moreover, at day 7 after UUO, mRNA expressions of Col I, PAI-1, and TGF-β1 were enhanced by 129%, 47%, and 40%, respectively, in Macro KO kidneys compared with WTs (Figure 1D). Additional experiments further confirmed our results (Supplemental Paragraph 2 and Supplemental Figures 7 and 8). These findings contrast starkly with studies in which global AT1A receptor deficiency ameliorates kidney fibrosis (14), but align with reports of exaggerated tissue damage in AT1A receptor–deficient BM chimeras (15, 16). Collectively, our data indicate that activation of AT1 receptors on infiltrating macrophages protects the kidney from progressive fibrosis.

The heterogeneity of macrophage polarization plays a critical role in the pathogenesis of kidney damage and fibrosis (6). In simplified terms, the balance between proinflammatory M1 and reparative M2 macrophages determines the local composition of cytokine reservoirs that orchestrate disruptions in renal architecture following an insult. Therefore, on days 3 and 7 after UUO, we profiled the renal expression of M1 and M2 markers, including M1 cytokines IL-1β and TNF-α, which have been implicated in the progression of renal fibrosis (Figure 1E). Obstructed Macro KO kidneys had significantly higher expression of M1 markers, including IL-1β, IL-1R1, TNF-α, IL-12p40, and CCL2, compared with obstructed WT kidneys 3 days after UUO, whereas expression of M2 markers Arg-1, FIZZ-1, YM-1, and IL-1R2 was virtually identical between cohorts. A similar pattern was noted at day 7 after UUO (Figure 1E), suggesting that activation of AT1 receptors on macrophages represses M1 cytokine expression during kidney fibrosis.

**Figure 1**

**AT1A receptor deficiency on macrophages exacerbates kidney fibrosis induced by UUO.** (A) Representative kidney sections from LysM-Cre+ mT/mG mice 7 days after UUO with contralateral unobstructed kidney on left and obstructed kidney on right. (B) Polarized images of representative sections from obstructed WT and Macro KO kidneys stained with picrosirius red for collagen fibrils at day 7 after UUO. Original magnification, ×20. (C) Western blot for Col I in whole kidney at day 7 after UUO. (D) mRNA expression of ColI, PaI1, and Tgfβ1 in obstructed kidney at day 7 after UUO. (E) Gene expression of M1 markers *Il1b*, *Il1r1*, *Tnfa*, *Il12b*, and *Ccl2* and M2 markers *Arg1*, *Fizz1*, *Ym1*, and *Il1r2* in obstructed kidneys at 3 and 7 days after UUO. At day 3, *P* < 0.02 vs. WT, †*P* = 0.02 vs. WT, ‡*P* = 0.01 vs. WT, §*P* = 0.03, ¶*P* < 0.02. At day 7, *P* < 0.03 vs. WT, †*P* = 0.02 vs. WT, ¶*P* = 0.05, ¶¶*P* < 0.001.

**AT1A receptors on macrophages limit their expression of M1 cytokines.** We next examined the effects of macrophage AT1 receptor activation on the phenotype of infiltrating macrophages at day 7 after UUO by histologic staining (Supplemental Paragraph 3 and Supplemental Figures 9 and 10) and, more precisely, by isolating CD45+CD11b+Ly6Chi and CD45+CD11b+Ly6Clo macrophages directly from the obstructed kidney (Supplemental Figure 11). Based on reports that CD45+CD11b+Ly6Chi macrophages promote tissue damage and fibrosis (4, 17), TaqMan low-density
array (TLDA) card–based microarray was performed to survey gene expression for inflammatory mediators and signaling molecules in the Ly6C\textsuperscript{hi}-activated macrophages (Figure 2A and Supplemental Table 1). Among the upregulated genes, IL-1\textbeta but not TNF-\alpha showed enhanced expression in Macro KO kidneys at both days 3 and 7 of UUO (Figure 1E). Moreover, by real-time PCR, Ly6C\textsuperscript{hi} and Ly6C\textsuperscript{lo} macrophages from the Macro KO kidneys had higher IL-1\textbeta expression levels than found in WT controls (Figure 2B). These data indicate that AT1A receptor activation constrains infiltrating macrophage activation by suppressing key inflammatory cytokines including IL-1 during kidney fibrosis.

To explore in vitro the role of the macrophage AT1A receptor in regulating macrophage activation, we cultured Macro KO and WT peritoneal macrophages and subjected them to M1- or M2-polarizing stimuli or vehicle (M0). M1 stimulation increased mRNA expression of M1 markers in both groups, confirming

Figure 2
AT1A receptor activation on macrophages suppresses their M1 proinflammatory polarization. (A) Heat map showing hierarchical cluster analysis of \Delta C\textsc{t} values of CD11b\textsuperscript{+}Ly6C\textsuperscript{hi} macrophages isolated from obstructed WT and Macro KO kidneys. (B) mRNA expression of IL-1\beta measured by real-time RT-PCR in Ly6C\textsuperscript{hi} and Ly6C\textsuperscript{lo} macrophages isolated from obstructed kidneys. (C) mRNA levels of proinflammatory cytokines II1b, II1a, Ccl5, Ccl2, II12a, and II12b in WT and Macro KO peritoneal macrophages following M1 stimulation. (D) Cll\textalpha and Ngal gene expression in WT (II1r1 WT) or IL-1R–deficient (II1r1 KO) renal tubular epithelial cells cocultured with M0- or M1-activated macrophages from WT or Macro KO (MKO) animals.
M1 polarization (Figure 2C). However, M1 macrophages lacking the AT1A receptor showed exaggerated expression levels for each marker. These findings together with M2 polarization studies (Supplemental Paragraph 4 and Supplemental Figures 12 and 13) suggest that AT1A receptor activation during M1 differentiation constrains M1 cytokine production without altering the macrophage’s susceptibility to M2 differentiation.

Enhanced IL-1 production by AT1 receptor–deficient macrophages drives renal cell injury in coculture. Proinflammatory cytokines including IL-1β have been implicated in renal tubular cell (RTC) damage (18). We therefore hypothesized that exaggerated production of inflammatory cytokines by AT1A receptor–deficient macrophages could alter gene expression programs for injury within the kidney tubular cell. Testing this hypothesis in vitro, we measured expres-
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sion of Col I and kidney injury marker neutrophil gelatinase-associated lipocalin (NGAL) in RTCs following coculture with M0- or M1-conditioned macrophages from our experimental animals (Figure 2D). At 6 hours, M1 macrophages from Macro WT or KO mice, but not M0 macrophages, upregulated Col I and Ngal mRNA expression in RTCs. Thus, cytokines from activated macrophages can mediate damage to kidney tubular cells without direct contact. Moreover, compared with WT M1 macrophages, M1 macrophages lacking AT1A receptors induced an even more profound increase in RTC expression of Col I and NGAL (Figure 2D). To explore whether AT1 receptor–deficient macrophages mediate this exaggerated tubular cell damage through enhanced IL-1 production acting on the renal cell IL-1 receptor (IL-1R1), we repeated our coculture experiments with Il1r1-deficient (KO) RTCs (Figure 2D).

In Il1r1 KO RTCs, coinoculation with M1-conditioned macrophages from WT or Macro KO animals induced severely blunted and equivalent increases in Col I and NGAL expression. Finally, recombinant IL-1β induced robust expression of Col I and NGAL in WT but not Il1r1 KO RTCs (Supplemental Figure 14). Thus, activation of the IL-1R on kidney cells by IL-1 secreted from activated macrophages triggers RTC damage. We therefore posited that AT1 receptor activation on infiltrating macrophages limits their IL-1 production, which in turn suppresses gene expression programs for injury in kidney parenchymal cells by preventing renal IL-1R stimulation.

The kidney crosstransplant UUO model. Testing this possibility in vivo requires a system that can separate the contribution of AT1 receptor activation on macrophages from the downstream effects of renal IL-1R activation during UUO-mediated renal fibrosis. We therefore developed a kidney transplantation–ureteral obstruction (KT-UO) model (Supplemental Paragraph 5 and Supplemental Figure 15). To examine the infiltration of macrophages from the kidney recipient into the donor KT-UO kidney, we transplanted a nonfluorescent WT kidney into a recipient containing only a rare GFP fluorescing macrophage (Figure 3A). Next, we transplanted a kidney from a LysM-Cre+ donor kidney in the KT-UO model are macrophages. The ability of AT1 receptor activation on macrophages to mitigate kidney damage has broad implications for the design of potent therapies to overcome the shortcomings of global angiotensin receptor blockade related not only to previously recognized side effects but also to patently detrimental effects of blocking angiotensin receptors on hematopoietic cells.

Methods

Detailed Supplemental Methods are available online.

Statistics. Data are expressed as mean ± SEM and analyzed by 2-tailed unpaired t test or ANOVA according to statistical methods described in detail in the Supplemental Methods. P < 0.05 was considered significant.

Study approval. Mice were bred and maintained in the Association for Assessment and Accreditation of Laboratory Animal Care–accredited animal facilities at the Durham Veterans’ Affairs Medical Center (DVAMC) per NIH guidelines. The animal studies were approved by the DVAMC Institutional Animal Care and Use Committee and conducted in accordance with the NIH Guide for the Care and Use of Laboratory Animals.
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