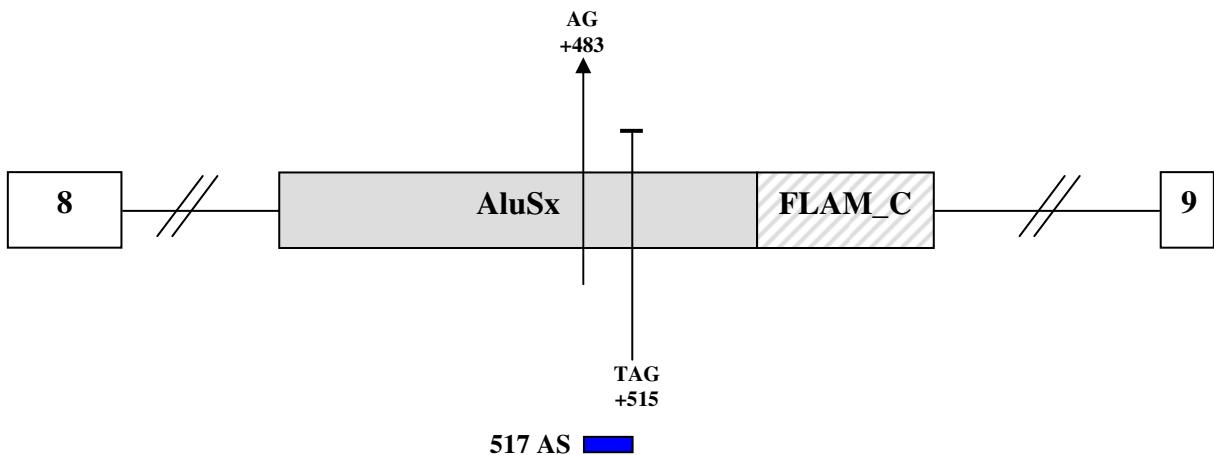


A

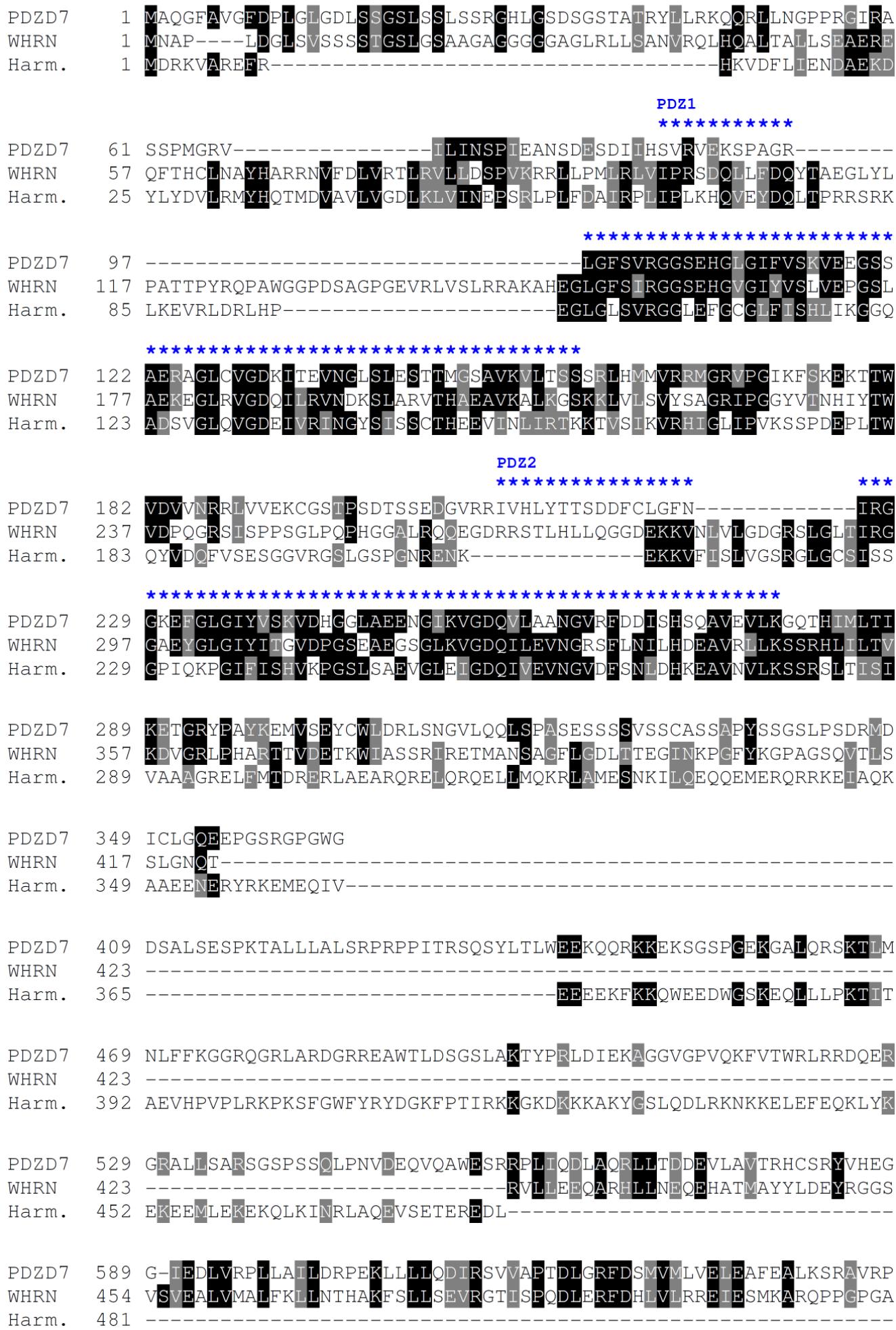
B

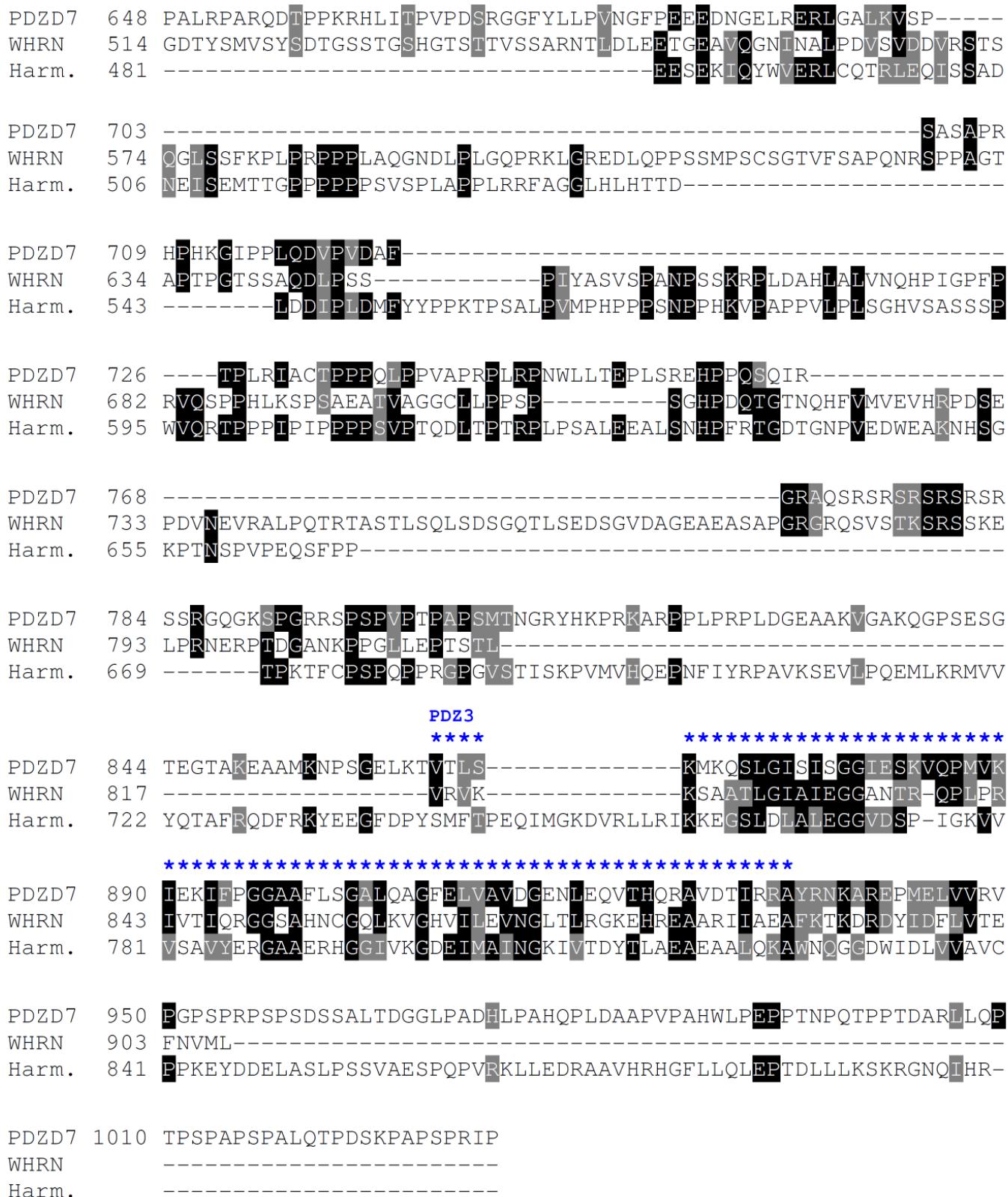
ctgcagaggagaagcagcagcggagaagaaggagaagtccgggtcccctggggagaagggtg Exon 8
 ccctgcagcgctccaagacgctgatgaacctttcaagggagggcggcagggaggg
 tagcgcggacggcgcagagaggctggacactggacagcggagcctggccaaaactt
 accctcgctggacatagagaagtaagttagtggctgatcaaaca... (433 bp) ...
transcribed Alu
 agctaatttttgcattttttagagatggggttcacccctgttgcccaggctag AluSx
 ctcaaattcctggactgaagctatctaccgccttagtctccaaagtgtctaggattaca
 gcgtgatctaccacacctggcaatccaaactactctggaaagctgaaggcagcaagatcac FLAM_C
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C

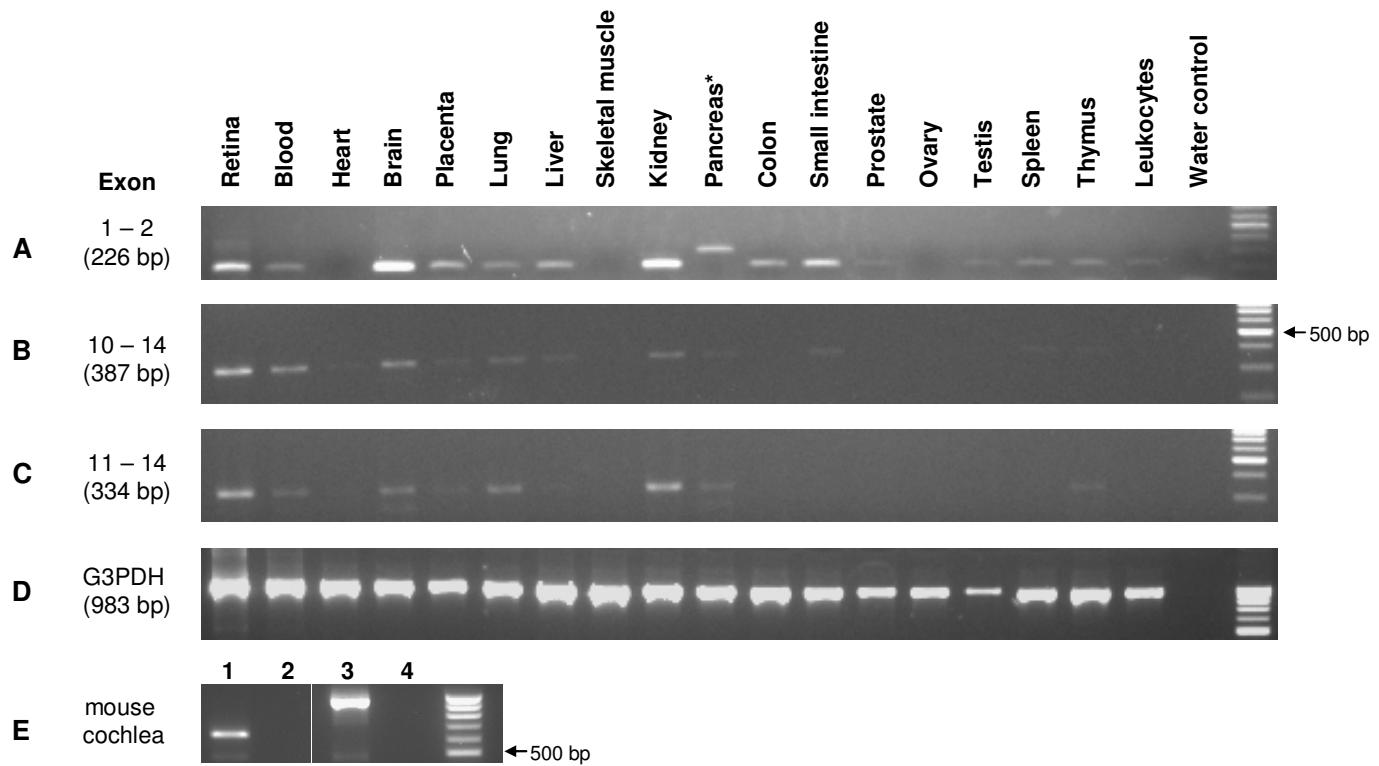
Supplemental Figure 1.

(A) Amino acid sequence of the long PDZD7 isoform (1,033 residues, GenBank acc. no. FJ617449), the Alu-derived 517 aa isoform (blue) and the 561 aa isoform that results from exclusion of exons 9 and 10 from the mRNA (grey). PDZ domains are highlighted in yellow. The proline-rich region is underlined. 5' ends of exons are indicated by arrows. The epitope chosen for production of the antibody is in green. (B) Genomic sequence of exon 8 (red) and intron 8. AluSx sequence in blue, transcribed Alu sequence in bold and underlined, FLAM_C in green. (C) Scheme of the Alu-specific splicing (517 aa isoform). Splicing occurs at an acceptor splice site located within the AluSx sequence (c.1522+483). Termination occurs at an *in-frame* stop codon (c.1522+515). The blue bar indicates the AluSx-derived sequence that is translated as C-terminus of the 517 aa *PDZD7* transcript.





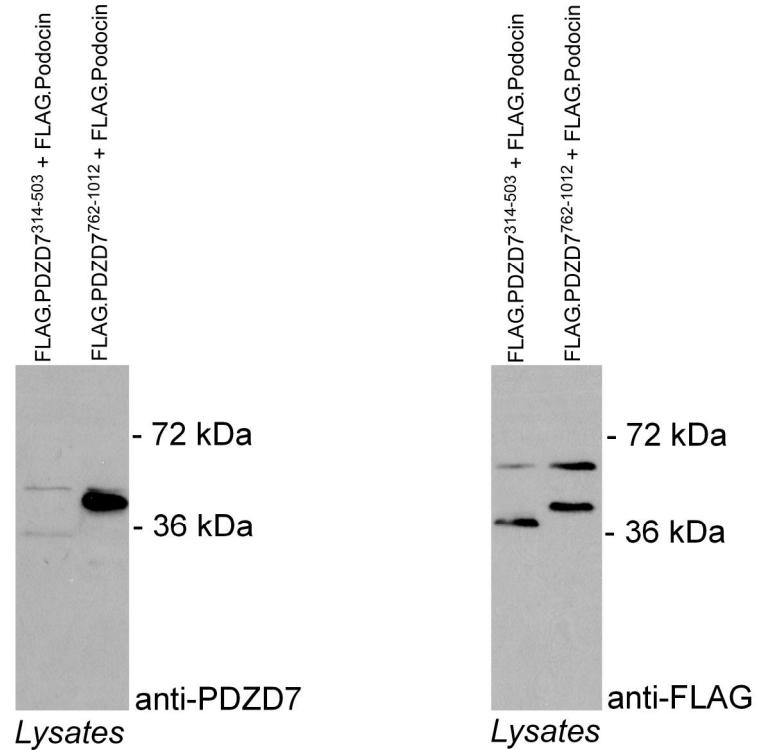
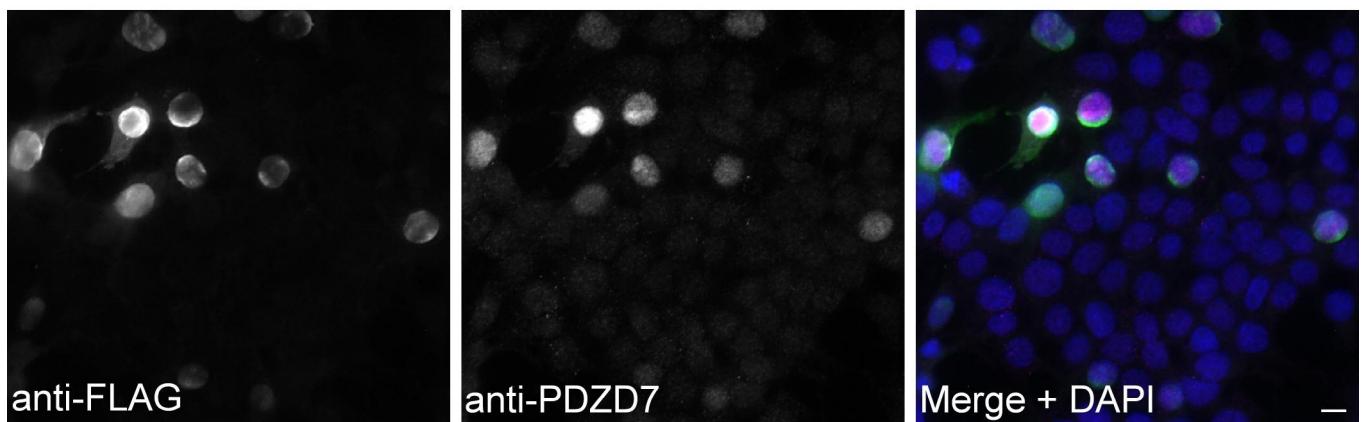
Supplemental Figure 2. Alignment of amino acid sequences of PDZD7 (1,033 residues isoform, GenBank acc. no. FJ617449), the USH2D protein whirlin (long isoform, GenBank acc. no. BC142614) and the USH1C protein harmonin (b3 isoform, GenBank acc. no. NM_153676).



Supplemental Figure 3

Expression profile of human and mouse *PDZD7* (RT-PCR analyses).

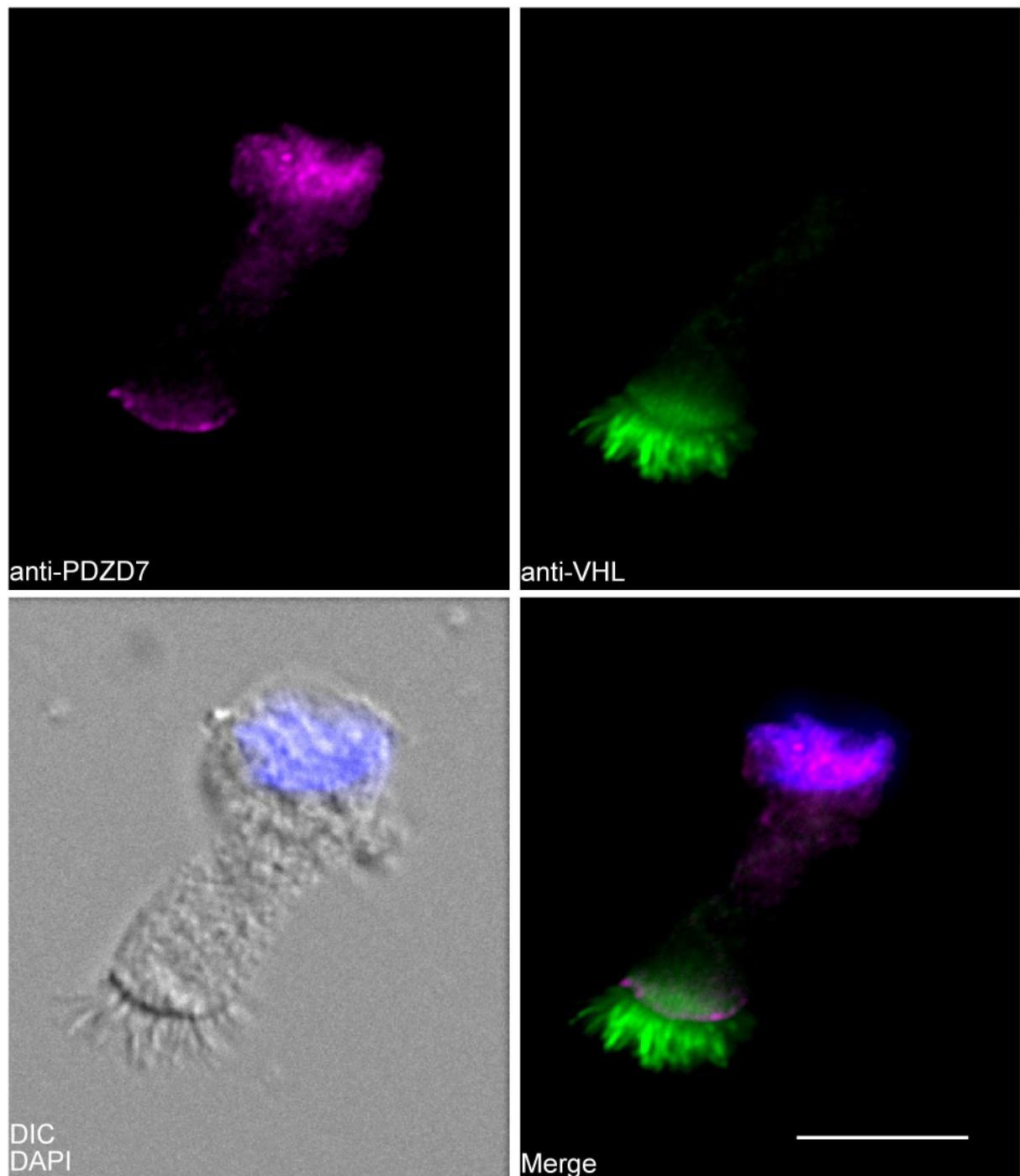
Samples were taken after 30 cycles. Bands were verified by sequencing as *PDZD7* amplicons. Amplification was carried out with primers located in **(A)** exon 1/2, **(B)** exons 10/14, and **(C)** exons 11/14. **(D)** The same cDNA samples were used as a template for PCR with primers for a housekeeping gene (*G3PDH*). *Unspecific PCR product: Pancreatic carboxypeptidase A1, CPA1 (7q32.2). **(E)** Expression of *pdzd7* in the mouse cochlea. 1: mouse *pdzd7* primers c.11F (5'-GTACTAGCAGTCACTCGCCAC-3') vs. c.15R (5'-CCTGCGCCCAGGAGACTTGCCTG-3'), 657 bp. 2: water control. 3: primers for a housekeeping gene (*G3PDH*). 4: water control. Lanes 1/2 and 3/4 were run on the same gel but noncontiguous.

A**B**

Supplemental Figure 4

A novel polyclonal antibody was raised against the newly predicted C-term of PDZD7.

(A) To prove specificity of the antibody, HEK293T cells were co-transfected with FLAG.Podocin and either FLAG.PDZD7³¹⁴⁻⁵⁰³ or FLAG.PDZD7⁷⁶²⁻¹⁰¹². Western blot analysis of cell lysates revealed that the antibody only recognizes the targeted protein truncation but neither FLAG.Podocin nor a control truncation of PDZD7. Comparable protein expression was proven by an anti-FLAG staining of the same lysates. (B) For further validation of the antibody, HEK293T cells were transfected with FLAG.PDZD7⁷⁶²⁻¹⁰¹². This PDZD7 truncation could be detected with either an anti-FLAG or an anti-PDZD7 antibody, resulting in an overlapping staining pattern. Scale bar: 10 µm.



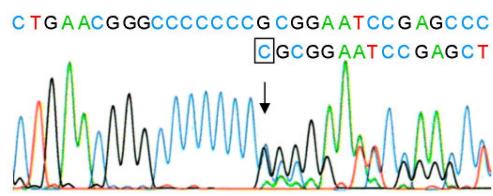
Supplemental Figure 5

PDZD7 localizes to the ciliary base.

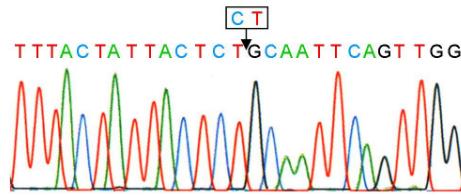
PDZD7 staining of human nasal cells demonstrates localization of PDZD7 at the ciliary base and at the nucleus. Cilia were visualized with an anti-VHL antibody, nuclei with DAPI. Scale bar: 10 μm .

FCa

PDZD7: c.166_167insC (p.R56PfsX24)

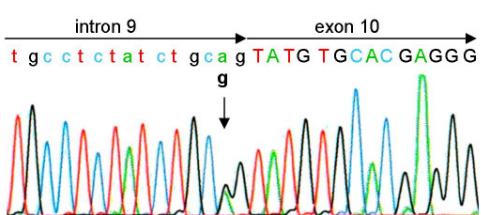


USH2A: c.4338_4339delCT (p.C1447QfsX29)

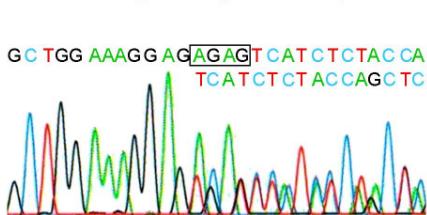


GER1

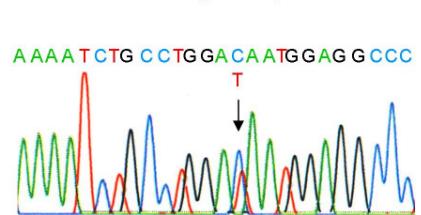
PDZD7: c.1750-2A>G



USH2A: c.4515_4518del (p.R1505SfsX7)

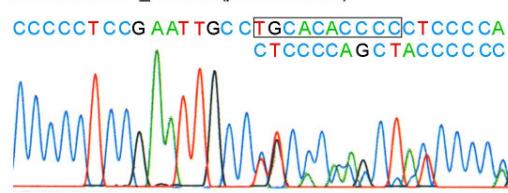


USH2A: c.13316C>T (p.T4439I)

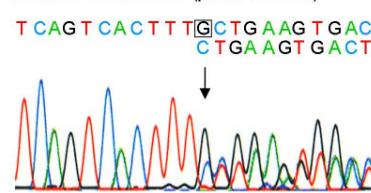


GER2

PDZD7: c.2194_2203del (p.C732LfsX18)

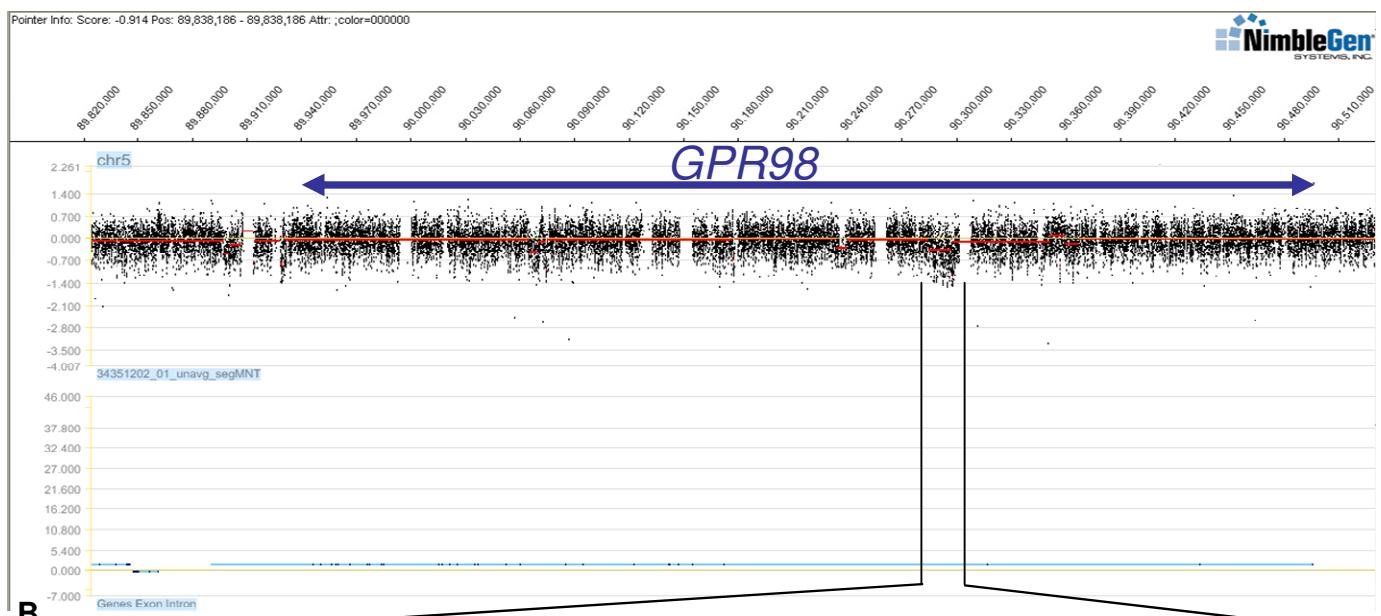
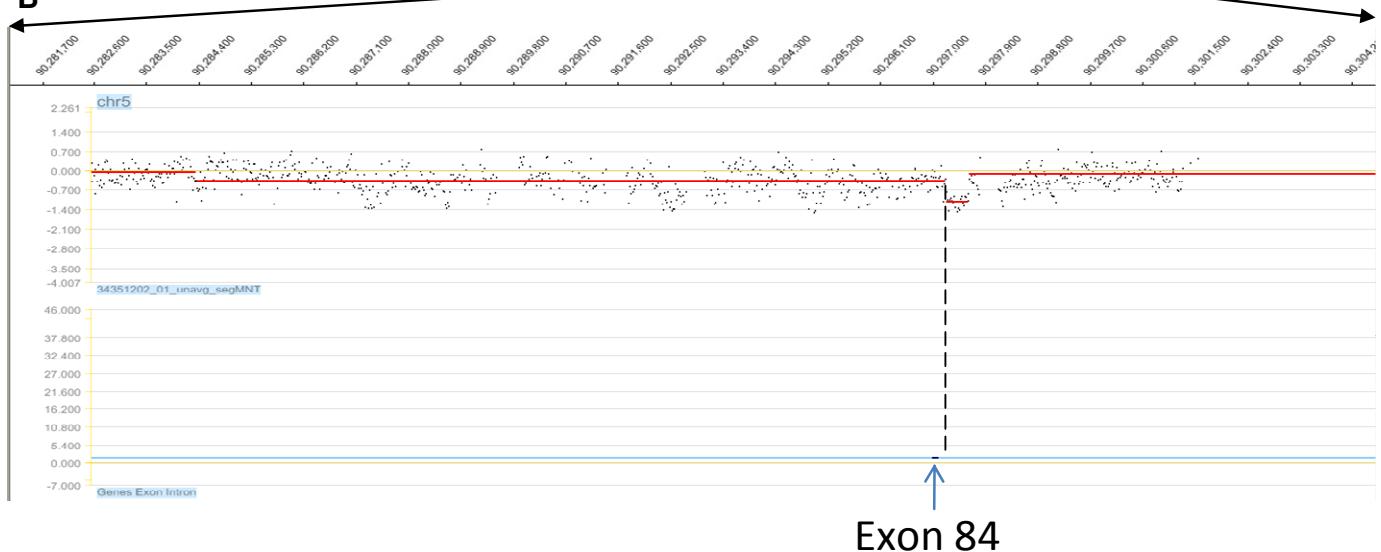


GPR98: c.17137delG (p.A5713LfsX3)



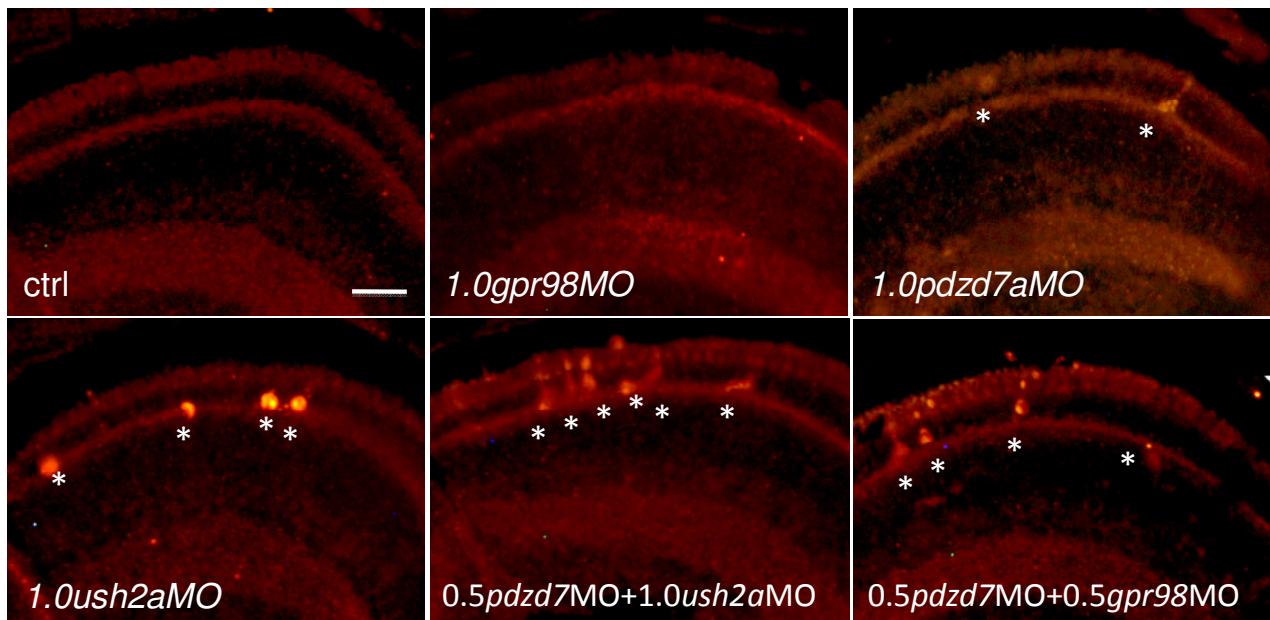
Supplemental Figure 6

Electropherograms of *PDZD7*, *USH2A* and *GPR98* mutations identified in this study.

A**B**

Supplemental Figure 7

Exclusion of large deletions or duplications affecting coding parts of *GPR98* by high-resolution array CGH using a customized NimbleGen array (black dots indicate oligonucleotide probes that densely cover the genomic *GPR98* region). Array CGH with a 6.0 Affymetrix SNP array likewise did not indicate any potentially deleterious copy number variation (not shown). **(A)** Overview of the array CGH data from the NimbleGen customized array. **(B)** Enlargement of a region with a questionable 12 kb deletion that would encompass *GPR98* exon 84. PCR experiments based on multiple primer combinations did not amplify junction fragments that would be expected if the 12 kb deletion was present.



Supplemental Figure 8

Anti-active Caspase-3 labeling of control and morpholino-treated retinas.

Exon	Forward primer	Reverse primer	Coding sequence (bp)	Fragment size (bp)
1	ggagctcacacacttctgagaggc	cctgccaccccccgtgactc	226	520
2	cttagaaatgggctgaccgc	gtccctgacagcagcatcc	141	384
3	gctctgacaatcctcaactcg	ctctccttaatttgaggtcag	175	453
4+5	ccagctaatacgAACCTATGC	ctacccTTCTAGTGGCTGTG	177/148	858
6	gcatgatgaattggaggac	ctgtactcaaggcgatcctc	61	341
7	cagcattcggagcagcggg	gtctgagcagcctggagcttg	396	694
8	ggaagaggaaccgcactctcac	ctcatgaggaaactgaggctc	198	489
9	cctcaggGCCAGGCCTTG	cataccattctagctgcctg	51	330
10	ggagcatataggctaggctag	cactagccctCTGGTGTCAAG	176	414
11-13	gaaccttccaaggcacgcag	gtggacaagagcttcccttc	92/92/72	848
14.1	gcgattctaatacAACATGTC	cctggagacttgcctgac	612	511
14.2	ctggctgctgacagaACCC	ggtgtgagcctctgcgcctg		471
15+16	gagtagtaacagaatgtagg	gatgctggagtcaGTTGGTG	101/384	1058

Supplemental Table 1

Primers for *PDZD7* mutation screening.

cDNA	Forward primer	Reverse primer	Amplicon (bp)
human			
1 – 763	ATGGCGCAGGGTTCGCAGTG	CCACCTTGATGCCATTCTCCTCG	763
512 – 1899	GCATCAAGTTCTCCAAGGAG	CTCCACAAGCATCACCATGC	1388
1753 – 3102+41 (3'-UTR)	GTGCACGAGGGAGGCATAGAGG	CACCCACTGACTCCAGCATC	1391
1391 – Alu in intron 8	CCAAGACGCTGATGAACCTC	GCTGGTCTTAAACTCCTGG	218/ 386
zebrafish			
100 bp of 5'-UTR – 247	CTCTGGACTGTAATCCTGCTAG	GTGAGGATGGCCATCTGG	347
1–646	ATGGCTCATTCTGCTGACACGG	CTTGCCGCCACGGATGTTGAAG	646
175–938	CCTGTCGATGGTGGAGATGAC	GTGGAGGAGTATGAATCTGAGC	764
757–1248	GACATCACACACAGCAATGCAG	CAGAAACTAGTCGAATGGTAGG	413
1209–1738	GACAGCCCATTCCGGACAAGG	CGACCAGAACCTTTACAGTGTG	530
1620–2237	GGAGCAAAGCCCGATGCCTAGC	CGTCCAGTGCCTGCTG	619
2167–2660	CGCTCTTGAGTCCTGCTCG	CTGCCCTTCAGGACATCAC	494
784–1930	TGGCGGCCTGGCAGAACAAATG	GGGGCGCACTAGATCCTCAACCAC	1146
1989–2907	TCATT CCTGCCACAGATCTGG	CTGAAAGCCCTCCGAATGG	918
1989–3'UTR	TCATT CCTGCCACAGATCTGG	CTTCGCTGTGAGTAAATCAGG	1033

Supplemental Table 2

Primers for cloning human full-length *PDZD7* and the isoform derived from intron 8 Alu insertion.

Position	Exon/ IVS	Change	Frequency in patients	Rel. frequency in patients	Frequency in controls	Rel. frequency in controls	Rel frequency total	rs number; carrier(s)
c.1-86A>G	IVS 0	-	27/474	0.0570	21/508	0.0413	0,0479	
c.159G>C	exon 1	p.G59G	1/474	0.0021	0/508	0	0,0010	1 x USH1
c.367+7A>G	IVS 2	-	153/474	0.3228	n. d.	n. d.	n. d.	rs6584410
c.367+51delT	IVS 2	-	3/452	0.0133	n. d.	n. d.	n. d.	
c.368-54A>G	IVS 2	-	6/460	0.0130	n. d.	n. d.	n. d.	
c.542+48C>G	IVS 3	-	48/446	0.1076	n. d.	n. d.	n. d.	rs3740496
c.539C>T	exon 3	p.T180M	1/474	0.0021	n. d.	n. d.	n. d.	1 x USH2
c.559C>T	exon 4	p.R187W	1/474	0.0021	n. d.	n. d.	n. d.	1 x USH2D (Ebermann et al., 2007)
c.572T>A	exon 4	p.V191E	7/474	0.0148	n. d.	n. d.	n. d.	
c.719+136C>A	IVS4	-	9/474	0.0190	n. d.	n. d.	n. d.	rs41291480
c.928+20delC	IVS 6	-	254/474	0.5359	n. d.	n. d.	n. d.	
c.928+63A>C	IVS 6	-	90/248	0.3629	n. d.	n. d.	n. d.	rs7075685
c.928+110C>T	IVS 6	-	117/248	0.4718	n. d.	n. d.	n. d.	rs7075659
c.936C>T	exon 7	p.N312N	8/474	0.0169	n. d.	n. d.	n. d.	rs35038258
c.1008C>T	exon 7	p.P336P	1/474	0.0021	n. d.	n. d.	n. d.	1 x USH2
c.1011C>T	exon 7	p.Y337Y	10/474	0.0211	n. d.	n. d.	n. d.	rs34705415
c.1388G>C	exon 8	p.R463P	13/474	0.0274	n. d.	n. d.	n. d.	
c.1522+88G>C	IVS 8	-	168/438	0.3836	n. d.	n. d.	n. d.	rs11190793
c.1613G>A	exon 10	-	6/474	0.0127	n. d.	n. d.	n. d.	
c.1749+43delA	IVS 10	-	253/468	0.5406	n. d.	n. d.	n. d.	rs34125357
c.1934-55C>T	IVS 12	-	88/474	0.1857	n. d.	n. d.	n. d.	rs807020
c.1934-40C>G	IVS 12	-	1/514	0.0019	n. d.	n. d.	n. d.	1 x USH2
c.2006-79A>G	IVS 13	-	1/454	0.0022	8/852	0.0069	0,0074	
c.2006-26_37ins17bp	IVS 13	-	8/474	0.0169	23/852	0.0234	0.0263	
c.2011C>A	exon 14	p.R671S	1/474	0.0021	0/808	0	0.0001	1 x USH2
c.2049G>A	exon 14	p.P683P	8/474	0.0169	3/808	0.0037	0.0086	rs34693310
c.2132A>G	exon 14	p.H711R	10/474	0.0211	2/808	0.0025	0.0094	rs34616847
c.2144C>T	exon 14	p.P715L	4/474	0.0084	7/808	0.0087	0.0086	
c.2250G>T	exon 14	p.W750C	1/474	0.0021	4/808	0.0050	0.0039	
c.2319C>T	exon 14	p.R773R	88/474	0.1857	127/808	0.1572	0.1677	rs807022
c.2340_2341ins6bp	exon 14	p.S780_R781insRS	249/474	0.5253	363/808	0.4493	0.4773	
c.2368A>G	exon 14	p.K790Q	7/474	0.0148	n. d.	n. d.	n. d.	
c.2564C>A	exon 14	p.T855N	82/474	0.1730	n. d.	n. d.	n. d.	rs807023
c.2718+146G>T	IVS 15	-	1/474	0.0021	n. d.	n. d.	n. d.	1x USH2
c.3092G>A	exon 16	p.R1031H	30/474	0.0633	n. d.	n. d.	n. d.	

Supplemental Table 3

Non-pathogenic variations in *PDZD7*.

237 patients (474 alleles) were analyzed. Differences in control numbers are due to heterozygosity of deletion or insertion polymorphisms upstream of SNP positions or otherwise illegible sequence at the respective position. For variants detected only once, the USH subtype of the patient is given in the last column.