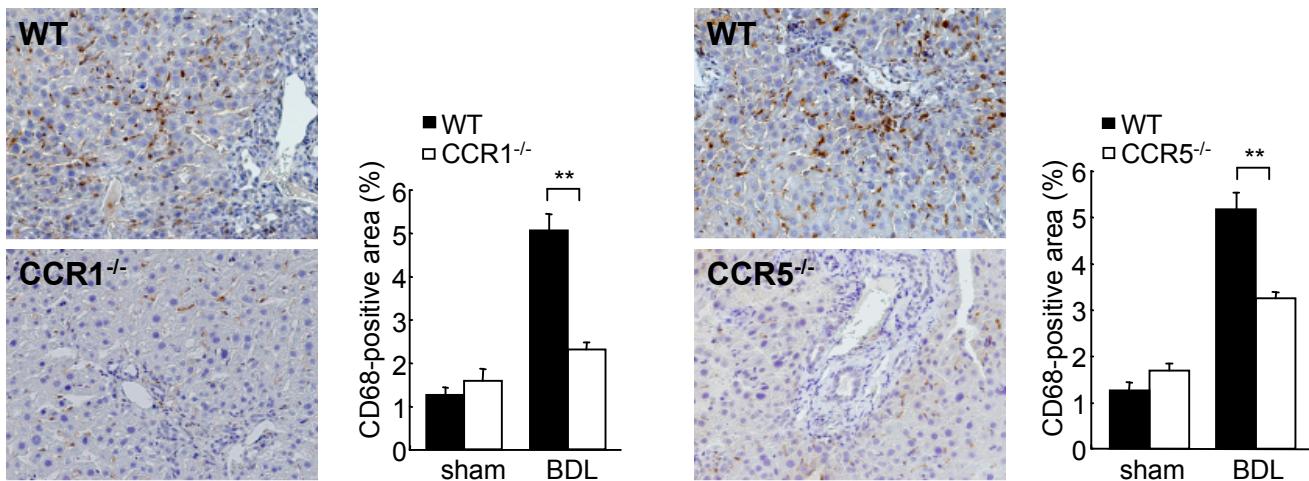
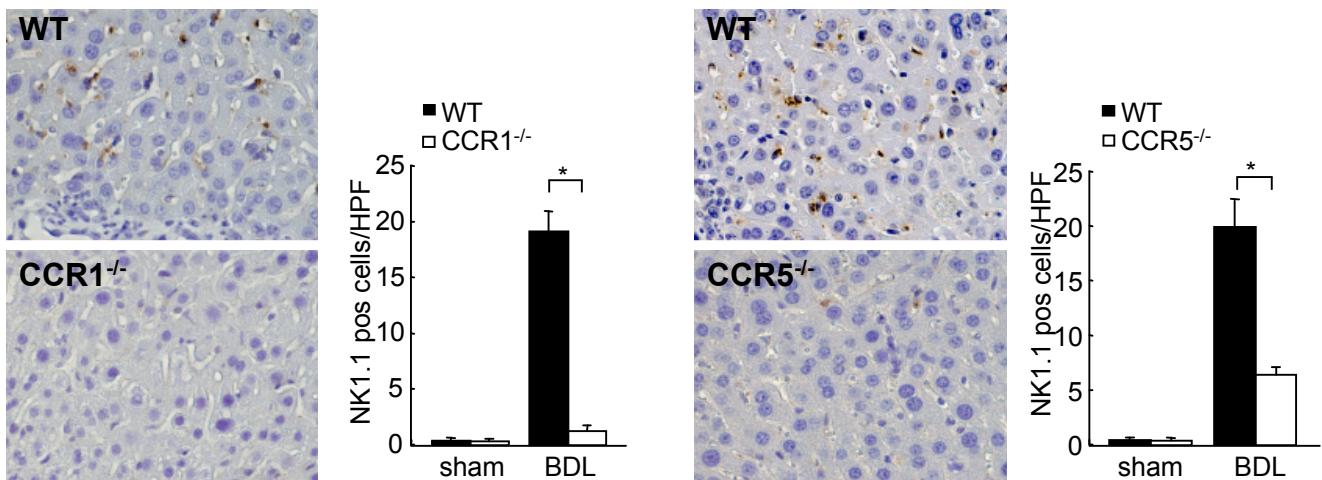
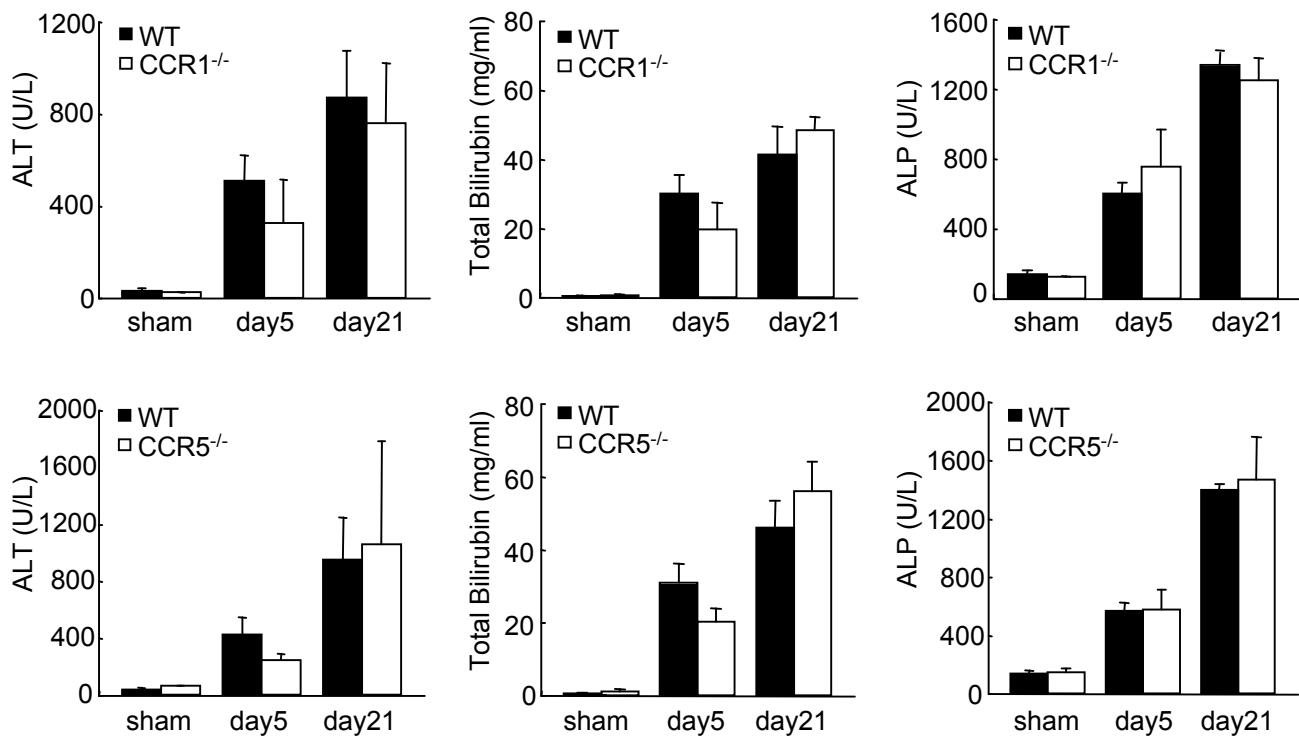
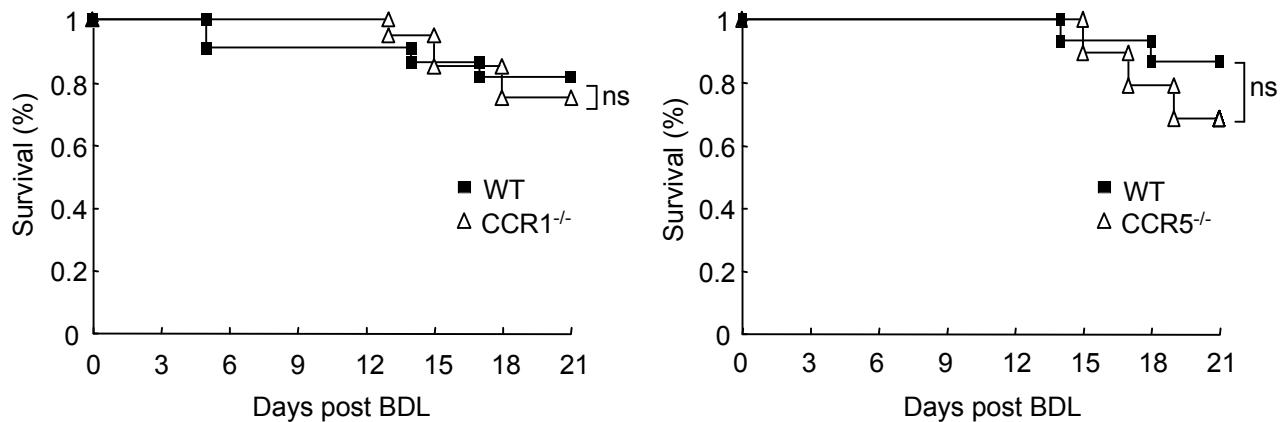


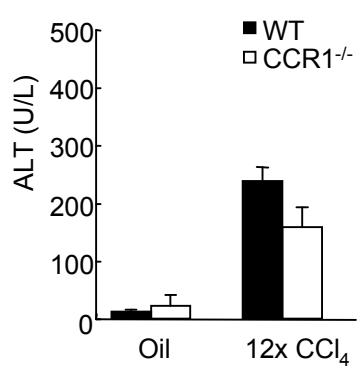
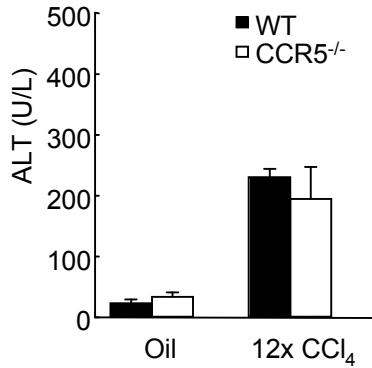
**Supplementary Figure 1. Expression of CCR1 and CCR5 in CCl<sub>4</sub>-induced liver fibrosis.** Mice underwent 12 injections of CCl<sub>4</sub> or oil. Livers were stained for CCR1 (green fluorescence, upper panel), CCR5 (green fluorescence, lower panel), F4/80, desmin, CD31 or panCK (all red fluorescence) followed by confocal microscopy.

**A.****B.**

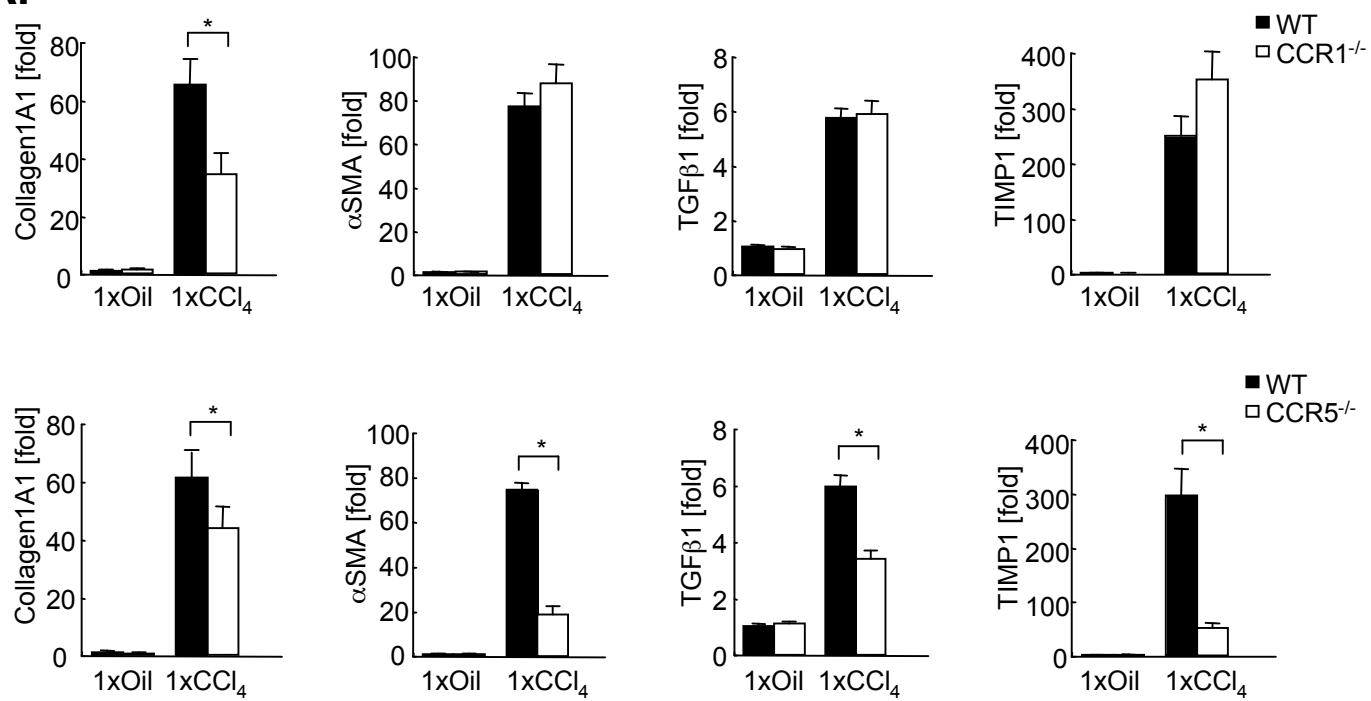
**Supplementary Figure 2. Inflammatory cell recruitment in CCR5- and CCR1-deficient mice after BDL.** A. Wild-type, CCR1<sup>-/-</sup> and CCR5<sup>-/-</sup> mice underwent BDL for 21 day followed by immunohistochemistry for CD68 (A) or NK1.1 (B). \*p<0.05, \*\*p<0.01

**A.****B.**

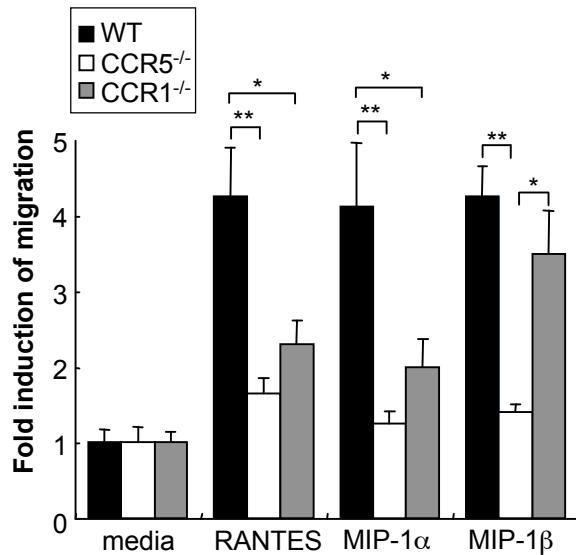
**Supplementary Figure 3. CCR1- and CCR5-deficient mice do not display increased liver injury or decreased survival after BDL.** A. CCR1-deficient (n=4) and wild-type mice (n=5) as well as CCR1-deficient (n=6) and wild-type controls (n=6) underwent BDL for 5 or 21 days followed by measurement of serum ALT, total bilirubin and alkaline phosphatase (ALP). B. Survival of CCR1<sup>-/-</sup> (n=15) and wild-type mice (n=22), as well as CCR5<sup>-/-</sup> (n=19) and wild-type mice (n=15) after BDL.

**A.****B.**

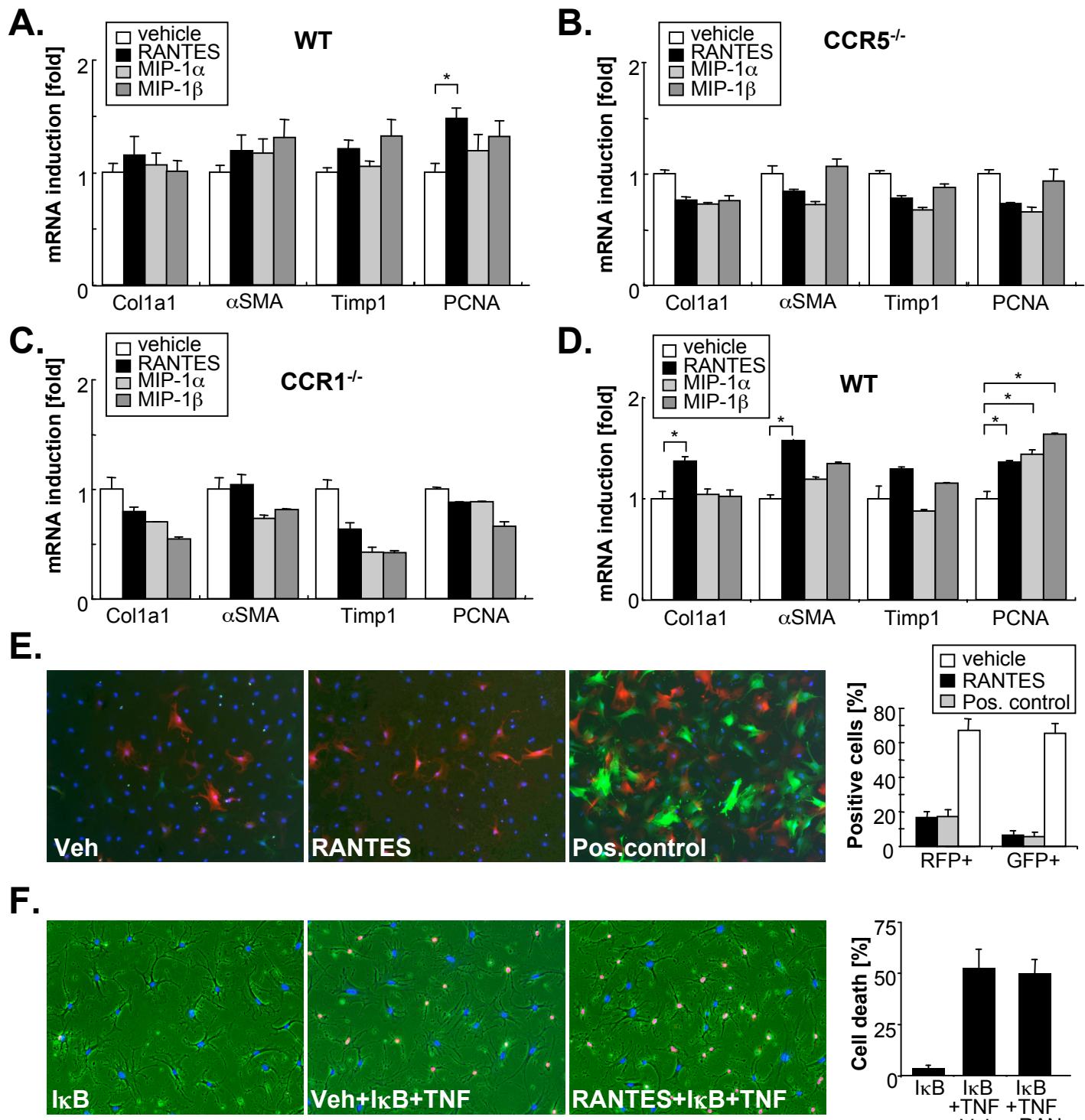
**Supplementary Figure 4. CCR1- and CCR5-deficient mice do not display increased liver injury after CCl<sub>4</sub>.** A. CCR1-deficient (n=5) and wild-type mice (n=5) were treated with 12 CCl<sub>4</sub>-injections followed by measurement of serum ALT levels. B. CCR5-deficient (n=5) and wild-type controls (n=5) were treated with 12 CCl<sub>4</sub>-injections followed by measurement of serum ALT levels.

**A.**

**Supplementary Figure 5. mRNA expression of fibrosis related genes in CCR1- and CCR5-deficient mice after CCl<sub>4</sub> treatment.** A-B. CCR1-deficient mice (n=4) and isogenic wildtype controls (n=4) as well as CCR5-deficient mice (n=4) and wild-type littermates (n=4) were treated with 1 injection of CCl<sub>4</sub> followed by quantitative real-time PCR for Colla1,  $\alpha$ SMA, TGF $\beta$ 1 and TIMP1. \*p<0.05



**Supplementary Figure 6. CCR5 and CCR1 induce Kupffer cell migration.** A. Kupffer cells from wild-type, CCR5- and CCR1-deficient mice were placed in a Boyden chamber and migration through an 8 micron filter was determined after stimulation with RANTES, MIP-1 $\alpha$  or MIP-1 $\beta$  (all 50 ng/ml). \* p<0.05 , \*\*p<0.01



**Supplementary Figure 7. CCR5 and CCR1 do not induce HSC activation and do not prevent cell death.** A-D. HSCs were isolated from wild-type, CCR5- and CCR1-deficient mice. HSCs were stimulated with RANTES, MIP-1 $\alpha$  or MIP-1 $\beta$  (all 100 ng/ml) or vehicle (0.1% BSA) for 24 hours (A-C) or 5 days (D). Following RNA extraction and reverse transcription, collagen  $\alpha$ 1(I),  $\alpha$ SMA, TIMP1 and PCNA mRNA levels were measured by quantitative real-time PCR. E. HSCs were isolated from double-transgenic mice expressing GFP under the collagen  $\alpha$ 1(I) promoter and RFP under the  $\alpha$ SMA promoter, and treated for five days with either vehicle (0.1% BSA) or RANTES (100 ng/ml). GFP and RFP expression were evaluated by fluorescent microscopy and quantified. F. Culture-activated HSCs were infected with AdIkBsr, followed by pretreatment with RANTES (100 ng/ml) or vehicle for 12h, and treatment with TNF $\alpha$  (30 ng/ml) for 8h. Cell death was determined by propidium iodide staining, and normalization to nuclei stained by cell-permeable Hoechst. \* p<0.05