

Pin1 has opposite effects on wild-type and P301L tau stability and tauopathy

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Tau pathology is a hallmark of many neurodegenerative diseases including Alzheimer disease (AD) and frontotemporal dementia with Parkinsonism linked to chromosome 17 (FTDP-17). Genetic tau mutations can cause FTDP-17, and mice overexpressing tau mutants such as P301L tau are used as AD models. However, since no tau mutations are found in AD, it remains unclear how appropriate tau mutant mice are as an AD model. The prolyl isomerase Pin1 binds and isomerizes tau and has been implicated in protecting against neurodegeneration, but whether such Pin1 regulation is affected by tau mutations is unknown. Consistent with earlier findings that Pin1 KO induces tauopathy, here we demonstrate that Pin1 knockdown or KO increased WT tau protein stability in vitro and in mice and that Pin1 overexpression suppressed the tauopathy phenotype in WT tau transgenic mice. Unexpectedly, Pin1 knockdown or KO decreased P301L tau protein stability and abolished its robust tauopathy phenotype in mice. In contrast, Pin1 overexpression exacerbated the tauopathy phenotype in P301L tau mice. Thus, Pin1 has opposite effects on the tauopathy phenotype depending on whether the tau is WT or a P301L mutant, indicating the need for disease-specific therapies for tauopathies.

Introduction

Tau-related phenotypes are prominent common features of many neurodegenerative diseases, called tauopathies, that include Alzheimer disease (AD), frontotemporal dementia with Parkinsonism linked to chromosome 17 (FTDP-17), corticobasal degeneration, progressive supranuclear palsy, and Pick disease (reviewed in refs. 1–3). Tau binds microtubules and promotes their polymerization, thereby playing a crucial role in maintaining the physiological function of healthy neurons (4). Phosphorylation is a key regulatory mechanism that disrupts the ability of tau to bind microtubules and to promote their assembly (4). Dynamic tau phosphorylation occurs during embryonic development and cell division and to some extent in postmitotic neurons and may play important roles in refining or maintaining neuronal structure and function (5). However, aberrant tau phosphorylation in mature neurons is believed to be harmful to the neuron (6). Although it has been debated whether tau hyperphosphorylation and aggregation in the neuron are the cause or the effect of the tauopathies, recent identification of tau genetic mutations in FTDP-17 patients has provided evidence that tau dysfunction can cause the tauopathy phenotype (1–3).

While not found in AD, genetic tau mutations have been identified in FTDP-17 patients (1–3). Moreover, Tg mice overexpressing human P301L tau, one of the most frequent FTDP-17 tau mutants, develop the robust tauopathy phenotype and thus are widely used as models of AD by itself or in combination with A β -related mouse models (7–14). There have been various studies comparing WT tau and P301L mutant tau to better understand their differences

in binding to microtubules or regulatory enzymes, forming tau filaments or causing neurotoxicity (1–3). However, it is not fully clear what impact these mutations have on tau regulation, and it is unclear how appropriate such tau mutant mice are as AD models.

Characterization of Tg mice overexpressing tau or its mutants reveals a major impact of tau protein levels on tau-related phenotypes (7–9, 15–21). For example, low levels affect tau compartmentalization and its phosphorylation, similar to the pathological changes characteristic of the so-called pretangle stage of AD (16, 17). However, high levels can also cause age-dependent neurodegenerative phenotypes (18, 20), as occurs in Tg mice overexpressing tau mutants, which are usually expressed at much higher levels (7, 19, 22). A key factor in the development of tau-related phenotypes appears to be tau phosphorylation, especially on Ser or Thr residues preceding a Pro (Ser/Thr-Pro) by Pro-directed kinases (23, 24) or phosphatases such as PP2A (25, 26). Indeed, overexpression of GSK3 β or CDK5 activator p25 or inhibition of PP2A enhances or induces tau-related phenotypes in mice (11, 27–30). In addition, tau phosphorylation has been shown to promote tau degradation via chaperone-interacting protein/Hsp70 (31–33) or to inhibit tau degradation (34). These results indicate that tau phosphorylation is intricately regulated, but nothing has been known of whether tau is further regulated between phosphorylation and dephosphorylation until recently.

We have previously shown that specific pSer/Thr-Pro motifs in certain proteins such as tau can exist in 2 distinct *cis* and *trans* conformations, whose conversion can be greatly accelerated by a unique prolyl isomerase, Pin1 (protein interacting with NIMA 1) (35–38). Subsequent studies indicate that such Pin1-catalyzed conformational changes, which can now be visualized by NMR (39), have a profound impact on phosphorylation signaling, achieved by regulating a spectrum of target activities (40–42). Importantly, Pin1 is tightly regulated and its deregulation contributes to pathological conditions, notably AD and cancer (39–45).

Nonstandard abbreviations used: AD, Alzheimer disease; APP, amyloid precursor protein; FTDP-17, frontotemporal dementia with Parkinsonism linked to chromosome 17; KD, knockdown; NFT, neurofibrillary tangles; Pin1, protein interacting with NIMA 1.

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In AD, Pin1 binds to and isomerizes the pThr231-Pro motif in tau to restore its ability to bind microtubules and to promote their assembly (46) as well as to facilitate tau dephosphorylation by PP2A since the phosphatase dephosphorylates only the *trans* pSer/Thr-Pro motifs (47, 48). Pin1 also binds to and isomerizes the pThr668-Pro motif in amyloid precursor protein (APP) to promote nonamyloidogenic APP processing and to reduce A β production (39). Therefore, Pin1 acts on both tau and APP, 2 key molecules in AD. Importantly, Pin1 expression is induced during neuronal differentiation and is highly expressed in most neurons in the brain (46, 48, 49). However, in the AD neuron, Pin1 is downregulated or inactivated by oxidative modifications, promoter polymorphisms, or other mechanisms (48, 50–52). Importantly, ablation of the *Pin1* gene alone (Pin1 KO, *Pin1*^{-/-}) in mice is sufficient to induce tau- and A β -related phenotypes and neurodegeneration in an age-dependent manner (39, 48). These results indicate that loss of Pin1 function contributes to the development of the tauopathy phenotype in AD (42). However, it remains to be determined whether Pin1 overexpression can inhibit tau-related phenotypes in AD and whether Pin1 regulates FTDP-17 tau mutants in the same way as the WT protein. Answers to these questions are important because they may facilitate the development of new therapies for AD and related disorders.

In this paper, we investigated the role of Pin1 in the development of tau-related phenotypes induced by WT tau and its P301L tau mutant both in cell cultures and animal models and surprisingly uncovered the diametrically opposite impacts of Pin1 on tau protein stability and tauopathy phenotype depending on whether the tau is WT or P301L mutant. These results indicate the importance of using relevant animal models for studying AD and FTDP-17 and also suggest that Pin1 upregulation might be beneficial for AD, whereas its inhibition might be helpful for patients carrying the P301L tau mutation.

Results

Pin1 knockdown in cells affects protein stability of WT tau and P301L tau in opposite manners via acting on tau proteins. Pin1 binds to and isomerizes the pThr231-Pro motif in tau to regulate tau dephosphorylation and its microtubule-binding activity (46–49). Interestingly, it has been shown that tau phosphorylation regulates its degradation (31–34) and that protein degradation is modulated by Pin1 (42). These results suggest that Pin1 might act on the pThr231-Pro motif in WT tau and P301L tau to regulate their protein stability since the P301L mutation does not affect tau binding to Pin1 (46).

To examine this possibility, we used retroviruses or lentiviruses carrying 2 different Pin1 RNAi constructs to stably deplete Pin1 in 2 different human cell lines, HT1080 fibrosarcoma cells and SH-SY5Y neuroblastoma cells, as described (53). Both RNAi constructs efficiently depleted Pin1 protein in both cell lines (Figure 1 and data not shown). To determine whether Pin1 knockdown (KD) affects protein stability of WT tau and P301L tau, Pin1 KD and control cells were cotransfected with tau or its P301L mutant expression construct overnight, followed by the addition of cycloheximide to inhibit *de novo* tau synthesis, followed by determination of the steady-state levels of tau at various times, as described (54, 55). Under this cycloheximide chase analysis, the rate at which tau steady-state levels decline reflects the half-life of the proteins. In control SY5Y cells and HT1080 cells, WT tau was unstable but became highly stable in both types of Pin1 KD cells (Figure 1, A and B, and data not shown). In contrast, P301L tau was highly unstable in Pin1 KD cells, as compared with control cells, where it was much stabler (Figure 1, C and D, and data not shown). Importantly, these effects of Pin1 KD on

tau protein stability were completely abolished in the presence of the proteasome inhibitor MG132, where both WT tau and P301L tau were stable and their levels were not affected by Pin1 KD (Figure 1, A–D). These results demonstrate that Pin1 KD has opposite impacts on protein stability of WT tau and P301L tau both in nonneuronal and neuronal cells and that the impact depends on functional proteasome, as shown for other Pin1 substrates (42).

To exclude off-target effects of Pin1 KD, we transfected Pin1 RNAi stable SY5Y cells with a vector expressing the Pin1 coding sequence because the Pin1 RNAi sequence is directed against the 3' untranslated region about 600 bp downstream of the stop codon. As expected, the cells reexpressed Pin1 (Figure 1). Importantly, Pin1 addback completely rescued the opposite effects of Pin1 KD on protein stability of WT tau and P301L tau; it rendered WT tau unstable but P301L tau as stable in Pin1 KD cells (Figure 1, A–D). These results indicate that the observed effects on tau protein stability are specifically due to depletion of Pin1.

To determine whether the effects of Pin1 KD on tau stability depend on the presence of the Pin1-binding site in tau proteins, we used the Thr231Ala mutation, which abolishes the ability of Pin1 to act on tau (46, 47). Protein stability of both tau-T231A and P301L tau-T231A mutant was affected neither by Pin1 KD nor by reexpression of Pin1 in Pin1 KD cells (Figure 1, E–H). These results indicate that the effects of Pin1 on both WT tau and P301L depend on the presence of the Pin1-binding site in tau, as shown for other Pin1 targets (42).

To determine whether Pin1 KO has any effects on protein stability of other FTDP-17 tau mutants, we selected 3 other commonly studied FTDP-17 tau point mutants, P301S, V337M, and R406W (56–58). Similar to P301L tau (Figure 1, C and D, and Figure 2, A and B), P301S tau was less stable in Pin1 KD cells than in control cells (Figure 2, C and D). In contrast, both V337M tau and R406W tau were more stable in Pin1 KD cells than in control cells (Figure 2, E–H), as seen in WT tau (Figure 1, A and B). Although further experiments are needed to examine other FTDP-17 tau mutants, these studies suggest that the effects of Pin1 on FTDP-17 tau mutants likely depend on the identity of the tau mutations. Given that P301L tau probably is the most well-studied FTDP tau mutant, especially with widely used Tg mouse models (7–14), we will here focus our study on comparing the impact of Pin1 on WT tau and P301L tau.

Pin1 KO affects protein stability of WT tau and P301L tau in primary neurons and in brain tissues in an opposite manner. To confirm the above surprising observations, we examined the effects of Pin1 KO on protein stability of WT tau and P301L tau using primary neuron cultures derived from Pin1 WT or KO mice and using organotypic brain-slice cultures derived from bigenic mice generated from crosses between Pin1 KO mice and WT tau or P301L Tg mice.

To determine the effects of Pin1 KO on protein stability of WT tau and P301L tau in primary neurons, we established cortical neuron cultures derived from *Pin1*^{+/+} and *Pin1*^{-/-} embryos at day E17.5 and infected them with lentiviruses expressing WT tau or P301L tau, followed by cycloheximide chase analysis to determine their half-lives. To distinguish exogenous tau proteins from the endogenous protein, we inserted an N-terminal His tag to WT tau and P301L tau. As compared with the corresponding control in control *Pin1*^{+/+} neurons, transfected WT tau was significantly more stable, whereas transfected P301L tau was much less stable in *Pin1*^{-/-} neurons (data not shown). These results demonstrate that Pin1 KO has the opposite effects on protein stability of WT and P301L tau in primary neuron cultures.

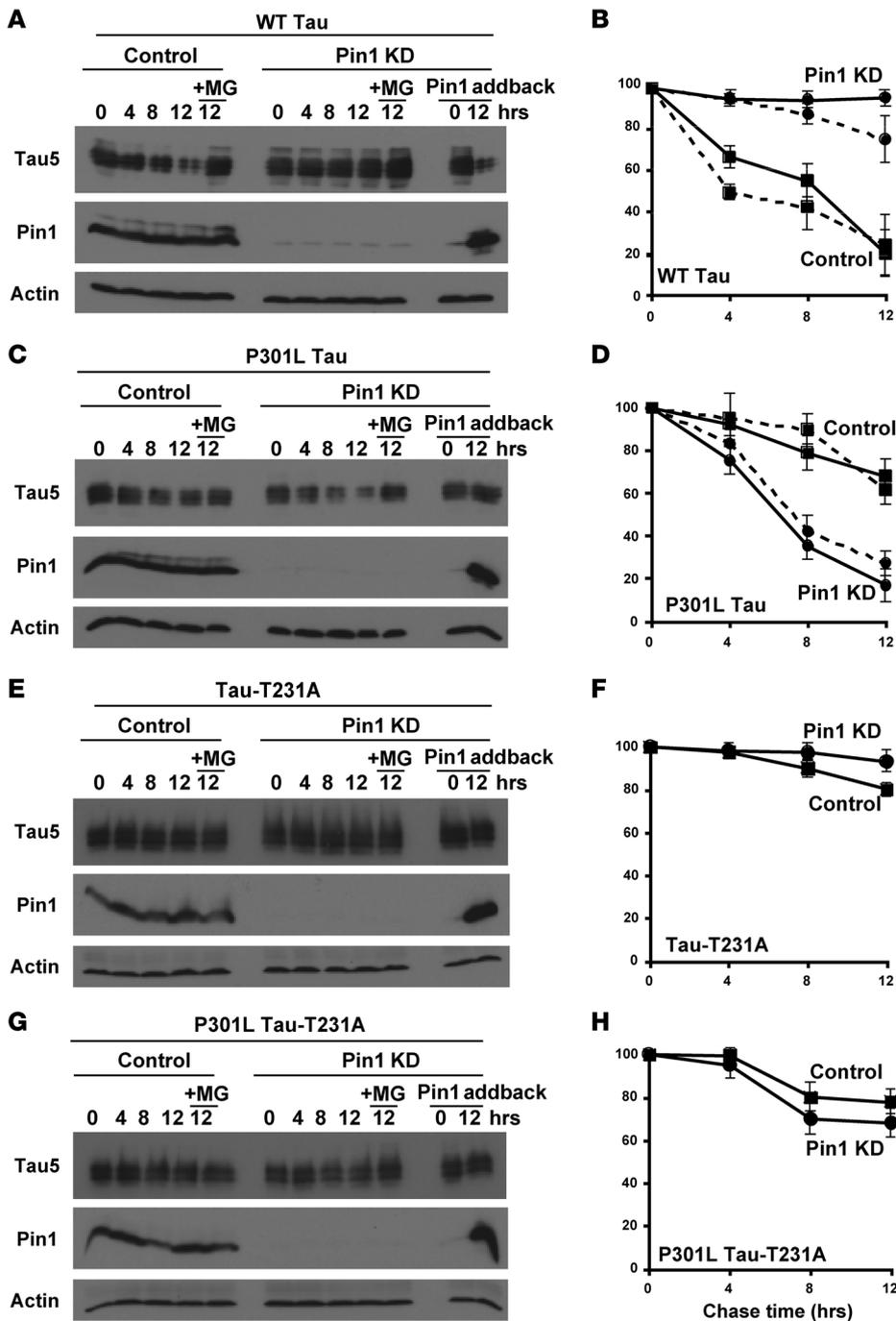


Figure 1

Pin1 KD in cells has opposite effects on the protein stability of WT tau and P301L tau; these effects depend on the presence of the Pin1-binding site in tau proteins. SH-SY5Y cells were infected with Pin1 siRNA or control siRNA lentiviruses, followed by selection for stable cells with puromycin. Stable Pin1 KD or control SY5Y cells were transfected overnight with a WT tau expression construct in the presence or absence of a Pin1 expression construct (+Pin1 addback) (A and B), with a P301L tau construct in the presence or absence of a Pin1 construct (C and D), with a tau-T231A construct in the presence or absence of a Pin1 construct (E and F), or with a P301L tau-T231A construct in the presence or absence of a Pin1 construct (G and H). Cycloheximide was added to stop new protein synthesis and chased for indicated times in the absence or presence of the proteasome inhibitor MG132. Cells were harvested and cell lysates were fractionated by SDS-PAGE and analyzed by immunoblotting with Tau5, anti-Pin1, or anti-actin antibodies (A, C, E, and G). Tau levels were semiquantified from 3 different experiments using ImageQuant and normalized using actin as an internal control (B, D, F, and H). For comparison, tau levels at 0 (before the cycloheximide addition) were defined as 100%. The opposite effects of Pin1 KD on protein stability of WT tau and P301L tau were also obtained in HT1080 cells, as shown by dashed lines in B and D. Error bars represent SD.

To examine the effects of Pin1 KO on protein stability of WT tau and P301L tau in brain tissues, we established brain-slice cultures from WT tau or Tg P301L tau mice in the presence or absence of Pin1, followed by cycloheximide chase analysis to examine the half-lives of WT tau and P301L tau in brain tissues. Tg WT tau had a half-life of approximately 12 hours in the presence of Pin1 but became almost completely stable in the absence of Pin1 (Figure 3, A and B). In contrast, the half-life of Tg P301L tau was significantly decreased to approximately 6 hours in the absence of Pin1 from approximately 24 hours in the presence of Pin1 (Figure 3, C and D). These results convincingly demonstrate that Pin1 has opposing effects on protein stability of WT tau and P301L tau

even in brain-slice cultures, a model that maintains the 3D architecture and synaptic connections of the brain closely resembling the in vivo situation. Although further experiments are needed to elucidate how Pin1 regulates tau protein stability and how P301L mutation of tau causes Pin1 regulation to move in the opposite direction, we decided to focus our efforts on determining the significance of this Pin1-dependent regulation, which we believe is novel, in the development of tau-related phenotypes induced by WT tau and P301L tau in vivo because it might have important therapeutic implications for treating human tauopathies.

Pin1 KO abrogates tau-related phenotypes in P301L tau Tg mice. The above findings that Pin1 KD or KO in cell cultures and in brain-slice

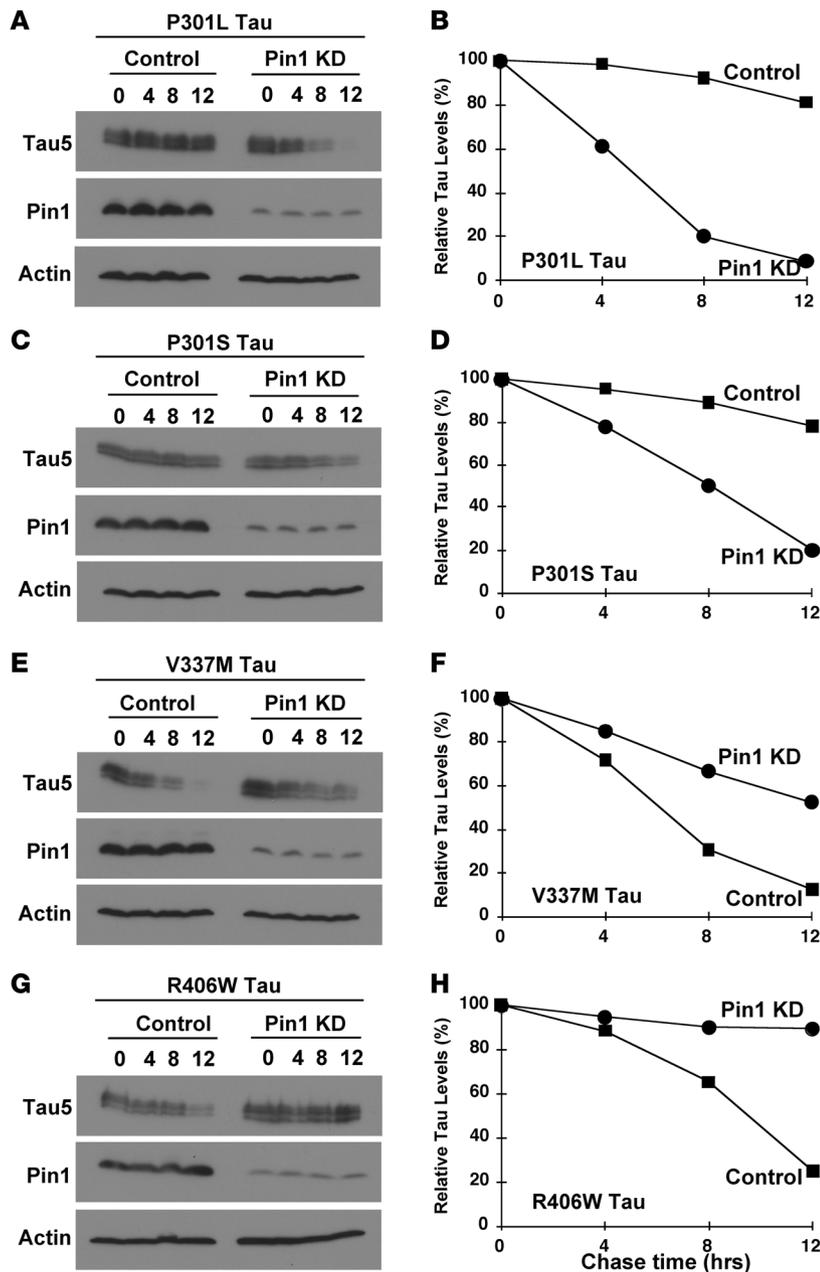


Figure 2

Effects of Pin1 KD on protein stability of other selected FTDP-17 tau mutants. Pin1 KD or control SY5Y cells were transfected with indicated FTDP tau mutants followed by cycloheximide chase analysis for indicated times. Cell lysates were fractionated by SDS-PAGE and analyzed by immunoblotting with Tau5, anti-Pin1, or anti-actin antibodies (A, C, E, and G). Tau levels were semiquantified from 3 different experiments using ImageQuant and normalized using actin as an internal control, with tau levels at 0 being defined as 100% (B, D, F, and H).

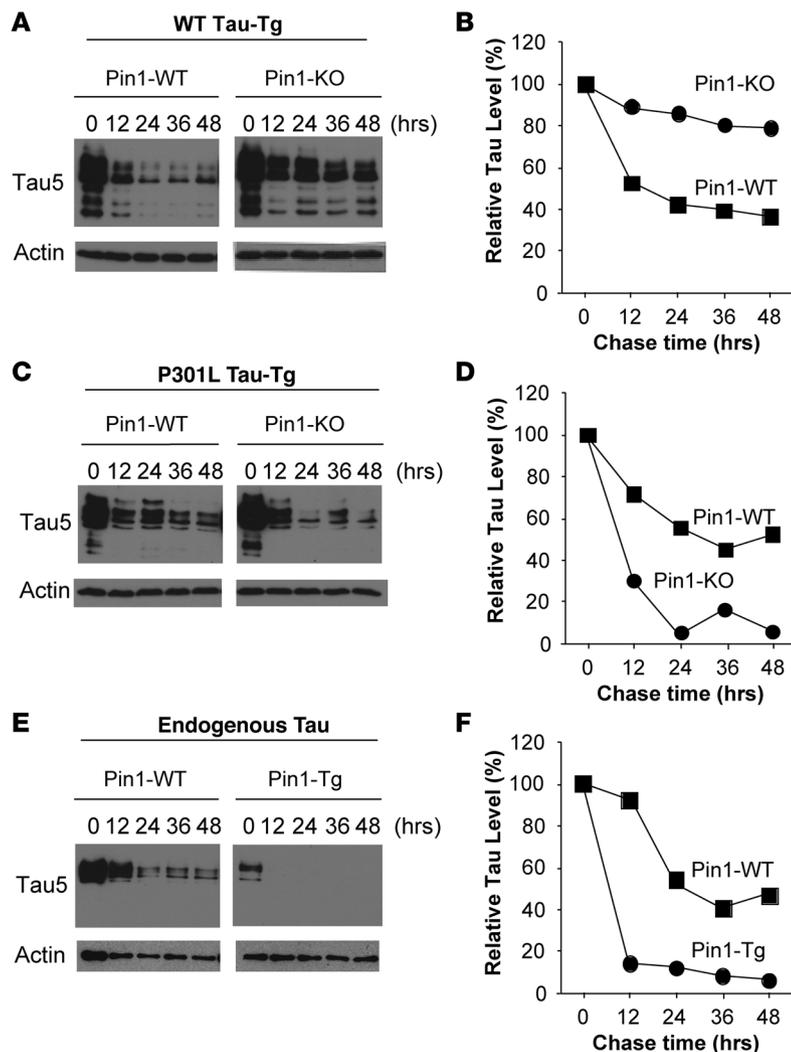
cultures increases protein stability of WT tau are consistent with the previous findings that Pin1 KO leads to increased levels, aggregation, and filament formation of endogenous tau in the mouse brain (48). However, the findings that Pin1 KD decreases protein stability of P301L tau were rather unexpected. To further confirm these surprising findings and to examine their pathological significance, we examined the effects of Pin1 KO on tau-related phenotypes induced

phosphorylation, NFT conformations, and tau aggregation (Figure 4, A and B). However, Pin1 KO in P301L Tg mice dramatically reduced the accumulation of total hyperphosphorylated tau as well as tau in the abnormal NFT-specific conformations (Figure 4A). Moreover, Pin1 KO almost completely abolished the aggregation of phosphorylated tau into the sarkosyl insoluble fraction (Figure 4B). For example, Pin1 KO reduced AT180 and AT100

by P301L tau by analyzing littermates generated from crosses between P301L tau Tg mice and Pin1 KO mice. Total brain lysates and sarkosyl-insoluble fractions were subjected to immunoblotting with mAbs that recognize all tau isoforms or specific tau phosphoepitopes and/or abnormal conformations present only in the neurofibrillary tangle (NFT) specific to AD (Figure 4, A and B), a procedure widely employed to characterize tau Tg mice (7–9, 15, 16, 18–20, 22, 48).

Since Pin1 KO in mice induces tau hyperphosphorylation and other tau-related phenotypes only after 8–9 months (48), we analyzed the effects of Pin1 KO on P301L tau at 6–8 months to avoid the complication of endogenous tau. As expected at this age, Pin1 KO did not have obvious effects on phosphorylation of endogenous tau, and P301L tau mice significantly overexpressed tau protein, as revealed by immunoblot with total tau mAb Tau5 (Figure 4A). Furthermore, tau protein was detected only in P301L tau mice when probed with human tau-specific mAb CP27 (Figure 4A). In total lysates, P301L tau overexpression resulted in aberrant tau hyperphosphorylation, as detected by pThr231-specific mAbs AT180, CP9, and CP17, and pSer202/pThr205-specific mAb AT8 (Figure 4A). Furthermore, the mutant tau also existed in the NFT-specific conformations, as detected by the conformation-specific mAbs MC1 and AT100 and by aberrant mobility shift (Figure 4A). As shown previously (7–9, 15, 16, 18–20, 22, 48), although some endogenous tau isoforms were extracted from WT mouse brains by the sarkosyl extraction procedure, endogenous tau was not recognized by many phospho-specific mAbs in spite of a weak recognition by AT8 and AT100 (Figure 4B). However, there was considerable enrichment of hyperphosphorylated P301L tau with markedly retarded mobility and aberrant NFT conformations in the sarkosyl insoluble fraction of P301L Tg mice (Figure 4B). These results indicate that P301L Tg mice develop robust tau-related phenotypes, as reported (7, 15, 22).

Pin1 KO greatly decreased overall levels of the P301L mutant (Figure 4A) and the aggregation of phosphorylated P301L tau into the sarkosyl insoluble fraction (Figure 4B) by approximately 4- to 5-fold, consistent with the above findings that Pin1 KD or KO reduced the half-life of the mutant tau (Figures 1 and 3). Furthermore, at the age of 8 months, Pin1 KO in non-tau Tg mice did not induce aberrant endogenous tau

**Figure 3**

Pin1 has opposite effects on the protein stability of WT tau and P301L tau in brain-slice cultures. Brain tissue slices of 230 μm were prepared and cultured from WT tau Tg mice in the presence (Pin1-WT) or absence (Pin1-KO) of Pin1 (A and B), from Tg P301L tau mice in the presence (Pin1-WT) or absence (Pin1-KO) of Pin1 (C and D), or from Thy1-Pin1 Tg mice (Pin1-Tg) or control non-Tg mice (Pin1-WT) (E and F). Cycloheximide was added to stop new protein synthesis and chased for indicated times. Brain slices were lysed in a buffer by sonication, followed by subjecting soluble protein lysates to immunoblotting with Tau5 or anti-actin antibodies (A, C, and E). Tau levels were semi-quantified from 2 independent experiments using ImageQuant and normalized using actin as an internal control (B, D, and F).

immunoreactivities of tau in total lysates by 2- to 3-fold and in the sarkosyl insoluble fraction by 5- to 10-fold. These results indicate that Pin1 KO dramatically reduces hyperphosphorylation, NFT conformations, and tau aggregation of P301L tau in mice.

To insure that the tau immunoreactivities observed on immunoblots are from neurons and also located in the expected subcellular compartments, we performed immunostaining analysis. Indeed, in contrast with non-Tg controls (data not shown), Tg P301L tau neurons exhibited strong immunoreactivities with AT180 in the soma region and axon of neurons in the cortex, brainstem, hippocampus, and spinal cord (Figure 4C and data not shown), as shown before (7, 9, 15, 19). Importantly, immunore-

activities with AT180 were dramatically reduced in P301L tau mice in the absence of Pin1 (Figure 4C). These results together indicate that Pin1 KO in mice almost completely suppresses the robust tauopathy phenotype induced by P301L tau.

Overexpression of Pin1 in postnatal neurons reduces protein stability of endogenous WT tau in mice. Together with the previous findings that Pin1 KO in mice is sufficient to cause the tauopathy phenotype (48), the above results indicate that Pin1 KD or KO has the opposite impact on tau protein stability and tau-induced phenotypes depending on whether the tau is WT or P301L in vitro and in vivo. Consequently, if Pin1 is indeed a critical rate-limiting regulator in the development of the tauopathy phenotype induced by WT tau and its FTDP-17 mutant, we would expect that Pin1 overexpression in the neuron would reduce the tauopathy phenotype in Tg WT tau mice but enhance the phenotypes in P301L Tg mice. To test this possibility, we generated Tg mice overexpressing Pin1 in neurons. Given that long-term overexpression of Pin1 in (proliferative) mammary glands disrupts the centrosome duplication checkpoint and induces breast cancer (44), we chose the Thy1.2 promoter to overexpress FLAG-tagged Pin1 in postnatal neurons (Figure 5, A–C) because it is active in neurons approximately 10 days after birth (59) so that the transgene does not interfere with the embryonic development of the brain.

To detect expression of FLAG-Pin1, we first performed immunoblotting analysis of various tissues, including brain, heart, liver, lung, pancreas, kidney, and testis. FLAG-Pin1 with expected molecular weight was mainly expressed in brain tissues at levels only approximately 1- to 2-fold higher than endogenous levels (Figure 5D). There was faint FLAG-Pin1 detected in testis, but none in many other tissues examined (Figure 5D). To determine whether Pin1 is indeed overexpressed in neurons in the central nervous system, we performed immunohistochemical staining using both FLAG antibody and Pin1 antibody on fixed tissue sections. Indeed, Pin1 was detected mainly in neurons in the hippocampus, cortex, spinal cord, and other regions in both WT and Pin1 Tg mice (Figure 5D and data not shown). In addition, the subcellular distribution of endogenous Pin1 detected in WT animals was generally distributed to cell bodies and filamentous skeletal structures of neuronal processes in addition to the nucleus (Figure 5E), as shown (48). Exogenous FLAG-Pin1 in Pin1 Tg animals exhibited a similar distribution pattern compared with the endogenous protein in WT animals albeit at higher intensities (Figure 5F). Therefore, Pin1 in these Tg mice is overexpressed only about 1–2 times over the endogenous levels and localized in the neuron as is endogenous Pin1.

To determine whether Pin1 overexpression affects endogenous tau in mouse brains, we examined Pin1 Tg mouse brains for any changes in tau phosphorylation and aggregation in the sarkosyl insoluble fraction and could not find any obvious differences between Pin1 Tg and non-Tg WT animals, even up to age

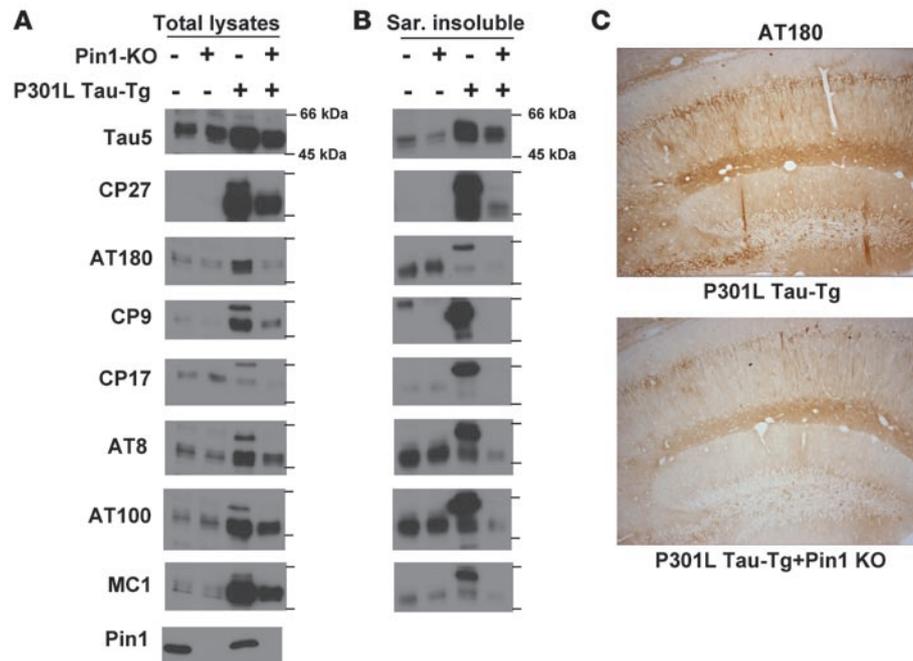


Figure 4 Pin1 KO in Tg P301L tau mice reduces tau hyperphosphorylation, NFT-specific conformations, and aggregation in the brain. **(A and B)** Brains of 6- to 8-month-old non-Tg or Tg P301L tau mice in the presence or absence of Pin1 KO, as indicated, were homogenized, followed by subjecting total lysates **(A)** or sarkosyl-insoluble (Sar. insoluble) fractions **(B)** to immunoblotting with different tau mAbs with the following specificities: Tau5, specific against total human and mouse tau; CP27, specific against total human tau; AT180, CP9, and CP17, specific against pThr231-tau; AT8, specific against pSer202/pThr205-tau; AT100, specific against pThr212-tau in the NFT-specific conformation; MC1, specific against tau in the NFT conformation. **(C)** Subcellular localization of phosphorylated tau. Mice were perfused and fixed, and the coronal hippocampal sections were subjected to immunohistochemical staining using AT180. Similar results were also obtained with AT100 (not shown). Original magnification, $\times 20$.

18 months, as assayed by immunoblotting and immunostaining using various antibodies (data not shown). However, we did observe that overall levels of endogenous tau were reduced in Pin1 Tg mice, as compared with non-Tg controls (Figure 6A), suggesting that Pin1 overexpression might reduce protein stability of endogenous mouse tau protein. To examine this possibility, we established brain-slice cultures from Thy1-Pin1 Tg mice and non-Tg littermates followed by cycloheximide chase analysis to examine the half-life of endogenous tau in brain tissues. Compared with control mice, the half-life of tau was significantly reduced in Thy1-Pin1 Tg brain tissues from approximately 24 hours to approximately 6 hours (Figure 3, E and F). These results indicate that Tg overexpression of Pin1 in postnatal neurons at a moderate level is functionally active and sufficient to reduce endogenous tau protein stability. Thus, these results demonstrate that Pin1 is a critical rate-limiting factor in controlling tau protein stability in vivo.

Neuronal Pin1 overexpression affects tau-related phenotypes in WT tau and Tg P301L tau mice in an opposite manner. To determine whether overexpression of Pin1 would affect the phenotypes of WT tau and P301L tau, we first crossed Pin1 Tg mice (in FVB genetic background) and WT tau (18) or Tg P301L tau mice (7) (both in C57BL/6 background) to generate double-Tg mice. In addition, to reduce the influence of genetic background, we generated double-Tg mice on a pure C57BL/6 background. We used mice carrying only 1 copy of the

respective transgenes and obtained similar results in both genetic backgrounds, although with slightly more variations in C57BL/6 mice (data not shown).

In 6-month-old Tg WT tau mice, Pin1 overexpression reduced tau phosphorylation on Thr231, as detected by AT180, but not on other sites as detected by AT8 and AT100 in brain lysates (data not shown). These results are consistent with previous studies showing that Pin1 selectively reduces the phosphorylation of the Thr231 site of tau in vitro and in cell cultures (47, 49). To monitor long-term effects of overexpressed Pin1 in WT tau phenotypes in mice, we analyzed the brain lysates of 1-year-old WT, Pin1 Tg, tau Tg, and Pin1/tau double-Tg mice by immunoblot (Figure 6A). As expected, tau levels were elevated in tau Tg mice by about 3-fold (Figure 6A). Furthermore, total tau was reduced by 50% in Pin1 and tau double-Tg mice, as compared with tau single-Tg mice (Figure 6A). These results indicate that Pin1 overexpression reduces total WT tau levels, consistent with the findings that Pin1 overexpression reduces endogenous mouse tau protein (Figure 3, E and F). More importantly, overexpression of Pin1 in tau Tg mice significantly reduced the accumulation of total hyperphosphorylated tau (Figure 6A). Furthermore, Pin1 overexpression in tau Tg mice also significantly reduced the accumulation of tau in the abnormal NFT-specific conformations

by about 3- to 4-fold (Figure 6A). These inhibitory effects of Pin1 overexpression on tau-related phenotypes are even more profound when analysis of sarkosyl insoluble fractions is considered (Figure 6B), with total tau5 immunoreactivity being reduced by 7- to 8-fold and AT180 and AT100 immunoreactivities by 3- to 5-fold. Moreover, immunohistochemical staining with mAb AT180 further confirmed that Pin1 overexpression reduced hyperphosphorylation of WT tau in neurons (Figure 6C and data not shown). These results together indicate that Pin1 overexpression protects against the accumulation of total tau, tau hyperphosphorylation, NFT-specific conformations, and insoluble aggregation of WT tau.

In contrast, Pin1 overexpression had the opposite effect on tau-related pathologies in P301L Tg mice (Figures 6 and 7). Young adult mice (6 months old) that were generated from Pin1 and Tg P301L tau mouse crosses were subject to analysis for tau-related phenotypes. Compared with non-P301L mice, there was an obvious increase in tau levels in P301L mice, and interestingly, tau levels further increased by 2- to 3-fold in the presence of overexpressed Pin1 (Figure 7A), which is consistent with the findings that Pin1 KD or KO increases P301L tau stability (Figures 1 and 3). More interestingly, Pin1 overexpression in Tg P301L tau mice significantly increased the accumulation of total hyperphosphorylated tau as well as tau in the abnormal NFT-specific conformations (Figure 7A). Moreover, these effects of Pin1 on tau-related pheno-

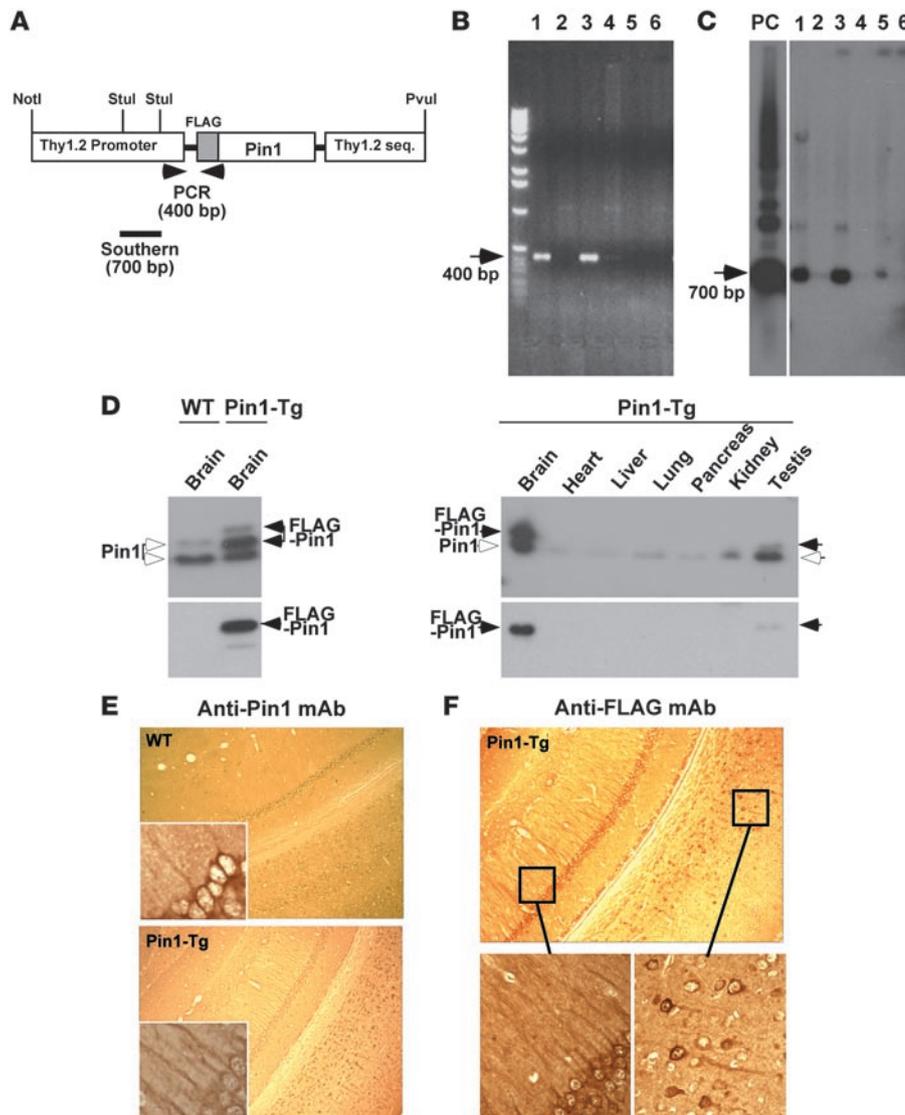


Figure 5 Generation of Tg mice that moderately overexpress Pin1 with subcellular localization similar to that of the endogenous protein. (A–C) Generation of Pin1 Tg mice. Pin1 cDNA with an N-terminal FLAG tag was subcloned into a murine Thy1.2 genomic expression vector (A) followed by removing vector sequences of this construct before pronuclear microinjection into pure FVB mouse embryos. Founders were genotyped by PCR analysis (B) and genomic Southern analysis after *StuI* digestion (C) of DNA isolated from tail biopsies. (D) Moderate overexpression of FLAG-Pin1 selectively in the brain. Various Pin1 Tg mouse tissues and WT mouse brains were dissected and frozen in liquid nitrogen, followed by homogenization in SDS sample buffer before submission to immunoblotting with antibodies against Pin1 and the FLAG epitope. (E and F) Similar subcellular localization of exogenous and endogenous Pin1 in the neuron. WT and Pin1 Tg mice were perfused and fixed, and the paraffin-embedded coronal sections were subjected to immunohistochemical staining with mAb against Pin1 (E) and the FLAG epitope (F). Original magnification, $\times 20$ (main photographs); $\times 40$ (insets).

types in Tg P301L tau mice were even more profound when the sarkosyl insoluble fractions were analyzed (Figure 7B). For example, Pin1 overexpression increased AT180 and AT100 immunoreactivity in total lysates by 2- to 3-fold and in the sarkosyl insoluble fractions by 3- to 6-fold. Finally, immunostaining confirmed that Pin1 overexpression in Tg P301L tau mice dramatically increased accumulation of P301L tau hyperphosphorylation and NFT-specific conformations (Figure 7C and data not shown). These results indicate that Pin1 overexpression greatly increases hyperphosphorylation, NFT conformations, and tau aggregation of P301L tau in mice, directly opposite of its effects in Tg WT tau mice.

Overexpression of Pin1 affects spinal and peripheral neuropathies in WT tau and Tg P301L tau mice in an opposite manner. Overexpression of both WT tau and P301L tau in mice has been reported to trigger neuropathological changes, especially in spinal and peripheral neurons (7, 18). Aged Tg WT tau mice develop accumulation of silver-positive intraneuronal inclusions and vacuolar lesions in spinal cords accompanied by neurodegeneration (18). Tg P301L tau mice accumulate silver-positive axonal spheroids and suffer from ventral horn cell loss and axonal degeneration in spinal

ventral roots, which correlates with their severe motor dysfunction (7). Given the opposite effects of Pin1 overexpression on tau-related biochemical phenotypes in WT tau and Tg P301L tau mice, we asked whether Pin1 overexpression has opposite effects on their spinal and peripheral neuropathies. We analyzed spinal cords and sciatic nerves isolated from age-matched WT, Tg WT tau, Pin1-Tg, and Pin1 Tg/tau Tg double-Tg mice, or Tg P301L tau and Pin1 Tg/Tg P301L tau double-Tg mice. In 18-month-old Pin1 Tg mice, we did not detect any obvious neuronal lesions either by Bielschowsky silver staining of spinal cord (Figure 8, A and B) or by analysis of the sciatic nerves (Figure 9, A and B). In 18-month-old Tg WT tau mice, we observed neuropathological changes, as demonstrated by the presence of some silver-positive neurons in the ventral horn of spinal cord (Figure 8C) and several degenerating or demyelinated axons within the sciatic nerves, which were often engulfed by macrophages (Figure 9C). However, we observed neither silver-positive neurons in the spinal cord (Figure 8D) nor degenerated neurons within the sciatic nerve (Figure 9D) in Pin1 Tg/Tg WT tau double-Tg mice. The silver stain and neurodegeneration of Pin1 Tg/Tg WT tau double-Tg mice were no different from those in age-matched

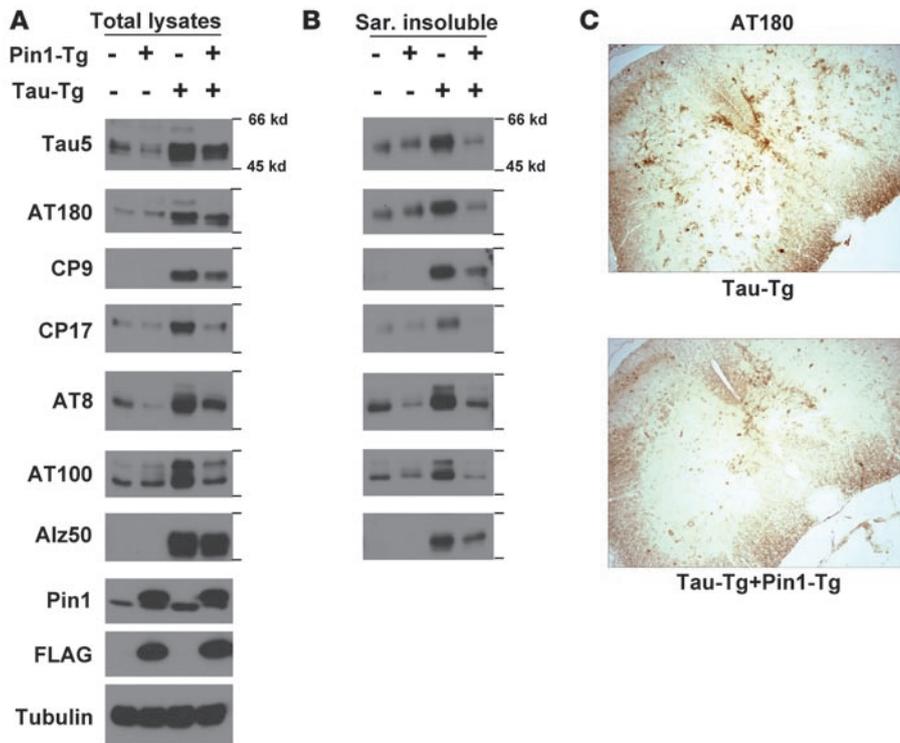


Figure 6

Pin1 overexpression in WT tau mice reduces tau hyperphosphorylation, NFT-specific conformations, and aggregation in the brain. (A and B) Brains of 1-year-old non-Tg or Tg WT tau mice in the presence or absence of Pin1 overexpression, as indicated, were homogenized, followed by subjecting total lysates (A) or sarkosyl-insoluble fractions (B) to immunoblotting with different tau mAbs with the following specificities: Tau5, specific against total human and mouse tau; AT180, CP9, and CP17, specific against pThr231-tau; AT8, specific against pSer202/pThr205-tau; AT100, specific against pThr212-tau in the NFT-specific conformation; Alz50, specific against tau in the NFT conformation. (C) Subcellular localization of phosphorylated tau. Mice were perfused and fixed; this was followed by subsection of coronal spinal cord sections to immunohistochemical staining with AT180. Original magnification, $\times 20$.

non-tau Tg controls (Figures 8, A and B, and 9, A and B). These results indicate that Pin1 overexpression effectively inhibits spinal and peripheral neuropathies induced by WT tau.

Since we observed the onset of the motor dysfunction of the Tg P301L tau mice at around 12 months of age in the C57BL/6 background, we analyzed Tg P301L tau mice at 7 months of age and compared them with the age-matched Pin1 Tg/Tg P301L tau double-Tg mice. At this early age, we detected relatively few silver-positive neurons in spinal cords of Tg P301L tau mice (Figure 8E) and few degenerating axons within their sciatic nerves (Figure 9E). However, we observed massive degeneration in age-matched Pin1 Tg/Tg P301L tau double-Tg mice, as demonstrated by Bielschowsky silver staining (Figure 8F) and by analysis of their sciatic nerves (Figure 9F). In addition, the peripheral neuropathy of the double-Tg mice was accompanied by their severe motor dysfunction (data not shown). These results indicate that Pin1 overexpression greatly enhances spinal and peripheral neuropathies induced by P301L tau. Furthermore, the opposite effects of Pin1 overexpression on hyperphosphorylation, NFT conformations, and tau aggregation of WT tau and P301L tau in Tg mice correlate well with the appearance of neurodegenerative phenotypes in spinal cords of the mice.

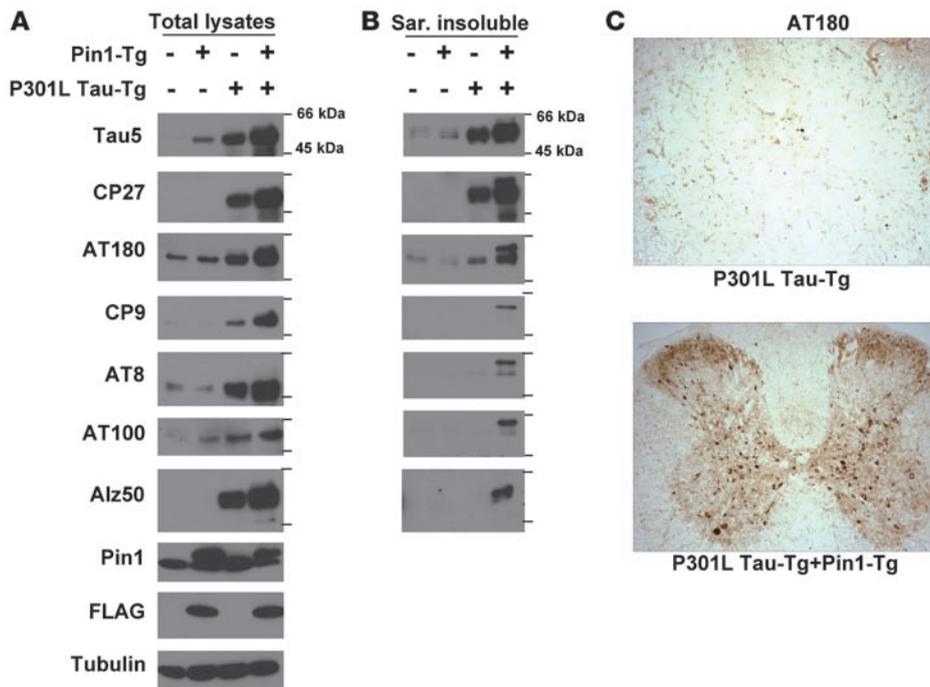
Discussion

We have shown that Pin1 KD or KO increases protein stability of WT tau in vitro and in vivo and that neuronal Pin1 overexpression greatly inhibits the tauopathy phenotype in Tg WT tau mice. These findings are consistent with the previous findings that Pin1 KO alone is sufficient to induce the tauopathy phenotype in mice. However, unexpectedly, we found that Pin1 KD or KO reduces protein stability of P301L tau in vitro and in vivo. This finding has been further substantiated by our demonstration that Pin1 KO almost completely abrogates the robust tau-related phenotypes in Tg P301L tau mice. In contrast, neuronal Pin1 overexpression dra-

cally exacerbates tau-related phenotypes and neurodegeneration in Tg P301L tau mice. These results have demonstrated for what we believe is the first time that Pin1 has diametrically opposite impacts on the tauopathy phenotype depending on whether the tau is WT or P301L mutant. Given that most human tauopathies such as AD do not have tau mutations, these results indicate that it is critical to use relevant tau models for investigating the development and treatment of different tauopathies. Furthermore, our results suggest that Pin1 upregulation might be beneficial for AD, whereas its inhibition might be helpful for patients carrying the P301L tau mutation.

It has been shown that Pin1 is greatly induced during neuronal differentiation and is highly expressed in most neurons in the brain but is especially low in the neurons that are most vulnerable to neurodegeneration in AD (48, 49). Furthermore, Pin1 has been shown to be inactivated or downregulated by oxidative modifications or genetic polymorphisms in human AD (48, 50–52). Importantly, Pin1 binds to and isomerizes phosphorylated tau to restore tau biological function and to promote tau dephosphorylation (46, 47). Moreover, Pin1 KO in mice induces the tauopathy phenotype, A β -related phenotypes, and neurodegeneration in an age-dependent manner, as seen in human AD (39, 48). These results indicate that loss of Pin1 function contributes to the tauopathy phenotype in AD, but whether increasing Pin1 function can suppress tau-related phenotypes is unknown.

To address this question, we generated Tg mice that overexpress Pin1 in postnatal neurons using the mouse Thy1.2 promoter. Exogenous Pin1 was expressed specifically in the neurons of the central nervous system at approximately 1- to 2-fold higher than endogenous levels and was localized to the same subcellular compartments as endogenous protein (Figure 5). This moderate level of Pin1 overexpression alone did not have any detectable deteriorative effects but was sufficient to reduce protein stability of endogenous mouse tau (Figure 3, E and F). More impressively, when

**Figure 7**

Pin1 overexpression in P301L tau mice increases tau hyperphosphorylation, NFT-specific conformations, and aggregation in the brain. (**A** and **B**) Brains of 6-month-old non-Tg or Tg P301L tau mice in the presence or absence of Pin1 overexpression, as indicated, were homogenized, followed by subjecting of total lysates (**A**) or sarkosyl-insoluble fractions (**B**) to immunoblotting with different tau mAbs with the following specificities: Tau1 and Tau5, specific against total human and mouse tau; CP27, specific against total human tau; AT180 and CP9, specific against pThr231-tau; AT8, specific against pSer202/pThr205-tau; AT100, specific against pThr212-tau in the NFT-specific conformations; Alz50, specific against tau in the NFT conformations. (**C**) Subcellular localization of phosphorylated tau. Mice were perfused and fixed; this was followed by subjecting coronal spinal cord sections to immunohistochemical staining with AT180. Original magnification, $\times 20$.

crossed to mice overexpressing human WT tau, Pin1 overexpression strongly suppressed tau-related phenotypes, including tau hyperphosphorylation, NFT conformations, and tau aggregation as well as neurodegeneration (Figures 6, 8, and 9). These results from Pin1 KO and Pin1 Tg mice have conclusively demonstrated that Pin1 is a critical rate-limiting factor in controlling WT tau protein stability and its tauopathy phenotype, which is found in AD and most other tauopathies, and also provide what we believe is the first *in vivo* evidence that upregulating Pin1 function is useful for inhibiting the tauopathy phenotype in AD.

The surprising findings of our current study are that the effects of Pin1 on P301L tau are diametrically opposite to those on WT tau *in vitro* and *in vivo*. Even though tau mutations have not been identified in AD, Tg P301L tau mice have been tacitly assumed to be a preferred AD model due to their robust tauopathy phenotype (7–14). Since the P301L mutation does not affect tau binding to Pin1 (46), we assumed that Pin1 could regulate P301L tau in the same way as WT tau. Indeed, Pin1 KD significantly affects protein stability of P301L tau, with the effects depending on the presence of the Pin1-binding site in tau, similarly to WT tau (Figure 1). However, unlike WT tau, Pin1 KD or KO did not increase but rather decreased P301L tau stability (Figures 1 and 3). As for other FTDP-17 tau mutants, Pin1 KD also decreased protein stability of P301S tau but increased protein stability of V337M tau and R406W tau (Figure 2), indicating that the effects of Pin1 on FTDP-17 tau mutants likely depend on the identity of the tau mutations, as expected. To confirm these findings, we focused on P301L by ablating the *Pin1* gene or overexpressed Pin1 in postnatal neurons in P301L tau mice and examined their effects on tau-related phenotypes, given that P301L tau mouse models are well established and widely used. Indeed, Pin1 KO in Tg P301L tau mice reduced protein stability and almost completely abolished hyperphosphorylation, NFT-specific conformations, and aggregation of the mutant tau (Figure 4), whereas Pin1 overexpression had the opposite effects and also greatly enhanced neu-

rodegeneration in P301L tau mice (Figures 7–9). These opposite effects of Pin1 overexpression in WT tau mice and P301L tau mice are unlikely due to 1 extra microtubule-binding repeat present in P301L tau mice because Pin1 KD also had the same opposite effects on both WT tau and P301L tau containing 4 microtubule-binding repeats (Figure 1). These *in vitro* and *in vivo* results together indicate that Pin1 has the diametrically opposite impact on the tauopathy phenotype depending on whether the tau is WT or P301L mutant (Figure 10). Given that most human tauopathies such as AD do not have tau mutations, our results indicate that it is critical to use Tg WT tau mice or to develop more relevant models for investigating the development and treatment of the tauopathy phenotype in AD. Moreover, our results suggest that inhibiting Pin1 function might be beneficial for FTDP-17 patients carrying P301L tau mutation.

Pin1 has been shown to affect a spectrum of protein activities, often in the opposite manner depending on substrates, such as decreasing or increasing protein stability, which might depend on whether specific pSer/Thr-Pro motifs are in *trans* or *cis* (42). Indeed, Pin1-catalyzed conformational changes between the 2 distinct *cis* and *trans* isomers have been visualized by NMR for APP and also shown to promote nonamyloidogenic APP processing and to reduce A β levels (39). In the case of tau, Pin1-catalyzed conformational changes have been shown to restore tau microtubule function and to promote tau dephosphorylation. Our results have shown that Pin1 also promotes tau degradation in WT protein, probably via inducing conformational changes (Figure 10A). This is consistent with a common theme that Pin1 often modulates its substrates via multiple mechanisms or acts on multiple targets to help drive certain cellular processes in one direction (42). However, in the case of P301L tau, Pin1 inhibits tau degradation (Figure 10B). Given the well-documented role of tau protein levels in the development of tauopathies, these differential effects of Pin1 on WT tau and P301L tau would be eventually manifested in their ability to aggregate and form NFTs and to induce neurodegeneration (Figure 10).

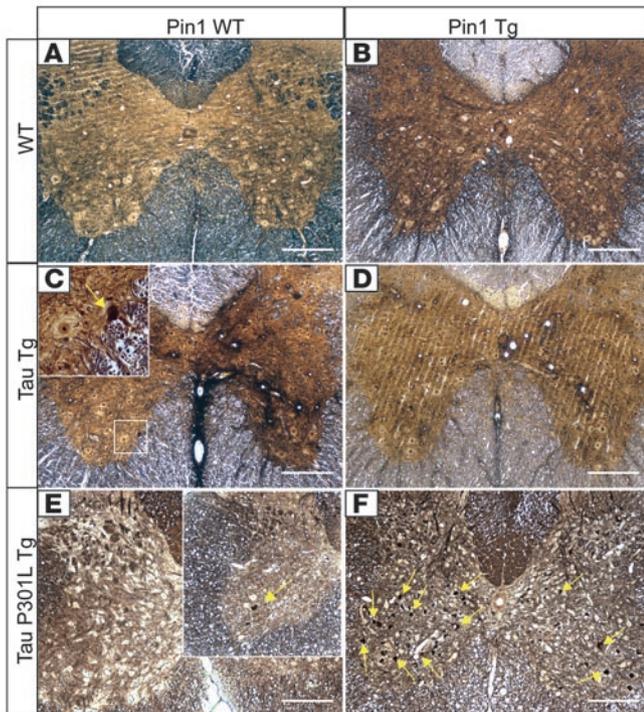


Figure 8

Opposite effects of Pin1 overexpression on spinal neuropathies of WT tau and Tg P301L tau mice. Bielschowsky silver staining revealed silver-positive neurons in spinal cords of 18-month-old Tg WT tau mice (C, inset) but not in the Pin1/tau double-Tg (D), WT (A), or Pin1 Tg mice (B). On the contrary, few neuronal lesions were detected in spinal cords (E) of 7-month-old Tg P301L tau mice, while overexpression of Pin1 and the P301L tau caused massive silver-positive neurodegeneration in the spinal cord in the age-matched double-Tg mice (F). Arrows point to silver-positive degenerated neurons. Scale bars: 100 μ m.

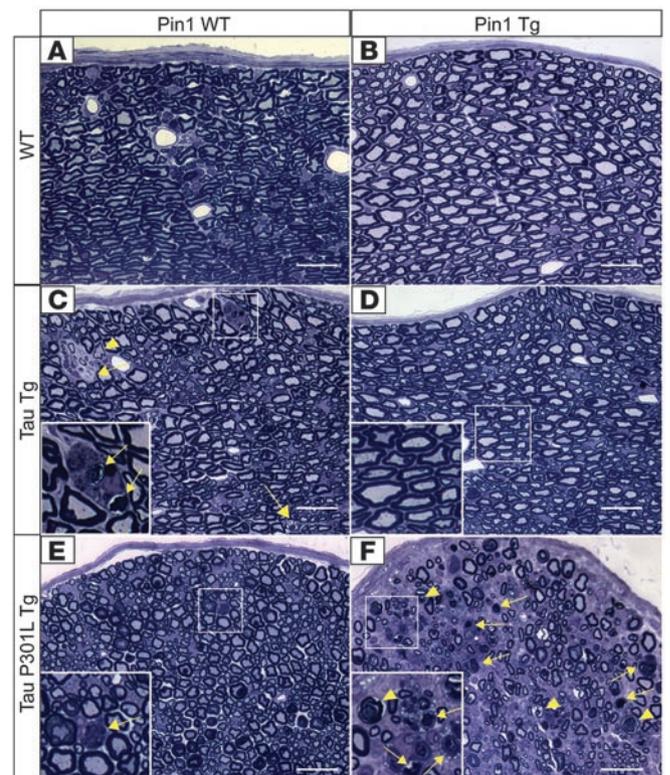
We do not yet know how the P301L mutation alters tau regulation so dramatically that the regulation of P301L tau by Pin1 becomes opposite to that of WT tau due to the lack of available tau structures. However, this is likely related to the fact that the P301L mutation would have a significant impact on the tau structure and that certain pSer/Thr-Pro motifs can exist in 2 completely distinct *cis* and *trans* conformations due to a 5-carbonyl ring of Pro in the peptide bond (42, 60). Tau phosphorylation has been shown to promote tau degradation via chaperone-interacting protein/Hsp70 (31–33) or inhibit tau degradation (34). Interestingly, F-box proteins, which target phosphorylated proteins to ubiquitin-mediated proteolysis, bind to pSer/Thr-Pro motifs only in *trans*, but not *cis* (61). Given the dramatic and opposite effects of manipulating Pin1 function on protein stability of WT tau and P301L tau and their ability to induce the tau-related phenotypes in cell cultures and animals, we hypothesize that proteins targeting phosphorylated tau for degradation might bind to certain pSer/Thr-Pro motif(s) in tau only in 1 conformation and Pin1-catalyzed *cis/trans* isomerization would promote this interaction to promote degradation of WT tau (Figure 10, A and B). However, in the case of P301L tau, the mutation might somehow render the pSer/Thr-Pro motifs in tau to be favored in the other conformation, and Pin1-catalyzed

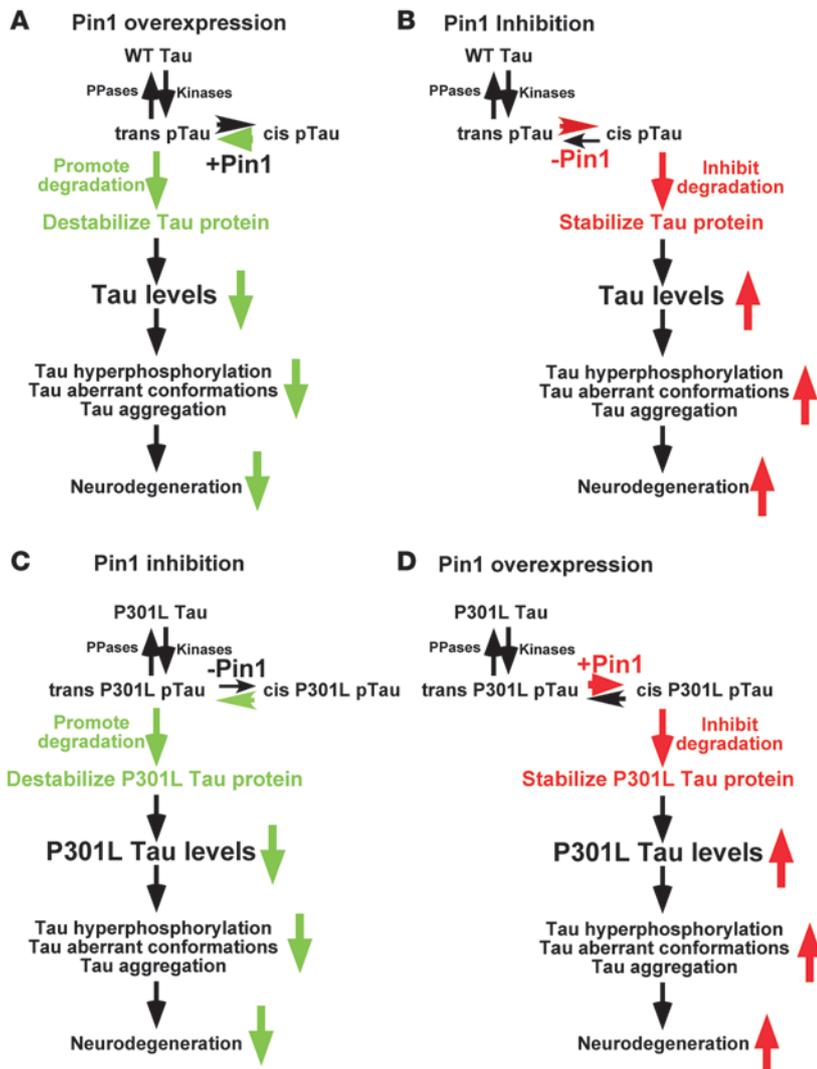
trans/cis isomerization would prevent phosphorylated P301L tau from being targeted for degradation (Figure 10, C and D).

In this model, although both WT tau and P301L tau are probably phosphorylated on certain Ser/Thr-Pro motifs such as the Thr231-Pro motif in *trans* due to the conformation specificity of kinases (42), the pThr231-Pro motif in WT tau might have a tendency to be in the *cis* conformation due to local structural constraints after phosphorylation, as shown for APP (62). However, Pin1 would rapidly reestablish equilibrium if the *trans* (or *cis*) population were suddenly depleted by certain degradation pathways due to the extremely slow uncatalyzed isomerization rate. Therefore, in the nonequilibrium cellular environment, Pin1 overexpression might greatly accelerate the *cis* to *trans* isomerization to promote tau degradation (Figure 10A), whereas Pin1 inhibition might allow a higher concentration of *cis* pThr231-Pro motif to be present for a longer time, which might inhibit tau degradation (Figure 10B). However, in the case of P301L tau, the mutation might somehow render certain pSer/Thr-Pro motifs in tau such as the pThr231-Pro motif to have a tendency to be in the *trans* conformation after phosphorylation. Therefore, Pin1 inhibition might allow a higher concentration of *trans* pThr231-Pro motif to be present for a lon-

Figure 9

Opposite effects of Pin1 overexpression on peripheral neuropathies of WT tau and Tg P301L tau mice. Analysis of semithin sections of the sciatic nerves revealed presence of degenerating axons (arrows) often engulfed by macrophages (C, inset) in 18-month-old Tg WT tau mice (C) but not in the age-matched Pin1/tau double-Tg mice (D), non-Tg WT (A), or Pin1 Tg mice (B). On the contrary, few neuronal lesions were detected in sciatic nerves (E) of 7-month-old Tg P301L tau mice, while overexpression of Pin1 and P301L tau caused massive neurodegeneration in the sciatic nerves of the age-matched double-Tg mice. (F). Arrowheads point to demyelinated axon, and arrows point to macrophages clearing myelin debris. Scale bars: 20 μ m.



**Figure 10**

A model for the diametrically opposite impact of Pin1 on tau levels and tauopathy phenotype depending on whether the tau is WT or P301L mutant. Our results have shown that Pin1 has the opposite impact on the ability to induce the tauopathy phenotype. Although both WT tau and P301L tau are likely phosphorylated on certain Ser/Thr-Pro motifs, such as the Thr231-Pro motif in *trans* by upstream kinases, the pThr231-Pro motif in WT tau might have a tendency to be in the *cis* conformation due to local structural constraints. Pin1 overexpression might greatly accelerate the *cis* to *trans* isomerization to promote tau degradation (A). When Pin1 is inhibited, the *cis* pThr231-Pro motif might not be isomerized to *trans* in a timely manner, which might inhibit tau degradation (B). However, in the case of P301L tau, the mutation might somehow change tau conformation so that the pThr231-Pro motif might have a tendency to be in *trans*, which might promote P301L tau degradation when Pin1 is inhibited (C). However, Pin1 overexpression might accelerate the *trans* to *cis* isomerization to inhibit P301L tau degradation (D). PPases, protein phosphatases.

ger time, which might promote P301L tau degradation (Figure 10C), whereas Pin1 overexpression might accelerate the *trans* to *cis* isomerization to inhibit P301L tau degradation (Figure 10D).

This model is consistent with our findings that overexpression of Pin1 reduces WT tau stability and abolishes its ability to induce the tauopathy phenotype but increases P301L tau stability and enhances its ability to induce the tauopathy phenotype. In contrast, Pin1 KO or KD has the opposite effects both on WT tau and P301L tau. Furthermore, the findings that Pin1 KD increased protein stability of WT, V337M, or R406W tau but decreased protein stability of P301L or P301S tau are coincidentally consistent with the fact that both P301L and P301S but not V337M tau and R406W tau mutations would allow P301 to exist only in the *trans*, but not *cis*, conformation, although further experiments are needed to determine the impact of the P301 mutations on tau structure, especially related to its Pin1-targeting site. Moreover, this is also consistent with the previous and current findings that manipulating Pin1 specifically affects the accumulation of an AD pathological conformation of the pThr231-Pro-containing tau in brain that is recognized by TG3, a mAb that probably recognizes only the *cis* pThr231-Pro motif (48, 63). Therefore, *cis* and *trans* conformations of the pThr-Pro motif in tau and APP are distinct

not only in their structures but also in their biological function or pathological consequences. Therefore, the events subsequent to tau phosphorylation such as degradation and dephosphorylation may be dynamically regulated, which are required under physiological conditions for cytoskeletal reorganization or vesicle transport along microtubule. The distinct properties of WT tau and P301L mutant tau may be magnified when Pin1 is added into the equation because of its pivotal role in catalyzing tau between 2 distinct conformations after phosphorylation (46–48). Further studies on Pin1-catalyzed conformational regulation of tau and its mutants and its biological and pathological significance should help elucidate the molecular events leading to tauopathies and might also lead to the development of new therapies.

In summary, we have shown that the prolyl isomerase Pin1 acts on both WT tau and P301L tau but has completely opposite results. Pin1 KD or KO greatly increases WT tau protein stability and its ability to induce the tauopathy phenotype but drastically decreases P301L tau protein stability and its ability to induce the tauopathy phenotype in mice, whereas Pin1 overexpression has the opposite effect. These results indicate that although WT tau deregulation, as seen in AD and most tauopathies, and P301L tau mutation, as seen in some FTDP-17 patients, cause similar tau-related pathologies,



the regulatory mechanisms that modulate the development of these tau-related phenotypes are quite different, pointing to the importance of using relevant models for investigating the development and treatment of human tauopathies. Moreover, our results suggest that Pin1 upregulation might be beneficial for AD, whereas its inhibition might be helpful for patients with P301L tau.

Methods

Generation of Pin1 KD cells and expression constructs. HT1080 cells were infected with Pin1 siRNA or control siRNA retroviruses, as described (53). For generating stable Pin1 KD SH-SY5Y cells, Pin1 shRNA lentiviral vector (Open Biosystems) was transfected together with VSV-G- and gag-pol-expressing plasmids (Open Biosystems) into 293T cells and stable clones selected using 0.5–1 µg/ml puromycin (53). To generate lentiviruses expressing WT tau and P301L tau, WT tau and P301L tau were cloned into the self-inactivating internal ribosome entry site–GFP lentiviral vector through SpeI, Sall restriction sites, as described (64).

Analysis of tau protein stability in cell cultures. Pin1 KD or control cells were transfected with WT tau or P301L tau constructs for 12 hours, and cycloheximide (100 µg/ml) was added. Cells were harvested at each time point, and total lysates were analyzed by immunoblotting with mAb against total tau or actin. For assaying tau stability in primary neurons, cortical neurons were isolated from E17.5 WT and Pin1^{-/-} embryos and cultured in Neurobasal Medium (Invitrogen) (65). After 2 days in culture, primary neurons were infected with lentiviruses expressing either WT tau or P301L tau. Tau stability assay was performed 3 days after infection after addition of 100 µg/ml cycloheximide, followed by assaying tau stability by immunoblotting. The immunoblots were scanned and semiquantitated using the software NIH Image 1.6.2, as described (66).

Generation of Pin1 Tg mice and mouse crosses. To generate Pin1 Tg mice, Pin1 cDNA with an N-terminal FLAG tag was subcloned into a murine Thy1.2 genomic expression vector pR5 (provided by Li-Huei Tsai, The Picower Institute, Massachusetts Institute of Technology, Boston, Massachusetts, USA) (67), followed by removal of vector sequences of this construct before microinjection. Tg mice were produced by pronuclear injection of pure FVB mouse embryos. Founders were identified by PCR analysis of lysates from tail biopsies and then intercrossed with pure FVB mice to establish lines. Expression of FLAG-Pin1 was confirmed by immunoblotting and immunostaining analyses. To generate Pin1 Tg mice in pure C57BL/6 background, Pin1 Tg mice in FVB background were backcrossed with C57BL/6 mice for 6 generations. Tg tau 3R mice in C57BL/6 background (18) and Tg P301L tau 4R mice in C57BL/6 background (7) (purchased from Taconic) were as described, respectively. Animal care and use for these experiments was approved by the Standing Committee on Animals at Harvard Medical School.

To generate P301L tau mice in the presence or absence of Pin1, Pin1^{+/-} mice heterozygous on a mixed 129/Svj and C57BL/6 background (68) were first crossed with P301L tau mice, which were subsequently crossed with Pin1^{+/-} heterozygous mice to obtain P301L tau Tg in Pin1 WT or KO mice with the expected controls. Since Pin1^{+/-} heterozygous mice are indistinguishable from WT mice (68, 69), Pin1 WT or homozygous KO mice were used. All the Tg mice analyzed carried only a single copy of the respective transgenes, and littermates from the same crosses were used, with the exception of some experiments involving Pin1 KO mice, in which different litters at the same ages were pooled due to difficulties in breeding Pin1 KO

mice. Similar phenotypes were observed in at least 3 different mice of the same genotypes in multiple experiments.

Organotypic brain-slice cultures. Organotypic brain-slice cultures were prepared essentially as described (70). In brief, brains from adult mice were removed and cut into coronal slices of 230-µm thickness with a vibratome. Slices containing the cerebral cortex and hippocampus were chosen. The slices were placed onto Millicell-CM culture inserts (Millipore) in 6-well culture plates with culture medium.

Immunoblotting analysis of tau in the mouse brain. Total lysates and sarkosyl-insoluble extracts were obtained as described (7, 15, 18, 48). Alz50, MC1, CP9, CP17, and CP27 mAbs against tau were gifts from P. Davies (Albert Einstein College of Medicine, Yeshiva University, New York, New York, USA). AT8, AT100, and AT180 (Endogen; Pierce Biotechnology), Tau1 (Chemicon), and Tau5 (BioSource) antibodies were purchased.

Immunohistochemical and histological analysis. Immunohistochemical analyses of mice were as described (7, 15, 18, 48). Bielschowsky silver staining was performed on 10-µm transversal sections of paraffin-embedded lumbar spinal cord as described previously (71). For analysis of the sciatic nerves, the dissected nerves were embedded in EPON, and 1-µm semithin sections were stained with toluidine blue.

Statistics. Values are presented as mean ± SD, and differences between groups were analyzed by 2-sided Student's *t* test. A *P* value of less than 0.05 was considered statistically significant.

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