Rheostat regulation of integrin-mediated leukocyte adhesion

Ivor S. Douglas1 and Themistocles Dassopoulos2

1Division of Pulmonary Sciences and Critical Care Medicine, University of Colorado at Denver and Health Sciences Center, and Denver Health Medical Center, Denver, Colorado, USA. 2Meyerhoff Inflammatory Bowel Disease Center, Johns Hopkins University Medical Center, Baltimore, Maryland, USA.

The homing of activated T lymphocytes to the gut in inflammatory bowel diseases is dependent on their coordinated, integrin-mediated adhesion and de-adhesion to substrates and blood vessel walls. In this issue of the JCI, Park and colleagues reveal a key modulatory role of a binding site within β integrins, known as the ADMIDAS domain, in controlling integrin de-adhesion in mice (see the related article beginning on page 2526). These observations add to our growing understanding of how integrin adhesiveness is regulated and raise the notion of the existence of a biological rheostat for lymphocyte homing. Disturbed migratory rheostat tone could account for variations in interindividual immune responses observed in patients with inflammatory bowel disease or other lymphocyte-mediated inflammatory disorders. These findings will inform future strategies to design small molecules for the treatment of a spectrum of chronic inflammatory conditions.

Multiple lines of evidence from animal models and diseased humans have defined the central role of gut-homing effector and regulatory T lymphocytes in the pathogenesis of inflammatory bowel diseases (IBDs) (1).

Integrins are cell surface–expressed, heterodimeric glycoproteins that play a prominent role in diverse immune cell interactions as well as in regulating lymphocyte migration, homing, survival, and proliferation (2, 3). The addressin receptors are expressed on gut lamina propia postcapillary venules and on high endothelial venules (HEVs; specialized postcapillary endothelial structures in mesenteric lymph nodes and Peyer’s patches) and function as organ-specific molecular “zip codes” for lymphocytes by facilitating integrin-mediated lymphocyte homing and transendothelial migration in a classical 4-step process. After initial tethering and rolling (step 1), lymphocytes are activated (step 2) and then undergo an activation-dependent change in conformational state of the α and β integrins, resulting in the upregulation of integrin adhesiveness (step 3). Ligand-receptor avidity and affinity interactions between lymphocyte integrins and addressin receptors are regulated by inside-out signaling in response to cytoskeletal stress, G protein–coupled receptor activation, or receptor tyrosine kinase receptor activation (3). This high-affinity binding is essential for lymphocyte migration across the endothelium (step 4) and into target organs (4).

Integrin-mediated adhesion and de-adhesion in leukocyte migration

Transendothelial lymphocyte diapedesis into target tissues such as the gut is...
regulated both by integrin-receptor adhesion and by de-adhesion. This tightly regulated shuffling of leading and trailing edges of transendothelial migratory cells is mediated largely by cytoskeletal plasticity signaled by the integrin receptors in response to microenvironment matrix proteins (such as fibronectin via the extracellular RGD domains) and integrin receptors of the immunoglobulin superfamily. This process of sticking and un-sticking facilitates remarkably efficient migration along chemokine gradients. In this issue of the JCI, Park and coworkers (5) present, in meticulous detail, the effects on lymphocyte de-adhesion and migration of a targeted activating mutation of one of three conformational regulatory extracellular domains of the integrin β subunit, a ligand-binding regulatory domain known as the adjacent to metal ion–dependent adhesion site (ADMIDAS; Figure 1). Multiple cation-binding sites are found in both α and β integrin subunits. The ADMIDAS domain regulates conformational activation by binding cations (Ca\(^{2+}\), Mg\(^{2+}\), and Mn\(^{2+}\)), thereby allosterically stabilizing alternative conformations of the integrin and affecting cellular adherence and release during adhesion and migration. The authors demonstrate, in genetically engineered mice carrying a targeted, activating mutation of the ADMIDAS domain, that lymphocyte migration is paralyzed both in vivo and in vitro because of impaired de-adhesion and excessive lymphocyte adhesiveness during migratory shuffling through HEVs in the gut. The impairment in de-adhesion was affected by fewer cell surface–expressed integrin molecules than in wild-type control animals, suggesting a particular powerful regulatory role for ADMIDAS-mediated conformational activation in regulating lymphocyte de-adhesion.

**Figure 1**
The interaction of the 3 β-chain cation-binding sites of lymphocyte α\(_4\)β\(_7\) integrin in the unliganded state and in the liganded conformation with endothelial cell MAdCAM-1. Inside-out signaling coordinates cation-dependent allosteric conformational activation of the α\(_4\)β\(_7\) integrin with the transition from a low-affinity binding state to an “open-hinge,” high-affinity binding state. This facilitates MAdCAM-1 binding and firm adhesion to α\(_4\)β\(_7\). The orientation in the MAdCAM-1–liganded form of the linear cluster of 3 metal-binding sites (ligand-induced metal-binding site [LIMBS], metal ion–dependent adhesion site [MIDAS], and ADMIDAS) in the ligand-binding head region of the integrin β subunit von Willebrand factor type A (VWF A or I-like) domain are shown in green, red, and yellow, respectively. ADMIDAS and ligand-induced metal-binding site are regulators of adhesion and de-adhesion and contribute to adhesive rheo-tone. As reported by Park et al. (5) in this issue of the JCI, ADMIDAS regulates de-adhesion of the lymphocyte’s trailing edge integrins, thus facilitating transendothelial migration along chemokine gradients.

**Role of integrin-dependent T lymphocyte migration in IBDs**
Integrin α\(_4\)β\(_7\) is the major lymphocyte-expressed ligand for addressins in the postcapillary venules of the intestinal lamina propria and in the HEVs of the gut secondary lymphoid organs, such as Peyer’s patches and mesenteric lymph nodes (Figure 2). Integrin α\(_4\)β\(_7\) is also a well-characterized gut-homing molecule and is important for the transendothelial migration of activated T cells in IBDs (6). Expression of mucosal addressin cell adhesion molecule-1 (MAdCAM-1), a cognate ligand for α\(_4\)β\(_7\), is increased in the inflamed mucosa of patients with ulcerative colitis (UC) and Crohn disease (CD) (7, 8) (Figure 3). The alternative α chain partner for β\(_7\) is α\(_E\), also known as CD103. α\(_E\)β\(_7\) is expressed on CD25+ Treg, is a potential MAdCAM-1 ligand, and is implicated in lymphocyte binding to intestinal epithelial cells (9).

Expression of endothelial cell adhesion molecules (CAMs) such as MAdCAM-1 is upregulated in several lymphocyte-dependent models of IBD (10, 11). The Cotton-top tamarin (CTT) is a New World primate that experiences a spontaneous, chronic colitis marked by periodic relapses of acute inflammation. This colitis model closely mimics human UC in its histologic features and response to pharmacologic agents. In landmark studies, administration of anti-α\(_E\) and anti-α\(_4\)β\(_7\)-monoclonal antibodies attenuated acute and chronic colitis in the CTT model.
Treatment with anti–MAdCAM-1 antibodies ameliorates intestinal inflammation in several IBD models, including dextran sodium sulfate–induced colitis (14, 15) and the ileitis of SAMP1/Yit mice (16). Antisense MAdCAM-1 oligonucleotides suppress the development of trinitrobenzene sulfonate–induced colitis in mice, a T cell–mediated animal model of IBD (17). Neutralizing monoclonal antibodies directed against other endothelial CAMs have also shown efficacy in some animal models by decreasing inflammation and mucosal damage (18–20). CAMs have also been demonstrated to be of importance in non-IBD models of bowel inflammation, such as graft-versus-host disease (21).

In humans with CD and UC, therapeutic targeting of gut integrins and addressins may represent a treatment option, particularly for patients with refractory disease. Natalizumab, a humanized monoclonal antibody against \( \alpha_4 \) integrin, is efficacious in inducing remission in patients with moderate to severe CD and elevated levels of C-reactive protein (22). However, therapeutic integrin modulation for the treatment of autoimmune diseases may have adverse effects on adaptive immune responses to infection: patients with multiple sclerosis and CD that were treated with natalizumab (23) developed progressive multifocal leukoencephalopathy, possibly as a result of inhibition of lymphocyte trafficking and impaired immune surveillance, presumably leading to reactivation of a clinically latent JC polyomavirus infection. MLN02, an anti-\( \alpha_4\beta_7 \) humanized monoclonal antibody, has shown promise in reducing gut inflammation without the attendant risk of more widespread immunosuppression in UC (24).

The CD4–CD45RB\(^{hi} \) adoptive transfer model of inflammatory colitis has provided important insights into the pathogenesis of gut inflammation. SCID or Rag–/– mice and athymic rats reconstituted with CD4–CD45RB\(^{hi} \) T lymphocytes from normal animals develop a Th1-type chronic colitis (25). As in human IBD, mucosal microbiota are critically important in this model: the immunodeficient mice do not develop colitis in a germ-free environment, and antibiotic therapy ameliorates the inflammation. The power of this model lies in the demonstration that cotransfer of CD4–CD45RB\(^{lo} \) naive T cells together with mature T cells prevents colitis. Elegant work has demonstrated that this cell population contains Tregs that antagonize the proinflammatory effector T cells via production of antiinflammatory cytokines such as IL-10 and TGF-\( \beta \) (26, 27). The findings of Park et al. (5) do not indicate whether the striking paralysis of lymphocyte migration in ADMIDAS mutants results in functional impairment of Tregs as well as effector T cells. This...
Rheostatic adhesive tone may regulate leukocyte homing and migration to target organs

The observations of Park et al. (5) raise several questions about the relevance of integrin-mediated adhesive/de-adhesive balance to human disease both in the gut and in other target organs affected by lymphocyte- and autoimmune-mediated inflammation, including bronchiolitis, interstitial pneumonitis, systemic sclerosis, rheumatoid disease, autoimmune hepatitis, cholangitis, autoimmune endocrinopathies, and neurological disorders.

The genetically engineered, constitutively active ADMIDAS domain phenotype (5) represents an extreme form of impaired de-adhesion during lymphocyte migration. It is possible that naturally occurring conformational variants of integrin ADMIDAS, whether regulated by SNPs or by epigenetic or posttranslational modifications, could contribute to a spectrum of lymphocyte “migratory tone” that contributes to both the risk for autoimmune disease and/or the magnitude of host adaptive immune responses to environmental stimuli. The interplay of integrin adhesive/de-adhesive resistance could function as a biological rheostat for target organ homing by activated effector and/or regulatory lymphocytes. In future studies it will be important to address whether constitutively active ADMIDAS...
domain—containing Tregs are also compromised. Conceptually, this hypothesis might be extended to explain variations in interindividual responses to intraluminal gut flora or inhaled environmental antigens. Individuals with high de-adhesive rheo-tone may be at a lower risk for developing diseases such as IBDs, asthma, or interstitial pneumonia.

Supportive of the integrin-addressin rheostat concept are associations between an ICAM-1/CD54 (K469E) SNP and the occurrence, disease localization, and severity of human IBD (31–33). Migratory rheostat—ILK-1 (integrin-linked kinase–1; ILK-1). ILK-1

Acknowledgments

This work was supported in part by NIH grant HL070940 to Ivor S. Douglas.

Address correspondence to: Ivor S. Douglas, Pulmonary and Critical Care Medicine, Denver Health and University of Colorado Health Sciences Center, Department of Medicine, MC 4000, 777 Bannock Street, Denver, Colorado 80204, USA. Phone: (303) 436-5905; Fax: (303) 436-7249; E-mail: idouglas@dhha.org.