Ewing sarcoma gene EWS is essential for meiosis and B lymphocyte development

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Ewing sarcoma gene EWS encodes a putative RNA-binding protein with proposed roles in transcription and splicing, but its physiological role in vivo remains undefined. Here, we have generated Ews-deficient mice and demonstrated that EWS is required for the completion of B cell development and meiosis. Analysis of Ews<sup>−/−</sup> lymphocytes revealed a cell-autonomous defect in precursor B lymphocyte (pre-B lymphocyte) development. During meiosis, Ews-null spermatocytes were deficient in XY bivalent formation and showed reduced meiotic recombination, resulting in massive apoptosis and complete arrest in gamete maturation. Inactivation of Ews in mouse embryonic fibroblasts resulted in premature cellular senescence, and the mutant animals showed hypersensitivity to ionizing radiation. Finally, we showed that EWS interacts with lamin A/C and that loss of EWS results in a reduced lamin A/C expression. Our findings reveal essential functions for EWS in pre-B cell development and meiosis, with proposed roles in DNA pairing and recombination/repair mechanisms. Furthermore, we demonstrate a novel role of EWS in cellular senescence, possibly through its interaction and modulation of lamin A/C.

Introduction

Ewing sarcoma gene EWS was first identified from the Ewing sarcoma chromosomal breakpoint t(11;22)(q24;q12) region as a translocation-generated fusion gene product between EWS and FLI1, a member of the ETS family of transcription factors (1). It was soon discovered that EWS was frequently rearranged in chromosomal translocations in other types of cancer and invariably led to a fusion of EWS to various transcription factor genes (reviewed in refs. 2, 3). The transcription factor partners include ATFI, a bZIP transcription factor, in malignant melanoma of soft parts (4); C/EBP-homologous protein (CHOP) in myxoid liposarcoma (5); Wilms tumor gene 1 (WT1) in desmoplastic small round cell tumor (DSRCT) (6); and NR4A3/CHN/NOR-1 in extraskeletal myxoid chondrosarcoma (7). In all cases, the aminoterminal domain (NTD) of EWS is fused, in frame, to a DNA-binding domain of various transcription factors, creating an aberrant transcription factor that is not present in normal cells. It is thus likely that the different pathophysiology of various EWS-fusion driven sarcomas is largely determined by a particular DNA-binding domain encoded by the EWS translocation partner.

EWS is a member of the TET family of proteins, which includes 3 other members, translocated in liposarcoma/fusion (TLS/FUS), TAF15/hTAFII68 (TATA-binding protein–associated factor 15/human TATA-binding protein–associated factor IIE68), and the Drosophila protein cabeça/SARF1, all of which contain a putative RNA-binding domain (8). The NTDs of EWS, TLS, and TAF15 contain degenerate repeats of the SYGQ motif and mediate potent transcriptional activation when fused to heterologous DNA-binding domains of different transcription factors (9–11). This domain is also essential for the transforming activity of EWS/Fli1 (12). Interestingly, the NTDs of EWS and TLS appear to be functionally interchangeable, as chromosomal translocation of either EWS-CHOP or TLS-CHOP is found in myxoid liposarcoma, with an indistinguishable pathophysiology (10, 13). Similarly, in extraskeletal myxoid chondrosarcoma, rearrangements of either EWS or TAF15 NTD to NR4A3 transcription factor have been found (7, 14, 15). The carboxyterminal domain (CTD) of EWS contains an RNA recognition motif (RRM), 3 RGG box domains, and a Ran-binding protein 2–like (RanBP2-like) zinc finger motif (1). All the TET proteins have been shown to bind RNA in vitro in a nonconsensus manner as well as single-strand DNA (ssDNA) (8), (16–18). Surprisingly, in vitro RNA binding by EWS only required the RGG box and not the RRM domain (16). In addition, EWS contains an IQ domain that is phosphorylated by PKC and is implicated in binding to calmodulin (19). Interestingly, PKC phosphorylation of EWS or EWS-caldulin interaction led to a decrease in RNA binding by EWS, suggesting a possible regulatory mechanism for EWS-RNA interaction.

EWS has been shown to interact with subunits of transcription factor II D and RNA polymerase II complexes (20) and various splicing factors (21, 22), but its role in basal transcription or splicing has yet to be demonstrated. A transcriptional role of EWS has been inferred from studies that demonstrated EWS interaction with RNA polymerase II, TFIIID, and CBP (23); subsequently, EWS was shown to influence transcriptional activities of HNF3 (24), BRN3A (25), and OCT4 (26) in reporter assays. A role of EWS in splicing was suggested from its interactions with various splicing factors, SF1/ZFM1 (27), U1C (21), YB1 (28), and TAR-S1 and -2 (22). However, EWS did not show any effects on splicing in vitro.
assays, while EWS-Fli1 inhibited U1C-, YB1-, and TASR-mediated splicing (21, 22, 28). To determine the in vivo role of EWS, we used gene targeting in mouse ES cells to inactivate EWS in mice. Our analysis of Ews-deficient mice revealed that EWS is essential for pre–B lymphocyte (pre–B lymphocyte) development and meiosis. Furthermore, studies with mouse embryonic fibroblasts (MEFs) demonstrated an involvement of EWS in cellular senescence, possibly through its interaction with nuclear lamin A/C.

Results

Inactivation of Ews by gene targeting. (A) Scheme for Ews targeting strategy. The triangles indicate loxP sequences flanking the stop cassette. WT1, WT1 cDNA; PA, poly(A) sequence. The probe used for Southern blot analysis is indicated. (B) Southern blot analysis of AseI-digested tail DNA. Wild-type (9.4-kb) and targeted (6.9-kb) alleles are marked. (C) Western blot analysis of EWS and TLS using whole-cell extracts isolated from MEFs. Ews genotype and the molecular weight markers (kDa) are indicated.

Inactivation of Ews in mouse ES cells. During the generation of conditional knock-in mice expressing the EWS/WT1 translocation gene product, we generated an Ews-null allele by inserting a stop sequence flanked by 2 loxP sequences (29) in intron 6 of Ews (Figure 1A). In addition, Ews exons 8–11, encoding a part of the RRM motif, were deleted by homologous recombination (Figure 1A). Targeted mouse ES clones were identified and used to generate chimeras, which were subsequently bred with Black Swiss outbred mice to establish F1 heterozygotes (Ews+/tmStop) and homozygotes (Ews−/−) mice. Homozygous Ews−/− mice were intercrossed to generate Ews−/− animals (Figure 1B). To determine whether the insertion of stop resulted in a complete loss of EWS, we examined EWS expression in MEFs by Western blot analysis using rabbit polyclonal antibodies raised against the first 74 amino acids of mouse Ews. As shown in Figure 1C, EWS expression was completely absent in the Ews−/− cells. A truncated EWS was not detectable even after a long exposure of the immunoblot. In addition, loss of EWS did not result in a compensatory increase in the expression of its homolog TLS/FUS (Figure 1C). Northern blot analysis revealed a complete absence of full-length Ews transcripts but an accumulation of short truncated transcripts in Ews-null MEFs (data not shown). Although the possibility remains that an undetectable amount of truncated Ews protein may exist, Ews-heterozygous mice were indistinguishable from wild-type littermates and developed normally, suggesting that the targeting strategy resulted in an Ews-null allele.

Ews-deficient mice are runted and show high postnatal lethality. EWS is a ubiquitously expressed protein, and based on its predicted functions in basal transcription and splicing (3), we presumed that a complete absence of Ews would interfere with normal development. Surprisingly, Ews-null animals were born at the expected Mendelian ratio (Table 1). However, all of the homozygous mutants were born smaller than their littermates and remained small through weaning (Supplemental Figure 1; supplemental material available online with this article; doi:10.1172/JCI31222DS1). We also observed a high rate (about 90%) of postnatal mortality in Ews−/− pups prior to weaning. A small number of the mutant animals did not suckle and may have died from lack of feeding. However, most Ews mutant pups were found dead or disappeared despite evidence of feeding. Examination of early postnatal Ews mutant pups revealed normal
appearance of all major organs; therefore, the exact cause of death is not known. We obtained a few surviving Ews–/– animals for further characterization. It is worth noting that we did not get any surviving homozygotes in 129SvEv or C57BL/6 inbred mice.

Severe lymphopenia and a cell-autonomous pre–B cell development defect in EWS mutant mice. Peripheral blood analysis of 3- to 4-week-old EWS mutant animals compared with littermate controls revealed severe lymphopenia (Figure 2A). Erythropoiesis seemed to be normal, since both wild-type and mutant mice showed similar red blood cell count and hematocrits (data not shown). All of the Ews-null animals examined displayed disproportionately small thymi and spleens compared with their littermates, and this was reflected by the markedly reduced cellularity of these organs (Figure 2B). To examine the lymphocyte development in detail, we performed flow cytometric analysis of T and B lymphocytes. Flow cytometry of T cells in thymus indicated only a slight decrease in the proportion of CD4+CD8+ cells in the mutants compared with controls (Supplemental Figure 2). Further analysis of CD4 CD8- precursor T cells with CD25 and CD44 markers also revealed no major alterations. However, analysis of B lymphocytes revealed marked reductions in both immature (B220+IgMhi) and more mature (IgMhiIgDhi) B cell populations from bone marrow and spleen of EWS mutant animals (Figure 2, C and D), suggesting a developmental defect in B lymphocyte maturation. Flow cytometry using markers to identify pre–B lymphocytes (B220-CD43-) from the Ews–/– bone marrow demonstrated a marked reduction in the pre–B cell compartment (Figure 2D). In contrast, progenitor B cell (pro–B cell) populations (B220-CD43+) appeared to be normal. These results suggest that there might be a developmental arrest in the pro–B to pre–B cell transition in the absence of EWS. To confirm this observation, we performed an in vitro pre-B CFU assay using bone marrow cells isolated from wild-type and mutant animals. The results showed that progenitor cells isolated from Ews–/– bone marrow failed to form pre-B colonies in vitro but differentiated as efficiently into other lineages (GM-CFU) as the control (data not shown; also see Figure 2F).

To further demonstrate the intrinsic nature of the pre–B cell defect in vivo, we performed a fetal liver transplantation experi-

Table 1
Genotype analysis of newborn and 3-week-old mice

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<tr>
<th>Age</th>
<th>Genotype</th>
<th>+/+</th>
<th>+/-</th>
<th>-/-</th>
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<tr>
<td>P0.5</td>
<td>29</td>
<td>71</td>
<td>29</td>
<td></td>
</tr>
<tr>
<td>P21</td>
<td>27</td>
<td>62</td>
<td>1 (17)</td>
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The number in parentheses indicates dead animals. The decrease in the number of animals at P21 was due to the use of 3 litters from P0.5 animals for histological analysis (after genotyping).

Figure 2
EWS is intrinsically required for pre–B cell development. (A) A complete blood count was performed with peripheral blood from 3-week-old littermates (n = 3/genotype). White blood cell differential data indicate a marked reduction of the lymphocyte population in Ews–/– mice. (B) Reduced cellularity of lymphoid organs in Ews–/– mice. Total cell count is shown for indicated organs of 3-week-old mice (n = 3/genotype). Flow cytometry of splenocytes (C) and bone marrow–derived cells (D) from Ews–/– mice and littermate controls using B cell markers B220, CD43, IgM, and IgD. (E) Fetal liver chimera analysis. Bone marrow cells were harvested from fetal liver chimeras and analyzed by flow cytometry with antibodies against CD43 and B220 along with CD45.1 and CD45.2. Presented are representative data from 3 independent experiments with similar results (n = 3 for Ews–/– and n = 3 for Ews–/– littermate controls). (F) CFU assay. Bone marrow cells from fetal liver chimeras were plated in duplicate in either pre-B or multilineage differentiation (GM-CFU) media. The number of colonies is presented as the mean colony number from 3 independent experiments.
To this end, lethally irradiated Rag2<sup>−/−</sup> hosts (B6.SJL strain, CD45.1<sup>+</sup>) were reconstituted with E14.5 fetal liver–derived hematopoietic precursors of all 3 Ews genotypes (129/Black Swiss hybrids, CD45.2<sup>+</sup>) by tail-vein injection and analyzed 6 weeks later by flow cytometry. Analysis of donor-derived T lymphocytes revealed a relatively normal reconstitution of all T cell subsets by the EWS mutant fetal liver cells (Supplemental Figure 2C). However, reconstitution of the pre–B cell population (B220<sup>−/CD43<sup>−</sup></sup>) was markedly reduced in animals injected with the EWS mutant cells, whereas the B220<sup>−/CD43<sup>+</sup></sup> pro–B cell population appeared normal compared with that of control (Figure 2E). Consistent with the results of our pre-B colony assay, bone marrow progenitors reconstituted from Ews<sup>−/−</sup> fetal liver cells were unable to differentiate into pre-B colonies in vitro but developed efficiently into myeloid/granulocytic lineages (Figure 2F). Together, these results clearly demonstrate that EWS is essential for pre-B lymphocyte development.

**Homozygous inactivation of Ews leads to meiotic arrest and apoptosis.** Homozygous EWS mutant male and female mice failed to produce any pups when mated with wild-type animals. Upon gross examination, testis and ovary were found to be greatly reduced in size in all EWS mutant animals examined (data not shown). Histologic analysis of testes from 12-week-old EWS mutants revealed a complete maturation arrest of spermatocytes (Figure 3A). The seminiferous tubule contained primary spermatocytes, but there were no condensing spermatids in the seminiferous tubules or any maturing spermatooza in the epididymis. Spermatogenesis were present, but primary spermatocytes and round spermatids were disorganized and markedly reduced in number. In Ews<sup>−/−</sup> testes, we also observed spermatocytes that were enlarged and multinucleated, possibly as a result of abnormal segregation of meiotic chromosomes and cell division (Figure 3A, inset, arrow). The wild-type testes contained normal primary spermatocytes, round spermatids, and condensing spermatids but did not contain any abnormal multinucleated spermatocytes. These results suggest that the Ews<sup>−/−</sup> primary spermatocytes were undergoing spermatogenic arrest at the point of or just prior to the first meiotic division. Consistent with this, TUNEL staining showed the occurrence of massive apoptosis of primary spermatocytes in the EWS mutant testes, but TUNEL-positive apoptotic spermatocytes were rarely found in the controls (Figure 3B).

Twelve-week-old female homozygous mice were devoid of maturing ovarian follicles and showed mild proliferation of interstitial cells (Figure 3C). There were no corpora lutea present in the ovaries of most EWS<sup>−/−</sup> female mice examined. In younger Ews<sup>−/−</sup> female mice (3 weeks), however, we observed immature ovarian follicles, suggesting that the defects probably occurred during the later stages of oocyte maturation (data not shown). These results demonstrate that EWS is essential for gametogenesis in both sexes.

**Ews deficiency leads to XY asynapsis and reduction in meiotic recombination.** Meiotic prophase I in spermatogenesis proceeds sequentially in morphologically distinguishable stages: leptotene, zygotene, pachytene, diplotene, and diakinesis (30). Each stage can be distinguished by immunostaining of synaptonemal proteins (e.g., synaptonemal complex protein 3 [SCP3]) present at the synaptonemal complex formed between homologous chromosomes (31). To examine the spermatogenesis defects in Ews<sup>−/−</sup> null mice in detail, we analyzed spermatocytes from 4- to 6-week-old mice by immunostaining of meiotic chromosome spreads with antibod-
ies against SCP3 and γ-H2AX, which is a marker of the sex body containing XY bivalent in pachynema (32). During the pachynema stage, sex chromosomes form a partial synopsis through their respective pseudoautosomal regions (Figure 4A, arrowhead). However, in Ews+/− spermatocytes, more than one-third of pachynema nuclei (60 of 164) showed asynapsis of XY chromosomes (Figure 4A, arrow). In comparison, only 9 of 195 nuclei from Ews−/− spermatocytes contained univalent XY (Figure 4B).

After the formation of bivalents, synapsed chromosomes undergo programmed double-strand breaks (DSBs) and initiate meiotic recombination by homologous recombination. Meiotic recombination is essential for the proper segregation of chromosomes during meiosis (30). Meiotic crossovers, which are sites of DSB and homologous recombination, can be visualized by immunostaining with an antibody against MLH1 (33), a homolog of the E. coli DNA mismatch repair gene mutM. To determine whether EWS mutant spermatocytes undergo normal meiotic recombination, we doubled stained spermatocyte nuclei with antibodies against SCP3 and MLH1. In control pachynema nuclei, all bivalents contained at least 1 MLH1 foci, with an average of 22 MLH1 foci per genome (Figure 4C, left panel, and Table 2); this is consistent with the approximately 23 MLH1 foci per genome present in normal spermatoocytes (Figure 4C, arrows, right panel, and indicated univalent XY. Original magnification, ×40).

Table 2

<table>
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<th>Genotype</th>
<th>No. of cells</th>
<th>No. of MLH1 foci</th>
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<tr>
<td>+/−</td>
<td>74</td>
<td>22.37 ± 0.87</td>
</tr>
<tr>
<td>−/−</td>
<td>91</td>
<td>18.10 ± 0.53</td>
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The mean ± SEM number of MLH1 foci was determined by dividing the total number of MLH1 foci counted by the total number of nuclei examined from 3 independent preparations (3 mice/genotype).
8 of 9 littermate controls (Ews<sup>+/–</sup> and Ews<sup>–/–</sup>) survived beyond 120 days. We observed a similar hypersensitivity to various types of DNA damage in Ews<sup>–/–</sup> MEFs compared with MEFs derived from litter-matched controls (Supplemental Figure 3).

**EWS physically associates with nuclear lamin A/C.** Recent reports have shown that mutations in LMNA, which encodes A-type lamins, are responsible for a variety of diseases collectively termed laminopathies (reviewed in ref. 39), including Hutchinson-Gilford progeria syndrome (HGPS) (40, 41). The observed aging-like features and asynapsis of XY in Ews<sup>–/–</sup> mice are characteristics shared by Lmna<sup>–/–</sup> mice (42, 43). Therefore, we considered a possible link between EWS and lamin A/C by first examining their subcellular localization. Cells transfected with GFP-EWS fusion constructs showed mostly diffuse expression of GFP-EWS in the nucleoplasm, and although rare, some cells also showed colocalization of GFP-EWS with endogenous lamin A/C in the perinuclear region (Figure 6A, arrow). GFP-EWS was also found in punctate nuclear structures, which did not colocalize with lamin A/C. Lamin A/C was expressed in both perinuclear and intranuclear regions (Figure 6A). To demonstrate a direct in vivo association between EWS and lamin A/C, endogenous EWS was immunoprecipitated from HeLa nuclear extracts, and the presence of lamin A/C in the complex was examined by immunoblotting. As shown in Figure 6B, endogenous EWS and lamin A/C physically associated in vivo. We next examined the expression of endogenous lamin A/C in Ews<sup>–/–</sup> MEFs. Ews-deficient cells contained enlarged nuclei and showed markedly reduced intranuclear staining of lamin A/C compared with controls (Figure 6C, arrows). There was a consistent reduction in lamin A/C expression in all independently derived Ews-null fibroblasts compared with MEFs derived from litter-matched controls (Figure 6, D and E). The level of lamin A/C transcripts was similar in both control and mutant MEFs (data not shown), demonstrating that inactivation of EWS results in a reduction of lamin A/C at a posttranscriptional level.

**Discussion**

Ubiquitous RNA-binding protein EWS has been suggested to play fundamental roles in both transcription and splicing and is thus presumed to be essential for normal development. In the present study, we show that EWS is dispensable for embryonic development. Our characterization of Ews-null mice demonstrates that EWS is essential for pre–B lymphocyte development and meiosis. A severe reduction in cellularity of thymus in the EWS mutant animals also suggests a thymic development defect. Yet the relatively normal ratios of thymic subpopulations in the Ews<sup>–/–</sup> and Ews-null fetal liver–transplanted animals (Supplemental Figure 2) suggest that the reduction in thymic cellularity may be due to stromal or other defects. Furthermore, we have uncovered an unexpected role for EWS in cellular senescence, possibly through its interaction with nuclear lamin A/C. Although our results do not preclude a role for EWS in the fundamental processes of transcription and splicing, as other TET proteins could serve compensatory roles, our work clearly demonstrates that EWS has unique functions that other TET proteins cannot compensate.

**EWS has essential and nonredundant functions during B lymphocyte development and meiosis.** The members of the TET family, EWS, TLS/FUS, and TAF15/hTAFII68, are highly related proteins that can bind RNA as well as ssDNA in vitro (3). Evidence from human cancer studies suggests that at least the NTDs of EWS, TLS, and TAF15 are functionally interchangeable (5, 7, 14, 15, 44, 45). However, our analyses of Ews<sup>–/–</sup> mice clearly demonstrate a lack of functional redundancy between EWS and TLS. Similar to deletion of Ews, inactivation of Tls in mice leads to defects in B cell development (46), but the molecular details of EWS and TLS deficiencies...
are different. The B cell defect in Tls-null mouse is non–cell autonomous as demonstrated by fetal liver reconstitution experiments in which the B cell populations were fully reconstituted by the Tls-null donor cells (46). The reconstituted Tls−/− mature B cells, however, are dysfunctional in the primary antibody production in response to antigen stimulation. In contrast, Ews deficiency results in an intrinsic defect in pre–B cell development, specifically during the pro–B to pre–B cell transition, as demonstrated by flow cytometry, in vitro pre–B colony formation assay, and fetal liver chimera analysis (Figure 2).

The functional differences between EWS and TLS are also evident during meiosis. Ews−null spermatocytes show a high frequency of unsynapsed XY chromosomes but show normal formation of autosomal bivalents (Figure 4). In contrast, Tls−deficient spermatocytes display defects in the formation of autosomal bivalents (47). Expression of EWS is diffuse throughout spermatocytes (Supplemental Figure 4), while TLS expression is excluded from autosomal bivalents (Figure 3). It has been well established that absence or reduced numbers of crossovers can lead to improper segregation of homologous chromosomes and arrest in meiosis (30). Thus, reduction in crossover formation and XY asynapsis probably result in the massive apoptosis and meiotic arrest observed in Ews−deficient spermatocytes. Finally, loss of EWS leads to infertility in both sexes, but the TLS deficiency results only in male sterility (47). These observations demonstrate that despite sharing highly homologous sequences and the same interacting proteins (such as TFIID and RNA polymerase II), EWS and TLS serve nonredundant functions in vivo during B lymphocyte development and meiosis.

A role for the TET family of proteins in DNA recombination or repair? Ews deficiency in mice leads to developmental defects in meiosis and pre–B cell formation. Pleiotropic defects in lymphocyte development, meiosis, and cellular senescence (or aging) are features also observed in mice deficient in Atm and c-abl (reviewed in ref. 38), 2 genes that are critical to the DNA damage/repair pathway. These observations implicate a possible role of EWS in the DNA recombination and/or repair process. Consistent with this, Ews−null mice and MEFs showed increased sensitivity to DNA damage (Figure 5G and Supplemental Figure 3). Similarly, Tls−/− mice are also radiosensitive (47), and Tls−/− MEFs display genomic instability (46). In vitro, both EWS and TLS can promote homologous DNA pairing activity (47).

Another meiotic defect observed in the Ews−/− spermatocytes is a reduced number of meiotic crossovers (Figure 4 and Table 2), which has not been reported in the Tls−/− mice. Decreased meiotic crossovers in Ews−null spermatocytes may be an indirect consequence of XY asynapsis. However, we observed reduced crossovers even when a normal XY bivalent was present. Furthermore, some bivalents showed a complete absence of crossovers (Figure 4C), suggesting that EWS may play a direct role in meiotic recombination. We also observed an aberrant segregation of chromosomes in Ews−/− spermatocytes (Figure 3A, arrow). It has been well established that absence or reduced numbers of crossovers can lead to improper segregation of homologous chromosomes and arrest in meiosis (30). Thus, reduction in crossover formation and XY asynapsis probably result in the massive apoptosis and meiotic arrest observed in Ews−deficient spermatocytes. Finally, loss of EWS leads to infertility in both sexes, but the TLS deficiency results only in male sterility (47). These observations demonstrate that despite sharing highly homologous sequences and the same interacting proteins (such as TFIID and RNA polymerase II), EWS and TLS serve nonredundant functions in vivo during B lymphocyte development and meiosis.
Loss of EWS leads to cellular senescence and identification of lamin A/C as an EWS-interacting protein. We uncovered an unexpected role of EWS in cellular senescence in Ews−/− fibroblasts (Figure 5). Cellular senescence can be provoked by telomere attrition when the telomere shortening reaches a critical threshold (35). However, this is unlikely given the early onset of senescence in EWS mutant cells and the presence of long telomeres in mice. The 2 major pathways involved in cellular senescence are p53 and RB, but we did not find any alterations in the levels of p53, p21, p19ARF, and p16INK4a in Ews−/− MEFs. We did observe a marked reduction in the pRB levels in the mutant cells, but this is more likely to result in proliferation than cellular senescence. These results indicate that the cellular senescence in Ews-null cells may be independent of the p53 or RB pathways or may affect these pathways in subtle ways.

Our observations suggest another mechanism by which loss of EWS leads to early onset of cellular senescence. We provide evidence for an endogenous association of EWS with nuclear lamin A/C, an intermediate-filament protein of nuclear lamina. Nuclear laminas are located in the inner nuclear membrane and form the basis of the nuclear matrix that maintains the physical structure of the nuclear envelope as well as playing important roles in transcription, DNA replication, recombination and repair, RNA splicing, and nuclear export (39). It has been well established that the mutations in Lmna, which encodes lamin A/C, result in a wide spectrum of diseases including premature aging syndromes, HGPS (40, 41), and atypical Werner syndrome (50–52). Mice with targeted mutations in Lmna show a premature aging phenotype (42) and impaired spermatogenesis with XY asynapsis defect (43). Furthermore, a recent report has demonstrated that human fibroblasts derived from old donors as well as fibroblasts derived from HGPS show greatly reduced intranuclear lamin A/C expression compared with fibroblasts from young donors (53). We observed a strikingly similar reduction in intranuclear lamin A/C expression in Ews-null fibroblasts (Figure 6C), indicating that the early onset of senescence may be due to decreased lamin A/C expression and function. Finally, cell fractionation studies have shown that EWS is a component of nuclear matrix (19). Thus, we hypothesize that EWS inactivation causes early onset of cellular senescence by reducing lamin A/C expression, which may lead to nuclear lamina dysfunction.

In conclusion, we demonstrate that EWS is essential for B cell development and meiosis. Moreover, our findings reveal previously unanticipated roles for EWS in cellular senescence and in nuclear lamin A/C expression. The presence of EWS and TLS in the nuclear matrix (19, 54) raises an intriguing possibility that other TET family proteins, especially TLS, might also be involved in nuclear lamina function. Our findings may also be relevant to EWS-related cancers in which EWS haploinsufficiency is generated by the EWS chromosomal translocation, and the potential dominant-negative effects of the EWS-fusion products on the endogenous EWS activity may contribute to tumorigenesis.

Methods

Generation of the targeting vector and Ews-deficient mouse. 129SvEv mouse genomic library was screened to obtain Ews genomic DNA. We inserted a loxp-flanked transcriptional stop cassette (29) into Ews intron 6 (Figure 1A). A human WT1(KO) cDNA sequence was fused to Ews exon 7 to create EWS/WT1 knock-in allele. A 5.5-kb SalI and BglII fragment containing Ews exons 12 to 14 was used as the other targeting arm, resulting in a deletion of Ews exons 8 to 11 following homologous recombination. The targeting construct was electroporated into TC-1 mouse ES cells (55). Ganciclovir- and G418-resistant clones were isolated, screened by PCR for homologous recombination, and confirmed by Southern blot analysis with a probe derived from Ews intron 5, which was PCR amplified using the primers 5’-CATGGCTAGTTAGTAGATAAG-3’ and 5’-GGGGATACATGCGT- GACTTACG-3’. Positive ES clones were injected into C57BL/6j blastocysts, and the resulting chimeras were crossed with Black Swiss females (Taconic) to generate F1, heterozygotes, which were interbred to generate homozygous Ews−/− mice. Genotyping was performed by PCR analysis of tail DNA using 2 sets of primers: wild-type (411-bp PCR product): 5’-TGGATCCTTA- CAGCGACCTCC-3’, 5’-TGTCCTGATGCTGTTGAGGAC-3’; mutant (237-bp PCR product): 5’-TGGATCCTCACGCAAGCCTCC-3’, 5’-CTCTGATGATGCCTGGTGGTGTGC-3’ (40 cycles of 94°C for 45 seconds, 58°C for 45 seconds, and 72°C for 45 seconds). Animal care and use was approved by the NIH Animal Research Advisory Committee.

Generation of rabbit polyclonal antibodies against EWS. Two N-terminal regions of mouse Ews were amplified by PCR and subcloned into pGEX3X (Amersham) to generate GST-EWS fusion proteins containing either residues 1–74 or 68–171 of EWS. Both GST-EWS fusion proteins were expressed in E. coli and purified using GST-Sepharose beads (Amersham). Two rabbit polyclonal antibodies were raised by immunizing rabbits with either purified GST-EWS1–74 or GST-EWS68–171 fusion proteins (Strategic Biosolutions). The specificity of crude sera and affinity-purified antibodies was verified by immunoblotting (data not shown and ref. 56).

Flow cytometry. Thymocytes, splenocytes, and bone marrow cells were harvested from 3-week-old animals using standard methods. Red blood cells were removed using NH4Cl lysis buffer (Sigma-Aldrich). Total cell number was determined using a hemocytometer. For flow cytometry, cells were suspended in 1% BSA-PBS and incubated with anti-FcR antibody (BD Biosciences). Cells were then incubated with indicated antibodies on ice for 30 minutes and washed and analyzed by flow cytometry using a FACS-Calibur (BD Biosciences). The following antibodies (BD Biosciences) were used: FITC-labeled anti-CD43, FITC-labeled anti-IgD, FITC-labeled anti-CD45.2, R-PE-labeled anti-CD45.1, PE-labeled anti-IgM, peridinin chlorophyll-a protein–labeled anti-B220, and allophycocyanin-labeled anti-IgM.

Fetal liver transplantation and in vitro CFU assays. For fetal liver transplantation experiments, embryos were obtained at E14.5 from Ews−/− intercrosses (CD45.2+, 129SvEv/Black Swiss hybrid), and fetal liver cell suspensions were injected (5 × 106) into tail veins of irradiated Rag−/−CD45.1− recipient mice (B6.SJL, Taconic) housed under pathogen-free conditions with medicated water. The genotypes were determined retrospectively by PCR. Six weeks later, tissues were harvested and analyzed by flow cytometry for donor-derived CD45.2+ B and T cell lineages. For CFU assays, fetal liver–transplanted bone marrow cells were plated onto 35-mm dishes in duplicate in a semisolid methylcellulose medium containing either pre-B differentiation cocktail (M3630) or multilineage differentiation cocktail (M3534; StemCell Technologies Inc.). Cells were plated at 2 × 104 per plate for the CFU-pre-B assay or at 1 × 104 per plate for the GM-CFU assay. The cultures were placed in an incubator (37°C, 5% CO2) for 1 week, and colonies were counted under a microscope.

Histological analysis and TUNEL assay. Mouse tissues, including testes and ovaries, were isolated, fixed in 10% formalin solution (Sigma-Aldrich), embedded in paraffin, and sectioned for histology (H&E). The TUNEL assay was performed with the TdT-FragEL DNA Fragmentation Detection Kit (EMD Biosciences) following the manufacturer’s instructions.

Spermatocyte preparations and immunofluorescence microscopy. For spermatocyte preparations, 4- to 6-week-old mice were used. Spermatocyte surface–spread preparations and immunofluorescent staining were performed as described previously (57). For double staining, mouse or rabbit antibodies against SCP3 (Abcam) were used with either rabbit antibody against γ-H2AX
Bone x-ray imaging and mouse irradiation. Mice were euthanized by CO₂, and the x-ray images of whole animals or the long bones were obtained using MX-20, a Faxitron X-ray Corp. instrument. X-ray doses used were 20 kV for 100 seconds for whole-body images and 16 kV for 100 seconds for the dissected long bones. For whole-body mouse irradiation, 6- to 10-month-old animals were γ-irradiated at a dose of 7 Gy (GammaCell 1000 irradiation) and the animals were monitored for survival. Moribund animals were euthanized and were considered dead on the day of the sacrifice.

Transfection, immunoprecipitation, and immunoblot analyses. PC3 prostate cancer cells were transfected with either pEGFP-C1 (BD Biosciences) or pEGFP-C1-EWS using Lipofectamine 2000 (Invitrogen). After 48 hours, cells were fixed and immunostained with goat anti–lamin A/C antibody (Santa Cruz C1-EWS) using Lipofectamine 2000 (Invitrogen). After 48 hours, cells were counted and replated at 5 × 10⁷ cells/plate. Cumulative cell numbers were calculated at each passage. For cellular senescence, passage 3 MEFs were seeded at a density of 5 × 10⁷ MEFs, and 3 days later, cells were fixed and stained for the senescence-associated acidic β-galactosidase activity as described previously (34).

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