Pancr...
stem cells (26). The expression of such a diversity of intermediate filament proteins highlights that PaSCs have a broad range of potential properties, including contractility, the presence of cellular extensions to sense their environment, the potential to elaborate ECM components, and the potential to proliferate. However, it is important to note that these markers have clear limitations (see below) and that there are species differences (Table 1). Activation of quiescent PaSCs, which occurs when primary PaSCs are cultured and in the pancreas as a consequence of pancreatic injury, is associated with several morphologic changes (3, 4), including nuclear enlargement and enhanced prominence of the ER network (Table 1). Furthermore, in situ hybridization and immunohistochemical studies indicated that activated PaSCs express α-SMA (also known as ACTA2) (Figure 2, C–E) and collagen type I, therefore marking these cells as a source of fibrosis in chronic pancreatitis and pancreatic adenocarcinoma (14, 27, 28).

Initial efforts to isolate PaSCs produced cells that expressed α-SMA and collagen I, collagen III, and collagen IV (29). However, use of the isolation protocols used in HSC studies, in particular density gradient centrifugation, allowed the isolation of quiescent PaSCs (3, 4). Isolated PaSCs are characterized as quiescent by the presence of desmin, GFAP, and intracellular fat droplets, but the absence of α-SMA (Table 1). Primary PaSCs become activated during culture and attain a myofibroblast-like phenotype characterized by the disappearance of intracellular fat droplets, and the expression of α-SMA and ECM proteins (collagen I, collagen III, and fibronectin) (3, 4) (Table 1). PaSCs also seem to increase their expression of nestin upon activation (30). One potential problem in the identification of PaSCs is that the markers used can also be expressed by other cell types, including PaSC precursors, (myo)fibroblasts, vascular cells, and neural cells. However, on the basis of these markers, PaSCs and HSCs are estimated to constitute nearly 4% and 8% of total pancreatic and hepatic cells, respectively (3, 21).

Although much has been learned from cultures of primary PaSCs, one of the dilemmas in the study of these cells is whether the transformation observed in culture is equivalent to what happens in pancreatic tissue (31). In culture, primary PaSCs continually change from a quiescent to an activated phenotype, and during this change they pass through a series of temporal states of transformation (32). For example, rapidly proliferating PaSCs in culture can either die by apoptosis or acquire a (myo)fibroblastic differentiated state that is more resistant to apoptosis. But how these observations relate to PaSC activation in vivo is unclear, which emphasizes the need for more information about the phenotypic states of PaSCs during disease progression and the mechanisms underlying the conversions between these states.

The propagation of immortalized PaSCs from rat and human pancreata provides additional experimental models to study PaSC biology (33–35) and provides a tool for overexpression and RNA interference studies, as well as a tool for high-throughput screening for compounds that affect PaSC activation. Immortalized cell lines have been generated by expression of either SV40 large T antigen alone in rat PaSCs or SV40 large T antigen and human telomerase in human PaSCs. The resultant immortalized cell lines possess a phenotype consistent with activated PaSCs, which includes expression of α-SMA and ECM proteins. DNA microarrays have been used to compare the gene expression profile of immortalized and primary cultures of rat PaSCs. These revealed only a few overall differences, including differences in the expression of genes encoding ECM-related proteins, cytokines, integrins, and intermediate filament proteins (34). In addition, both rat and human immortalized cell lines responded to TGF-β1,
review series

Table 1

<table>
<thead>
<tr>
<th>General features of quiescent (inactive) and activated PaSCs</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Morphologic features</strong></td>
</tr>
<tr>
<td>Vitamin A autofluorescence, + (in fat droplets)</td>
</tr>
<tr>
<td>Endoplasmic reticulum, +</td>
</tr>
<tr>
<td>Nucleus, Basal size, +</td>
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<tr>
<td><strong>Molecular markers</strong></td>
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<tr>
<td>Vimentin, +</td>
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<tr>
<td>Desmin, +</td>
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<tr>
<td>GFAP, +</td>
</tr>
<tr>
<td>Nestin, +</td>
</tr>
<tr>
<td>α-SMA, +</td>
</tr>
<tr>
<td><strong>Properties</strong></td>
</tr>
<tr>
<td>ECM production, Limited</td>
</tr>
<tr>
<td>Cell proliferation, Limited</td>
</tr>
<tr>
<td>Cell migration, Limited</td>
</tr>
<tr>
<td><strong>Absence</strong> (-), presence (+), present at high levels (+)</td>
</tr>
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Note: PaSCs, pancreatic stellate cells; HSCs, hepatic stellate cells; α-SMA, α-smooth muscle actin; ECM, extracellular matrix.

PDGF, and the PPARγ ligand PGJ2, in a manner similar to that of their cultured primary cell counterparts (34–36). The combined use of cultured primary PaSCs and immortalized cells, coupled with the use of coculture systems (for example, coculture of acinar cells and PaSCs), is likely to provide additional mechanistic insights into the biology of PaSCs.

Mediators of PaSC activation

The critical regulatory events that orchestrate the persistent activation of PaSCs in vivo are likely to be similar, at least in part, to the events that regulate the activation of primary PaSCs in culture. Studies of human and rodent primary PaSCs in culture have identified numerous growth factors, cytokines, hormones, intracellular signaling molecules, and transcription factors as regulators of PaSC activation. Potential activators of PaSCs in vivo include paracrine factors, such as cytokines (IL-1, IL-6, IL-8, and TNF-α), growth factors (PDGF and TGF-β1), angiotensin II, and reactive oxygen species released by damaged neighboring cells and leukocytes recruited in response to pancreatic injury (37–43) (Figure 3). Activated PaSCs, in turn, can produce autocrine factors, such as PDGF, TGF-β1, cytokines (IL-1, IL-6, and TRAIL), and proinflammatory molecules (COX-2), that can perpetuate the activated phenotype (40, 42, 44, 45). In addition, activin-A, a member of the TGF-β family of soluble factors, also functions in an autocrine manner, increasing collagen secretion and augmenting TGF-β1 expression and secretion (46). Also, endothelin-1 is expressed by rat PaSCs in primary culture and can stimulate their migration and contraction (47). Although several inflammatory mediators released during pancreatitis have the potential to regulate PaSCs, abundant experimental evidence supports major roles for PDGF (which induces the proliferation of PaSCs and contributes to the migration potential of PaSCs) and TGF-β1, and angiotensin II (which induce PaSCs to express α-SMA and ECM proteins) as modulators of the persistently activated and profibrotic phenotype of these cells (16–18, 37, 41) (Figure 3).

Oxidative stress and ethanol metabolites have also been suggested as potential perpetuators of the activated PaSC phenotype. Ethanol can be metabolized in pancreatic acinar cells, resulting in toxic metabolites and oxidative stress that can induce pancreatic damage (48). Cultured rat PaSCs exhibit ethanol-induced alcohol dehydrogenase activity in vitro, suggesting that PaSCs can also metabolize ethanol (48). In vitro studies indicate that ethanol and its metabolite acetaldehyde promote the activation of rat PaSCs and cause lipid peroxidation in these cells (48, 49). Moreover, the antioxidant vitamin E prevents ethanol- and acetaldehyde-induced activation of PaSCs, thereby suggesting that oxidative stress regulates PaSC activation. For example, 4-hydroxy-nonenal, a highly reactive product of lipid peroxidation, activates primary rat PaSCs in culture. These in vitro findings were supported by histological analysis of pancreatic sections from patients with chronic pancreatitis, which showed 4-hydroxy-nonenal staining localized to activated PaSCs within fibrotic areas and to acinar cells adjacent to areas of fibrosis (28).

Multiple studies have identified several major signaling pathways involved in the regulation of PaSC function (27, 43, 47, 50–52). MAPKs are key mediators of activating signals initiated by growth factors, angiotensin II, and ethanol (51, 52). Other signaling pathways regulating PaSC activation include PI3K, RHO kinase, the JAK/STAT pathways, the activator protein-1 and NF-κB pathways, and the TGF-β1/SMAD-related pathways (47,
In addition, studies using PPARγ ligands have implicated this pathway in the downregulation of PaSC activation (27). These signaling pathways are potential therapeutic targets for the modulation of PaSC function.

**PaSCs and pancreatic inflammation**

Extensive studies have shown that activated PaSCs in culture express several growth factors and cytokines, as well as the receptors for these molecules, which are known to participate in inflammatory and fibrotic processes. Although the cellular and molecular mechanisms underlying the development of pancreatitis are not completely understood, it is generally accepted that pancreatitis is initiated by damage to acinar, ductal, and/or mesenchymal cells in the pancreas (7, 54, 55). Following damage, pathologic processes that occur in the pancreas include interstitial edema, necrosis of parenchymal cells, intrapancreatic trypsin activation, inflammatory cell infiltration, and activation and proliferation of PaSCs. In human and rodent pancreatic tissues, activated PaSCs are usually found in areas of extensive necrosis and inflammation where an environment rich in cytokines, growth factors, and reactive oxygen species prevails (56–58). Time-course studies using several animal models of experimental pancreatitis (59–61) indicate that parenchymal necrosis and inflammation precede PaSC activation, thereby suggesting that a necrotic, inflammatory process is a prerequisite for activation of these cells. Hence, autocrine and paracrine mediators are probably involved in PaSC activation. In turn, activation facilitates PaSC proliferation, migration, and ECM deposition, which leads to fibrosis or ECM remodeling as part of a repair process (Figure 3).

Evidence of a pivotal role for activated PaSCs in the development of pancreatic fibrosis is based, in part, on the analysis of pancreatic sections from patients with chronic pancreatitis and from experimental pancreatic fibrosis in rodents. For example, α-SMA–expressing cells are abundant in areas of fibrosis in pancreatic tissue sections from patients with chronic pancreatitis of different etiologies (4, 27, 28, 58). In these fibrotic areas, only α-SMA–expressing cells produce mRNA encoding procollagen α1(I), indicating that activated PaSCs are probably the predominant source of collagens during pancreatic fibrosis (27). Further evidence for this conclusion comes from the observation that spontaneous chronic pancreatitis in the Wistar Bonn/Kobori (Wbn/Kob) rat is characterized by inflammation and PaSC activation in areas of pancreatic fibrosis (50). In addition, in rats, administration of trinitrobenzene sulfonic acid into the pancreatic duct, as well as i.v. injection of dibutylin dichloride, induces necrosis and inflammation of the pancreas followed by PaSC activation and fibrosis (27, 62). Similarly, mouse pancreatitis induced with the use of cerulein (a cholecystokinin analogue and pancreas secretagogue) is also accompanied by PaSC activation and fibrosis (63, 64).

The involvement of PaSCs in the regulation of the inflammatory response during pancreatitis occurs, at least in part, through their production of chemokines and cytokines, as has been reported for myofibroblast-type cells in other tissues (65–68). PaSCs have also been shown to have phagocytic activity in vivo and in cell culture (69) and therefore might also function as resident phagocytic cells during pancreatitis. PaSC phagocytic activity is regulated by PPARγ and expression of CD36, a scavenger receptor that promotes phagocytosis. Interestingly, profibrogenic cytokines, such as...
as TGF-β, TNF-α, and IL-1, decreased the phagocytic capacity of PaSCs activated in culture (69). These results highlight the complex regulation of the profibrotic and proinflammatory properties of PaSCs during pancreatic inflammation.

**Inflammation and PaSC modulation of the ECM**

PaSCs have a key role in the extensive tissue fibrosis that accompanies chronic pancreatitis and leads to destruction of the pancreas and loss of exocrine function (17, 56, 58). Conversely, observational studies have shown that activated PaSCs participate in tissue repair processes after acute necrotizing pancreatitis in humans and experimental acute pancreatitis in rodents (59–61, 70). Although the exact role of PaSCs in pancreatic repair and regeneration remains to be elucidated, activated PaSCs and other myofibroblast cells resident in the pancreas probably contribute to the formation of a provisional matrix at the site of injury that allows cell proliferation, migration, and assembly of new parenchymal cells (60, 61, 71). In addition, PaSCs can regulate ECM remodeling during pancreatic tissue repair through the production of ECM-degrading proteases and their inhibitors, such as tissue inhibitor of metalloproteinase 1 (TIMP-1) (40, 72). In this context, resolution of mouse cerulein-induced pancreatitis involves transient activation of PaSCs and deposition of ECM proteins, as well as transient upregulation of MMPs and TIMP-1 (61). Similar regulation of ECM remodeling by HSCs is also likely to mediate liver regeneration after experimental injury (20, 73).

In most studies in which PaSCs are activated after damage to the pancreas, the inflammatory process resolves and activated PaSCs progressively disappear after the cessation of the injurious agent. However, repeated pancreatic damage and failure of the mechanisms regulating tissue repair can both lead to chronic inflammation, persistent activation, proliferation of PaSCs, and finally fibrosis. Consistent with this hypothesis, repeated episodes of acute experimental pancreatitis produce changes that resemble those found in chronic pancreatitis (63, 74). In fact, fibrosis in the pancreas and other organs can be considered the consequence of a wound-healing process responding to chronic injurious stimuli. In humans, repeated damage to the pancreas is associated with chronic alcohol consumption, pancreatic duct obstruction, metabolic disorders, and genetic defects (7, 75). The chronic injury results in perpetuation of the activated PaSC phenotype. Furthermore, chronic pancreatitis is associated with reduced production of MMPs by PaSCs, which probably helps promote and sustain the fibrotic phenotype (40).

In addition to the multiple factors described above, other mechanisms can explain the persistent activated state of PaSCs during pancreatitis. For example, activated PaSCs express protease-activated receptor-2 (PAR-2), which is cleaved by trypsin (a key pathogenic protease in pancreatitis) to become active. Active PAR-2 stimulates PaSC proliferation and collagen synthesis (76). Changes in the composition of the ECM during repair processes can also modulate PaSC activation. For example, activated PaSCs revert to a quiescent state when cultured on a basal membrane–like matrix, thereby suggesting that ECM composition regulates PaSC behavior (35, 40). Studies of liver fibrosis have shown that extensive ECM degradation is accompanied by apoptosis of HSCs, as a result of either increased proapoptotic signaling or reduced survival signals from the ECM (77), but it remains to be shown whether this is also true in the pancreas. More recently, impaired extracellular proteolysis of the ECM in mice lacking plasminogen has been associated with persistent PaSC activation and accumulation of pancreatic collagens during the recovery phase of mouse cerulein-induced pancreatitis (61). Moreover, levels of pancreatic TGF-β₁, plasminogen activator inhibitor-1, and TIMP-1, factors that promote fibrosis (37, 77), were elevated persistently in plasminogen-deficient mice after treatment with cerulein but only transiently in similarly treated plasminogen-sufficient mice (61).

**PaSCs and pancreatic cancer**

Ductal pancreatic adenocarcinoma, which is the most common form of pancreatic cancer, is the fourth leading cause of cancer-related death, with a 5-year survival of less than 5% (78). Several tumors, and in particular pancreatic adenocarcinomas, are characterized by “tumor desmoplasia,” a remarkable increase in connective tissue that infiltrates and envelopes the neoplasm (79). Activated PaSCs in the tumor desmoplasia of human pancreatic cancers express α-SMA and colocalize with mRNA encoding procollagen α₁(14) and are probably major contributors of the ECM proteins that constitute the desmoplasia (12, 14, 80–84). Of note, analysis of gene expression in human pancreatic adenocarcinoma, chronic pancreatitis, normal pancreas, and pancreatic cancer cell lines demonstrated that 107 genes predicted to be expressed in the stromal compartment were found in both pancreatic adenocarcinoma and chronic pancreatitis (81). Furthermore, isolation and characterization of stromal cells from human pancreatic adenocarcinoma and alcohol-induced chronic pancreatitis samples demonstrated that cells from both sources had the same characteristic morphology, cytofilament expression, and capacity to synthesize ECM proteins (84). These results demonstrate that these 2 disorders contain common stromal elements and suggest similar mechanisms underlying the development of fibrosis in chronic pancreatitis and the desmoplasia in pancreatic adenocarcinoma.

Evidence is emerging that there is a symbiotic relationship between pancreatic adenocarcinoma cells and PaSCs that results in an overall increase in the rate of growth of the tumor (Figure 4). For example, culture supernatants from human pancreatic tumor cell lines stimulate PaSC proliferation and production of ECM proteins (14, 82–84). In addition, the growth rate of tumor cells injected s.c. into nude mice (mice that lack T cells and are severely immunocompromised) is markedly increased when PaSCs are included in the inoculum (84). In contrast to the tumors that form when only cancer cells are injected, the tumors that form when both cancer cells and PaSCs are used have a desmoplasia similar to that observed in human pancreatic adenocarcinoma (84). Furthermore, pancreatic tumor cells induce the proliferation of PaSCs by secreting PDGF and induce PaSC production of ECM proteins by secreting TGF-β₁ and FGF-2 (84).

Although studies using animals indicate that pancreatic tumor cells and PaSCs promote each other’s proliferation, supporting data from human pancreatic tumors are limited and inconsistent. One report shows that more extensive intratumoral fibroblastic cell proliferation correlates with a poorer disease outcome (85). Another report shows that patients with a better outcome have increased expression of connective tissue growth factor (CTGF) in fibroblasts surrounding the human pancreatic tumor cells (86). CTGF has been implicated in the pathogenesis of fibrotic diseases, and its expression is predominant in PaSCs and is regulated by TGF-β. These early findings are intriguing and suggest that further work is needed to delineate the role of PaSCs and their regulators in pancreatic cancer.
The mechanisms by which PaSCs and the desmoplasia enhance the growth of tumor cells in the adenocarcinomas are complex and only partly understood. One role of the desmoplasia is to promote survival and prevent apoptosis of the tumor cells through a direct action of ECM proteins on the tumor cells (87, 88) (Figure 4). The prosurvival effects of the ECM proteins laminin and fibronectin are mediated through their integrin receptors, which are expressed by the tumor cells. In addition, the effects of fibronectin seem to be mediated through transactivation of IGF-1. Overall, these interactions lead to the activation of prosurvival and progrowth signaling pathways in pancreatic tumor cells (88, 89).

Another possible mechanism by which tumor desmoplasia might promote pancreatic adenocarcinoma cell growth is that PaSCs and tumor cells produce MMPs and tissue serine proteases (Figure 4), such as members of the plasminogen activator system, that degrade ECM proteins (61, 72, 90). In this context, MMPs and tissue serine proteases might promote tumor cell invasion and metastasis, as has been postulated in other cancers (90–93).

**Comparison of PaSCs and HSCs**

Although PaSCs and HSCs share several morphologic and marker features, some differences exist. Unique, context-related features of HSCs include the portal perfusion and generally abundant vascular flow in the liver, and their proximity to encounter endothelial cell cross-talk due to their hepatic location within the subendothelial space (19). In addition, a genome-wide assessment of gene expression that included 21,329 genes identified 29 that were differentially expressed between cultures of primary HSCs and PaSCs (94). However, the fact that the overwhelming majority of genes showed no difference in expression supports the notion that these 2 cell types are very similar. Recent studies have revealed that the bone marrow is the source of 68% of HSCs and 70% of myofibroblast cells in mouse models of carbon tetrachloride- and thioacetamide-induced fibrosis (95). Whether the same holds true in the pancreas remains to be determined, but a common origin in the bone marrow could account for the strong similarities between HSCs and PaSCs.

Although this Review has concentrated on stellate cells, other potential sources of fibrosis are present in both the pancreas and the liver. For example, hepatic myofibroblasts, which are distinguished from HSCs by their location within the liver and their expression of the ECM component fibulin-2 but not GFAP (96, 97), and tissue fibroblasts, which express vimentin but not GFAP and desmin, represent an additional source of ECM proteins (96–100). Subpopulations of fibroblastic cells, which might reflect distinct levels of fibroblast activation, can be distinguished in cirrhotic livers on the basis of differing levels of cellular retinol-binding protein-1 and α-SMA (101), and these cells might also produce ECM proteins. The origin of these liver myofibroblast-type cells and similar cells in the pancreas remains to be conclusively determined. Bone marrow transplantation in a mouse model of pancreatic insulinoma demonstrated that the bone marrow is the source of 25% of pancreatic myofibroblasts and is likely to contribute to tumor-associated fibrosis (102). In addition, nestin-positive and nestin-negative precursors isolated from adult mouse pancreas are able to differentiate in vitro to yield multiple lineages of the pancreas, including PaSCs (103). The relative contributions of bone marrow and resident precursors to the PaSC population, as well as to the other myofibroblast- and fibroblast-related cell populations, remain to be determined.

The function of quiescent PaSCs is still being investigated, but clues can be gleaned from studies involving HSCs (20–22). For example, HSCs have been implicated in several important liver physiologic roles, under basal conditions, that include vitamin A storage; production and turnover of normal ECM proteins; and production of paracrine factors to promote hepatocyte differentiation, and to regulate ductal and vascular pressures. Similar roles can be envisioned for PaSCs in the inactive state. For example, perivascular and periductal PaSCs (2) might regulate the pressure within these compartments by contraction mediated by endothelin-1 (47). The functions of PaSCs, by analogy to HSCs, should be envisioned not only in isolation but also in the context and under the influence of their microenvironment.

Although alcohol and/or its metabolites have been shown to similarly activate stellate cells of the pancreas and liver, it is unclear why substantial differences exist in an individual’s sus-
ceptibility to liver and pancreas fibrosis (Figure 5). Potential differences between the pancreas and the liver after injury, in addition to the apparent difference in the cellular microenvironment, include edema formation during pancreatitis; different ethanol metabolism in acinar cells and hepatocytes; and different lipid metabolism between acinar cells and hepatocytes (for example, “fatty” pancreas has not been described). Given that the stellate cell is an integral element of the onset of these diseases, microenvironment differences and other genetic and epigenetic modifiers related to stellate and/or other resident and infiltrating cells are likely to be important (Figure 5), but this hypothesis remains to be tested. For example, comparison of the response of PaSCs and HSCs isolated from the same animal with parallel comparative responses in different genetic backgrounds has not been reported. Furthermore, attempts at assessing genetic polymorphisms (for example, in enzymes such as cytochrome P450 2E1 and aldehyde dehydrogenase-2) that may promote susceptibility to alcoholic chronic pancreatitis or cirrhosis have not been overly revealing (104). However, an understanding of genetic polymorphisms that predispose to chronic pancreatitis (8, 75) or liver fibrosis (103, 106), or even protect from chronic pancreatitis (107), is being pursued. A systematic and broad-based genetic screening will be needed, using well-defined animal models and patient cohorts. Proteomic comparisons between PaSCs and HSCs, which are likely to be forthcoming, should also provide additional insights.

**Potential treatments for PaSC disorders**

In concept, treatments for disorders involving PaSCs (that is, chronic pancreatitis and pancreatic cancer) should target the key mechanisms involved in their activation and proliferation. For example, blockade of the receptors for PDGF, TGF-β, and angiotensin II, as well as blockade of the intracellular signaling pathways downstream of these receptors, is likely to provide therapeutic benefit (17, 51, 76, 108). In this regard, there are in vivo experiments showing the importance of the angiotensin II system in the development of pancreatic fibrosis (64, 108). There are also several reports from in vitro experiments using PaSCs that show key roles in the activation and/or proliferation process for MAPK pathways, in particular, ERK1/2, p38 kinase, and JNK (50, 52, 53, 66, 109, 110); PI3K and PKC (111); PPARγ (36, 112); NADPH oxidases (our unpublished observations); and ethanol metabolism to acetaldehyde (18). In these studies, inhibition of most of these pathways results in attenuation of the activation and proliferation of PaSCs, but activation of PPARγ seems to block PaSC activation (36, 112). It has also been reported that activated rat PaSCs express COX-2 when stimulated with TGF-β1 and other cytokines (45), as well as when stimulated with conditioned media from human pancreatic tumor cells (113). Pharmacological inhibition of COX-2 and blockade of the TGF-β1 signaling pathway decrease the expression of COX-2, α-SMA, and collagen I, suggesting that COX-2 might be a relevant therapeutic target for chronic pancreatitis and pancreatic cancer. Furthermore, efforts aimed at inducing PaSC transdifferentiation from an activated to a quiescent state and at inducing PaSC apoptosis are also attractive modalities. For example, administration of vitamin A (retinol and its metabolites) induces culture-activated rat PaSCs to become quiescent (114).

In vivo studies using animal models of chronic pancreatitis have demonstrated that several agents have beneficial effects (115), but few in vivo studies have assessed whether targeting PaSCs in the...
36. Jaster, R. et al. 2005. Periosome proliferator-activated receptor gamma overexpression inhibits pro...
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