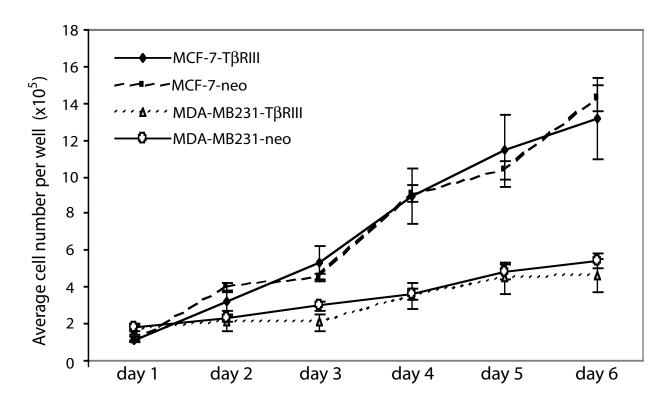


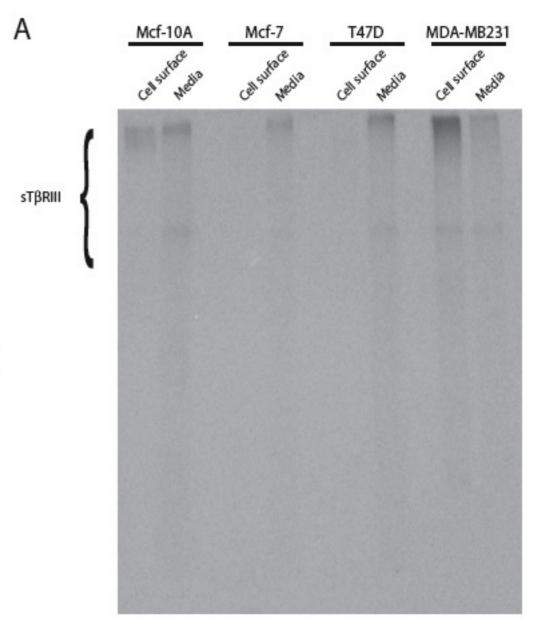
Supplementary Figure 1. Expression of T $\beta$ RIII in 4T1 cells. A [ $^{125}$ I]TGF- $\beta$ 1 binding and cross-linking assay followed by immunoprecipitation with anti-T $\beta$ RIII antibody was performed to confirm the expression of T $\beta$ RIII.

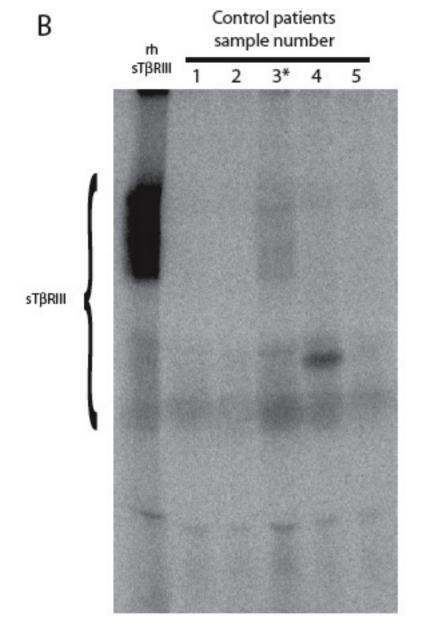
## Supplementary Figure 2. Dong et al., 2006

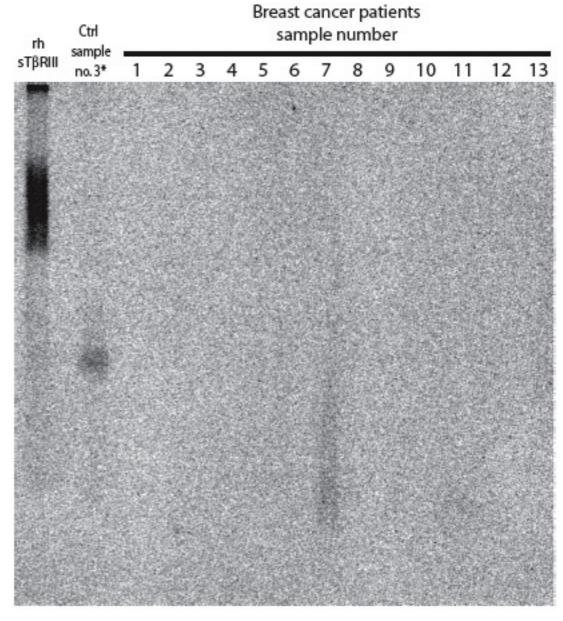


Supplementary Figure 2. Effect of T $\beta$ RIII on proliferation of breast cancer cells. 5,000 cells were seeded in a 6-well plate. After a 48-hour incubation, cells were collected and counted using a Coulter Counter every day for 6 days. This experiment was repeated in triplicate three times. Data is presented as mean  $\pm$  S.D.

Supplementary Figure 3. Soluble TβRIII expression in human breast epithelial and breast cancer cell lines; and in human plasma. A. 250,000 cells were seeded. After a 48-hour incubation, cells were placed in serum free media for 24 hours, conditioned media from each cell line collected, cross-linked with iodinated TGFβ-1 and sTβRIII specifically immunoprecipitated with an antibody to the extracellular domain.sTβRIII was detected in all breast epithelial and breast cancer cell lines tested. B. Human plasma (2.0 ml in left panel, 0.75 ml in right panel) from normal donors or breast cancer patients was cross-linked with iodinated TGFβ-1 and sTβRIII specifically immunoprecipitated with an antibody to the extracellular domain. Recombinant human sTβRIII (rh sTβRIII) was run as a positive control for both sets of samples. In addition, normal human donor plasma sample number 3 was run as a positive control for the breast cancer plasma samples (on right). TβRIII was detected in plasma from normal donors, but not in plasma from breast cancer patients.







exon 2	forward	CTCCAGCCTGGGTAACAGAG
	reverse	TGAAGTGACTGGACGACG
exon 3	forward	CCCCAGAAGAAAAGGA
	reverse	AGTGGTTTGATCACCCTTGC
exon 4	forward	TTTGGACTCTGGCATTATTTCA
	reverse	TTCCAGAGGCTGCTCTGAGT
exon 5	forward	TGATTCCTTGCCCTAACCAC
	reverse	TTATGATTCAGCTCCGGTGA
exon 6	forward	TGTAGGACGGCTACACCTC
exon 6	reverse	CTGCCATTTGCTCAGTTTCA
exon 7	forward	GAGAGTCCAAAGAGGCAGGA
exuii /	reverse	GAAGACTTGGAAGAGGGGAGA
exon 8	forward	TTCCTCACTGGAAAAGCTTCA
CAUITO	reverse	TCGTTCTTAGCCCAAGGAAA
exon 9	forward	AGATGCAGACTAGGGCCAGA
CAUII 3	reverse	GCAGAGGAGATGA
exon 10	forward	GGGGTGAAAACCCTCTTCAT
CAUIT IU	reverse	CCTTGGAAGACCAGGCAAT
exon 11	forward	TGAAGGATGAAGGCCCTGTA
CAUITII	reverse	GCAGAACCAAACACATGG
exon 12	forward	GAGCCAATCCCTCATCAGAC
CAUIT 12	reverse	CAGGGCCTTCATCCTTCATA
exon 13	forward	CCTGCAAGACCTTGGGATTA
CVOILID	reverse	AAAGCAGCGTGTCATCTGAA
exon 14	forward	CCTCCCAAAGCACACCTTTA
CAUIT 14	reverse	TCTCCCAGGTTATTTGGTGTTC
exon 15	forward	TTAGGAAAGCCCAGGAGGTT
CAUITIS	reverse	TGGAGAGCCTATGCAACTGA
exon 16	forward	GAGTGTCTATTGTGTGGCAGGA
CAUIT IU	reverse	TCTTCATTGCATTCTCCGATT
exon 17	forward	GAACCAAAATCCAGCCCTCT
CAUTI 17	reverse	AATGGCAAATGCGGAATAAA

Supplementary Table 1. Sequence of primers used to PCR amplify the 16 coding exons of human T  $\!\beta RIII$  .

## Supplementary Table 2, Dong et al., 2006

ΤβRΙ	forward reverse	ACGGCGTTACAGTGTCTG GGTGTGGCAGATATAGACC
ΤβRΙΙ	forward	AGCAACTGCAGCATCACCTC
	reverse	TGATGTCTGAGAAGATGTCC
ΤβΡΙΙΙ	forward	CTGTTCACCCGACCTGAAAT
	reverse	CGTCAGGAGGCACACTTA
GAPDH	forward	GAGTCAACGGATTTGGTCGT
	reverse	TTGATTTTGGAGGGATCTCG

Supplementary Table 2. Sequence of primers used for RT- PCR.