FGF-21 as a novel metabolic regulator

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Diabetes mellitus is a major health concern, affecting more than 5% of the population. Here we describe a potential novel therapeutic agent for this disease, FGF-21, which was discovered to be a potent regulator of glucose uptake in mouse 3T3-L1 and primary human adipocytes. FGF-21–transgenic mice were viable and resistant to diet-induced obesity. Therapeutic administration of FGF-21 reduced plasma glucose and triglycerides to near normal levels in both ob/ob and db/db mice. These effects persisted for at least 24 hours following the cessation of FGF-21 administration. Importantly, FGF-21 did not induce mitogenicity, hypoglycemia, or weight gain at any dose tested in diabetic or healthy animals or when overexpressed in transgenic mice. Thus, we conclude that FGF-21, which we have identified as a novel metabolic factor, exhibits the therapeutic characteristics necessary for an effective treatment of diabetes.

Introduction

While the majority of the 22 known members of FGF family have been primarily associated with mitosis, development, transformation, angiogenesis, and survival (1–5), recent data shows that they may play important roles in defining and regulating functions of some endocrine-relevant tissues and organs, as well as modulating various metabolic processes. For example, FGF-10 is implicated in the differentiation processes in white adipose tissue (6) and pancreas (7, 8), while FGF-16 (9) is considered to be a specific factor for brown adipocytes. Another recently characterized molecule, FGF-19 (10, 11), has been shown to cause resistance to diet-induced obesity and insulin desensitization and to improve insulin, glucose, and lipid profiles in diabetic rodents. Since these effects, at least in part, are mediated through the observed changes in metabolic rates, FGF-19 can be considered as a regulator of energy expenditure (12, 13).

FGFs modulate cellular activity via at least 5 distinct subfamilies of high-affinity FGF receptors (FGFRs): FGFR-1, -2, -3, and -4, all with intrinsic tyrosine kinase activity and, except for FGFR-4, multiple splice isoforms (1–3); and FGFR-5 (14, 15), which lacks an intracellular kinase domain. There is growing evidence that FGFRs can be important for regulation of glucose and lipid homeostasis. The overexpression of a dominant negative form of FGFR-1 in β cells leads to diabetes in mice, which thus implies that proper FGF signaling is required for normal β cell function and glycemia maintenance (16). FGFR-2 appears to be a key molecule during pancreatic development (17–19). Moreover, FGFR-4 has been implicated in cholesterol metabolism and bile acid synthesis (20).

FGF-21 (21) is a novel member of FGF family. It is preferentially expressed in liver, but an exact knowledge of FGF-21 bioactivity and its mode of action have been lacking to date. Here we show that FGF-21 is a potent activator of glucose uptake on adipocytes, protects animals from diet-induced obesity when overexpressed in transgenic mice, and lowers blood glucose and triglyceride levels when therapeutically administered to diabetic rodents. Thus, we believe our report to be the first to document a clear biological function of this protein and its potential therapeutic application.

Results

Identification of FGF-21 in vitro bioactivity. Using a glucose uptake assay to search for novel proteins with therapeutic potential to treat diabetes mellitus, we found that human recombinant FGF-21 stimulated glucose incorporation in differentiated mouse 3T3-L1 adipocytes, as well as in human primary adipocytes after 24-hour treatment of the cells with the protein (Figure 1, A and B). Since FGF-21 did not induce glucose uptake in undifferentiated 3T3-L1 fibroblasts, human primary preadipocytes, muscle L6–glucose transporter-4myc (L6–GLUT-4myc) myoblasts and myotubes (22), or liver clone 9 cells, the FGF-21 effect appeared to be adipocyte specific. The effects of FGF-21 on glucose uptake in adipocytes were insulin independent, additive to the activity of insulin upon cotreatment (Figure 1C), and not modulated by addition of exogenous heparin. In contrast to the rapid response elicited by insulin, the predominant effect of FGF-21 on glucose uptake required at least 4 hours of cell treatment, and it was substantially diminished in the presence of cycloheximide (1 μg/ml), a protein synthesis inhibitor (23) (Figure 1D). These observations led us to hypothesize that the mode of action for FGF-21 requires transcriptional activation.

To identify a potential mechanism at the molecular level by which FGF-21 increases glucose uptake, we examined whether it modulates the expression levels of the glucose transporters GLUT1 and GLUT4 in 3T3-L1 adipocytes. FGF-21 treatment (1 μg/ml) led to a significant increase in GLUT1 mRNA and protein but not those of GLUT4 (Figure 1, E and F). FGF-21–dependent GLUT1 upregulation was further demonstrated in vivo following a bolus injection in ob/ob mice. Four hours after s.c. administration of FGF-21 (500 μg/animal), an increase in GLUT1 mRNA was specifically detected in white adipose tissue but not in muscle, liver, kidney, and brain (Figure 1G).

Additional studies revealed that early FGF-21–induced signaling in 3T3-L1 adipocytes included heparin-independent tyrosine phos-
Phosphorylation of FGFR substrate-2 (FRS-2), a docking protein linking FGFRs to the Ras/MAPK pathway (24), and transient activation of MAPK (Figure 2A). All these in vitro activities are typically initiated upon activation of FGFR-mediated pathways (25).

FGFR-1 and FGFR-2 may represent FGF-21–corresponding receptors. The sequence characteristics and signaling profile of FGF-21 indicate that its receptor may belong to the FGFR superfamily. The expression of FGFR-1 and FGFR-2, including several splice variants of these receptors, was readily detected in 3T3-L1 cells by RT-PCR and immunoblot analysis. While FGFR-5 was also detected by RT-PCR, significantly lower levels of FGFR-3 and no expression of FGFR-4 were observed. We immunoprecipitated FGFR-1 and FGFR-2 from 3T3-L1 adipocytes with specific antibodies and detected in both cases 2 tyrosine-phosphorylated proteins of approximately 120–150 kDa in FGF-21–stimulated cells (Figure 2B), while no phosphorylation was observed in FGFR-3 immunoprecipitates under similar conditions (data not shown). These tyrosine-phosphorylated bands were later reprobed with anti–FGFR-1 and anti–FGFR-2 antibodies, respectively (Figure 2B), which indicates that they may represent activated forms of FGFR-1 and FGFR-2. Importantly, the observed FGF-21–dependent phosphorylation was adipocyte specific, since it was not detected in 3T3-L1 preadipocytes.

FGF-21 does not induce in vitro mitogenicity. As a class, FGFs are generally known to induce cell proliferation. Therefore, we examined the mitogenic potential of FGF-21 in cells typically sensitive to FGFs. FGF-21 did not induce proliferation of 3T3-L1, NIH 3T3, or BALB/c 3T3 fibroblasts, monkey epithelial 4MBr5 cells, primary human mammary epithelial cells (HMECs), or human umbilical vein endothelial cells (HUVECs), either in the absence or presence of exogenous heparin. In contrast, FGF-7, FGF-1, and FGF-2 stimulated the growth of these cells. Moreover, in costim-
FGF-21 administration (data not shown). At doses up to 4 mg/kg/d in mice and 8 mg/kg/d in rats, these molecules were shown to induce biological effects in vivo, we administered the protein to ob/ob mice, a model of hyperglycemia and insulin resistance. Mice were injected s.c. once daily with 125 or 750 μg/kg/d of FGF-21 for 7 days. Fed glucose levels were determined 1 hour after administration on days 3 and 7. Both doses significantly lowered blood glucose compared with vehicle treatment after 3 days of administration. The effect was even more pronounced after 7 days, with fed glucose levels being normalized in both dose groups (Figure 4A). Moreover, the plasma triglyceride levels exhibited a dose-dependent reduction after 7 days of FGF-21 injections (Figure 4B). Similar glucose-lowering effects were also observed upon FGF-21 administration in db/db mice (Figure 4C) and 8-week-old obese Zucker diabetic fatty (ZDF) rats (Figure 4D).

Remarkably, FGF-21 administration to ob/ob mice for 7 days had a sustained blood glucose lowering effect. Fed glucose levels in treated animals were 25–35% lower than in the control group 24 hours after last dose (Figure 4F). This in vivo observation was unexpected, as pharmacokinetic studies in ob/ob mice determined the elimination half-life of FGF-21 to be only 0.7–1.1 hours and indicates a mechanism for the observed effects in vivo, we generated FGF-21–transgenic mice that overexpressed the human protein from the liver using the apoE promoter. As measured by an FGF-21–specific ELISA, the plasma concentrations of FGF-21 in the transgenic animals ranged between 70 and 150 ng/ml.

These FGF-21–transgenic mice were viable and at 2 months of age had glucose levels similar to those of their wild-type littermates. However, at 9 months, differences between transgenic and wild-type mice became apparent. FGF-21–transgenic animals weighed significantly less, had lower fasted glucose levels (Table 1) and less fat in liver, retained more brown adipose tissue, had subcutaneous adipocytes of smaller size (Figure 5, A and B), and 8-week-old obese ZDF rats upon FGF-21 administration (data not shown).

Several members of FGF family have been shown to induce therapeutically undesirable in vivo proliferation of various cell types (1–5, 26). Therefore, we examined FGF-21–transgenic mice for their potential to develop tumors throughout their lifespan. As evidenced by histological analysis, transgenic mice overexpressing FGF-21 did not develop liver tumors or show evidence of any other tissue hyperplasia up to 10 months of age (Figure 5C).

To evaluate the potential for FGF-21 to induce hypoglycemia in rodents, we administered FGF-21 s.c. twice or once daily for 7 days to ob/ob, db/db, and C57BL/6 mice and 8-week-old obese and lean ZDF rats and measured blood glucose 1 hour after the last injection. At doses up to 4 mg/kg/d in mice and 8 mg/kg/d in rats, these diabetic or normal animals displayed no evidence of hypoglycemic effects in fed or even in the fasted state (Figure 4, A, C, F, and H).

FGF–21 transgenic mice. In order to evaluate the effect of enforced expression of FGF-21 in vivo, we generated FGF-21–transgenic mice that overexpressed the human protein from the liver using the apoE promoter.
We challenged FGF-21 transgenic animals by feeding them a high-fat/high-carbohydrate (HFHC) diet for 15 weeks. Intriguingly, FGF-21–transgenic mice consumed almost twice as much food as wild-type littermates when the amounts were normalized to body weights and calculated as the actual amount of food eaten per animal per day (Table 1). Despite the significant increase in caloric intake, they did not gain as much weight as wild-type controls and were also resistant to diet-induced obesity (Figure 6, A and B).

Discussion
Several secreted polypeptides, including insulin, glucagon-like peptide–1 (GLP-1), adiponectin, and others are ultimately involved in the regulation of glucose homeostasis, which thus makes them clinically relevant pharmacological agents or attractive candidates for novel medicines for the treatment of diabetes mellitus (27). Recent reports on bone morphogenic protein–9 (BMP-9) (28) and FGF-19 (12, 13), and our findings with FGF-21, have shown that the other proteins also constitute this list of promising biomolecules.

FGF-21 bioactivity was discovered through a cell-based functional screen aimed at identifying novel secreted molecules that affect glucose uptake on mouse 3T3-L1 adipocytes and was found to be very potent in this assay (EC50, ∼0.5 nM) (Figure 1A). With comparable potency, FGF-21 was active on differentiated human primary adipocytes (Figure 1B), which indicates that FGF-21 bioactivity is not limited to murine adipocytes.

The follow-up analysis on the initial observation in the glucose uptake assay revealed what we believe to be a novel and unique mechanism of the FGF-21 mode of action. FGF-21 effects appeared to be insulin independent and additive to insulin activity upon coadministration (Figure 1C). FGF-21 needed to be present on cells for several hours to produce a robust response in glucose uptake, and the effect was significantly diminished by the protein synthesis inhibitor cycloheximide (Figure 1D). While insulin is known to work in a rapid, hormone-like manner, we hypothesized that FGF-21 activity is likely to be mediated through changes in gene expression. Indeed, we showed that FGF-21 induced a signifi-
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The fact that FGF-21 induced tyrosine phosphorylation of FGFR-1 and FGFR-2 (Figure 2B) suggests that these molecules may function as FGF-21 receptors. Although they carry readily detectable and functional FGFR-1 and/or FGFR-2 molecules, it is, however, currently unclear why several FGF-sensitive cells that were tested for FGF-21 bioactivity do not respond to FGF-21 stimulation. Moreover, in preliminary experiments with all commercially available FGFR extracellular domain–Fc fusion proteins (R&D Systems), we were unable to demonstrate direct interaction between any of these FGFR variants and FGF-21, despite the fact that we clearly observed binding for both FGFR-1 and FGFR-2 (data not shown). Thus, FGF-21 may be physically interacting with different splice variants of FGFR-1 and FGFR-2 that are induced upon adipocyte differentiation. Alternatively, FGF-21–dependent activation of these receptors may require a cell–specific modification or additional cofactor.

Table 1
Metabolic parameters in FGF-21–transgenic and control mice

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control (M)</th>
<th>Tg (M)</th>
<th>P value</th>
<th>Control (F)</th>
<th>Tg (F)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight (g)</td>
<td>50.4 ± 0.7</td>
<td>29.1 ± 3.37</td>
<td>&lt;0.001</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Fasted glucose (mg/dl)</td>
<td>102 ± 5.5</td>
<td>67 ± 8.1</td>
<td>&lt;0.003</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Food intake (g/g body wt/wk)</td>
<td>0.42 ± 0.02</td>
<td>0.76 ± 0.03</td>
<td>&lt;0.001</td>
<td>0.55 ± 0.01</td>
<td>0.88 ± 0.04</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Leptin (ng/ml)</td>
<td>63.0 ± 1.97</td>
<td>15.6 ± 4.12</td>
<td>&lt;0.001</td>
<td>61.4 ± 3.64</td>
<td>18.26 ± 6.97</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Glucagon (pg/ml)</td>
<td>116 ± 7.12</td>
<td>93.8 ± 8.54</td>
<td>0.08</td>
<td>141 ± 9.5</td>
<td>93 ± 6.55</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Body temperature (°C)</td>
<td>35.58 ± 0.09</td>
<td>35.4 ± 0.27</td>
<td>NS</td>
<td>36.2 ± 0.23</td>
<td>36.5 ± 0.17</td>
<td>NS</td>
</tr>
</tbody>
</table>

*Parameters measured in 9-month-old mice on a regular chow. Parameters measured in diet-induced obese mice during or after HFHC feeding for 15 weeks. The values (± SE) shown are the average of the measurements of at least 5 animals in a group. M, males; F, females; Tg, FGF-21–transgenic mice; ND, not determined.

Figure 5
Histological analysis of FGF-21–transgenic and FGF-21–infused animals. (A) H&E staining of brown fat. Notice an increase in intensity of brown fat in the FGF-21–transgenic mouse compared with the wild-type mouse. (B) H&E staining of subcutaneous white fat. Notice the smaller adipocytes in the FGF-21 mouse compared with the wild type. (C) H&E staining of livers from FGF-21–transgenic and wild-type mice. PCNA immunostaining shows very low proliferation (less than 5%) of hepatocytes (brown staining, arrows) in both the control and treated groups. Magnification, ×400.
FGF-21 did not stimulate glucose uptake on insulin-sensitive liver clone 9 and muscle L6-GLUT-4myc cells and on 3T3-L1 fibroblasts. Also, we were unable to detect FGF-21 activity in proliferation assays on several cell lines and primary cells of a different nature, which further indicates that FGF-21 effects might be adipocyte specific. Nevertheless, we recently observed a clear FGF-21 response on cells of nonfat origin. Unexpectedly, FGF-21 (1 μg/ml) showed efficacy in modulating glucagon secretion from isolated rat islets (Figure 7), while no effect on insulin secretion was observed. Thus, the specificity of FGF-21 bioactivity remains to be further studied.

The administration of FGF-21 to diabetic ob/ob and db/db mice and obese ZDF rats led to significant lowering of circulating glucose and triglycerides, as well as a reduction in fasted insulin levels and improved glucose clearance during an OGTt (Figure 4, A–H). All these effects were observed after at least 3 days of injections and were more pronounced after 7 days of administration. Since no changes in levels of fed and fasted glucose, circulating lipid levels, insulin levels, and glucose disposal during OGTt were observed after a single s.c. injection of FGF-21, it appears that beneficial FGF-21-dependent effects require that animals be exposed to the protein multiple times. However, once the reduction in circulating glucose was achieved, FGF-21–induced changes were sustained for at least 24 hours (Figure 4F). Thus, despite its short elimination half-life, FGF-21 induced an extended pharmacodynamic effect in these diabetic animals. Taken together, these observations are remarkable in highlighting the difference in time action between FGF-21 and insulin.

Although potent in correcting elevated glucose levels in ob/ob and db/db mice, FGF-21 did not induce hypoglycemia in normal or diabetic rodents in either fasted or fed states (Figure 4, A, C–F, and H) at efficacious or significantly higher doses, and no hypoglycemia was seen in fasted FGF-21–transgenic mice (Table 1). This further distinguishes the effects of FGF-21 from those of insulin, which induced a significant reduction of blood glucose in lean animals (Figure 4E). Moreover, FGF-21 did not affect food intake or body weightcomposition of diabetic or lean mice and rats over the course of 2 weeks of administration (doses ranging from 25 μg/kg/d to 8 mg/kg/d).

Further insights into the FGF-21 mechanism of action can be gleaned from the phenotype of FGF-21–transgenic mice. These animals are viable and are not metabolically distinguishable from wild-type littermates at 2 months of age. However, they appeared to be resistant to the age-related impairment of glucose metabolism since they had lower plasma glucose levels at 9 months (Table 1). Moreover, when challenged on HFHC diet for 15 weeks, FGF-21–transgenic mice were resistant to diet-induced weight gain and fat accumulation (Figure 6, A and B), even though they consumed more food when the amounts were normalized to body weights (Table 1). We also observed lower levels of circulating leptin (Table 1). The reduction in leptin is consistent with lower adiposity in the transgenic animals and may be a primary cause of the increased food intake in the transgenic mice (31). However, these changes in feeding behavior are unlikely to have been induced by a direct effect of FGF-21, since no impact on food intake in rodents administered the protein was observed. There was also a decrease in circulating glucagon levels (Table 1), which is consistent with the in vitro observations made with rat pancreatic islets (Figure 7).

There was no evidence of poor nutrient absorption in FGF-21–transgenic animals. Thus, another potential reason for the observed resistance to diet-induced obesity may be an effect of FGF-21 on energy expenditure. However, if this is true, it was not reflected in any changes in rectal body temperatures (Table 1). Whether or not BAT activation may contribute to the effects of FGF-21 remains to be evaluated in future studies.

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The FGF family member most closely related to FGF-21 is FGF-19, with 31% amino acid sequence identity between these 2 molecules (21). The phenotype of transgenic mice overexpressing FGF-19 (12, 13) is strikingly reminiscent of that of mice overexpressing FGF-21. However, we have uncovered a fundamental in vivo difference that clearly distinguishes these 2 molecules from each other. While transgenic mice overexpressing FGF-19 were resistant to high-fat diet-induced obesity, they also developed histologically detectable liver tumors. Furthermore, wild-type mice that were injected with FGF-19 for 6 days had an increase in hepatocellular proliferation (26). In contrast to FGF-19–overexpressing animals, FGF-21–transgenic mice did not form tumors in liver or show histological evidence of hyperplasia in any other tissue after 10 months of age (Figure 5C). Nevertheless, in order to further determine the potential of FGF-21 to stimulate liver mitogenicity in vivo, we administered FGF-21 to db/db mice via ALZET pumps at an efficacious dose once daily with 125 μg/kg/day of FGF-21 for 7 days and measured the levels of several secreted polypeptides in circulation (Table 2). While we were able to achieve a clear glucose-lowering effect in the study (Figure 4A), only insulin and glucagon levels were changed in a statistically significant manner. The reduction of insulin levels is consistent with our observation in ob/ob mice during OGTT (Figure 4G) and is suggestive of improvements in insulin sensitivity in FGF-21–treated mice.

GLUT1 and glucagon potentially mediate the mode of action of FGF-21. We were able to detect FGF-21–dependent upregulation of GLUT1 message specifically in white fat upon bolus injection into ob/ob mice (Figure 1G), which thus confirms our in vitro observations on 3T3-L1 adipocytes. The increase in GLUT1 may mechanistically be linked to FGF-21–dependent glucose lowering in diabetic rodents. Alternatively, or in concert, the glucose lowering effect of FGF-21 is likely to result from reduced glucagon secretion from pancreatic α cells, since FGF-21 inhibits glucagon release in vitro (Figure 7) and is lowered in FGF-21–transgenic mice (Table 1). The observation of glucagon lowering in FGF-21–injected ob/ob mice (Table 2) further strengthens the hypothesis that this hormone is an important mediator of FGF-21 in vivo effects. There is accumulating evidence supporting a pathophysiological role of glucagon in the development and progression of type 2 diabetes. Basal glucagon is inappropriately elevated and its suppression is impaired following food consumption, which leads to increased hepatic glucose production and aggravation of the hyperglycemia associated with the disease (33, 34). Interestingly, attenuation of signaling through the glucagon receptor leads to normalization of plasma glucose and triglyceride levels in diabetic animals (35).

Despite substantial progress in understanding the pathophysiology of diabetes mellitus and the development of new drugs to treat diabetic patients, this disease remains a major health problem (36). New treatments are required that will allow an efficacious regulation of glycemia and reduce the risk of the side effects associated with current therapies (27, 37, 38). Here we demonstrate that FGF-21, as a single agent, can be used to provide efficient and durable glucose control and triglyceride lowering in diabetic animals, without apparent mitogenicity, hypoglycemia, or weight gain. FGF-21 thus holds promise as an effective therapeutic agent for the treatment of diabetes.

### Methods

Expression and purification of FGF-21. A pET30a vector was used to express human FGF-21 in the Escherichia coli strain BL21(DE3) (Novagen; EMD Biosciences Inc.). FGF-21 product accumulated in the insoluble fraction. Inclusion bodies were prepared by standard centrifugation method. We solubilized inclusion bodies by bringing granule pellets to 10 times the original volume in 50 mM Tris-HCL, pH 9.0, 7 M urea and homogenizing the material. The protein mixture was adjusted to pH 11, stirred for 1 hour, readjusted to pH 9.0, and loaded onto a Q Sepharose Fast Flow (Amersham Biosciences). Anion-exchange (AEX) chromatography was done in 50 mM Tris-HCL, pH 9.0, 7 M urea, 1 mM DTT and with a 0–400 mM NaCl gradient elution. The eluted AEX pool was treated with 10 mM DTT for 2 hours at room temperature and diluted 10-fold with 10 mM cysteine/7 M urea. The protein was refolded by dialysis against 20 mM glycine, pH 9.0, for 48 hours, from insulin, which thus rules out a possibility that FGF-21 functions as an insulin mimetic and/or sensitizer.

The beneficial in vivo effects of FGF-21 may be mediated through changes in circulating levels of endocrine-relevant hormones, in particular, adipokines, since adipokines appear to be a target for FGF-21 bioactivity. To explore this possibility, we dosed ob/ob mice s.c. once daily with 125 μg/kg/day of FGF-21 for 7 days and measured the levels of several secreted polypeptides in circulation (Table 2). While we were able to achieve a clear glucose-lowering effect in the study (Figure 4A), only insulin and glucagon levels were changed in a statistically significant manner. The reduction of insulin levels is consistent with our observation in ob/ob mice during OGTT (Figure 4G) and is suggestive of improvements in insulin sensitivity in FGF-21–treated mice.

### Table 2

**Administration of FGF-21 in ob/ob mice affects serum levels of glucagon and insulin but not of other secreted polypeptides**

<table>
<thead>
<tr>
<th>Substance</th>
<th>Vehicle control</th>
<th>FGF-21</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucagon (pM)</td>
<td>252 ± 40</td>
<td>156 ± 15</td>
<td>0.04</td>
</tr>
<tr>
<td>Insulin (pM)</td>
<td>2551 ± 81.7</td>
<td>2,344 ± 46.3</td>
<td>0.01</td>
</tr>
<tr>
<td>Adiponectin (μg/ml)</td>
<td>12.3 ± 0.82</td>
<td>13.9 ± 0.87</td>
<td>NS</td>
</tr>
<tr>
<td>MCP-1 (pg/ml)</td>
<td>111.8 ± 18.9</td>
<td>137.8 ± 16.6</td>
<td>NS</td>
</tr>
<tr>
<td>IL-6 (pg/ml)</td>
<td>11.9 ± 0.5</td>
<td>12 ± 0.7</td>
<td>NS</td>
</tr>
<tr>
<td>TNF-α (pg/ml)</td>
<td>11 ± 0</td>
<td>11.4 ± 0.4</td>
<td>NS</td>
</tr>
<tr>
<td>PAI-1 (pg/ml)</td>
<td>2,104 ± 339</td>
<td>1,689 ± 291</td>
<td>NS</td>
</tr>
<tr>
<td>Amylin (pM)</td>
<td>150 ± 15</td>
<td>113 ± 10</td>
<td>NS</td>
</tr>
<tr>
<td>Leptin (pM)</td>
<td>12 ± 3.3</td>
<td>8.5 ± 1.4</td>
<td>NS</td>
</tr>
<tr>
<td>GLP-1 (pM)</td>
<td>10.6 ± 1.2</td>
<td>13.3 ± 0.9</td>
<td>NS</td>
</tr>
</tbody>
</table>

The values (± SE) shown are the average of the measurements of 8 animals in a group. MCP-1, monocyte chemotactic protein–1; PAI-1, plasminogen activator inhibitor–1.
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at 4°C. Further purification was carried out with reversed-phase high-performance liquid chromatography (RP-HPLC) performed with a Grace Vydac C18 column run in H₂O/0.1% trifluoroacetic acid/acetonirole mobile phase with a 0–50% acetonirole gradient; size-exclusion chromatography on Superdex 75 (Amersham Biosciences) in PBS, pH 7.4; and AEX chromatography on MonoQ (Amersham Biosciences) in 50 mM Tris, pH 8.0, with 0–300 mM NaCl gradient. The final FGF-21 pool was diazylized into PBS, pH 7.4, sterile filtered, and stored at −80°C.

Cell culture, adipocyte differentiation, glucose uptake, and mitogenicity experiments. 3T3-L1, NIH 3T3, BALB/c 3T3, 4MRbr5, and clone 9 cells were from American Type Culture Collection; HMECs and HUECs from Clonetech Corp.; and human primary preadipocytes from Zen-Bio Inc. A previously described 3T3-L1 adipocyte differentiation protocol (39) was adapted for Cytoplastar T 96-well plates (Amersham Biosciences). 3T3-L1 fibroblasts were seeded at 25,000 cells/well; the differentiation was induced 2 days later in DMEM supplemented with 10% FBS, 0.25 μM dexamethasone, 0.5 mM 3-isobutyl-1-methylxanthine, and 5 μM insulin (48 hours); and the medium was changed to DMEM/10% FBS/5μg/ml insulin (48 hours). Thereafter, the cells were incubated for an additional 9–20 days in DMEM/10% FBS (changed every other day). Primary human adipocytes were seeded in Cytoplastar T 96-well plates at 15,000 cells/well; differentiated in Adipocyte Medium (AM; Zen-Bio Inc.) with 0.05 mM IBMX, 0.1 μM dexamethasone, 10 mM insulin, and 1 μM rosiglitazone (14 days); and thereafter kept in AM (changed every other day). For glucose uptake, adipocytes were starved for 3 hours in DMEM/0.1% BSA, stimulated with FGF-21 for 24 hours, and washed twice with KRP buffer (15 mM HEPES, pH 7.4, 118 mM NaCl, 4.8 mM KCl, 1.2 mM MgSO₄, 1.3 mM CaCl₂, 1.2 mM KH₂PO₄, 0.1% BSA), and 100 μl of KRP buffer containing 2-deoxy-6-[³²³³H]glucose (2-DOG) (0.1 μCi, 100 μM) was added to each well. Control wells contained 100 μl of KRP buffer with 2-DOG (0.1 μCi, 10 mM) to monitor for nonspecificity. The uptake reaction was carried out for 1 hour at 37°C, terminated by addition of cytochalasin B (20 μM), and measured using Wallac 1450 MicroBeta counter (Perkin Elmer). For mitogenicity experiments, cells were grown to confluence, starved for 24 hours in DMEM/0.5% FCS, stimulated for 18 hours, and incubated with 0.25 μCi [³²³³H]thymidine per well for 2 hours. Cell lysates were then harvested and counted.

RNA extraction, cDNA synthesis, quantitative PCR. We used an RNeasy 96 Kit (Qiagen Inc.) to extract RNA from cells and TRizol reagent (Invitrogen Corp.) to extract RNA from tissues. We performed reverse transcription using a SuperScript First-Strand Synthesis kit (Invitrogen Corp.). The forward and reverse primer sequences for GLUT1 were 5′-GCCCCCAAGATTTATAA-3′ and 5′-GCTGTTGAGTGTTGAGATG-3′, respectively. The probe sequence was 5′-TTCTTACATCAACACATGGAGACCCGGCA-3′. To normalize for differences in the amount of total RNA added to each reaction, we performed amplification of 18S ribosomal RNA as an endogenous control.

Immunoblotting, immunoprecipitation, and multiplex assays. 3T3-L1 adipocytes were starved for 18 hours, stimulated with FGF-21 (1 μg/ml) for 10 minutes or for the indicated times (Figures 1F and 2A), and lysed (40), and soluble fractions were analyzed. Antibodies were: anti-phospho-MAPK (Thr202/Tyr204), anti-MAPK, anti-phospho-FRS-2 (Y196) (Cell Signaling Technology); anti-FRS-2 (H-91) and anti-FGFR-2 (C-17) (Santa Cruz Biotechnology Inc.); anti-phosphotyrosine (4G10; Upstate); rabbit polyclonal: anti-GLUT1 against the 29 C-terminal amino acids of the human sequence, anti-GLUT4 (41), and anti–FGFR-1 against the 15 C-terminal amino acids of the mouse sequence. For immunodetection, goat anti-mouse and anti-rabbit HRP conjugates (Bio-Rad Laboratories) and ECL detection system (Amersham Biosciences) were used. We measured hormone and adipokine levels in circulation of vehicle-treated and FGF-21–injected ob/ob mice using Multiplex assay kits from LInCco Research Inc.

Tissue preparation, histology analysis, and immunostaining. Tissues were fixed overnight in zinc-buffered formalin and then transferred to 70% ethanol prior to processing through paraffin. Five-micrometer sections were stained with H&E. Adjacent 5-micrometer sections were placed on positively charged slides. The slides were then baked overnight at 60°C in an oven and then deparaffinized in xylene and rehydrated through graded alcohols to water. Antigen retrieval was performed by immersing the slides in Target Retrieval Solution for 20 minutes at 90°C, cooling at 25°C for 10 minutes, and washing in water; we then proceeded with immunostaining.

All subsequent staining steps were performed on the Autoimmunostainer; incubations and all washes were done at 25°C in 50 mM Tris-HCl, pH 7.4, containing 0.05% Tween-20. Slides were blocked with protein blocking solution for 25 minutes, and the PCNA antibody (PC10 clone) was incubated at a dilution of 1:10 for 1 hour. A biotinylated antibody plus streptavidin-HRP kit was then applied and followed with 3,3′-diaminobenzidine (DAB) staining. The slides were briefly counterstained with hematoxylin. All immunoreagents and the Autoimmunostainer were from Dako Corp.

Pancreatic islet isolation and hormone release studies. Pancreatic islets from male Wistar rats (200 g; Harlan Winkelmann GmbH) were isolated and cultured as described previously (35). For measurements of glucagon and insulin release, islets were starved in Earle’s balanced salt solution medium containing 1 mM glucose for 30 minutes. Groups of 10 (glucagon) or 3 (insulin) islets were selected and transferred into 0.3 ml of EBSS medium with tested compounds. Islets were further incubated for 90 minutes at 37°C with vehicle or FGF-21 (1 μg/ml), supernatants were collected, and hormone content was measured.

In vivo protocols. The protocols used in these studies were approved by the Eli Lilly Research Laboratories Institutional Animal Care and Use Committee. Mice were maintained in a controlled environment (21 ± 2°C, 50–60% humidity, 12-hour light-dark cycle, lights on at 6 am). Male ob/ob and db/db mice were from Harlan Teklad, fed Purina 5008 Chow, and had free access to food and water. FGF-21 was administered by s.c. injection in saline. For OGTT, the animals were fasted 16 hours and challenged by an oral glucose load (2.5 g/kg) 1 hour after the last injection. Blood samples were taken from conscious, fed animals by tail snip, and glucose and plasma triglyceride levels were determined using Precision G Blood Glucose Testing System (Abbott Laboratories) and Hitachi 912 Clinical Chemistry analyzer (Roche Diagnostics Corp.), respectively. Insulin and leptin levels were determined with murine ELISA kits (Crystal Chem Inc.).

Male obese or lean ZDF rats were obtained from Charles River Laboratories Inc. and housed singly in a humidity and temperature-controlled environment (21 ± 2°C, 50–60% humidity, 12-hour light-dark cycle, lights on at 6 am) with free access to food (Purina 5008 Chow) and water for 2 weeks prior to start of the experiment. The day before the study, all animals were tail bled, and plasma glucose was analyzed on a Hitachi 912 Clinical Chemistry analyzer (Roche Diagnostics Corp.). Rats were then randomized based on their glucose levels and body weights and placed into groups. Animals were dosed with vehicle (0.9% saline), insulin (Humulin; Eli Lilly and Company), or FGF-21 for 7 continuous days of twice-daily administration. On days 3 and 7, fed rats were bled (by tail snip) at 1-hour after administration of the last dose, and plasma glucose was assayed as described above.

The human apoE promoter including its hepatic control region (42) was used to express the human FGF-21 cDNA in transgenic mice. The transgenic vector was linearized and microinjected into C57BL/6NTac eggs by the Eli Lilly Research Laboratories Institutional Animal Care and Use Committee. Mice were maintained in a controlled environment (21 ± 2°C, 50–60% humidity, 12-hour light-dark cycle, lights on at 6 am) with free access to food (Purina 5008 Chow) and water for 2 weeks prior to start of the experiment. The day before the study, all animals were tail bled, and plasma glucose was analyzed on a Hitachi 912 Clinical Chemistry analyzer (Roche Diagnostics Corp.). Rats were then randomized based on their glucose levels and body weights and placed into groups. Animals were dosed with vehicle (0.9% saline), insulin (Humulin; Eli Lilly and Company), or FGF-21 for 7 continuous days of twice-daily administration. On days 3 and 7, fed rats were bled (by tail snip) at 1-hour after administration of the last dose, and plasma glucose was assayed as described above.

The human apoE promoter including its hepatic control region (42) was used to express the human FGF-21 cDNA in transgenic mice. The transgenic vector was linearized and microinjected into C57BL/6NTac eggs by standard methods (43). We identified transgenic mice by PCR using transgene-specific primers and confirmed transgene expression by real-time quantitative PCR on RNA isolated from livers as well as by ELISA on plasma obtained from the transgenic mice using an FGF-21 polyclonal antibody.

For high-fat feeding, all mice were housed individually in Micro-Isolator cages (Lab Products Inc.) and maintained from age 24 days on high-fat TD95217 chow (40% fat; Harlan Teklad) with free access to food and water.

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A wide-line nuclear magnetic resonance (NMR) instrument (Bruker BioSpin Corp.) was used to quantify tissue mass. Unanesthetized mice were placed into a 5-cm diameter glass cylinder that was lowered into the instrument. Data was obtained for 4.5 minutes (3 determinations at 1.5-minute intervals) and analyzed by the manufacturer’s software. The mean of triplicate determinations for apparent muscle mass, fat mass, and free water mass was calculated for each mouse. Coefficient of variation of all three masses determined for a live moving mouse was less than 3%.

Acknowledgments

We wish to thank P. Atkinson, G. Kelly, T. Black, B. Pies, and B. Strifer for supporting FGF-21 protein production; D. Bruce Baldwin for the generation of FGFR-Fc fusion constructs; S. Bright and J. Dunbar for FGF-21/FGFR binding experiments; S. Sissons and W. Roell for assistance with glucose uptake and proliferation assays; N. Fox and K. Brune for generation of FGF-21 transgenic animals; K. Coble for help with pharmacokinetic studies; M. Brenner and A. Efano for assistance with glucagon secretion experiments; D. Ballard and K. Mintze for supporting histochemistry work; C. Shrike for in vivo assistance; J. Manetta and L. Sliker for generation of polyclonal anti–FGF-21, anti–GLUT1, and anti–GLUT4 antibodies; A. Klip (Hospital for Sick Children, Toronto, Ontario, Canada) for L6-GLUT-4myc cells; J. Caro, S. Jacobs, and G. Etgen for critically reading the manuscript; and T. Bumol, B. Grinnell, A. Glasebrook, R. Smith, and S. Taylor for helpful discussions.

Received for publication October 12, 2004, and accepted in revised form March 23, 2005.

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