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## The multiple causes of human SCID

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#### Commentary

SCID, a syndrome characterized by the absence of T cells and adaptive immunity, can result from mutations in multiple genes that encode components of the immune system. Three such components are cytokine receptor chains or signaling molecules, five are needed for antigen receptor development, one is adenosine deaminase — a purine salvage pathway enzyme, and the last is a phosphatase, CD45. In this issue of the *JCI*, a report describes how complete deficiency of the CD3c chain of the T cell antigen receptor/CD3 complex causes human SCID.



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skin conditions, confirming the different developmental patterns of ectoderm and mesoderm (3). The disparate development of ectodermal and mesodermal structures emphasizes the need to study affected tissues when seeking evidence of gene mosaicism. Patterned epidermal and dermal mosaic disorders (such as morphea and focal dermal hypoplasia) provide an ideal focus for future investigations that will elucidate the mechanisms of embryologic development of both ectodermal and mesodermal tissues.

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1. Blaschko, A. 1901. Die Nervenverteilung in der Haut in ihrer Beziehung zu den Erkrankungen der Haut. Wilhelm Braunmuller. Vienna, Austria and Leipzig, Germany.

- Happle, R. 1993. Mosaicism in human skin. Understanding the patterns and mechanisms. Arch. Dermatol. 129:1460–1470.
- Paller, A.S., et al. 1994. Genetic and clinical mosaicism in a type of epidermal nevus. *N. Engl. J. Med.* 331:1408–1415.
- Rothnagel, J.A., et al. 1992. Mutations in the rod domains of keratins 1 and 10 in epidermolytic hyperkeratosis. *Science*. 257:1128–1130.
- Colman, S.D., Rasmussen, S.A., Ho, V.T., Abernathy, C.R., and Wallace, M.R. 1996. Somatic mosaicism in a patient with neurofibromatosis type 1. *Am. J. Hum. Genet.* 58:484–490.
- Munro, C.S., and Wilkie, A.O. 1998. Epidermal mosaicism producing localised acne: somatic mutation in FGFR2. *Lancet.* 352:704–705.
- Sakuntabhai, A., Dhitavat, J., Burge, S., and Hovnanian, A. 2000. Mosaicism for ATP2A2 mutations causes segmental Darier's disease. *J. Invest. Dermatol.* 115:1144–1147.
- Terrinoni, A., et al. 2000. A mutation in the V1 domain of K16 is responsible for unilateral palmoplantar verrucous nevus. *J. Invest. Dermatol.* 114:1136–1140.
- 9. Jonkman, M.F., et al. 1997. Revertant mosaicism in epidermolysis bullosa caused by mitotic gene conversion. *Cell.* **88**:543–551.
- 10. Smith, F.J., Morley, S.M., and McLean, W.H. 2004.

Novel mechanism of revertant mosaicism in Dowling-Meara epidermolysis bullosa simplex. J. Invest. Dermatol. **122**:73–77.

- Arin, M.J., Longley, M.A., Wang, X.J., and Roop, D.R. 2001. Focal activation of a mutant allele defines the role of stem cells in mosaic skin disorders. J. Cell Biol. 152:645–649.
- Happle, R. 1991. Somatic recombination may explain linear porokeratosis associated with disseminated superficial actinic porokeratosis. *Am. J. Med. Genet.* 39:237.
- Happle, R. 1996. Segmental forms of autosomal dominant skin disorders: different types of severity reflect different states of zygosity. *Am. J. Med. Genet.* 66:241–242.
- Itin, P.H., and Buechner, S.A. 1999. Segmental forms of autosomal dominant skin disorders: the puzzle of mosaicism. *Am. J. Med. Genet.* 85:351–354.
- Smith, C.G., Glaser, D.A., and Leonardi, C. 1998. Zosteriform multiple leiomyomas. J. Am. Acad. Dermatol. 38:272-273.
- Vakilzadeh, F., and Kolde, G. 1985. Relapsing linear acantholytic dermatosis. Br. J. Dermatol. 112:349–355.
- Poblete-Gutiérrez, P., et al. 2004. Allelic loss underlies type 2 segmental Hailey-Hailey disease, providing molecular confirmation of a novel genetic concept. J. Clin. Invest. 114:1467–1474. doi:10.1172/ JCI200421791.

# The multiple causes of human SCID

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SCID, a syndrome characterized by the absence of T cells and adaptive immunity, can result from mutations in multiple genes that encode components of the immune system. Three such components are cytokine receptor chains or signaling molecules, five are needed for antigen receptor development, one is adenosine deaminase — a purine salvage pathway enzyme, and the last is a phosphatase, CD45. In this issue of the *JCI*, a report describes how complete deficiency of the CD3 $\varepsilon$  chain of the T cell antigen receptor/CD3 complex causes human SCID (see the related article beginning on page 1512).

Human SCID was first reported by Glanzmann and Riniker in 1950 (1). Swiss infants with the condition were profoundly lymphopenic and died of infection before their first or second birthdays. In the ensuing years, differences were noted in inheritance patterns for SCID. This indicated that there was more than one cause for this fatal syndrome characterized by an absence of T cells and all adaptive immunity. In many families there was an X-linked recessive mode of inheritance while in others an autosomal recessive mode of inheritance was observed. The first discovered molecular cause of human SCID, adenosine deaminase deficiency, was reported in 1972 (2). However, it was not until 21 years later, in 1993, that a second fundamental cause of the condition was found, i.e., the molecular basis of X-linked human SCID (3, 4). Over the past 11 years, remarkable progress has been made in elucidating several other causes of this syndrome (5). Advances in molecular biology and the Human Genome Project as well as increased knowledge of various components of the immune system through studies of mutant mice and humans with genetically determined immunodeficiencies have all contributed to this understanding. It is now known that SCID can be caused in humans by mutations in at least 10 different genes (Table 1) (6-11), and the likelihood is that there are other causes yet to be discovered. The gene products of 3 of these mutated genes are components of cytokine receptors (the IL-2 receptor  $\gamma$  chain that is also shared with 5 other cytokine receptors [IL-4R, IL-7R, IL-9R, IL-15R, and IL-21R], JAK3, the primary signal transducer from the common  $\gamma$  chain, and the  $\alpha$  chain of the IL-7 receptor); the products of 5 more genes (RAG1, RAG2, Artemis, CD3 $\delta$ , and CD3 $\epsilon$ ) are necessary for antigen-receptor development;

#### Table 1

Ten abnormal genes in human SCID

#### Cytokine-receptor genes

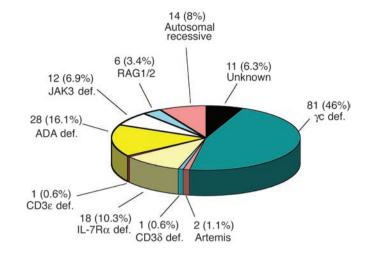
IL-2RG JAK3 IL-7Rα Antigen-receptor genes RAG1 RAG2 Artemis CD3δ CD3ε Other genes ADA CD45

**Conflict of interest:** The author has declared that no conflict of interest exists.

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#### commentaries



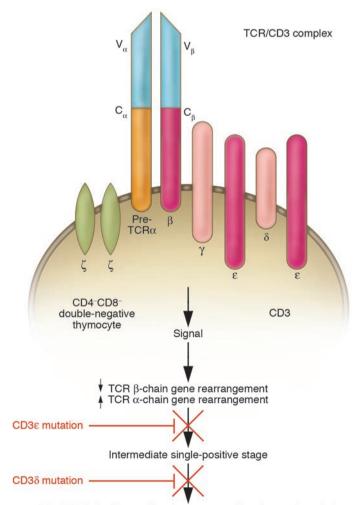


#### Figure 1

Relative frequencies of the various molecular defects found in 174 consecutive cases of human SCID evaluated at Duke University Medical Center over the past 3 decades. The most common type is X-linked SCID, due to mutations in the gene encoding the common  $\gamma$  chain for multiple cytokine receptors; the second most common cause is adenosine deaminase deficiency (ADA def.), and the third most common cause is IL-7R $\alpha$ -chain deficiency. In 25 cases the molecular defect remains unknown (those in the groups labeled autosomal recessive and unknown). No cases of CD45 deficiency have been seen at this institution. Def., deficiency.

the product of one gene (adenosine deaminase) is necessary for detoxification of metabolic products of the purine salvage pathway that cause lymphocytes to apoptose; and the final gene encodes CD45, a phosphatase that serves as a critical regulator of signaling thresholds in immune cells (Table 1) (12). The most common form of human SCID is the X-linked type, caused by mutations in *IL-2RG*, which accounts for 46% of cases at the author's institution, the Duke University Medical Center

(Figure 1). This is followed by adenosine deaminase deficiency in 16.1% of cases and IL-7R $\alpha$ -chain deficiency in 10.3% of cases. The relative frequencies of these different molecular types of human SCID may vary in other geographic areas.



#### CD4+CD8+ double-positive thymocyte proliferation and survival

Schematic of the T cell receptor/CD3 (TCR/ CD3) complex on the surface of a normal CD4-CD8- double-negative thymocyte. All but one of the normal chains of the CD3 complex, including the  $\beta$  chain, are present, but instead of the CD3 $\alpha$  chain, the pre–T cell receptor  $\alpha$ gene is expressed at this stage. It is postulated, based on the CD3<sub>E</sub> murine knockout studies, that the block in T cell development caused by mutations in CD3 coccurs at or just after this stage. In their study in this issue, de Saint Basile and colleagues (13) report histologic data which suggests that, in the case of mutations in the CD36 gene, the block in T cell maturation occurs at the next stage, i.e., at the intermediate single-positive stage.



## A newly discovered cause of human SCID

The tenth known cause of human SCID is reported in this issue of the ICI by de Saint Basile et al. (13). The authors identified mutations in the gene encoding the  $\epsilon$  component of the T cell receptor/CD3 complex (Figure 2). Building on the discovery of Dadi et al. (11), that mutations in a gene encoding the  $\delta$  chain of the CD3 complex cause human SCID, de Saint Basile and coworkers used segregation analysis of polymorphic markers for chromosome 11q23 - the location of the CD3 locus - to study 3 families with fetuses or infants who had SCID of unknown molecular type (13). All of the fetuses or infants were characterized by the T<sup>-</sup>B<sup>+</sup>NK<sup>+</sup> lymphocyte phenotype, i.e., they had no T cells but did have phenotypically normal B cells and NK cells, the same phenotype observed in the SCID infants described by Dadi et al. (11). There was a homozygous haplotype segregation of polymorphic markers for 11q23 in all 3 families studied by de Saint Basile et al. However, mutations in the gene encoding the CD3 $\delta$  chain were found in only 2 of the families. The investigators then searched for mutations in other components of the CD3 complex and found a homozygous mutation in the gene encoding CD3<sup>ε</sup> that created a premature stop codon near the start of the extracellular domain, resulting in the absence of CD3 expression in those individuals affected in the 3rd family. In 1993, members of this research group reported a child with low, but detectable, expression levels of CD3E on circulating T cells and identified a splice site mutation on one allele that did not totally abrogate the normal intron 7 splicing (14). That child did not have impaired T cell development and had only a mild immunodeficiency (14). In the present report (13), the authors show that an absence of CD3ɛ completely blocks T cell development. In the CD3Eknockout mouse, this block appears to occur at the pre-T cell receptor  $\alpha$ , doublenegative stage. Dadi et al. (11) had concluded, based on studies of thymic material from their patients, that an absence of CD38 also causes a block in T cell development at the pre-T cell receptor  $\alpha$ , doublenegative stage of intrathymic development. In the present report, de Saint Basile et al. (13) confirm that an absence of CD3 $\delta$  completely abrogates T cell development but conclude that the block is slightly later – at the intermediate single-positive stage, just before the double-positive (i.e., CD4<sup>+</sup>CD8<sup>+</sup>) stage. Thus both CD3 $\delta$  and CD3 $\epsilon$  appear to be essential for intrathymic development of T cells (Figure 2) whereas CD3 $\gamma$  deficiency does not (15). We have also found mutations in these two genes in our human SCID population (J.L. Roberts et al., unpublished results; Figure 1).

#### Importance

SCID is a pediatric emergency. If the diagnosis is made at birth or shortly thereafter, definitive therapy in the form of HLAidentical or haploidentical allogeneic bone marrow stem cell transplantation can result in a survival rate as high as 97%, regardless of the molecular type of SCID (5). However, if the diagnosis is made later, serious infections develop for which antibiotics are poorly effective or nonexistent, and the survival rate is significantly less. Allogeneic stem cell transplantation is not a perfect therapy, however, as many recipients experience incomplete B cell and/or NK cell immune reconstitution. The hope is that gene therapy could effect more complete immune reconstitution, and there has already been remarkable success in achieving this in a few patients (16-18). However, gene therapy cannot be performed unless the abnormal gene for a specific patient is known. Knowledge of the specific molecular defect is also essential for genetic counseling and prenatal diagnosis. Of the 174 SCID infants evaluated at the Duke University Medical Center over the past 3 decades, there are still 25 infants for whom the molecular basis of their disease is unknown (Figure 1). It is possible that other genes yet to be discovered can result in human SCID when mutated. Thus knowing all of the abnormal genes that result in human SCID is an important ongoing goal for all who work in this area.

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- Glanzmann, E., and Riniker, P. 1950. Essentielle Lymphocytophtose. Ein neues Krankeitsbild aus der Sauglingspathologie. Ann. Paediat. 174:1–5.
- Giblett, E.R., Anderson, J.E., and Cohen, I. 1972. Adenosine deaminase deficiency in two patients with severely impaired cellular immunity. *Lancet.* 2:1067–1070.
- Noguchi, M., et al. 1993. Interleukin-2 receptor gamma chain mutation results in X-linked severe combined immunodeficiency in humans. *Cell.* 73:147–157.
- 4. Puck, J.M., et al. 1993. The interleukin-2 receptor gamma chain maps to Xq13.1 and is mutated in X-linked severe combined immunodeficiency, SCIDX1. *Hum. Mol. Genet.* **2**:1099–1104.
- Buckley, R.H. 2004. Molecular defects in human severe combined immunodeficiency and approaches to immune reconstitution. *Annu. Rev. Immunol.* 55:625–656.
- Russell, S.M., et al. 1995. Mutation of Jak3 in a patient with SCID: Essential role of Jak3 in lymphoid development. *Science*. 270:797–800.
- Puel, A., Ziegler, S.F., Buckley, R.H., and Leonard, WJ. 1998. Defective IL7R expression in T(-)B(+)NK(+) severe combined immunodeficiency. *Nat. Genet.* 20:394–397.
- 8. Schwarz, K., et al. 1996. RAG mutations in human B cell-negative SCID. *Science*. **274**:97–99.
- Moshous, D., et al. 2001. Artemis, a novel DNA double-strand break repair/V(D)J recombination protein, is mutated in human severe combined immune deficiency. *Cell.* 105:177–186.
- Kung, C., et al. 2000. Mutations in the tyrosine phosphatase CD45 gene in a child with severe combined immunodeficiency disease. *Nat. Med.* 6:343-345.
- 11. Dadi, H.K., Simon, A.J., and Roifman, C.M. 2003. Effect of CD3delta deficiency on maturation of alpha/beta and gamma/delta T-cell lineages in severe combined immunodeficiency. *N. Engl. J. Med.* 349:1821–1828.
- Hermiston, M.L., Xu, Z., and Weiss, A. 2003. CD45: a critical regulator of signaling thresholds in immune cells. *Annu. Rev. Immunol.* 21:107–137.
- 13. de Saint Basile, G., et al. 2004. Severe combined immunodeficiency caused by deficiency in either the  $\delta$  or the  $\epsilon$  subunit of CD3. *J. Clin. Invest.* **114**:1512–1517. doi:10.1172/JCI200422588.
- Soudais, C., De Villartay, J.P., Le Deist, F., Fischer, A., and Lisowska-Grospierre, B. 1993. Independent mutations of the human CD3-epsilon gene resulting in a T cell receptor/CD3 complex immunodeficiency. *Nat. Genet.* 3:77–81.
- Arnaiz-Villena, A., et al. 1993. T lymphocyte signalling defects and immunodeficiency due to the lack of CD3 gamma. *Immunodeficiency*. 4:121-129.
- 16. Cavazzana-Calvo, M., et al. 2000. Gene therapy of human severe combined immunodeficiency (SCID)-X1 disease. *Science*. **288**:669–672.
- Hacein-Bey-Abina, S., et al. 2002. Sustained correction of X-linked severe combined immunodeficiency by ex vivo gene therapy. *N. Engl. J. Med.* 346:1185–1193.
- Aiuti, A., et al. 2002. Correction of ADA-SCID by stem cell gene therapy combined with nonmyeloablative conditioning. *Science*. 296:2410–2413.