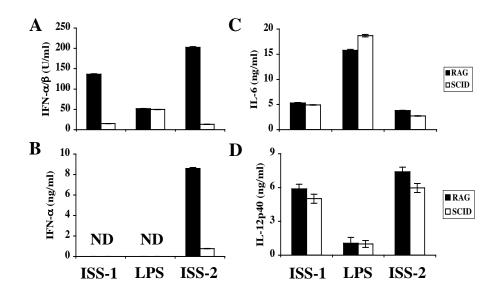
Supplemental Material

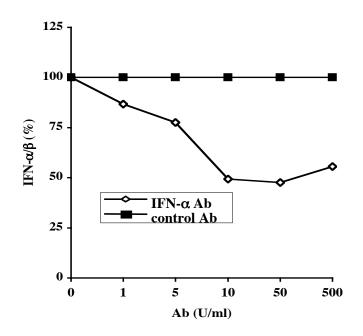
We evaluated the production of type 1 IFN by BM-PDC derived from $RAG1^{-/-}$ and SCID mice. BM-PDC were generated from bone marrow cells cultured with 100 ng/ml of hFlt3 ligand (PeproTech Inc. #300-19), and the levels of type 1 IFN generated upon stimulation with ISS-1 were determined by a bioassay and by an IFN- α ELISA kit (PBL Biomedical Laboratories). As shown in Supplemental Figure 1 below, we did not detect any IFN- α in the RAG- or SCID-derived BM-PDC supernatants after stimulation with ISS-1 and LPS (Supplemental Figure 2B) when tested by ELISA. In contrast, different levels of type 1 IFN in these supernatants were detected when tested by the bioassay. IFN- α levels were detectable by ELISA in the supernatants of BM-PDC upon stimulation with ISS-2, a class A CpG ODN that is known to elicit very high levels of this cytokine. As expected, the levels of IFN- α were by far higher in the supernatants of RAG-derived BM-PDC than in the supernatants of SCID-derived BM-PDC.



Supplemental Figure 1: Comparison of assays for the measurement of type-1 IFN: Bioassay vs. ELISA.

BM-PDC (10⁶/ml) from 3 RAG1^{-/-} and 3 SCID mice were pooled and stimulated with ISS-1 (10 μ g/ml), LPS (50 ng/ml), and ISS-2 (10 μ g/ml). Cytokine levels in the supernatants were measured 24 hrs later (triplicates). (A) Type 1 IFN (IFN α/β) levels (U/ml) by bioassay. (B) IFN- α levels (pg/ml) by ELISA (PBL). (C) IL-6 and (D) IL-12p40 levels (ELISA). ND - not detected.

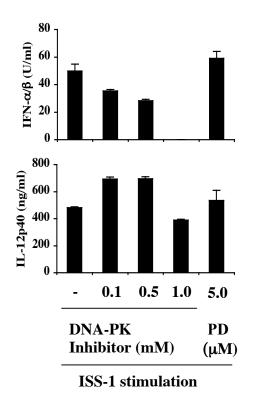
The bioassay detects type 1 IFN activity, which consists of multiple IFN- α gene products and a single IFN- β gene product. In contrast, the ELISA kit (PBL) detects only a few species of IFN- α . To verify and to measure the relative IFN- α and IFN- β activities in the PDC supernatants, we neutralized IFN- α from the supernatants of ISS-stimulated BM-MDC. Thus, increasing concentrations of rabbit anti-mouse IFN- α antibody (PBL) and rabbit IgG (Jackson Immuno-Research Laboratories) were added to the supernatants and the resulting type 1 IFN was analyzed by bioassay. As shown below, IFN- α neutralization inhibited up to 50% of the total type-1 IFN activity in the supernatants.



Supplemental Figure 2: IFN-α neutralization does not abolish type-1 IFN activity when tested by bioassay

BM-MDC (10⁶/ml) generated from 3 RAG1^{-/-} mice were pooled and stimulated with ISS-1 (10 μ g/ml). After 24 hrs the supernatants were collected and co-incubated with various concentrations of rabbit anti-mIFN- α IgG or control rabbit IgG overnight at 37^oC. Type 1 IFN levels were measured in the supernatants by bioassay. The level obtained for each control IgG concentration was considered as 100% of type 1 IFN (IFN- α/β) activity.

We performed a type 1 IFN bioassay using BM-MDC made from RAG deficient mice. These cells were stimulated with ISS-1 in the presence or absence of a novel DNA-PK inhibitor NU7026 (Calbiochem). As presented in the Figure below type 1 IFN production was inhibited by the DNA-PK inhibitor, while the IL-12 (p40) production was intact.



Supplemental Figure 3: DNA-PK inhibitor suppresses type-1 IFN production from BMDC (MDC).

BM-MDC (10⁶/ml) from 2 *RAG1*^{-/-} mice were pooled and co-incubated with or without various concentrations of DNA-PK or MEK inhibitor (control). Cells were stimulated with ISS-1 (10 μ g/ml). Cytokine levels in the supernatants were measured 24 hrs later in triplicate. IL-12 (p40) levels were measured by ELISA (BD-Pharmingen) and type 1 IFN (IFN- α/β) levels (U/ml) were measured by bioassay.

We analyzed the response of BM-MDC and BM-PDC from RAG1 deficient and DNA-PK deficient mice (provided by Dr. Gloria Li, Memorial Sloan Kettering, NY). As shown in Supplemental Tables 1 and 2, after ISS or LPS stimulation the levels of IFN- α in the supernatants of these cells were below detection when measured by ELISA (PBL). In contrast, we detected type 1 IFN activity in these supernatants when tested by bioassay (Tables 1 and 2). The levels of type 1 IFN in the supernatants of BM-MDC and BM-PDC derived from RAG1 deficient animals were higher than those detected in BM-MDC and BM-PDC supernatants derived from DNA-PK deficient mice when tested by bioassay (Tables 1 and 2 below). These data combined with the SCID BM-MDC data presented in the main text indicate that DNA-PK is involved in type 1 IFN production by these two DC subsets upon TLR9 stimulation.

	IFN-α/β (U/ml)		IFN-α (pg/ml)		IL-6 (ng/ml)		IL-12p40 (ng/ml)	
	ISS-1	LPS	ISS-1	LPS	ISS-1	LPS	ISS-1	LPS
<i>RAG</i> ^{-/-}	190 <u>+</u> 10	40 <u>+</u> 5	ND	ND	13 <u>+</u> 2	28 <u>+</u> 2	284 <u>+</u> 21	96 <u>+</u> 8
DNA-PK-/-	35 <u>+</u> 5	35 <u>+</u> 5	ND	ND	12 <u>+</u> 1	26 <u>+</u> 2	272 <u>+</u> 17	98 <u>+</u> 10

Supplemental Table 1: Reduced type-1 IFN production in BM-MDC from DNA-PK KO mice.

BM-MDC (10⁶/ml) from 3 RAG1 and 3 DNA-PK KO mice were pooled and stimulated with ISS-1 (10 μ g/ml) or LPS (50 ng/ml). Cytokine levels in the supernatants were measured 24 hrs later in triplicate. Type 1 IFN levels (IFN α/β) were measured by bioassay (U/ml), IFN- α levels were measured by ELISA (pg/ml, PBL) and IL-6 and IL-12p40 levels were measured by ELISA (BD-Pharmingen). ND - not detected.

	IFN-α/β (U/ml)		IFN-α (pg/ml)		IL-6 (ng/ml)		IL-12p40 (ng/ml)	
	ISS-1	LPS	ISS-1	LPS	ISS-1	LPS	ISS-1	LPS
<i>RAG</i> -/-	527 <u>+</u> 10	15 <u>+</u> 0.5	ND	ND	25 <u>+</u> 2	13 <u>+</u> 2	227 <u>+</u> 14	44 <u>+</u> 15
DNA-PK-/-	89 <u>+</u> 5	22 <u>+</u> 2	ND	ND	22 <u>+</u> 1	10 <u>+</u> 2	200 <u>+</u> 20	29 <u>+</u> 9

Supplemental Table 2: Reduced type-1 IFN production in BM-PDC from DNA-PK KO mice.

BM-PDC (10^6 /ml) from 3 RAG1 and 3 DNA-PK KO mice were pooled and stimulated with ISS-1 ($10 \mu g$ /ml) or LPS (50 ng/ml). Cytokine levels were detected as described above for Table 1. ND - not detected.