

## Supplementary Figure Legends:

### Supplementary Figure 1. Caloric Restriction in Rats:

- a. Representative photo of F344 rats: young, CR- Caloric restricted, Ad Lib- *Ad Libitum* fed.
- b. Weights of old CR and AL F344 rats normalized to the mean weight of sex-matched young adult rats.
- c. Representative H&E-stained sections of the testis of the young, old AL and old CR F344 rats, showing extensive Leydig cell hyperplasia in AL-fed old rats. This neoplasm is largely attenuated by CR, but small foci of hyperplasia are still detectable (arrows).

### Supplementary Figure 2. Effect of CR and GHR deficiency on p16<sup>INK4a</sup> expression with aging

The absolute copy number (log 10 scale) of *p16<sup>INK4a</sup>* mRNA molecules per 90 ng total RNA RT-PCR from lung and kidney of young (5 months) and old (21 months) *GHR* *+/+* and *-/-* mice with or without caloric restriction is graphed +/- SEM. Each estimate represents the mean of 8-16 quantitative RT-PCR reactions on independent RNA samples derived from organs of 22 mice.

### Supplementary Figure 3. Expression of regulators of p16<sup>INK4a</sup> with aging

- a. The ratios of the expression of p16<sup>INK4a</sup> regulators *Ets-1* and *Bmi-1* in (Old (26 months) / Young (2.5 months)) mice from 15 tissues is graphed +/- SEM. Each estimate represents the mean of 4-8 quantitative RT-PCR reactions on independent RNA samples derived from 4-6 mice.
- b. The ratios of the expression of p16<sup>INK4a</sup> regulators *Ets-1* and *Id1* from 12 tissues derived from (Old (28 months) / Young (3 months)) *ad libitum* fed (AL) or calorically restricted

(CR) F344 rats is graphed +/- SEM. Each estimate represents the mean of 4-8 quantitative RT-PCR reactions on independent RNA samples derived from 4 rats.

**Supplementary Figure 4. Comparison of different housekeeping genes used for normalization in RT-PCR experiments.**

The expression of housekeeping genes in different tissues (four independent assays of each housekeeping gene per tissue) expressed as ratio of expression in individual tissue to the mean expression seen in all tissues. Based on this analysis, each housekeeping gene has limitations. Using Tata-Binding protein (TATA) expression in the testis would overestimate RNA quality, as would normalizing to 18S expression in the pancreas. While GAPDH appears to correlate best across all tissues with the other two housekeeping genes, it demonstrates the lowest reproducibility ( $r^2$ ) when repeated on the same sample. Of note, none of the housekeeping genes appeared to change expression in any tissue with aging, and all three housekeeping genes correlated strongly with one another except in testis and pancreas. As these variations in housekeeping gene expression were tissue-specific, they would not bias O/Y ratios (which measure expression of old versus young within the same tissue), but would affect absolute copy number determination. Therefore, 18S was used for normalization in most tissues, except in the pancreas and testis; where TATA and/or GAPDH were used.

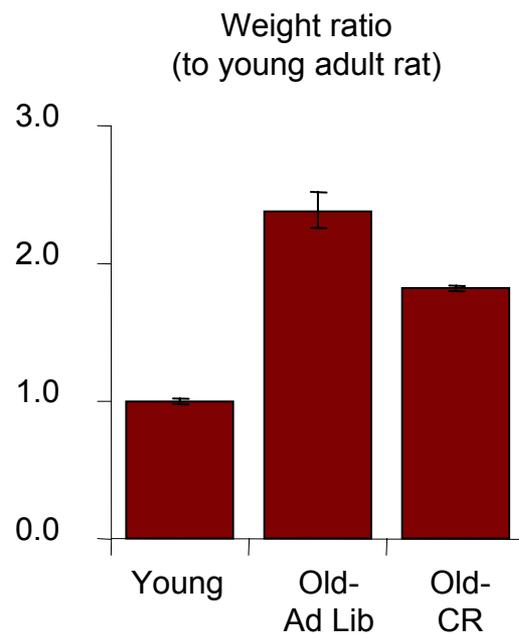
**Supplementary Figure 5. Quantitation of  $p16^{INK4a}$  and  $p19^{ARF}$  in tissues**

- a. Standard curve for the quantitation of the expression of  $p16^{INK4a}$  and  $Arf$  by quantitative RT-PCR. Similar curves were generated for  $p21^{CIP}$ ,  $p18^{INK4c}$ , and  $p19^{INK4d}$ . For all assays tested, the PCR reaction was linear over the range studied (19 to 40 cycles of amplification).

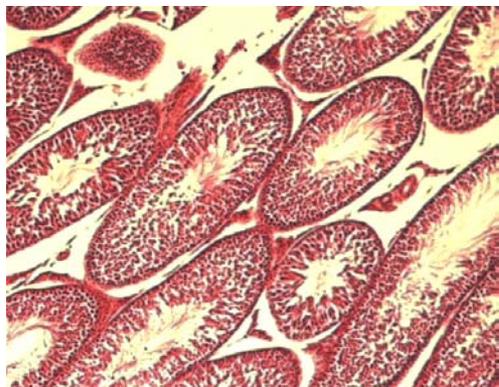
b, c, and d. Real-time p16<sup>INK4a</sup> amplification curves from cDNA from old versus young murine lung (b), cecum (c), and uterus (d). Change in TaqMan fluorescence per reaction cycle versus cycle number is plotted, and threshold for determination of Ct is indicated by a solid horizontal line.

**Supplementary Table I:** List of primers and probes used for TaqMan Real-time quantitative PCR.

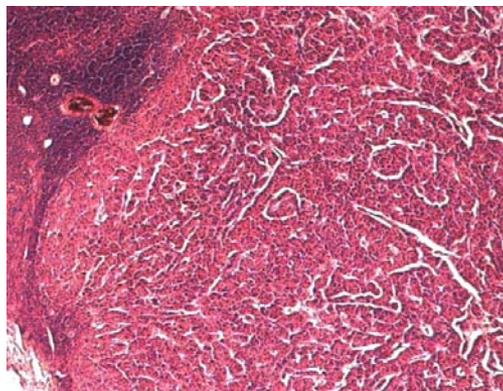
Gene	Forward primer	Reverse primer	Probe (5'-Tet, 3'-Tamra)	Reference sequence
<b>Mouse</b>				
<i>p16<sup>INK4a</sup></i>	CCCAACGCCCCGAACT	GCAGAAGAGCTGCTACGTGAA	TTCGGTCGTACCCCGATTGAGGTG	AF044336
<i>p19<sup>ARF</sup></i>	TGAGGCTAGAGAGGATCTTGAGA	GCAGAAGAGCTGCTACGTGAA	CCGCACCGGAATCCTGGACC	NM_009877
<i>Bmi-1</i>	AGAAGAGATTTTTATGCAGCTCA	CAACTTCTCCTCGGTCTTCA	AGCTGATGCTGCCAATGGCTCCA	NM_007552
	<b>ABI assay ID</b>			
<i>p15<sup>INK4b</sup></i>	Mm00483241_m1			NM_007670
<i>p18<sup>INK4c</sup></i>	Mm00483243_m1			NM_007671
<i>p19<sup>INK4d</sup></i>	Mm00486943_m1			NM_009878
<i>p21<sup>CIP</sup></i>	Mm00432448_m1			NM_007669
<i>p27<sup>KIP</sup></i>	Mm00438167_g1			NM_009875
<i>Ets-1</i>	Mm00468970_m1			NM_011808
<i>TBP</i>	Mm00446973_m1			NM_013684
<i>18S rRNA</i>	4333760F			X03205
<i>GAPDH</i>	Mm99999915_g1			NM_008084
<b>Rat</b>				
<i>p16<sup>INK4a</sup></i>	ACCAAACGCCCCGAACA	GAGAGCTGCCACTTTGACGT	TCGGTCGTACCCCGATACAGGTGA	L81167
<i>p19<sup>ARF</sup></i>	GAGGGCCGCAGCCACAT	CACCATAGGAGAGCAGGAGAGCT	CGTTGCCCATCATCATCACC TGGT	AF474975
	<b>ABI assay ID</b>			
<i>p21<sup>CIP</sup></i>	Rn00589996_m1			NM_080782
<i>Ets-1</i>	Rn00561167_m1			NM_012555
<i>Id1</i>	Rn00562985_s1			NM_012797
<i>18S rRNA</i>	4333760F			X03205

**a****b****c**

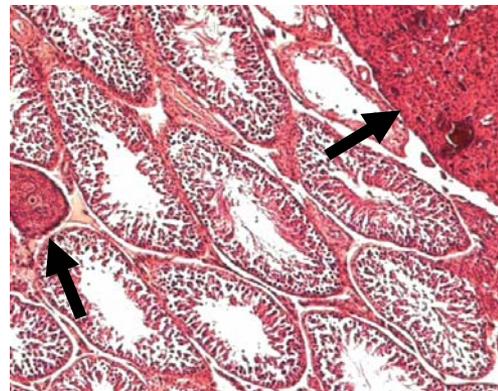
### Leydig Cell tumors in aging rat testis



YOUNG



OLD-AL



OLD-CR

Arrows = Leydig Cell Hyperplasia. Photos with 10x objective.

