

## Plasma chondroitin sulfate predicts the effectiveness of fluid resuscitation strategies in patients with sepsis

Kaori Oshima, Bailu Yan, Ran Tao, Gustavo Amorim, Chiara Di Gravio, Sarah A. McMurtry, Ryan C. Burke, Yunbi Nam, Ina Nikolli, Max S. Kravitz, Daniel Stephenson, Aaron Issaian, Kirk C. Hansen, Angelo D'Alessandro, Ivor S. Douglas, Wesley H. Self, Christopher J. Lindsell, Carolyn Leroux, Angelika Ringor, Michael A. Matthay, Jonathan S. Schildcrout, Nathan I. Shapiro, Eric P. Schmidt

*J Clin Invest.* 2026. <https://doi.org/10.1172/JCI202480>.

**Clinical Research and Public Health** **In-Press Preview** **Clinical Research** **Infectious disease** **Inflammation**

**BACKGROUND.** Plasma heparan sulfate, a glycosaminoglycan released during endothelial glycocalyx degradation, predicts sepsis mortality. Chondroitin sulfate is a circulating glycosaminoglycan not specific to glycocalyx degradation; its relevance to sepsis is unknown.

**METHODS.** We studied the associations of plasma chondroitin sulfate with (a) mortality in patients with sepsis-associated hypotension and (b) the relative effectiveness of a randomly-assigned liberal versus restrictive intravenous fluid resuscitation strategy. We selected 574 patients enrolled in the Crystalloid Liberal or Vasopressors Early Resuscitation in Sepsis trial using an outcome-enriched sampling strategy. We used liquid chromatography-mass spectrometry to quantify plasma chondroitin sulfate. In comparison, we measured hyaluronic acid as a glycocalyx degradation marker and IL-6 as an inflammatory biomarker. We conducted Cox proportional hazards regression analyses to examine associations of baseline biomarker concentrations with mortality and resuscitation strategy effectiveness. We used inverse probability of selection weights and generalized raking to account for the non-representative sampling design.

**RESULTS.** Plasma chondroitin sulfate, hyaluronic acid, and IL-6 were associated with mortality within 90 days. As baseline chondroitin sulfate increased, subsequent randomization to a restrictive strategy was increasingly beneficial ( $p = 0.022$ ): treatment effect hazard ratio (restrictive versus liberal) for mortality was estimated as 1.49 (95% CI 0.98–2.27), 1.30 (1.00–1.69), 1.09 (0.82–1.44), 0.88 (0.66–1.16), and 0.71 (0.52–0.97) for 10th, 25th, 50th, 75th and 90th percentiles of baseline chondroitin sulfate.

**CONCLUSIONS.** Plasma chondroitin [...]

**Find the latest version:**

<https://jci.me/202480/pdf>



1 Plasma chondroitin sulfate predicts the effectiveness of fluid resuscitation strategies in  
2 patients with sepsis

3 \*Kaori Oshima<sup>1</sup>, \*Bailu Yan<sup>2</sup>, Ran Tao<sup>2,3</sup>, Gustavo Amorim<sup>2</sup>, Chiara Di Gravio<sup>4</sup>, Sarah A.  
4 McMurtry<sup>5</sup>, Ryan C. Burke<sup>6</sup>, Yunbi Nam<sup>2</sup>, Ina Nikolli<sup>1</sup>, Max S. Kravitz<sup>6</sup>, Daniel  
5 Stephenson<sup>7</sup>, Aaron Issaian<sup>7</sup>, Kirk C. Hansen<sup>7</sup>, Angelo D'Alessandro<sup>7</sup>, Ivor S.  
6 Douglas<sup>5,8</sup>, Wesley H. Self<sup>9</sup>, Christopher J. Lindsell<sup>10</sup>, Carolyn Leroux<sup>11</sup>, Angelika  
7 Ringor<sup>11</sup>, Michael A. Matthay<sup>11</sup>, Jonathan S. Schildcrout<sup>2</sup>, †Nathan I. Shapiro<sup>6</sup>, †Eric P.  
8 Schmidt<sup>1</sup>

9 \*Contributed equally to the manuscript (first author)

10 †Contributed equally to the manuscript (last author)

11 Affiliations

12 <sup>1</sup>Department of Medicine, Mass General Brigham, Boston, MA, USA

13 <sup>2</sup>Department of Biostatistics, Vanderbilt University Medical Center, Nashville, TN, USA

14 <sup>3</sup>Vanderbilt Genetics Institute, Vanderbilt University Medical Center, Nashville, TN, USA

15 <sup>4</sup>Department of Epidemiology and Biostatistics, School of Public Health, Imperial  
16 College London, London, UK

17 <sup>5</sup>Department of Medicine, University of Colorado, Aurora, CO, USA

18 <sup>6</sup>Department of Emergency Medicine, Beth Israel Deaconess Medical Center and  
19 Harvard Medical School, Boston, MA, USA

20 <sup>7</sup>Department of Biochemistry and Molecular Genetics, University of Colorado, Aurora,  
21 CO, USA

22 <sup>8</sup>Department of Medicine, Denver Health Medical Center, Denver, CO, USA

23 <sup>9</sup>Vanderbilt Institute for Clinical and Translational Research and Department of  
24 Emergency Medicine, Vanderbilt University Medical Center, Nashville, TN, USA  
25 <sup>10</sup>Department of Biostatistics and Bioinformatics and the Duke Clinical Research  
26 Institute, Duke University Medical Center, Durham, NC, USA  
27 <sup>11</sup>Department of Medicine, Cardiovascular Research Institute, University of California  
28 San Francisco, San Francisco, CA, USA

29  
30 Since completion of this work, ISD affiliation has changed to Department of Medicine,  
31 Westchester Medical Center, Valhalla, New York, USA

32  
33 Corresponding Author:  
34 Eric P. Schmidt, MD  
35 Chief of Pulmonary and Critical Care Medicine, Mass General Brigham  
36 Andrew M. Tager MD Endowed Chair, Massachusetts General Hospital  
37 Associate Professor of Medicine, Harvard Medical School  
38 55 Fruit Street, Boston, MA, 02114, USA

39 Email: [eschmidt1@mgh.harvard.edu](mailto:eschmidt1@mgh.harvard.edu)  
40 Phone: 617-643-8692  
41 Fax: 617-726-6878

42  
43 **Author contribution:**

44 KO, CJL, JSS, NIS, EPS conceived and designed the study; KO, SAM, IN, DS, AI,  
45 KCH, AD, CL, AR, MAM acquired and analyzed the data; BY, GA, RT, CDG, RCB, YN,  
46 CJL, JSS performed statistical analyses; KO, MSK, BY, JSS, NIS, EPS drafted the  
47 manuscript; all authors contributed to the critical review and interpretation of the data  
48 and manuscript and intellectual content. KO is listed as first co-author, as she directly  
49 contributed to the conception and scope of the project.

50 **Conflict of interest:** AD is the founder and chief scientific officer of Omix Technologies,  
51 Inc, whose work is unrelated to the topic of this manuscript. ISD received a research  
52 grant from OpT, unrelated to the topic of this manuscript. WHS received consulting fees  
53 from Regeneron, Merck, and Pfizer, all unrelated to the topic of this manuscript. MAM  
54 received research grants from Roche-Genentech and Quantum Therapeutics, as well  
55 as consulting fees from Healios Pharmaceuticals and Merck and Co, all unrelated to the  
56 topic of this manuscript.

58 **Abstract**

59 **Background:** Plasma heparan sulfate, a glycosaminoglycan released during  
60 endothelial glycocalyx degradation, predicts sepsis mortality. Chondroitin sulfate is a  
61 circulating glycosaminoglycan not specific to glycocalyx degradation; its relevance to  
62 sepsis is unknown.

63 **Methods:** We studied the associations of plasma chondroitin sulfate with (a) mortality in  
64 patients with sepsis-associated hypotension and (b) the relative effectiveness of a  
65 randomly-assigned liberal versus restrictive intravenous fluid resuscitation strategy. We  
66 selected 574 patients enrolled in the Crystalloid Liberal or Vasopressors Early  
67 Resuscitation in Sepsis trial using an outcome-enriched sampling strategy. We used  
68 liquid chromatography-mass spectrometry to quantify plasma chondroitin sulfate. In  
69 comparison, we measured hyaluronic acid as a glycocalyx degradation marker and IL-6  
70 as an inflammatory biomarker. We conducted Cox proportional hazards regression  
71 analyses to examine associations of baseline biomarker concentrations with mortality  
72 and resuscitation strategy effectiveness. We used inverse probability of selection  
73 weights and generalized raking to account for the non-representative sampling design.

74 **Results:** Plasma chondroitin sulfate, hyaluronic acid, and IL-6 were associated with  
75 mortality within 90 days. As baseline chondroitin sulfate increased, subsequent  
76 randomization to a restrictive strategy was increasingly beneficial ( $p = 0.022$ ): treatment  
77 effect hazard ratio (restrictive versus liberal) for mortality was estimated as 1.49 (95%  
78 CI 0.98–2.27), 1.30 (1.00–1.69), 1.09 (0.82–1.44), 0.88 (0.66–1.16), and 0.71 (0.52–  
79 0.97) for 10<sup>th</sup>, 25<sup>th</sup>, 50<sup>th</sup>, 75<sup>th</sup> and 90<sup>th</sup> percentiles of baseline chondroitin sulfate.

80 **Conclusions:** Plasma chondroitin sulfate predicts sepsis mortality and may modify the  
81 response to a subsequent liberal vs. restrictive intravenous fluid resuscitation strategy.

82 **Trial:** ClinicalTrials.gov NCT03434028

83 **Funding:** R01HL149422, R01HL094786

84 **Introduction**

85 Randomized controlled trials comparing liberal and restrictive intravenous fluid  
86 resuscitation strategies in sepsis have found that neither strategy confers mortality  
87 benefit over the other in an unselected patient population (1, 2). However, it remains  
88 unclear if these strategies are equally effective for all patients, or if benefits experienced  
89 by a specific patient subtype for a given fluid resuscitation strategy are offset by harms  
90 experienced by a different patient subtype for that same strategy. Increasing  
91 appreciation of the biological heterogeneity of sepsis suggests that the ideal intravenous  
92 fluid resuscitation strategy for a patient with sepsis may vary based on the specific  
93 pathobiology responsible for the development of critical illness in that patient (3). To  
94 achieve such a precision medicine approach to fluid resuscitation, there is a need to  
95 identify predictive biomarkers that can identify an individual's responsiveness to a  
96 subsequent resuscitation strategy. To date, no such biomarker has been identified.

97 Circulating glycosaminoglycans such as heparan sulfate and hyaluronic acid  
98 serve as blood biomarkers of pathological shedding of the endothelial glycocalyx, a  
99 glycosaminoglycan-enriched endovascular layer necessary for vascular homeostasis  
100 (4). Accordingly, elevated plasma glycocalyx degradation products have been  
101 independently associated with worsened outcomes in patients with sepsis (5–7).  
102 Chondroitin sulfate, the most abundant glycosaminoglycan in human plasma (8), is  
103 similarly a glycocalyx glycosaminoglycan that is shed during endothelial glycocalyx  
104 degradation (9–11). However, the majority of plasma chondroitin sulfate is not derived  
105 from the glycocalyx but rather exists as components of circulating proteoglycans such  
106 as bikunin, a constituent of the circulating antiprotease inter- $\alpha$ -trypsin inhibitor (12, 13).

107 Reflecting these multifaceted origins, the value of plasma chondroitin sulfate as a  
108 prognostic and/or predictive biomarker in sepsis is uncertain.

109 To study the prognostic or predictive abilities of chondroitin sulfate as a  
110 biomarker in sepsis, we leveraged plasma specimens and patient data collected as part  
111 of a pre-specified ancillary study of the Crystalloid Liberal or Vasopressors Early  
112 Resuscitation in Sepsis (CLOVERS) trial, which randomized 1563 patients with sepsis-  
113 induced hypotension to a liberal intravenous fluid resuscitation strategy or a restrictive  
114 fluid strategy accompanied by early vasopressor administration (ClinicalTrials.gov  
115 NCT03434028) (2). We sought to determine if plasma chondroitin sulfate concentrations  
116 were associated with (a) mortality within 90 days of study randomization and (b)  
117 differential treatment effectiveness of the liberal and restrictive fluid strategies. We  
118 compared these prognostic and predictive abilities of chondroitin sulfate to hyaluronic  
119 acid, a plasma glycosaminoglycan studied as a specific biomarker of endothelial  
120 glycocalyx degradation (9, 14), and interleukin 6 (IL-6), a marker of systemic  
121 inflammation (15).

122

123

124 **Results**

125 *Study cohorts*

126 CLOVERS inclusion criteria included patients aged >18 years with sepsis-induced  
127 hypotension, defined as systolic blood pressure <100 mmHg after 1 liter of intravenous  
128 fluid, who were early in their hospital course. The complete inclusion and exclusion  
129 criteria for the trial are available in the primary CLOVERS report (2). Plasma specimens  
130 were collected at enrollment (a median of 61 min after study eligibility) and 24 and 72  
131 hours after randomization.

132 Since liquid chromatography-mass spectrometry (LC-MS/MS) quantification of  
133 chondroitin sulfate and hyaluronic acid is cost-prohibitive to perform on all CLOVERS  
134 participants, we selected a subset of 600 participants from the 1563 participants for  
135 inclusion into the present substudy. As previously described (7), we enriched this subset  
136 with patients who died within 90 days of randomization and those who had Acute  
137 Respiratory Distress Syndrome (ARDS) at baseline or within seven days of  
138 randomization. By enriching the subset with those who died, we observed a relatively  
139 informative sample compared to a random sample and therefore can estimate the  
140 parameters in our models with lower uncertainty (i.e., smaller standard errors) and  
141 higher power. We then randomly selected from the remaining participants to reach a  
142 total of 300 participants from each of the liberal and restrictive groups. Twenty  
143 participants did not have sufficient baseline plasma sample volume available for mass  
144 spectrometry, and six participants were missing baseline characteristics. Our analyses  
145 were therefore performed using samples collected from the remaining 574 participants.  
146 In those who were still alive and hospitalized, repeat measurements were made 24

147 hours (N=476) and 72 hours (N=326) after study randomization. Since the selected sub-  
148 samples were not random samples from the original CLOVERS cohort, the analyses  
149 were weighted using inverse probability of selection weighting and generalized raking  
150 so that results from our selected cohort generalize to the population represented by the  
151 entire CLOVERS cohort (Table 1, Supplemental Table 1).

152 IL-6 measurements were performed as part of the trial protocol on 1371 of the  
153 1563 subjects enrolled in CLOVERS. There were 2 participants missing baseline  
154 characteristics, so our analyses were performed on the remaining 1369 participants. IL-  
155 6 was measured at enrollment (N=1369) and 72 hours (N=844). Like the analyses of  
156 chondroitin sulfate and hyaluronic acid, we used inverse probability of selection  
157 weighting and generalized raking estimation strategies to ensure that results from IL-6  
158 analyses generalize to the population represented by the full CLOVERS cohort  
159 (Supplemental Table 2).

160

161 *Association of chondroitin sulfate with indices of endothelial glycocalyx degradation and*  
162 *systemic inflammation*

163 In each plasma sample collected for mass spectrometry analysis of circulating  
164 glycosaminoglycans, we compared concentrations of chondroitin sulfate, hyaluronic  
165 acid, and IL-6 with previously measured concentrations of heparan sulfate and  
166 syndecan-1 (7), canonical markers of endothelial glycocalyx shedding that we have  
167 previously shown to be prognostic for 90-day mortality in CLOVERS (7). We found  
168 moderate associations between circulating chondroitin sulfate and canonical glycocalyx  
169 shedding markers hyaluronic acid ( $r = 0.33$ ), heparan sulfate ( $r = 0.42$ ) and the

170 glycocalyx proteoglycan syndecan-1 ( $r = 0.28$ ) in samples collected at study enrollment,  
171 confirming that circulating chondroitin sulfate is not solely a marker of endothelial  
172 glycocalyx degradation (Figure 1). Chondroitin sulfate was not well correlated with IL-6  
173 ( $r = 0.07$ ), a canonical marker of systemic inflammation in sepsis (Figure 1). Taken  
174 together, these findings indicate that plasma chondroitin sulfate reflects biological  
175 processes distinct from endothelial glycocalyx injury and/or systemic inflammation.

176

177 *Plasma chondroitin sulfate, hyaluronic acid, and IL-6 at study enrollment are*  
178 *independently associated with sepsis mortality*

179 At each time point measured, plasma concentrations of chondroitin sulfate (Figure 2A)  
180 were elevated in CLOVERS patients who died within 90 days of study randomization.  
181 Patients in the highest tertile of plasma chondroitin sulfate concentration (9275 – 32147  
182 ng/ml) at the time of study enrollment (i.e., prior to randomization) had a significantly  
183 higher 90-day mortality compared to those in the medium (7022 – 9263 ng/ml) or low  
184 (1272 – 7017 ng/ml) plasma chondroitin sulfate tertiles (log-rank  $p < 0.001$ ) (Figure 2B).  
185 The association of elevated plasma chondroitin sulfate at study enrollment with  
186 increased 90-day mortality persisted in our fitted Cox model that adjusted for severity of  
187 illness, patient demographics, randomized fluid resuscitation strategy, and chronic co-  
188 morbidities (Figure 2C, entire model shown in Supplemental Figure 1). For example, the  
189 hazard ratio for mortality comparing the 10<sup>th</sup> percentile (5303 ng/ml) to the 50<sup>th</sup>  
190 percentile (7861 ng/ml) of baseline chondroitin sulfate was 1.06 (95% CI: 0.84 – 1.33),  
191 while the hazard ratio comparing the 90<sup>th</sup> percentile (12202 ng/ml) to the 50<sup>th</sup> percentile  
192 was 1.98 (95% CI: 1.65 – 2.39).

193 Plasma concentrations of hyaluronic acid (a canonical endothelial glycocalyx  
194 degradation marker) and IL-6 (a canonical marker of systemic inflammation) similarly  
195 were associated with 90-day survival. Patients who went on to die within 90 days of  
196 study enrollment had higher levels of plasma hyaluronic acid at all timepoints (Figure  
197 2D) in comparison to 90-day survivors. Patients in the highest tertile of plasma  
198 hyaluronic acid concentration (370 – 17812 ng/ml) at the time of study enrollment  
199 demonstrated higher 90-day mortality than those in the medium (90 - 368 ng/ml) and  
200 low (7 – 89 ng/ml) tertiles, log rank  $p < 0.001$  (Figure 2E). Our adjusted Cox model  
201 accordingly showed that baseline hyaluronic acid was independently associated with  
202 90-day mortality (Figure 2F, entire model shown in Supplemental Figure 2). The hazard  
203 ratio comparing the 10<sup>th</sup> percentile (31 ng/ml) to the 50<sup>th</sup> percentile (142 ng/ml) of  
204 baseline hyaluronic acid was 0.28 (95% CI: 0.17 – 0.47), while the hazard ratio  
205 comparing the 90<sup>th</sup> percentile (1321 ng/ml) to the 50<sup>th</sup> percentile was 2.05 (95% CI: 1.58  
206 – 2.66). Similarly, patients who went on to die within 90 days of study enrollment had  
207 higher levels of plasma IL-6 at all timepoints (Figure 2G) in comparison to 90-day  
208 survivors. Patients in the lowest tertile of baseline plasma IL-6 (0 – 29 pg/ml) at study  
209 enrollment had lower mortality than those in the highest tertile (170 – 34519 pg/ml) or  
210 medium tertile (29 – 170 pg/ml) tertiles (log-rank  $p < 0.001$ , Figure 2H). This association  
211 of baseline IL-6 with 90-day mortality persisted in covariate-adjusted analysis (Figure 2I,  
212 entire model shown in Supplemental Figure 3). The hazard ratio comparing the 10<sup>th</sup>  
213 percentile (8 pg/ml) to the 50<sup>th</sup> percentile (63 pg/ml) of baseline IL-6 was 0.60 (95% CI:  
214 0.44 – 0.81), while the hazard ratio comparing the 90<sup>th</sup> percentile (2382 pg/ml) to the  
215 50<sup>th</sup> percentile was 1.08 (95% CI: 0.83 – 1.41).

216

217 *Post-resuscitation plasma chondroitin sulfate, hyaluronic acid, and IL-6 are not altered*  
218 *by antecedent randomization to a liberal vs. restrictive fluid resuscitation strategy*

219 Previous observational studies suggested that a liberal intravenous fluid resuscitation  
220 strategy may induce iatrogenic endothelial glycocalyx degradation, leading to an  
221 increase in circulating glycocalyx fragments (16). Such iatrogenic endothelial glycocalyx  
222 injury, however, was not observed in randomized, prospective trials of fluid resuscitation  
223 (7). In contrast to glycocalyx-derived glycosaminoglycans, the impact of antecedent  
224 intravenous fluid resuscitation on circulating chondroitin sulfate concentrations was  
225 unstudied. We found that plasma chondroitin sulfate concentrations 24 hours after  
226 enrollment were unaffected by antecedent randomized resuscitation strategy (Figure 3,  
227 Supplemental Figure 4), suggesting that choice of fluid resuscitation strategy did not  
228 induce changes in circulating chondroitin sulfate. Similarly, plasma hyaluronic acid (a  
229 canonical endothelial glycocalyx degradation biomarker) 24 hours after enrollment (i.e.,  
230 at the completion of the study fluid protocol) was not affected by the antecedent  
231 randomization to either fluid resuscitation strategy (Figure 3, Supplemental Figure 5),  
232 supporting previous studies' findings that fluid resuscitation strategy does not drive  
233 glycocalyx injury.

234 As IL-6 levels were not immediately measured after completion of the 24-hour  
235 period of protocolized fluid resuscitation, we used plasma IL-6 measured 72 hours after  
236 enrollment to determine if antecedent early fluid resuscitation strategy influenced  
237 persistent systemic inflammation. Using our adjusted linear regression model, we found  
238 that early assignment to a liberal or restrictive intravenous fluid resuscitation strategy

239 had no association with plasma IL-6 72 hours after study enrollment, suggesting no  
240 impact of early (first 24 hours) resuscitation strategy on persistent indices of systemic  
241 inflammation (Figure 3, Supplemental Figure 6).

242

243 *Plasma chondroitin sulfate, but not hyaluronic acid or IL-6, modified the effectiveness of*  
244 *a subsequent, randomly assigned fluid resuscitation strategy on 90-day mortality*

245 We sought to determine whether baseline plasma chondroitin sulfate concentration  
246 modified the relative effectiveness of a subsequent, randomly assigned liberal versus  
247 restrictive intravenous fluid resuscitation strategy on mortality within 90 days. Our  
248 covariate-adjusted Cox model demonstrated a differential treatment effect (Figure 4A;  $p$   
249 = 0.022) according to chondroitin sulfate concentrations at study enrollment. As  
250 baseline plasma chondroitin sulfate concentrations increased, randomization to a  
251 restrictive (as compared to liberal) intravenous fluid resuscitation strategy was  
252 estimated to be increasingly beneficial to patients. For example, the treatment effect  
253 hazard ratio (restrictive versus liberal fluid resuscitation strategy) for mortality was  
254 estimated to be 1.49 (95% CI 0.98 – 2.27), 1.30 (95% CI 1.00 – 1.69), 1.09 (95% CI  
255 0.82 – 1.44), 0.88 (95% CI 0.66 – 1.16), and 0.71 (95% CI 0.52 – 0.97) for baseline  
256 chondroitin sulfate concentrations of 5329 ng/ml, 6418 ng/ml, 7937 ng/ml, 10112 ng/ml,  
257 and 12779 ng/ml, respectively. These reference values of baseline chondroitin sulfate  
258 concentrations represent the 10<sup>th</sup>, 25<sup>th</sup>, 50<sup>th</sup>, 75<sup>th</sup> and 90<sup>th</sup> percentiles of the observed  
259 distribution. Associations of chondroitin sulfate concentrations with hazard of mortality  
260 are shown separately for the liberal and restrictive resuscitation strategy arms in Figure  
261 4B.

262 In contrast to chondroitin sulfate, we observed no evidence of differential  
263 treatment effect for 90-day mortality as a function of baseline plasma hyaluronic acid  
264 (Figure 4C, D) or IL-6 (Figure 4E, F) and subsequent randomization to liberal or  
265 restrictive fluid resuscitation strategy.

266

267 *Disaccharide analyses suggest the importance of unsulfated chondroitin sulfate in  
268 modifying response to fluid resuscitation strategy*

269 To explore potential mechanisms underlying the predictive ability of circulating  
270 chondroitin sulfate on an individual patient's response to a subsequent, randomly  
271 assigned fluid resuscitation strategy, we investigated the sulfation characteristics of  
272 plasma chondroitin sulfate in CLOVERS subjects. Chondroitin sulfate is a linear  
273 polysaccharide, composed of repeating glucuronic acid-galactosamine disaccharides.  
274 Each disaccharide unit may be unsulfated ("0S") or sulfated at either the 6-O or 4-O  
275 position of galactosamine. The relative abundance of these sulfated disaccharides  
276 within plasma chondroitin sulfate, as measured by mass spectrometry, can provide  
277 insight into the potential binding partners of circulating chondroitin sulfate during sepsis.  
278 For example, the circulating proteoglycan bikunin, a component of the endogenous  
279 antiprotease inter-alpha-trypsin inhibitor, is decorated with chondroitin sulfate that is  
280 enriched in 4-O sulfated disaccharides (12, 13). Interestingly, we found that the  
281 differential treatment effect of baseline plasma chondroitin sulfate on the response to a  
282 fluid resuscitation strategy was largely driven by unsulfated chondroitin sulfate (Figure  
283 5A) and not by 4-O or 6-O sulfated disaccharides (Figure 5B, C). Therefore, the lack of  
284 predictive ability of 4-O chondroitin sulfate suggests that the observed differential

285 response of fluid resuscitation strategy in CLOVERS might not be mediated by  
286 circulating bikunin. Patient-level factors associated with circulating concentrations of  
287 chondroitin sulfate and its subtypes are provided in Supplemental Figures 7, 8.  
288 Intriguingly, 0S chondroitin sulfate concentrations at study entry were inversely  
289 correlated with an antecedent history of liver disease, potentially suggesting a hepatic  
290 source of this chondroitin sulfate subtype (Supplemental Figure 8A).

291

292

293 **Discussion**

294 In this secondary analysis of the CLOVERS trial, we found that plasma chondroitin  
295 sulfate was both a prognostic and predictive biomarker in patients with sepsis-  
296 associated hypotension. Plasma chondroitin sulfate concentrations early in the course  
297 of sepsis care (with CLOVERS patient enrollment occurring a median of 61 min after  
298 study eligibility, defined as sepsis-induced hypotension despite one liter of crystalloid  
299 treatment) independently predicted 90-day mortality, similar to other circulating  
300 glycosaminoglycans (such as hyaluronic acid or heparan sulfate (7)) more specific to  
301 endothelial glycocalyx degradation. Strikingly, plasma chondroitin sulfate concentrations  
302 at study enrollment predicted a patient's subsequent response to a randomly assigned  
303 liberal or restrictive intravenous fluid resuscitation strategy, using 90-day mortality as  
304 the outcome. This heterogeneity of treatment effect was unique to chondroitin sulfate  
305 and was not observed with hyaluronic acid (a canonical glycosaminoglycan marker of  
306 endothelial glycocalyx shedding) or IL-6 (a canonical marker of systemic inflammation).  
307 These findings suggest that plasma chondroitin sulfate levels could inform decision-  
308 making between different fluid resuscitation strategies in patients with sepsis-induced  
309 hypotension. Indeed, on adjusted analysis the "inflection point" of plasma chondroitin  
310 sulfate at which the benefits of liberal vs. restrictive fluid resuscitation flip is the 62<sup>nd</sup>  
311 percentile of patients, suggesting that the relevance of this predictive biomarker is not  
312 limited to a small subset of our cohort but rather applies to large numbers of patients  
313 with sepsis.

314 The mechanisms underlying the predictive capabilities of circulating chondroitin  
315 sulfate are uncertain. Unlike the glycosaminoglycans heparan sulfate and hyaluronic

316 acid, which are anchored within the endothelial glycocalyx and do not circulate at high  
317 concentrations during health, plasma chondroitin sulfate is abundant in normal blood  
318 (12, 17). Indeed, our findings are consistent with chondroitin sulfate not being simply a  
319 marker of endothelial glycocalyx degradation, as chondroitin sulfate was only  
320 moderately associated with traditional biomarkers of endothelial glycocalyx degradation  
321 such as heparan sulfate, hyaluronic acid, or the glycocalyx proteoglycan syndecan-1.  
322 Our findings also suggest that chondroitin sulfate is not simply a proxy for systemic  
323 inflammation, as circulating chondroitin sulfate level was minimally correlated with the  
324 canonical inflammatory marker IL-6. While the predictive capabilities of chondroitin  
325 sulfate persisted even after controlling for SOFA score (which includes measures of  
326 renal function) and pre-existing chronic kidney disease, it is possible that plasma  
327 chondroitin sulfate may be a biomarker of early, subclinical renal injury. As patients with  
328 renal disease demonstrated a non-significant trend towards worsened survival with  
329 liberal fluid resuscitation in the parent CLOVERS study (2), an association of high  
330 chondroitin sulfate levels with early, subclinical renal failure could potentially explain the  
331 predictive capabilities of chondroitin sulfate observed in our study.

332 We additionally considered that the heterogeneity of treatment effect associated  
333 with chondroitin sulfate may relate to its role as a component of circulating  
334 proteoglycans such as bikunin, a constituent of inter-alpha-trypsin inhibitor (18). Inter-  
335 alpha-trypsin inhibitors are protective antiproteases that inhibit serine proteases such as  
336 leukocyte elastase and cathepsin G (19). In sepsis, inter-alpha-trypsin inhibitor is  
337 proteolytically cleaved, increasing the circulating concentrations of free, chondroitin  
338 sulfate-decorated bikunin (20). As bikunin enriched from the plasma of patients with

339 septic shock was sufficient to induce endothelial damage *ex vivo* (21), an increase in  
340 circulating chondroitin sulfate could indicate an endothelial-injured state in which  
341 pathogenic edema might be worsened by a liberal fluid resuscitation approach.  
342 However, we found that the differential responses of fluid resuscitation strategy was  
343 largely associated with unsulfated chondroitin sulfate (Figure 5A, B), as opposed to the  
344 4-O sulfated chondroitin sulfates typically bound to bikunin (Figure 5C, D) (12, 13). This  
345 finding suggests that any ability of circulating chondroitin sulfate to modify the response  
346 to fluid resuscitation is unrelated to bikunin-associated chondroitin sulfate. Alternatively,  
347 our results may represent a potential shift in chondroitin sulfate sulfation in sepsis (22,  
348 23) or the exposure and circulation of cryptic undersulfated chondroitin sulfate sites by  
349 bikunin/inter-alpha-trypsin inhibitor fragmentation during sepsis (20).

350 Collectively, our findings identify an important yet unexplored line of mechanistic  
351 investigation in sepsis: how chondroitin sulfate modifies fluid resuscitation strategies.  
352 The lack of impact of randomized resuscitation strategy on plasma chondroitin sulfate  
353 measured at 24 hours (i.e., at the conclusion of protocolized resuscitation) or at 72  
354 hours (Figure 3, Supplemental Figure 4) suggests that increased plasma concentrations  
355 of chondroitin sulfate, while able to identify patient subtypes that are differentially  
356 responsive to fluid resuscitation strategy, are not part of the causal pathway of that  
357 differential responsiveness. This observation, however, requires exploration with  
358 additional mechanistic investigation.

359 Regardless of its underlying biological mechanism, the predictive capabilities of  
360 plasma chondroitin sulfate in CLOVERS suggest a role for this biomarker in a precision  
361 medicine approach to sepsis resuscitation. Our findings (Figure 4A, B) suggest that

362 patients with high concentrations of plasma chondroitin sulfate at sepsis presentation  
363 would benefit from assignment to a restrictive intravenous fluid resuscitation strategy.  
364 However, our findings were derived using state-of-the art mass spectrometry  
365 approaches that are expensive and infeasible for rapid implementation at the bedside.  
366 Future trials testing the value of plasma chondroitin sulfate in a sepsis treatment  
367 algorithm should therefore employ rapid, inexpensive point-of-care assays of circulating  
368 glycosaminoglycans. As circulating chondroitin sulfate arises from multiple non-  
369 glycocalyx sources, endothelial glycocalyx degradation markers such as plasma  
370 syndecan-1 (Figure 1) or sublingual intravital microscopy (4) are unlikely to serve as  
371 accurate proxies of plasma chondroitin sulfate. Future studies should test the bedside  
372 validity of direct assays of circulating glycosaminoglycans (such as dimethylmethyleno  
373 blue, DMMB (9)) as rapid indices of plasma chondroitin sulfate in patients with sepsis. It  
374 is unclear, however, if these assays (which typically target sulfated regions of  
375 glycosaminoglycans) will capture the heterogeneity of treatment effect observed in  
376 CLOVERS, which was largely driven by unsulfated chondroitin sulfate (Figure 5A, B).

377 Our work provides additional insights into the mechanisms and consequences of  
378 endothelial glycocalyx shedding during critical illness. Previous preclinical studies have  
379 suggested that endothelial glycocalyx sheddases are induced by inflammatory stimuli  
380 (24, 25). However, our study only found a weak association between circulating IL-6  
381 and biomarkers of endothelial glycocalyx shedding (heparan sulfate, hyaluronic acid, or  
382 syndecan-1) or circulating chondroitin sulfate. This finding suggests that circulating  
383 glycosaminoglycans are not simply a proxy for systemic inflammation, but likely  
384 represent distinct pathological processes such as vascular injury (potentially mediated

385 by angiopoietin-2 signaling (26)) or, for chondroitin sulfate, alterations in processes  
386 mediated by inter-alpha-trypsin inhibitor and/or bikunin signaling (18). Importantly,  
387 neither baseline hyaluronic acid nor IL-6 (or heparan sulfate and syndecan-1, as  
388 previously reported (7)) were associated with differential responses to a randomized  
389 fluid resuscitation strategy, suggesting that the presence (and magnitude) of endothelial  
390 glycocalyx injury and/or systemic inflammation did not determine an individual patient's  
391 subsequent response to a liberal or restrictive approach to intravenous fluids. Finally,  
392 our study suggests that sepsis-induced endothelial glycocalyx degradation is not simply  
393 the consequence of a single "sheddase". In contrast to heparan sulfate, which is  
394 anchored to the endothelial surface by heparan sulfate proteoglycans such as  
395 syndecans, hyaluronic acid intercalates throughout the endothelial glycocalyx and  
396 interacts with the endothelial surface via cell membrane hyaladherins such as CD44 (4).  
397 Thus, hyaluronic acid shedding is unlikely to occur via heparan sulfate-specific  
398 sheddases (such as heparanase) previously implicated in septic endothelial glycocalyx  
399 degradation (4). The similar associations of plasma hyaluronic acid, heparan sulfate,  
400 and syndecan-1—each of which is shed from the endothelial glycocalyx by distinct  
401 sheddases—with sepsis outcomes suggests the presence of coordinated, possibly  
402 redundant processes of endothelial glycocalyx shedding during human sepsis.

403 Our investigation has several limitations. As noted above, the biological  
404 mechanism responsible for the striking finding of heterogeneity of treatment effect for  
405 plasma chondroitin sulfate and intravenous fluid resuscitation strategy remains  
406 unknown. Further mechanistic investigation is needed to study the relative impact of  
407 plasma chondroitin sulfate and/or its associated proteins on the response to intravenous

408 fluids. The second limitation regards our weighted analyses: although we controlled for  
409 numerous variables that could confound our results, the possibility of residual  
410 confounding remains. Finally, this study did not stratify patients by the type of  
411 intravenous fluid received. While the majority of patients were given balanced  
412 crystalloids for resuscitation, some received normal saline, and this variation was not  
413 controlled for in our analyses or original CLOVERS study (2).

414 In conclusion, this investigation identifies that in patients with sepsis-induced  
415 hypotension, baseline chondroitin sulfate levels reflect pathobiological processes other  
416 than endothelial glycocalyx degradation and/or systemic inflammation. Strikingly,  
417 baseline chondroitin sulfate concentrations predict an individual's response (as judged  
418 by the outcome of 90-day mortality) to a subsequent liberal versus restrictive fluid  
419 resuscitation strategy. This finding represents a major advance towards precision  
420 medicine approaches to intravenous fluid resuscitation in sepsis. Finally, this study  
421 validates chondroitin sulfate, hyaluronic acid, and IL-6 as prognostic biomarkers for  
422 sepsis mortality.

423 **Methods**

424 *Sex as a biological variable*

425 The study cohort included both male and female patients with sepsis-associated  
426 hypotension. Inverse probability of selection weighting and generalized raking  
427 estimation strategies were used to ensure that this cohort reflected the entire CLOVERS  
428 population (Table 1, Supplemental Table 2).

429

430 *Analyses of circulating biomarkers*

431 We measured chondroitin sulfate and hyaluronic acid disaccharides (Supplemental  
432 Table 3A) with ultra high-performance liquid chromatography-mass spectrometry  
433 (UPLC-MS/MS) as described previously (7) with modifications. Briefly, we spiked all  
434 samples with C<sup>13</sup>-labeled heparan sulfate polysaccharide recovery calibrant (1 ng) prior  
435 to sample processing to later verify complete digestion of glycosaminoglycans and to  
436 account for any substantial sample loss during processing. We desalted 50 µL of  
437 plasma using 3 kDa molecular weight cut-off column (Millipore UFC500396) and  
438 performed overnight on-column digestion with heparinase I, II, III, and chondroitinase  
439 ABC. We removed enzymes with centrifugation and collected digested  
440 glycosaminoglycans. To quantify chondroitin sulfate and hyaluronic acid, we spiked in  
441 C<sup>13</sup>-labeled disaccharide calibrants, 1 ng each, (Supplemental Table 3B) and lyophilized  
442 samples. We then derivatized disaccharides with 2-aminoacridone (AMAC) in  
443 DMSO/glacial acetic acid (17:3, v/v) followed by addition of aqueous sodium  
444 cyanoborohydride. The reaction mixture was incubated at 45°C for 2 hours. We cleaned  
445 samples with C18 resin, washed with 0.1% formic acid solution and eluted with 80%

446 acetonitrile and 0.1% formic acid solution. We removed the organics with a speed  
447 vacuum concentrator overnight and reconstituted the samples with mobile phase A.

448 We quantified AMAC-derivatized disaccharides with UltiMate 3000 LC system  
449 (Thermo) in tandem with QTRAP 5500 mass spectrometry (AB Sciex). Chromatography  
450 was reverse-phase and performed with Aquity UPLC BEH-C18 (Waters, 150 x 1.0, 1.7  
451  $\mu$ m) and with Aquity UPLC BEH-C18 guard column (Waters, 5x 2.1, 1.7  $\mu$ m).

452 Disaccharides were separated using mobile phase A (95:5 water:methanol, 1 mM  
453 ammonium acetate, pH 8.6) and mobile phase B (95:5 methanol: water, 1 mM  
454 ammonium acetate) with the gradient as follows: 0–15 minutes, 0%–15% B; 15–17  
455 minutes, 15%–35% B; 17.01 minutes, 100% B; 20 minutes, 100% B; 20.01 minutes, 0%  
456 B; 23 minutes, 0% B, at a flow rate of 0.1 mL/min. The column temperature was held at  
457 45°C. The mass spectrometer was operated in electrospray negative ionization mode  
458 with multiple reaction monitoring. Ionization parameters common to all analytes were  
459 set as follows: curtain gas at 30 psi, nebulizing gas at 40 psi, drying gas at 20 psi, ion  
460 spray voltage at -4500V, source temperature at 400°C. Compound-specific parameters  
461 (entrance potential, declustering potential, collision energy, collision cell exit potential)  
462 are listed on Supplemental Table 3C. Dwell time for each analyte was 20 msec. The  
463 mass spectrometry data were acquired with Analyst 1.6.2. We determined the  
464 concentration of each analyte based on the internal calibrant as shown in Supplemental  
465 Table 3B.

466 Total chondroitin sulfate concentration within an individual sample was calculated  
467 as the sum of all eight possible chondroitin sulfate disaccharides (Supplemental Table  
468 3A) within that sample. In rare cases, select peaks for individual disaccharides were

469 unable to be measured within a sample. In this case, the concentration of the missing  
470 disaccharide was imputed using information from the other, non-missing disaccharides  
471 within that sample. At baseline, of the  $574 \times 8 = 4592$  disaccharide measurements, 6  
472 (0.1%) of them were missing and were singly imputed prior to calculating the baseline  
473 total chondroitin sulfate value. At 24 hours post-randomization, of the  $476 \times 8 = 3808$   
474 constituents, 4 (0.1%) of them were missing and were singly imputed prior to calculating  
475 the 24-hour total chondroitin sulfate value.

476 Plasma IL-6 was measured using Human Luminex Discovery Assays (R&D  
477 Systems, Minneapolis, MN, USA).

478

479 *Statistical Analysis*

480 We summarized the baseline and demographic characteristics of the study sample for  
481 chondroitin sulfate and hyaluronic acid, the study sample for IL-6, and the full  
482 CLOVERS study cohort with frequencies and percentages for discrete variables and  
483 with the median and interquartile range for quantitative variables. We used violin plots to  
484 display the distributions at baseline and at 24 hours (chondroitin sulfate and hyaluronic  
485 acid only) and 72 hours after study enrollment for those who did and did not die during  
486 the 90-day follow-up period, and for those assigned to liberal or restrictive fluid strategy.

487 Because our study included non-representative, stratified sampling from the  
488 original CLOVERS cohort, we conducted analyses using inverse probability of being  
489 sampled weighting (IPSW) (27) and generalized raking approaches (28, 29). IPSW  
490 weights each participant selected for plasma collection by the inverse of their probability  
491 of being sampled, while generalized raking further calibrates the sampling weights using

492 information observed in the full CLOVERS cohort. Importantly, even though the sub-  
493 study samples are not representative of the original CLOVERS cohort, by reweighting  
494 individuals selected for plasma collection, our findings generalize to the population  
495 represented by the original CLOVERS cohort (30). See Supplemental Material for more  
496 detail on the IPSW and generalized raking analysis strategies.

497 We applied IPSW with estimated weights for correlation estimates and tests and  
498 for Kaplan-Meier analyses. Sampling probabilities used to calculate the IPSW weights  
499 for chondroitin sulfate and hyaluronic acid are shown in Supplemental Table 4, and  
500 sampling probabilities for IL-6 are shown in Supplemental Table 5. The Kaplan-Meier  
501 analyses examined the unadjusted association between baseline biomarker tertiles and  
502 survival from randomization to 90 days post-randomization.

503 For all regression models, we sought to adjust for a pre-specified set of baseline  
504 covariates that might confound associations with mortality. This set includes:  
505 randomized treatment assignment, baseline SOFA score, sex, age, race and ethnicity,  
506 ARDS at randomization, history of diabetes, history of heart failure, and history of end-  
507 stage renal disease. SOFA score and age were entered into the models using restricted  
508 cubic splines with three knots to permit non-linear associations. After an initial model fit,  
509 the proportional hazards assumption was observed to be violated for associations with  
510 sex, heart failure history, and SOFA score. To address this concern, to avoid the  
511 uncertainty associated with making multiple decisions about proportional hazards  
512 assumptions across models, and to flexibly adjust for key potential confounders, we  
513 stratified all Cox model analyses by sex and heart failure history, and we included  
514 SOFA score interactions with follow-up time (0–4 days, 5–11 days, and 12–90 days)(7).

515 Biomarker measurements were centered in all regression models. Because patient data  
516 were collected at 51 sites in the CLOVERS study, we used design-adjusted robust  
517 standard errors to account for clustering by site and to acknowledge the stratified  
518 sampling design (30). We combined sites with less or equal to five subjects into one  
519 group with N=23 subjects to have sufficient data for each site.

520 We fit Cox proportional hazard regression models to examine 1) the baseline-  
521 covariate adjusted biomarker associations with mortality through 90 days and 2) the  
522 extent to which the treatment (restrictive versus liberal resuscitation strategy) effect on  
523 mortality through 90 days varied according to baseline biomarker concentrations. These  
524 models controlled for the baseline covariates mentioned above. To fit the latter model,  
525 we added the randomized treatment assignment by biomarker concentration interaction  
526 to the former model. To adjust for the non-representative sample, we used generalized  
527 raking.

528 To examine the randomized treatment assignment effect on biomarker  
529 concentrations at 24-hours (for chondroitin sulfate and hyaluronic acid) and the 72-  
530 hours (for IL-6), we performed linear regression after adjusting for the non-  
531 representative sample with IPSW. These models controlled for baseline covariates  
532 mentioned above as well as the baseline biomarker concentration.

533 For all baseline models, there were 574 subjects analyzed for chondroitin sulfate  
534 and hyaluronic acid, and 1369 for IL-6; For models with 24-hour (for chondroitin sulfate  
535 and hyaluronic acid) or 72-hour data (for IL-6), 476 subjects were analyzed for  
536 chondroitin sulfate and hyaluronic acid, and 844 subjects for IL-6. We summarized  
537 model results graphically.

538           Additional statistical details are provided in the Supplemental Statistical Methods.  
539   All analyses were performed using R version 4·4·1, and analyses were performed with  
540   the survey, survival, Hmisc, and packages (31–38). A  $p$  value less than 0.05 was  
541   considered significant.

542

543   *Study Approval.*

544   The CLOVERS study and this ancillary analysis were approved by the Vanderbilt IRB,  
545   which acted as a central IRB, and patients were enrolled using a written informed  
546   consent, as previously described (2).

547

548   *Data Availability.*

549   Data underlying all dot-plot, survival analyses, and violin figures are provided in the  
550   accompanying “Supporting Data Values” file. Details on the code used to fit the  
551   regression models are provided in the Supplemental R code for Model Fitting.  
552   Deidentified human data are available upon reasonable request to the corresponding  
553   author (EPS).

554

555

556 **Funding Support**

557 This work is the result of NIH funding (R01HL149422 to NIS and EPS, R01HL094786 to  
558 JSS), in whole or in part, and is subject to the NIH Public Access Policy. Through  
559 acceptance of this federal funding, the NIH has been given a right to make the work  
560 publicly available in PubMed Central.

561

562

563 **References**

564 1. Meyhoff TS, et al. Restriction of Intravenous Fluid in ICU Patients with Septic Shock.  
565 *N Engl J Med.* 2022;386(26):2459–2470.

566 2. The National Heart, Lung, and Blood Institute Prevention and Early Treatment of  
567 Acute Lung Injury Clinical Trials Network. Early Restrictive or Liberal Fluid Management  
568 for Sepsis-Induced Hypotension. *N Engl J Med.* 2023;388(6):499–510.

569 3. Pickkers P, Kox M. Towards precision medicine for sepsis patients. *Crit Care.*  
570 2017;21(1):11, s13054-016-1583-z.

571 4. Uchimido R, Schmidt EP, Shapiro NI. The glycocalyx: a novel diagnostic and  
572 therapeutic target in sepsis. *Crit Care Lond Engl.* 2019;23(1):16.

573 5. Johansen M, et al. Profound Endothelial Damage Predicts Impending Organ Failure  
574 and Death in Sepsis. *Semin Thromb Hemost.* 2015;41(01):016–025.

575 6. Piotti A, et al. Endothelial damage in septic shock patients as evidenced by  
576 circulating syndecan-1, sphingosine-1-phosphate and soluble VE-cadherin: a substudy  
577 of ALBIOS. *Crit Care.* 2021;25(1):113.

578 7. Oshima K, et al. Endothelial Glycocalyx Degradation in Sepsis: Analysis of the  
579 Crystalloid Liberal Or Vasopressors Early Resuscitation in Sepsis (CLOVERS) Trial, a  
580 Multicenter, Phase 3, Randomized Trial. *Ann Am Thorac Soc.* [published online ahead  
581 of print: May 2, 2025]. <https://doi.org/10.1513/AnnalsATS.202501-012OC>.

582 8. Staprans I, Felts JM. Isolation and characterization of glycosaminoglycans in human  
583 plasma. *J Clin Invest.* 1985;76(5):1984–1991.

584 9. Schmidt EP, et al. Urinary Glycosaminoglycans Predict Outcomes in Septic Shock  
585 and Acute Respiratory Distress Syndrome. *Am J Respir Crit Care Med.*  
586 2016;194(4):439–449.

587 10. Weinbaum S, Tarbell JM, Damiano ER. The structure and function of the endothelial  
588 glycocalyx layer. *Annu Rev Biomed Eng.* 2007;9:121–167.

589 11. Sullivan RC, et al. Endothelial glycocalyx degradation during sepsis: Causes and  
590 consequences. *Matrix Biol Plus.* 2021;12:100094.

591 12. Ly M, et al. The proteoglycan bikunin has a defined sequence. *Nat Chem Biol.*  
592 2011;7(11):827–833.

593 13. Enghild JJ, et al. Chondroitin 4-sulfate covalently cross-links the chains of the  
594 human blood protein pre-alpha-inhibitor. *J Biol Chem.* 1991;266(2):747–751.

595 14. Anand D, et al. Evolution of serum hyaluronan and syndecan levels in prognosis of  
596 sepsis patients. *Clin Biochem.* 2016;49(10–11):768–776.

597 15. Molano Franco D, et al. Interleukin-6 for diagnosis of sepsis in critically ill adult  
598 patients. *Cochrane Database Syst Rev.* [published online ahead of print: July 24, 2015].  
599 <https://doi.org/10.1002/14651858.CD011811>.

600 16. Hippensteel JA, et al. Intravenous fluid resuscitation is associated with septic  
601 endothelial glycocalyx degradation. *Crit Care Lond Engl.* 2019;23(1):259.

602 17. Zhuo L, Salustri A, Kimata K. A physiological function of serum proteoglycan  
603 bikunin: the chondroitin sulfate moiety plays a central role. *Glycoconj J.* 2002;19(4–  
604 5):241–247.

605 18. Lord MS, et al. The Inter- $\alpha$ -Trypsin Inhibitor Family: Versatile Molecules in Biology  
606 and Pathology. *J Histochem Cytochem Off J Histochem Soc.* 2020;68(12):907–927.

607 19. Bost F, Diarra-Mehrpoor M, Martin JP. Inter-alpha-trypsin inhibitor proteoglycan  
608 family--a group of proteins binding and stabilizing the extracellular matrix. *Eur J  
609 Biochem.* 1998;252(3):339–346.

610 20. Balduyck M, et al. Inflammation-induced systemic proteolysis of inter-alpha-inhibitor  
611 in plasma from patients with sepsis. *J Lab Clin Med.* 2000;135(2):188–198.

612 21. Kasper R, et al. Major endothelial damage markers identified from hemadsorption  
613 filters derived from treated patients with septic shock - endoplasmic reticulum stress and  
614 bikunin may play a role. *Front Immunol.* 2024;15:1359097.

615 22. Capon C, et al. In acute inflammation, the chondroitin-4 sulphate carried by bikunin  
616 is not only longer, it is also undersulphated. *Biochimie.* 2003;85(1–2):101–107.

617 23. Lord MS, et al. Sulfation of the bikunin chondroitin sulfate chain determines heavy  
618 chain·hyaluronan complex formation. *J Biol Chem.* 2013;288(32):22930–22941.

619 24. Yang X, et al. A disintegrin and metalloproteinase 15-mediated glycocalyx shedding  
620 contributes to vascular leakage during inflammation. *Cardiovasc Res.*  
621 2018;114(13):1752–1763.

622 25. Ali MM, et al. Role of matrix metalloproteinases and histone deacetylase in oxidative  
623 stress-induced degradation of the endothelial glycocalyx. *Am J Physiol-Heart Circ*  
624 *Physiol.* 2019;316(3):H647–H663.

625 26. Han S, et al. Amelioration of sepsis by TIE2 activation-induced vascular protection.  
626 *Sci Transl Med.* 2016;8(335):335ra55.

627 27. Horvitz DG, Thompson DJ. A Generalization of Sampling Without Replacement from  
628 a Finite Universe. *J Am Stat Assoc.* 1952;47(260):663–685.

629 28. Deville J-C, Särndal C-E. Calibration Estimators in Survey Sampling. *J Am Stat*  
630 *Assoc.* 1992;87(418):376–382.

631 29. Deville J-C, Särndal C-E, Sautory O. Generalized Raking Procedures in Survey  
632 Sampling. *J Am Stat Assoc.* 1993;88(423):1013–1020.

633 30. White H. Maximum Likelihood Estimation of Misspecified Models. *Econometrica*.  
634 1982;50(1):1.

635 31. Lumley T. *Complex Surveys: A Guide to Analysis Using R*. Wiley; 2010.

636 32. R Core Team. *R: A Language and Environment for Statistical Computing*. Vienna,  
637 Austria: R Foundation for Statistical Computing; 2024.

638 33. Lumley T. Analysis of Complex Survey Samples. *J Stat Softw.* 2004;9(8).

639 <https://doi.org/10.18637/jss.v009.i08>.

640 34. Lumley T. *Survey: Analysis of Complex Survey Samples*. 2024.

641 35. Therneau TM. *A Package for Survival Analysis in R*. 2024.

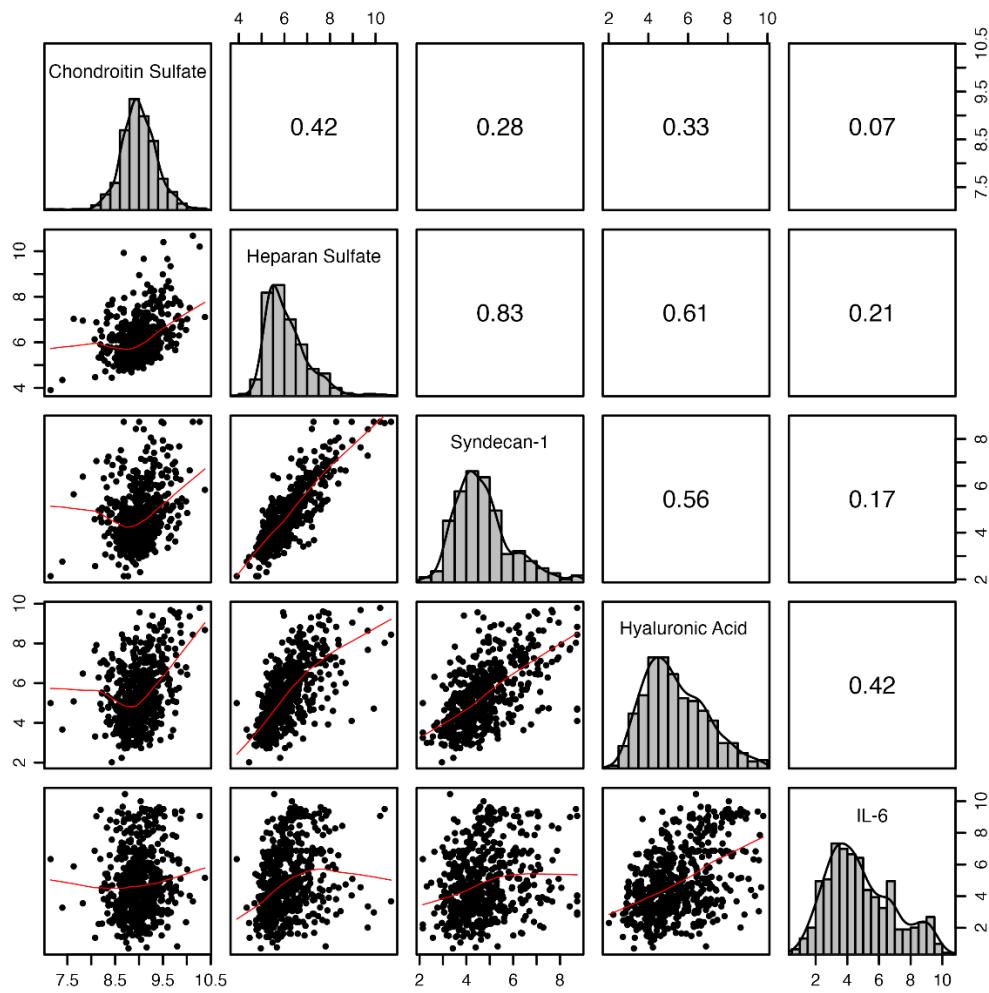
642 36. Therneau TM, Grambsch PM. *Modeling Survival Data: Extending the Cox Model*.

643 New York: Springer; 2000.

644 37. Harrell Jr Frank E. *Hmisc: Harrell Miscellaneous*. 2024.

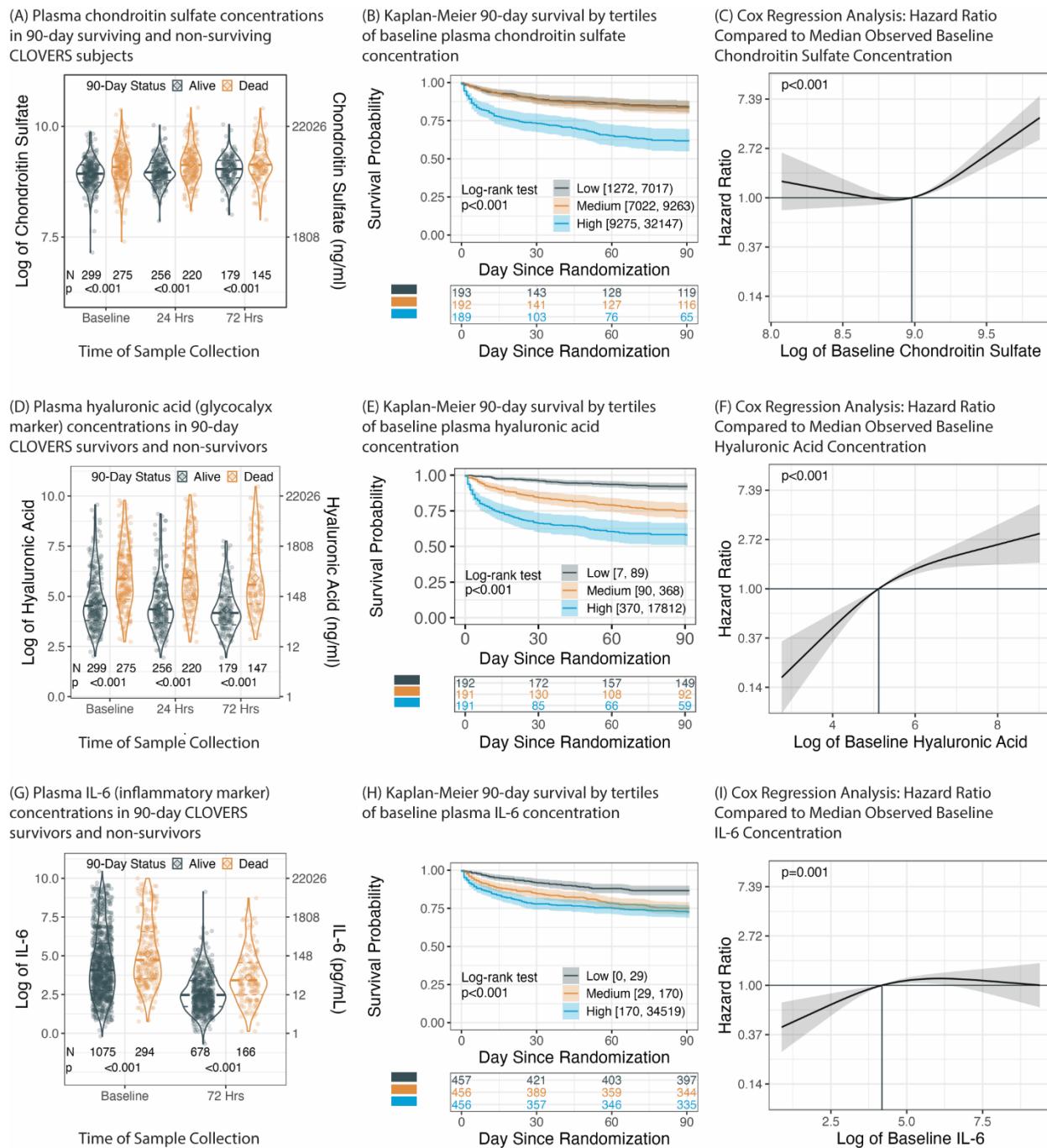
645 38. Pasek J. *Weights: Weighting and Weighted Statistics*. 2025.

646



647

648 **Figure 1. Correlation between chondroitin sulfate, indices of endothelial**  
 649 **glycocalyx degradation (hyaluronic acid, heparan sulfate, and syndecan-1), and**  
 650 **an index of systemic inflammation (IL-6) in plasma samples collected at**  
 651 **CLOVERS enrollment.** We display scatterplots of the observed joint distributions of the  
 652 log-transformed biomarker concentrations at baseline in the lower triangular paired  
 653 panels. The histogram and density curves of the log of biomarker concentrations are  
 654 shown on the diagonals. Weighted Pearson correlation coefficients are shown in the  
 655 corresponding upper triangular paired panels.



656

657 **Figure 2. Plasma biomarkers as predictors of 90-day mortality in CLOVERS. (A)**

658 Plasma **(A)** chondroitin sulfate, **(D)** hyaluronic acid, and **(G)** IL-6 (measured only at 72  
659 hours) were elevated at all timepoints in patients who died within 90 days of CLOVERS  
660 enrollment. The number of observations at each time point and p-values from the Wald

661 test of inverse probability weighted linear regression are shown under the violin plots.

662 **(B)** Kaplan-Meier plot of the unadjusted association between baseline plasma

663 chondroitin sulfate, divided into tertiles, and survival. Tertile ranges are listed in the

664 legend, with the table beneath the plot indicating the number of observed participants at

665 risk at each time. Kaplan-Meier estimates and the log rank test were calculated using

666 inverse probability of selection weights. **(C)** Covariate-adjusted association of baseline

667 (log-transformed) chondroitin sulfate with mortality rates within 90-days of

668 randomization. To address concerns about violations of the proportional hazards

669 assumption, this Cox model and all other Cox models are stratified by sex assigned at

670 birth and chronic heart failure. Additionally, we estimated associations between SOFA

671 score and log-hazard of death separately for 0–4 days, 5–11 days, and 12–90 days of

672 follow-up. Due to the stratified sampling study design, we used generalized raking when

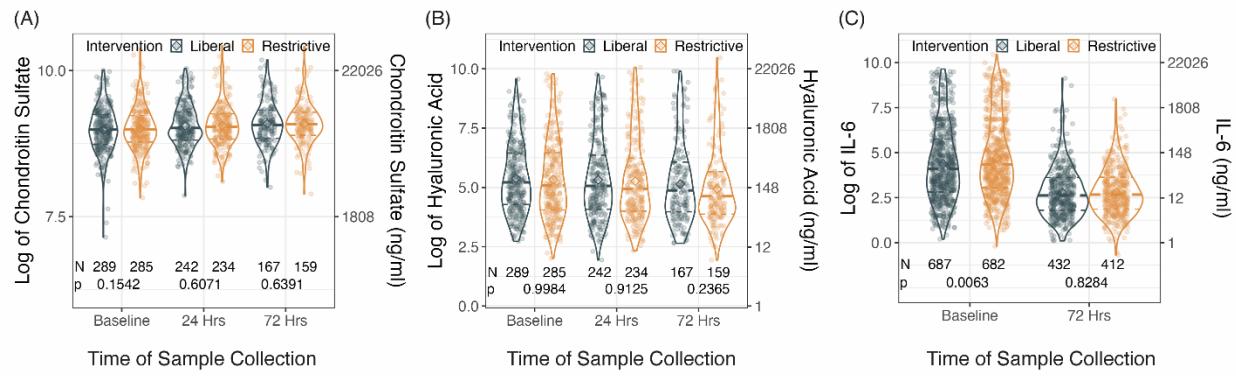
673 fitting the model. Identical analyses were conducted to estimate the association

674 between baseline plasma hyaluronic acid **(E, F)**, IL-6 **(H, I)**, and mortality rates within

675 90-days of randomization. Full covariate-adjusted models are demonstrated in

676 **Supplemental Figures 1, 2, and 3.**

677



678

Time of Sample Collection

679 **Figure 3. Plasma biomarker concentrations in CLOVERS patients randomized**

680 **(after day 0 sample collection) to 24 hours of protocolized liberal or restrictive**

681 **fluid resuscitation.** On unadjusted analyses, treatment approach had no impact on

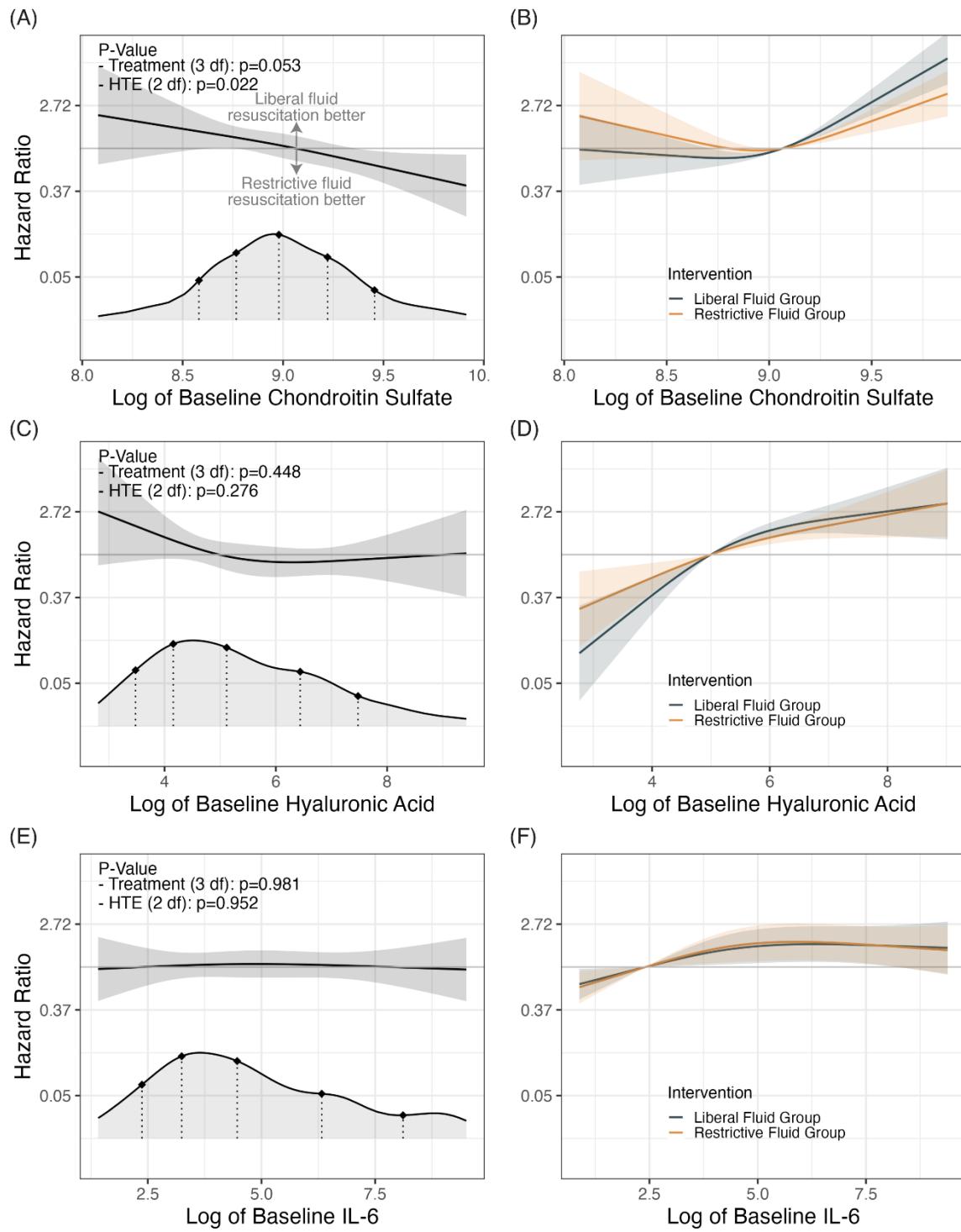
682 subsequent **(A)** chondroitin sulfate, **(B)** hyaluronic acid, or **(C)** IL-6 (measured only at 72

683 hours) plasma concentrations. The number of observations at each time point and p-

684 values from the Wald test of inverse probability weighted linear regression are shown

685 under the violin plots.

686



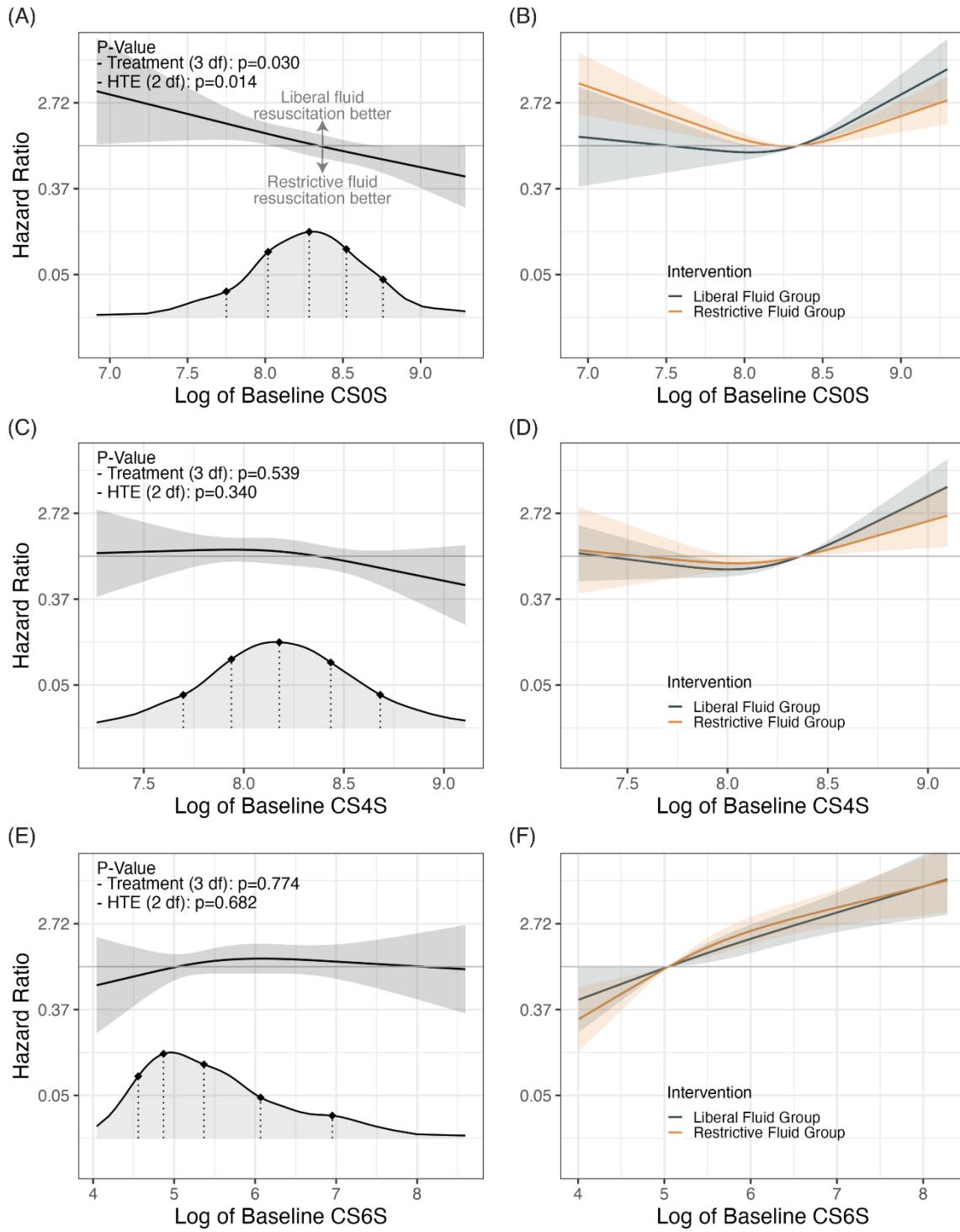
687

688 **Figure 4. The effectiveness of the randomized treatment assignment is modified**  
689 **by (log of) baseline chondroitin sulfate but not hyaluronic acid or IL-6 plasma**

690 **concentrations.** Panels display results from a model that added a randomized  
691 treatment assignment by baseline log plasma chondroitin sulfate (**A,B**), hyaluronic acid  
692 (**C,D**), or IL-6 (**E,F**) interaction to the models shown in Figures 2C, 2F, and 2I. Left-side  
693 panels demonstrate hazard ratios comparing restrictive versus liberal fluid resuscitation  
694 and pointwise 95% confidence interval across the distribution of log of baseline  
695 chondroitin sulfate (**A**), hyaluronic acid (**C**), or IL-6 (**E**) concentrations. The densities at  
696 the bottom of the figures refer to the distribution of baseline (log-transformed) plasma  
697 biomarker concentrations across the CLOVERS cohort, and for reference, we highlight  
698 the 10<sup>th</sup>, 25<sup>th</sup>, 50<sup>th</sup>, 75<sup>th</sup> and 90<sup>th</sup> percentiles. We show the p-values associated with (1)  
699 the overall treatment effect and (2) the heterogeneity of treatment effect. The  
700 heterogeneity of treatment effect test is a two degree of freedom test against the null  
701 that the treatment effect is constant across the distribution of the biomarker values. The  
702 overall treatment effect test includes the main effect of the intervention (randomized  
703 fluid resuscitation strategy) and the two terms for the interaction with biomarker values.  
704 It is a test against the null hypothesis that treatment has no effect on mortality rates  
705 within 90 days of randomization against the alternative that it has any effect. Right-side  
706 panels demonstrate estimated hazard ratio and pointwise 95% confidence interval  
707 comparing baseline log chondroitin sulfate (**B**), hyaluronic acid (**D**), or IL-6 (**F**) values to  
708 a reference value when the hazard ratio of the restrictive versus liberal fluid  
709 resuscitation strategies equal to one. The associations are displayed separately for  
710 those randomized to the restrictive and liberal fluid resuscitation strategies.

711

712



713

714 **Figure 5. The effectiveness of the randomized treatment assignment is modified**  
 715 **by (log of) baseline unsulfated chondroitin sulfate disaccharides, but not 4-O or**  
 716 **6-O sulfated disaccharides.** Left-side panels demonstrate hazard ratios comparing

717 restrictive versus liberal fluid resuscitation and pointwise 95% confidence interval across  
718 the distribution of log of baseline unsulfated (CS0S, **A**), 4-O sulfated (CS4S, **C**), or 6-O  
719 sulfated (CS6S, **E**) chondroitin sulfate concentrations, as per modeling detailed in  
720 Figure 4. The densities at the bottom of the figures refer to the distribution of baseline  
721 (log-transformed) plasma biomarker concentrations across the CLOVERS cohort, and  
722 for reference, we highlight the 10<sup>th</sup>, 25<sup>th</sup>, 50<sup>th</sup>, 75<sup>th</sup> and 90<sup>th</sup> percentiles. We show the p-  
723 values associated with (1) the overall treatment effect and (2) the heterogeneity of  
724 treatment effect. Right-side panels demonstrate estimated hazard ratio and pointwise  
725 95% confidence interval comparing baseline log unsulfated (**B**), 4-O sulfated (**D**), or 6-O  
726 sulfated (**F**) chondroitin sulfate values to a reference value when the hazard ratio of the  
727 restrictive versus liberal fluid resuscitation strategies equal to one.

728

729

730 **Table 1:** Baseline characteristics and sample weighting for chondroitin sulfate and  
 731 hyaluronic acid

	<b>Study Sample (N=574)</b>	<b>Overall (N=1563)</b>	<b>Reweighted Study Sample (N=1563)</b>
<b>Intervention</b>			
Liberal Fluid Group	289 (50.3%)	781 (50.0%)	786 (50.3%)
Restrictive Fluid Group	285 (49.7%)	782 (50.0%)	777 (49.7%)
<b>Age (years)</b>			
Mean (SD)	62.3 (15.0)	59.5 (15.8)	60.3 (15.5)
Median [IQR]	64.5 [53.0, 72.8]	61.0 [50.0, 71.0]	62.0 [50.0, 71.0]
<b>Sex</b>			
Female	252 (43.9%)	737 (47.2%)	725 (46.4%)
Male	322 (56.1%)	826 (52.8%)	838 (53.6%)
<b>Race</b>			
White	413 (72.0%)	1103 (70.6%)	1149 (73.5%)
Black	90 (15.7%)	246 (15.7%)	216 (13.8%)
Asian	21 (3.7%)	53 (3.4%)	55 (3.5%)
American Indian, Alaska Native, Native Hawaiian, or Pacific Islander	6 (1.0%)	16 (1.0%)	20 (1.3%)
Not Reported	44 (7.7%)	145 (9.3%)	123 (7.9%)
<b>Ethnicity</b>			
Hispanic or Latino	79 (13.8%)	226 (14.5%)	244 (15.6%)
Not Hispanic or Latino	473 (82.4%)	1274 (81.5%)	1261 (80.7%)
Not reported	22 (3.8%)	63 (4.0%)	58 (3.7%)
<b>End-stage Renal Disease</b>			
No	538 (93.7%)	1490 (95.3%)	1490 (95.3%)
Yes	36 (6.3%)	73 (4.7%)	73 (4.7%)
<b>Chronic Heart Failure</b>			
No	491 (85.5%)	1372 (87.8%)	1363 (87.2%)
Yes	83 (14.5%)	178 (11.4%)	200 (12.8%)
Missing	0 (0%)	13 (0.8%)	0 (0%)
<b>ARDS at Randomization</b>			
No	541 (94.3%)	1517 (97.1%)	1524 (97.5%)
Yes	33 (5.7%)	42 (2.7%)	39 (2.5%)

Missing	0 (0%)	4 (0.3%)	0 (0%)
<b>SOFA Score</b>			
Mean (SD)	4.11 (2.99)	3.44 (2.73)	3.46 (2.69)
Median [IQR]	4.00 [2.00, 6.00]	3.00 [1.00, 5.00]	3.00 [1.00, 5.00]
<b>Diabetes</b>			
No	399 (69.5%)	1104 (70.6%)	1091 (69.8%)
Yes	175 (30.5%)	446 (28.5%)	472 (30.2%)
Missing	0 (0%)	13 (0.8%)	0 (0%)
<b>Pre-Randomization Volume of Fluid Administered (mL)</b>			
Mean (SD)	1990 (630)	1970 (628)	1990 (618)
Median [IQR]	2080 [1500, 2400]	2050 [1450, 2450]	2080 [1500, 2400]
<b>Status at Day 90</b>			
Alive	299 (52.1%)	1222 (78.2%)	1221 (78.1%)
Dead	275 (47.9%)	341 (21.8%)	342 (21.9%)

732

733