Somatic mutations in *TBX3* promote hepatic clonal expansion by accelerating VLDL secretion

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SUPPLEMENTAL FIGURES

Supplemental Figure 1











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Supplemental Figure 1. Loss of *Tbx3* protects against WD induced MASLD.

- **A.** qPCR for fibrosis markers in *Tbx3* WT or KO mice fed WD for 3 months.
- **B.** Representative Sirius Red staining in mice from A.
- **C.** qPCR for fibrosis genes in *Tbx3* WT or KO mice fed WD for 6 months.
- **D.** Representative Sirius Red staining in mice from C.
- E. Plasma ALT in mice from C.
- **F.** Western blot for cleaved PARP in *Tbx3* KO or WT mice fed WD for 3 (left) and 6 (right) months.
- **G.** Liver weight (left), body weight (middle) and liver:body weight ratio from female *Tbx3* KO or WT mice fed WD for 3 months.
- **H.** Representative H&E image from mice in G.



Supplemental Figure 2. Loss of *Tbx3* protects against MASLD induced liver tumors.

- **A.** Liver weight (left), body weight (middle), and LW:BW ratio (right) of *Tbx3* WT or KO mice fed a WD for 48 weeks.
- **B.** Surface tumors from mice in A.
- **C.** Example of large surface tumor from mice in A.



Supplemental Figure 3. MASLD exerts negative selective pressures on *Tbx3* expressing hepatocytes.

- A. *Tbx3* expression in TBX3-V5 or GFP-V5 injected mice after 3 months of WD feeding.
- B. *Tbx3* expression in Tbx3-V5 or GFP-V5 injected mice after 3 months of NC feeding.



Supplemental Figure 4. *Tbx3* deletion is protective against MASLD associated with aging.

- **A.** Liver weight (left), body weight (middle), and liver:body weight ratio (right) of *Tbx3* WT or KO mice fed NC diet for 6 months.
- **B.** Representative H&E image from mice in A.
- **C.** Plasma ALT from mice in A.



Supplemental Figure 5. *Tbx3* does not alter de novo lipogenesis or free fatty acid uptake.

- A. LW:BW ratio of *Tbx3* KO or WT mice fed a WD for 4 weeks.
- **B.** Representative H&E images from mice in A.
- **C.** Fractional enrichment of M+16 palmitate in the plasma of *Tbx3* KO or WT mice infused with ¹³C potassium palmitate after 2 weeks of WD feeding.
- **D.** Fractional enrichment of M+16 palmitate in the liver from mice in C.



Supplemental Figure 6. Loss of Tbx3 protects from MASLD by increasing VLDL secretion.

- A. Plasma ALT in *Tbx3* KO or WT mice fed a CDAHFD for 3 months.
- B. Plasma AST from mice in A.
- C. qPCR for fibrosis marker genes from mice in A.
- D. Representative sirius red staining from mice in A.
- E. Liver triglyceride measurements from mice in A.
- F. Volcano plot of MOSAICS screen (11) showing depletion of *Mttp* and *Tm6sf2* KO clones in WD vs. NC fed mice.
- G. Relative secretion of Gaussia luciferase after TBX3 knockdown in HEK293T cells.
- H. qPCR of cholesterol biosynthesis genes in Tbx3 KO or WT mice 1 week after AAV injection.
- I. Liver triglyceride measurements from mice overexpressing GFP-V5, WT TBX3-V5, TBX3-V5 containing a point mutation after 12 weeks of WD feeding.

Relative secretion in G was calculated using a One-way ANOVA corrected for multiple

comparisons.

Bibliography

1. Li R, et al. A body map of somatic mutagenesis in morphologically normal human tissues. *Nature*. 2021;597(7876):398–403.

2. Yizhak K, et al. RNA sequence analysis reveals macroscopic somatic clonal expansion across normal tissues. *Science*. 2019;364(6444). https://doi.org/10.1126/science.aaw0726.

Kakiuchi N, Ogawa S. Clonal expansion in non-cancer tissues. *Nat Rev Cancer*.
2021;21(4):239–256.

4. Martincorena I, et al. Somatic mutant clones colonize the human esophagus with age. *Science*. 2018;362(6417):911–917.

5. Yoshida K, et al. Tobacco smoking and somatic mutations in human bronchial epithelium. *Nature*. 2020;578(7794):266–272.

Olafsson S, et al. Somatic Evolution in Non-neoplastic IBD-Affected Colon. *Cell*.
2020;182(3):672-684.e11.

7. Kakiuchi N, et al. Frequent mutations that converge on the NFKBIZ pathway in ulcerative colitis. *Nature*. 2020;577(7789):260–265.

8. Nanki K, et al. Somatic inflammatory gene mutations in human ulcerative colitis epithelium. *Nature*. 2020;577(7789):254–259.

9. Brunner SF, et al. Somatic mutations and clonal dynamics in healthy and cirrhotic human liver. *Nature*. 2019;574(7779):538–542.

10. Ng SWK, et al. Convergent somatic mutations in metabolism genes in chronic liver disease. *Nature*. 2021;598(7881):473–478.

11. Wang Z, et al. Positive selection of somatically mutated clones identifies adaptive pathways

in metabolic liver disease. Cell. 2023;186(9):1968-1984.e20.

12. Khan SF, et al. The roles and regulation of TBX3 in development and disease. *Gene*. 2020;726:144223.

13. Davenport TG, et al. Mammary gland, limb and yolk sac defects in mice lacking Tbx3, the gene mutated in human ulnar mammary syndrome. *Development*. 2003;130(10):2263–2273.

14. Frank DU, et al. Lethal arrhythmias in Tbx3-deficient mice reveal extreme dosage sensitivity of cardiac conduction system function and homeostasis. *Proc Natl Acad Sci USA*. 2012;109(3):E154-63.

15. Bamshad M, et al. The spectrum of mutations in TBX3: Genotype/Phenotype relationship in ulnar-mammary syndrome. *Am J Hum Genet*. 1999;64(6):1550–1562.

16. Jin Y, et al. Wnt signaling regulates hepatocyte cell division by a transcriptional repressor cascade. *Proc Natl Acad Sci USA*. 2022;119(30):e2203849119.

17. Coll M, et al. Structure of the DNA-bound T-box domain of human TBX3, a transcription factor responsible for ulnar-mammary syndrome. *Structure*. 2002;10(3):343–356.

18. Zhou Y, et al. DDMut: predicting effects of mutations on protein stability using deep learning. *Nucleic Acids Res.* 2023;51(W1):W122–W128.

19. Liang W, et al. Establishment of a general NAFLD scoring system for rodent models and comparison to human liver pathology. *PLoS ONE*. 2014;9(12):e115922.

20. Nogueira JP, Cusi K. Role of insulin resistance in the development of nonalcoholic fatty liver disease in people with type 2 diabetes: from bench to patient care. *Diabetes Spectr*. 2024;37(1):20–28.

21. Brown MS, Goldstein JL. Selective versus total insulin resistance: a pathogenic paradox. *Cell Metab.* 2008;7(2):95–96.

22. Ipsen DH, et al. Molecular mechanisms of hepatic lipid accumulation in non-alcoholic fatty liver disease. *Cell Mol Life Sci*. 2018;75(18):3313–3327.

23. Cole LK, et al. Phosphatidylcholine biosynthesis and lipoprotein metabolism. *Biochim Biophys Acta*. 2012;1821(5):754–761.

24. Wang X, et al. Metabolic inflexibility promotes mitochondrial health during liver regeneration. *Science*. 2024;384(6701):eadj4301.

25. Matsumoto M, et al. An improved mouse model that rapidly develops fibrosis in nonalcoholic steatohepatitis. *Int J Exp Pathol*. 2013;94(2):93–103.

26. Badr CE, et al. A highly sensitive assay for monitoring the secretory pathway and ER stress. *PLoS ONE*. 2007;2(6):e571.

27. Zinnall U, et al. HDLBP binds ER-targeted mRNAs by multivalent interactions to promote protein synthesis of transmembrane and secreted proteins. *Nat Commun*. 2022;13(1):2727.

28. Mobin MB, et al. The RNA-binding protein vigilin regulates VLDL secretion through modulation of Apob mRNA translation. *Nat Commun*. 2016;7:12848.

29. Abby E, et al. Notch1 mutations drive clonal expansion in normal esophageal epithelium but impair tumor growth. *Nat Genet*. 2023;55(2):232–245.

30. Colom B, et al. Mutant clones in normal epithelium outcompete and eliminate emerging tumours. *Nature*. 2021;598(7881):510–514.

31. Zhu M, et al. Somatic mutations increase hepatic clonal fitness and regeneration in chronic liver disease. *Cell*. 2019;177(3):608-621.e12.

32. Zhu M, et al. PKD1 mutant clones within cirrhotic livers inhibit steatohepatitis without promoting cancer. *Cell Metab*. 2024;36(8):1711-1725.e8.

33. Smagris E, et al. Inactivation of tm6sf2, a gene defective in fatty liver disease, impairs

lipidation but not secretion of very low density lipoproteins. *J Biol Chem*. 2016;291(20):10659–10676.

34. Roux C, et al. Role of cholesterol in embryonic development. *Am J Clin Nutr*. 2000;71(5 Suppl):1270S–9S.

35. Porter FD, Herman GE. Malformation syndromes caused by disorders of cholesterol synthesis. *J Lipid Res*. 2011;52(1):6–34.

36. Baardman ME, et al. The role of maternal-fetal cholesterol transport in early fetal life: current insights. *Biol Reprod*. 2013;88(1):24.

37. Samuel VT, et al. Lipid-induced insulin resistance: unravelling the mechanism. *Lancet*. 2010;375(9733):2267–2277.

38. Sniderman AD, et al. Apolipoprotein B particles and cardiovascular disease: A narrative review. *JAMA Cardiol*. 2019;4(12):1287–1295.

39. Wang Z, et al. Dual ARID1A/ARID1B loss leads to rapid carcinogenesis and disruptive redistribution of BAF complexes. *Nat Cancer*. 2020;1(9):909–922.