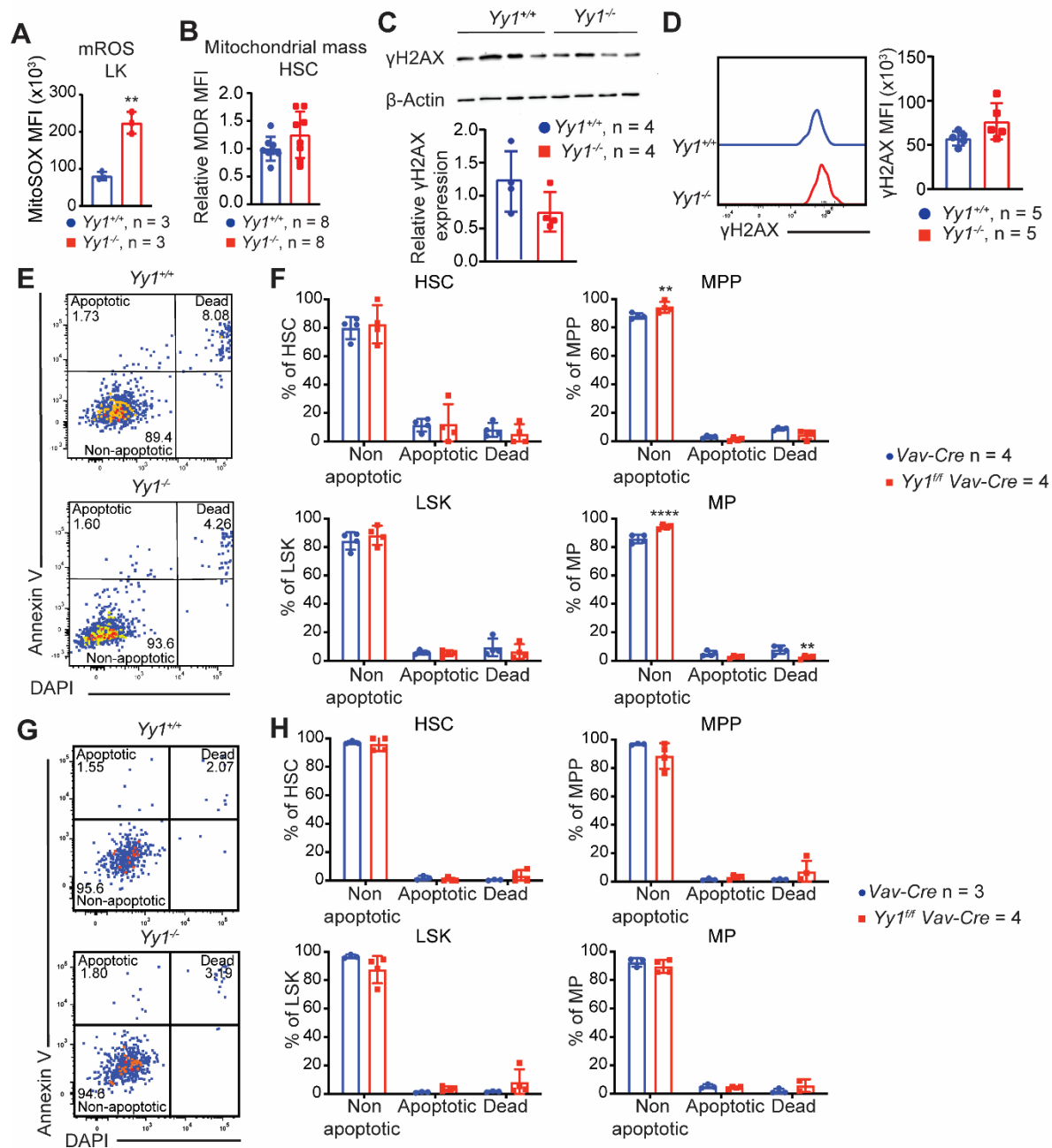
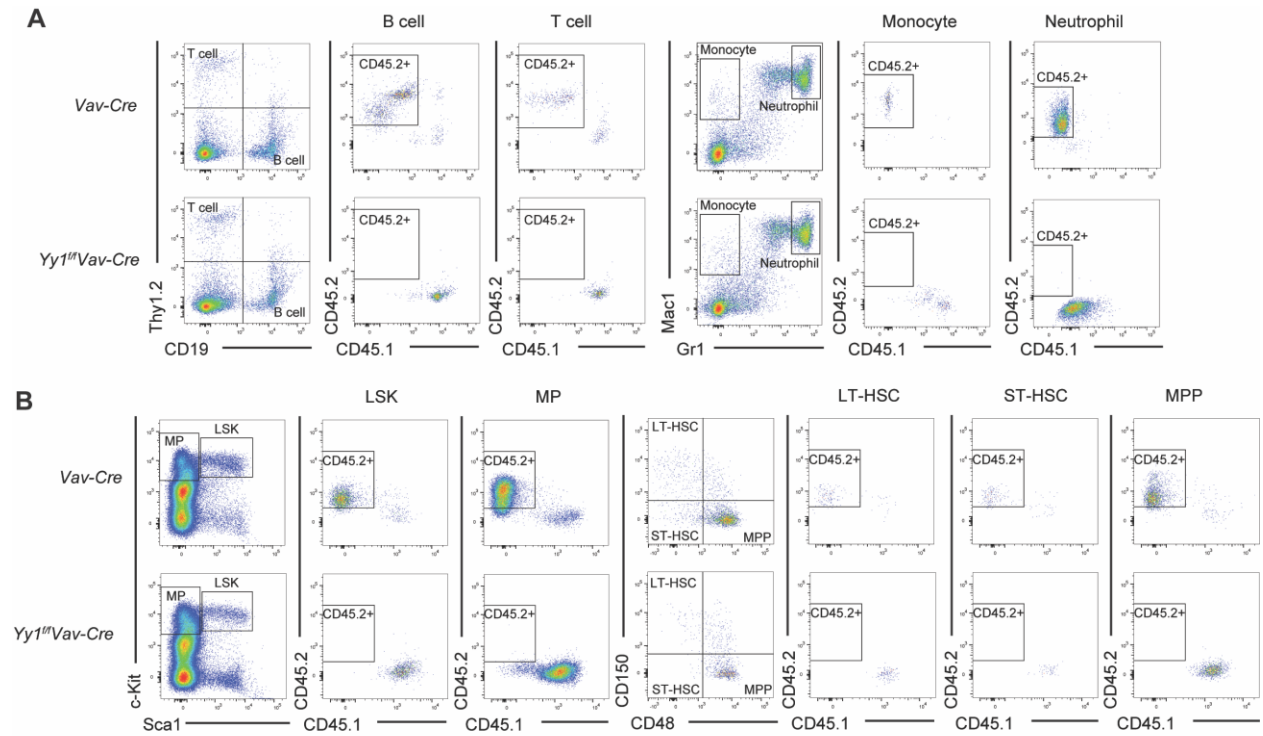


## Supplementary data

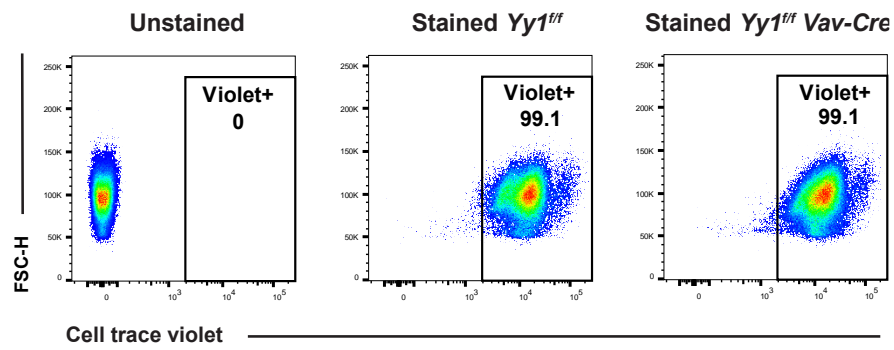


**Supplemental Figure 1.** Flow cytometry evaluation of (A) Mitochondrial ROS levels and (B) Mitochondrial mass in WT and YY1 KO HSCs. (C) WB and quantification of  $\gamma$ H2AX in Lin<sup>-</sup> FL cells. (D) Flowcytometry evaluation of  $\gamma$ H2AX expression in WT and YY1 KO HSCs. (E-H) In vivo and ex vivo survival assay of fetal HSPCs. (E) Representative flow gating strategy for non-

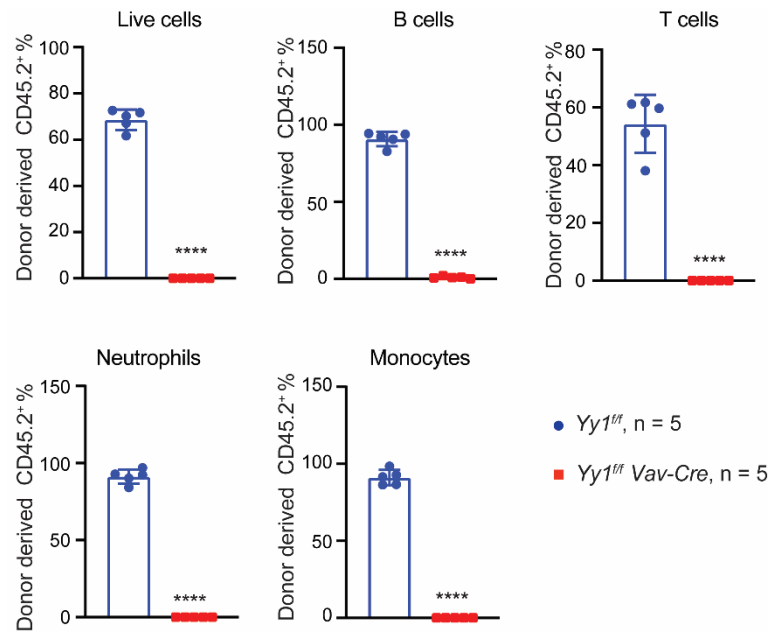
apoptotic (Annexin V<sup>-</sup>, DAPI<sup>-</sup>), apoptotic (Annexin V<sup>+</sup>, DAPI<sup>-</sup>) and dead cells (Annexin V<sup>+</sup>, DAPI<sup>+</sup>) in the freshly isolated FL HSPC cells. **(F)** Quantification of percentage of non-apoptotic, apoptotic, and dead cells in FL HSC, MPP, LSK, and MP cells. **(G)** Representative flow gating strategy and **(H)** quantification of non-apoptotic, apoptotic, and dead FL HSPCs cultured with serum free media overnight. N represents the number of fetuses; graphs show means  $\pm$  SD, \*\*P < 0.01, \*\*\*\*P < 0.0001 by unpaired t test (**A-D**), 2-way ANOVA (**F** and **H**).



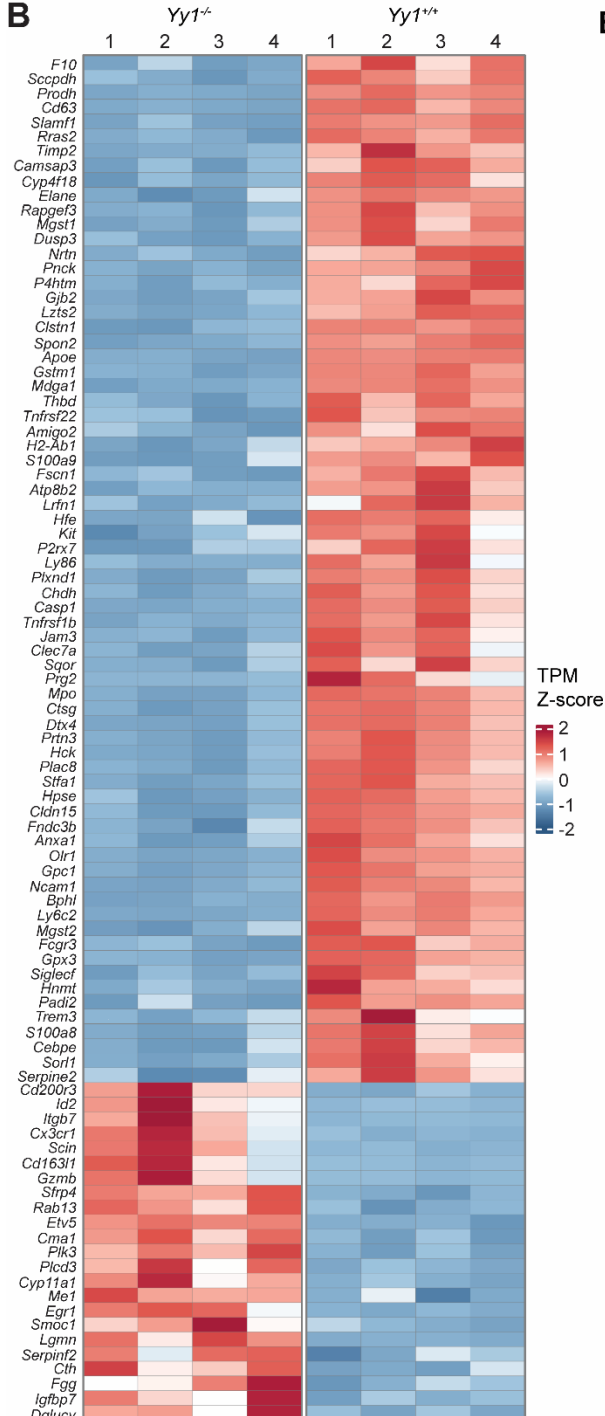
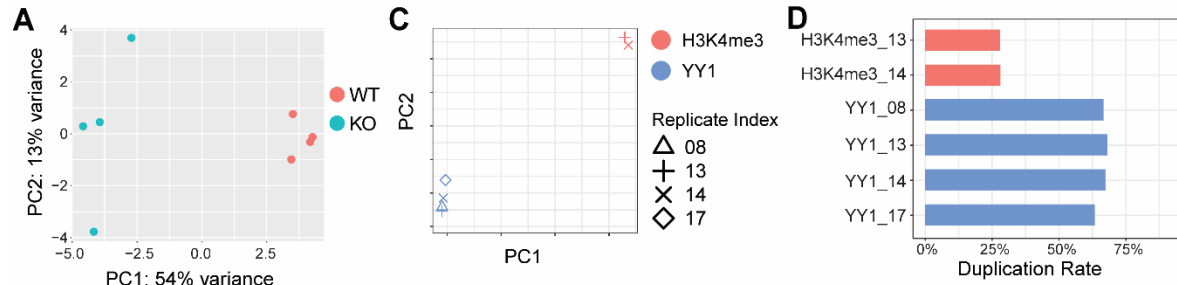
**Supplemental Figure 2. (A)** Representative flow gating strategy for donor derived BM B cells ( $CD45.2^{+}Thy1.2^{-}CD19^{+}$ ), T cells ( $CD45.2^{+}Thy1.2^{+}CD19^{-}$ ), neutrophils ( $CD45.2^{+}Mac1^{+}Gr1^{hi}$ ) and monocytes ( $CD45.2^{+}Mac1^{+}Gr1^{-}$ ) of FL transplant. **(B)** Representative flow gating strategy for donor derived BM LT-HSC ( $CD45.2^{+}Lin^{-}Sca1^{+}c-Kit^{+}CD48^{-}CD150^{+}$ ), ST-HSC ( $CD45.2^{+}Lin^{-}Sca1^{+}c-Kit^{+}CD48^{-}CD150^{-}$ ), MPP ( $CD45.2^{+}Lin^{-}Sca1^{+}c-Kit^{+}CD48^{+}CD150^{-}$ ), LSK ( $CD45.2^{+}Lin^{-}Sca1^{+}c-Kit^{+}$ ) and MP ( $CD45.2^{+}Lin^{-}Sca1^{-}c-Kit^{+}$ ) populations.



**Supplemental Figure 3.** CellTrace Violet dye labeled over 99% cells. Representative flow gating strategy for percentage of CellTrace Violet<sup>+</sup> *Yy1<sup>ff</sup>* and *Yy1<sup>ff</sup> Vav-Cre* Lin<sup>-</sup> fetal liver cells after staining.



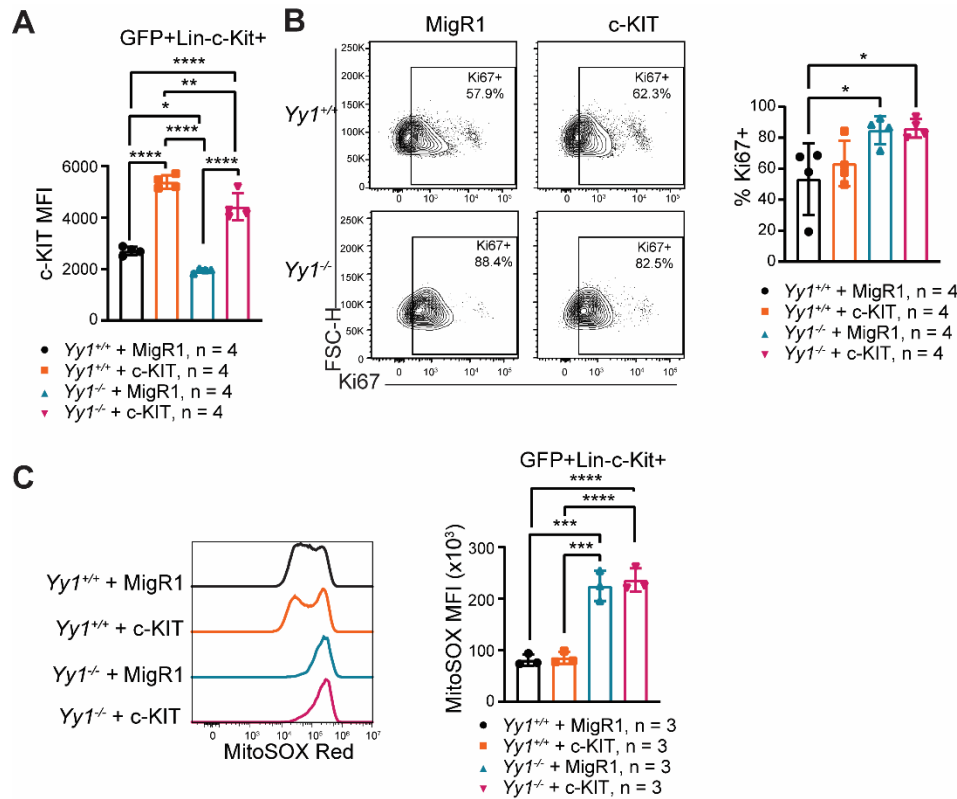
**Supplemental Figure 4.** Donor derived percentage of total live cells (CD45.2<sup>+</sup>), B cells (CD45.2<sup>+</sup>Thy1.2<sup>-</sup>CD19<sup>+</sup>), T cells (CD45.2<sup>+</sup>Thy1.2<sup>+</sup>CD19<sup>-</sup>), neutrophils (CD45.2<sup>+</sup>Mac1<sup>+</sup>Gr1<sup>hi</sup>) and monocytes (CD45.2<sup>+</sup> Mac1<sup>+</sup>Gr1<sup>-</sup>) in bone marrow of recipient mice that received intraosseous transplant of wild-type or YY1 deficient FL cells. N represents the number of mice; graphs show means  $\pm$  SD, \*\*\*\*P<0.0001 by unpaired t test.



**E**

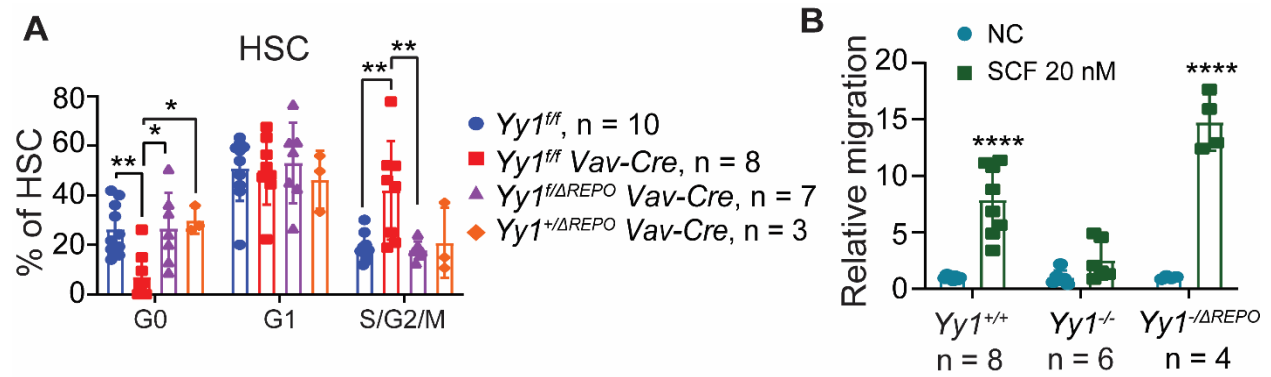
Rank	TF	Adjusted P	Motif Sequence
1	KLF3	0	GGGVDGGCGGGGCGYGGG
2	E2F4	0	RGGGCGGGGMMN
3	SP3	0	SGVVGCGGGCGGGCGRGS
4	SP2	0	GGSSVGGGGCGGGCCDGS
5	KLF1	2.4E-303	DGGVYKGGCYGG
6	E2F1	3.8E-272	GRGGCGGGRMV
7	KLF5	2.1E-269	RGGSWGGHGGGG
8	SP4	1.5E-259	SRGVARGGGGGRGCHDR
9	KLF4	3.7E-228	KRRVWGGGTGKGC
10	KLF6	2.9E-219	GSRRGGHGGGGMGGGRV
11	FOX11	2.7E-188	YTBGTATTGGY
12	SP1	1.1E-184	GVVGGGGVGGGGGMMGGGCGSGG
13	NRF1	3.1E-174	CWSTGCGCATGCGVRS
14	EGF1	1.4E-158	GKCGKGGGGGGG
15	NFYB	3.1E-154	YRRCATAGMR
16	KLF15	9.4E-147	RGCGMGRGVGGGGGRG
17	ZBT17	1.7E-140	GSGGGGGGGAGGGMG
18	NFYC	4.3E-140	BYARCCATCAGAR
19	EGF2	6.8E-119	KHGTGGHGGGG
20	E2F2	1.5E-109	GGCGCGAAG
21	ZFX	3.5E-109	GSCBVGGCGTSSSSCS
22	E2F3	1.3E-98	GGCGGGGARS
23	CTCF	4.4E-97	GCCRSCAGGGGCGCYVB
24	NFYA	1.4E-96	BYRCCATCAGAR
25	WT1	1.3E-94	RGVGGGGAGGGGGVGGRG
26	E2F7	4.4E-93	GDGGCGGAARDR
27	SP5	1.6E-86	RRGGRRGGRRGGGGGAGGRG
28	MBD2	4.0E-77	SSGCGGGMR
29	FLI1	9.8E-74	GGVCCGGAAGYGS
30	E2F6	3.8E-71	NKGGCGGARRG
31	MECP2	6.0E-71	SCCGGR
32	GABPA	1.2E-69	GGVRCGGAAGTV
33	ELF1	1.4E-69	SRACCGGAAGTGS
34	CTCF	2.5E-62	YGGCCASCAGRGGGCGCYVB
35	SALL4	2.2E-60	GSDGGGWWGG
36	YY1	2.1E-59	CAAVATGGCGG
37	ZN281	1.9E-54	GGGGTGGGGAGGGG
38	AP2B	1.3E-49	GCCYGVGGGS
39	PBX3	1.7E-48	YBYTGATTGRYTGR
40	ELK1	1.4E-47	RCCGGAAGTV
41	ARNT	3.6E-46	BYRCGTGC
42	AHR	6.0E-46	DTYCGTGM
43	ELK4	3.0E-43	SRRCGGAAGYRG
44	FEV	2.8E-41	GCVGGAGY
45	TFEB	2.9E-38	RGTACGTG
46	MAZ	5.0E-37	GGGGGAGGGGGDRRRRRRG
47	ATF1	3.3E-36	VTGACGTAS
48	ZFP57	2.2E-34	CTGCGGCARR
49	AP2C	3.0E-34	YRSCCYAGGSYN
50	MYC	3.3E-34	MGCCACGTGSBC
51	ETS1	5.0E-34	VRRRCMGAAGTGG
52	KAISO	4.5E-32	SARRYCTCGGAGAV
53	TAF1	4.5E-31	VAAAWGGCGMG
54	MAX	2.4E-30	SVSCAGTGGC
55	AP2A	6.1E-30	VTRSCGTGRGC
56	PKNX1	8.6E-30	TGABTGRGAS
57	USF2	9.3E-27	GGGTACGTGRBSVSSSB
58	RFX1	5.2E-26	SSSCBTTGCCAKGVRACSS
59	PBX1	7.3E-26	TGAKTGACAGS
60	KLF8	1.7E-24	CAGGGKGTG
61	ZIC1	7.0E-23	KGGWGGTS
62	ERG	5.9E-22	VVRCMGGAAGYRVV
63	SRBP2	6.3E-20	VSGTGGGGWABG
64	SRBP1	1.9E-18	GGTGGGGTGAB
65	USF1	3.7E-16	SRGTACGTGRCS
66	MYCN	4.5E-13	SCACGTGG
67	E2F5	1.5E-12	SGCGCSAAH
68	MXI1	1.8E-12	VCACGTGGSNGSBS
69	MTF1	2.2E-12	MKGCCGTGTGCAARS
70	HINFP	4.0E-12	DMSHHMCGGACGTTV
71	HEN1	8.8E-12	BGGKMKACGTGCKCCYY
72	INSM1	8.0E-11	TGTMAAGGGGCR
73	PAX5	6.8E-10	VDRVKAGYSRAGCGTGAC
74	ZFP42	1.2E-09	GGCMGCCATTTT
75	ELF2	3.4E-09	TDCAGGAAGTRRV
76	OVOL2	9.4E-09	RTAACGGD
77	EPAS1	1.8E-07	VTACGTGMC
78	NR1H4	3.8E-07	RRGGCAVTGRCCBSGVGG
79	HIF1A	4.2E-07	GDACGTGM
80	ZBT7A	3.4E-06	VRGGGKCKY
81	ZIC3	7.3E-06	GGCYHCCTGTGWGH
82	ZN143	1.2E-05	KGCMTKGTGGGARWGTAGTYY
83	GATA1	6.2E-05	TGSNSSNDSAGATAAGRV
84	TFE3	1.1E-04	RGTACGTGA
85	ZN322	9.1E-04	GAGCCTGSTACWSGCCGTR
86	ZIC2	1.1E-03	DGCVHCCTGTGWGH
87	COE1	3.4E-03	RGYCCWGGGGAS
88	RFX3	3.6E-03	CSGTTGCCATGGMRC
89	MITF	5.3E-03	YCWYGTGACY
90	TAL1	6.4E-03	CTGSNNNNRSAGATAAGRV
91	PRDM5	1.1E-02	VMTGGAGDCMRGGW
92	EHF	1.2E-02	VWACSGAGGAGTDS
93	ETV2	1.2E-02	RRARRCAGGAARYGS
94	MYBA	1.8E-02	VGYGGCVGTTTSVGV

**Supplemental Figure 5.** (A) PCA plot of RNA-seq from sorted *Yy1*<sup>+/+</sup> and *Yy1*<sup>-/-</sup> Lin<sup>-</sup> cKit<sup>+</sup> E14.5 FL cells. (B) Heatmap depicting selected upregulated and down-regulated genes based on TPM. (C) PCA plot of CUT&Tag. (D) Duplication rate of each CUT&Tag sample. (E) Motif analysis identified 94 significantly enriched (adjusted P < 0.05) transcription factor motifs, ranked based on adjusted P. TF motifs known to play essential roles in hematopoiesis, chromatin structure regulation, cell stress response, and cellular proliferation are highlighted.

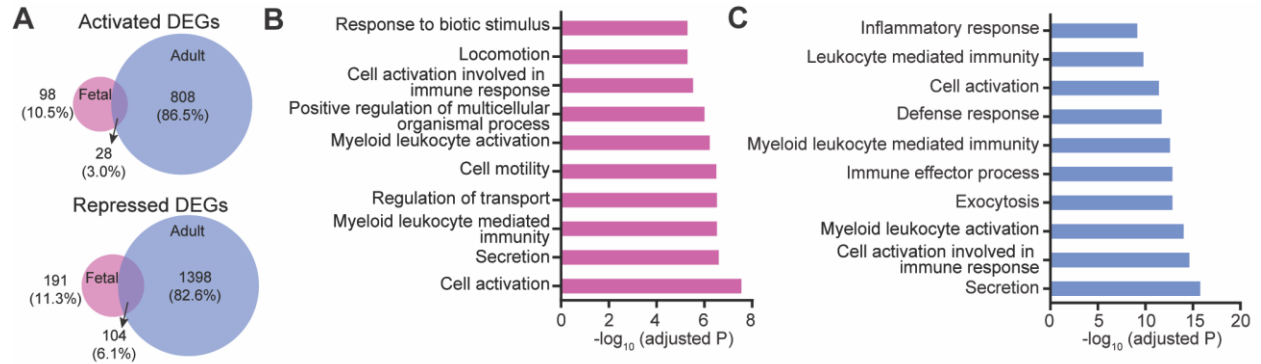


**Supplemental Figure 6. Ectopic expression of c-Kit did not rescue YY1-dependent cell cycle or mitochondrial ROS in FL LK cells. (A)** Ectopic expression of c-Kit rescued c-Kit MFI in YY1 deficient FL LK cells. **(B)** Representative gating strategy for Ki67 proliferation assay. E14.5 Lin<sup>-</sup> FL cells were infected with either MigR1 (control) or MigR1-c-Kit vectors. The percentages of Ki67<sup>+</sup> and Ki67<sup>-</sup> cells were quantified within GFP<sup>+</sup>Lin<sup>-</sup>c-Kit<sup>+</sup> populations. **(C)** Representative flow cytometry plots and quantification of mitochondrial ROS levels in GFP<sup>+</sup>Lin<sup>-</sup>c-Kit<sup>+</sup> cells infected with either MigR1-c-Kit or MigR1 vector control. N represents the number of fetuses; graphs show means  $\pm$  SD, \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001, \*\*\*\*P < 0.0001 by one way ANOVA.





**Supplemental Figure 7. The YY1 REPO domain deletion did not impact HSC proliferation and SCF-dependent cell migration.** (A) Quantification of percentage of fetal HSC in G0 (Ki67<sup>-</sup>DAPI<sup>-</sup>), G1 (Ki67<sup>+</sup>DAPI<sup>-</sup>) and S/G2/M (Ki67<sup>+</sup>DAPI<sup>+</sup>) phases. (B) Ex vivo migration assay of *Yy1<sup>-/-</sup>ΔREPO*, *Yy1<sup>-/-</sup>* and *Yy1<sup>+/+</sup>* Lin<sup>-</sup> FL cells in response to SCF stimulation. While YY1-deficient cells exhibit significantly impaired migration upon SCF stimulation, both wild type and *Yy1<sup>-/-</sup>ΔREPO* cells are responsive to SCF stimulation. N represents the number of fetuses; graphs show means ± SD, \*P < 0.05, \*\*P < 0.01, \*\*\*\*P < 0.0001 by 2-way ANOVA.



**Supplemental Figure 8.** (A) Venn diagram of activated and repressed DEGs associated with YY1 deletion in fetal versus adult HSPCs. (B-C) GSEA analysis of enriched GO pathways in fetal (B) and adult HSPCs (C).

**Table 1****qRT-PCR Primer list**

	Forward	Reverse
<i>Yy1</i>	TCAGACCCTAAGCAACTGGCAGAA	TTGAGCTCTCAACGAACGCTTTGC
<i>Kit</i>	AGTGGACGTACAGGTCCAGAA	GCCTGGATTTGCTCTTTGTTGT
<i>S100a8</i>	ATCACCATGCCCTCTACAAGAATG	GTCCAATTCTCTGAACAAGTTTTC
<i>S100a9</i>	AAGCTGCATGAGAACAACCCA	CCCAGAACAAAGGCCATTGA
<i>Gpc1</i>	GAATTTGGCCAACCACAGCC	GTAGTAGAGGCGCAGCTCAG
<i>Hck</i>	GCCAACCTGATGAAGTCGCT	TGTACTCATTGTCCTCGATGATTCTG
<i>Csf1r</i>	CAGTTCAGAGTGATGTGTGGTC	CTTGTTGTTCACTAGGATGCCG
<i>Padi2</i>	CAGCCGCCTATACGGGAAA	CTTCACACCTTCCGAGTGCT
<i>Jam3</i>	TACCGCAATGATGTGCCACT	GATGAAGCAGCCTCGTCTGT
<i>Cd63</i>	CTGTGGGCTGTGGGAATGAT	CCTCCACAAAAGCAATGCCC
<i>Cxcr2</i>	CATCCCGTTTGAGGGTCGTA	GCAGGAAGACAAGGACGACA
<i>Mpo</i>	CCTCCAACCGTGCCTTTGTA	GTCAGCTGATCGTTGGGGAA
<i>Olr1</i>	CTGCGAATGACGAGCCTGA	CCAGAGTCATAGCAGCAGGG
<i>Pnck</i>	ATCGCGGTACTTCGCAGAAT	CCAAGGACCTGCCCTACAAG
<i>Cxcr4</i>	GACTGGCATAGTCGGCAATG	AGAAGGGGAGTGTGATGACAAA
<i>Gapdh</i>	TCCTGCACCACCAACTGCTT	GTCTTCTGGGTGGCAGTGAT

**Table 2****Genotyping Primer list**

	Forward	Reverse
<i>Yy1</i> P1, P2	ACCTGGTCTATCGAAAGGAAGCAC	GCTTCGCCTATTCTCGCTCATAA
<i>Yy1</i> P3, P4	TAGAGAATAGGAACTTCGGCCGCCA	CCAAAGTTCGAAACCTGCTTTCCT
<i>Yy1<sup>AREPO</sup></i>	GACGGCTTCGAGGACCAGAT	GCGCCCATCACACACATAAAA