

Supplementary Fig. 1. (A and B) Representative PAS and H&E staining of the lung tissue from Sal, Alt, Nuc and Alt+Nuc treated B6 mice as per Fig. 2A. (C) AHR following Saline, Nuc, Aspergillus (Asp) or Asp+Nuc exposure in B6 mice as measured by flexiVent. \*(Sal vs Nuc) P=0.01;  $^{\circ}$ (Sal vs Asp) P<0.0001;  $^{\varepsilon}$ (Sal vs Asp+Nuc) P<0.0001; #(Nuc vs Asp) P<0.0001;  $^{\delta}$ (Nuc vs Asp+Nuc) P<0.0001;  $^{\psi}$ (Asp vs Asp+Nuc) P=0.002. (n=5; 2-way ANOVA, Tukey's multiple comparisons test).

(**D-G**) Differential counts of BAL macrophages, lymphocytes, eosinophils and neutrophils. (**H and I**) Morphometric quantification of lung inflammation and mucus production following H&E and PAS staining of the lung tissue (groups were color-coded as in C). (n=5; 2-way ANOVA, Tukey's multiple comparisons test). Data are presented as mean+/-SEM, \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 \*\*\*\*P<0.0001, ns=not significant.



**Supplementary Fig. 2**. Top Nuc up- and down-regulated genes in the lung from Nuc-treated mice and comparison with Sal, Alt and Alt-Nuc treated mice. (**A**) Genes that were selectively upregulated by Nuc but inhibited by Alt and Alt-Nuc. (**B**) Genes upregulated by all three treatment protocols--Nuc, Alt and Alt-Nuc. (**C**) Genes down-regulated by Nuc. (**D**) Top GO biological processes in Alt-Nuc vs Alt treatment groups.



**Supplementary Fig. 3**. Nuc-induced cytokines/chemokines. Human BAL macrophages were cultured with medium (Med), and Nuc for 4 and 24 hr and the culture supernatant was assayed for 80 cytokines/chemokines using the human cytokine array C5 from RayBiotec. The cytokine/chemokine that was upregulated by Nuc is shown in color-coded square symbol.



**Supplementary Fig. 4**. Nuc uptake and induction of cytokines. (**A**) Nuc uptake by human BAL macrophages. Macrophages were incubated with Nuc or medium for 4 hr, stained for cytosolic dsDNA (red) and IFI16 (green) and counterstained with DAPI. The anti-dsDNA antibody specifically detects cytosolic but not nuclear dsDNA. One representative image is shown. Scale bar, 5  $\mu$ m. (**B-D**) Cytokine production by monocytes. Blood monocytes from asthma patients were cultured with. Medium, Nuc (10  $\mu$ g/ml), Alt (10  $\mu$ g/ml) and Alt + Nuc (10  $\mu$ g/ml each) for 24 hr and

assayed for the indicated cytokines. Statistical analyses by Mann-Whitney U test. (E) The increase in mRNA for *il33* and *il1rl1* (ST2) in the lung from the study groups as compared to the Saline (S) group.



**Supplementary Fig. 5**. Flow cytometric measurement of CLEC2D following stimulation with Alt for 0, 24, 48, and 72 hrs (n=4).



**Supplementary Fig. 6**. CLEC2D immunostaining. Triple immunostaining of human BAL macrophages for CLEC2D (red), p-STING (blue) and IFI16 (green) and counterstaining with DAPI (pink) following incubation with Nuc for 4 hr. Co-localization (white) of internalized CLEC2D with p-STING and cytosolic IFI16 is shown in the merged image (n=3). Scale bar, 5 μm.



**Supplementary Fig. 7**. IFI16 and p-STING in the airway tissue from asthma patients. (A) Representative immunostaining of endobronchial biopsy specimens from a neutrophilic asthma patient and a disease control subjects for IFI16 (red). The tissue was counterstained with DAPI for DNA. Cytosolic IFI16 co-localizing with DAPI-staining extranuclear DNA is shown by white arrows (n=17 as per **Fig. 5A**). Scale bar, 10  $\mu$ m. (**B and C**) Representative immunostaining of cadaveric lung sections from a healthy subject and an age- and gender-matched fatal asthma patient

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for IFI16 and p-STING (red). Scale bar, 20  $\mu m.$ 



Supplementary Fig. 8. Autophagy in the asthma models. Representative immunostaining of lung tissue from Sal, Nuc, Alt and Alt-Nuc treated mice for the autophagy marker LC3B. The quantification of the LC3B+ cells from the 4 study groups is shown in the right panel. Scale bar, 20  $\mu$ m.



**Supplementary Fig. 9**. (**A-C**) B6 mice were pretreated i.p. with the STING inhibitor H151 (750 nmol H-151 per mouse in 200 µl 10% Tween-80 in PBS) or vehicle (10% Tween-80 in PBS) 1 hr before i.n. exposure to Alt-Nuc or Sal as per **Fig. 2A**. (A) AHR (lung resistance) in Veh-Sal, H151-Sal, Veh-A+N and H151-A+N groups as measured by flexiVent. <sup>\*</sup>(Veh-Sal vs Veh-A+N) *P*<0.0001; <sup>§</sup>(Veh-Sal vs H151-A+N) *P*<0.0001; <sup>§</sup>(Veh-A+N vs H151-A+N) *P*=0.006. (n=4; 2-way ANOVA, Tukey's multiple comparisons test). (**B and C**) Differential counts of BAL eosinophils and neutrophils. (**D and E**) The lung tissue from  $ALR^{+/+}$ ,  $ALR^{-/-}$ , *Sting*<sup>[/f]</sup> and *LysM*Cre:*Sting*<sup>[/f]</sup> mice (n=4 per group) that were exposed to Alt-Nuc (as per **Fig. 2A**) was immunostained for neutrophil myeloperoxidase (MPO) and the MPO+ cells were quantified. Comparison made by Student's 2 tailed t-test. Data are presented as mean+/-SEM, \**P*<0.005, \*\**P*<0.01, \*\*\**P*<0.001, \*\*\**P*<0.0001, ns=not significant.



**Supplementary Fig. 10.** TNFR2 and asthma. (**A and B**) TNFR2 induction by Alt. Isolated lung CD4+ and Lin- cells from the 4 mouse groups (Sal, Nuc, Alt, and Alt-Nuc) were analyzed by FCM (n=3 per group) (**C and D**) Immunostaining for TNFR2 expressing blood vessels and cells, and CD68+ macrophages in cadaveric lungs from a healthy subject and a fatal asthma patient. Scale bar, 20  $\mu$ m. (**E-I**) Effect of an anti-TNFR2 antibody and an isotype control antibody. B6 mice were pretreated with 100 $\mu$ g of anti-TNFR2 antibody (clone TR75-54.7) or an isotype control antibody on day -1, +2 and +4 and then treated with Alt-Nuc or Sal as per **Fig. 2A**. (**E**) AHR (lung resistance) in Iso ab-Sal, TNFR2 ab-Sal, Iso ab-A+N and TNFR2 ab-A+N groups as measured by flexiVent. \*(Iso ab-Sal vs Iso ab-A+N) *P*<0.0001; <sup>§</sup>(Iso ab-Sal vs TNFR2 ab-A+N) *P*<0.0001; <sup>w</sup>(Iso ab-A+N vs TNFR2 ab-A+N) *P*<0.0001. (n=4; 2-way ANOVA, Tukey's multiple comparisons test). (**F-I**) Quantification of eosinophils and neutrophils in BAL, morphometric quantification of lung inflammation and mucus production in mice pretreated with Iso ab or TNFR2 ab and treated with Sal or A+N (Groups were color coded as in **E**). (n=4; 2-way ANOVA, Tukey's multiple

comparisons test). Data are presented as mean+/-SEM, \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 \*\*\*\*P<0.0001, ns=not significant.



**Supplementary Fig. 11**. Flow cytometric analysis of TNF $\alpha$ + lymphoid cells. (**A-G**) The flow cytometry gating strategy for detection of TNF $\alpha$ + cells in Lin+CD4-, Lin+CD4+ and Lin-CD45+ cell populations in the lung cell digest. (**H**) Comparison of TNF $\alpha$ + cells between Lin+ and Lin-CD45+ lung cells from Alt-treated mice. (**I-L**) Frequency of ST2+TNFR2+ ILC2s in Lin-CD25+ lung cell population and their FMO controls. (**M-O**) Frequency of IL5+TNF $\alpha$ + cells

in lung ILCs (Lin-CD25+) and their isotype controls.