

The CoREST Repressor Complex Mediates Phenotype Switching and Therapy Resistance in Melanoma

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Supplemental Methods

RNA-Seq data analysis

Paired-end RNA-Seq reads were quality- and adapter-trimmed using trimmomatic (1) and gene expression was quantified using the salmon software package (2) against the GENCODE v32 transcriptome (3) summarized to the gene level. Differential expression analysis for pairwise cell type/treatment conditions was conducted using DESeq2 (4) modeling counts as a function of treatment vs the relevant control. Genes with a false discovery rate < 0.01 were considered significant. Gene set enrichment analysis was performed using the differential expression results ranked by decreasing effect size (i.e. log₂ fold change) for each pairwise comparison using the fgsea R package (5) with the KEGG pathways (6) and REACTOME pathways (7) gene set databases. Gene sets with a false discovery rate of < 0.01 were considered significant. The analysis code is available at https://bitbucket.org/bucab/corin_melanoma. Box plots and Volcano plots were generated using Prism 6 software (GraphPad Software). A workflow diagram is included as Supplemental Figure 12 for clarification.

HOMER Motif Analysis

We analyzed corin-upregulated genes (Corin+PLX vs PLX; p<0.01, LF>2) for enrichment of transcription factor motifs using HOMER v4.10.1 *findMotifs* against random background sequences (len = 8-10bp, promoters approximated to -400 to +100 TSS). Both “de novo” and “known” motif analyses were generated.

DUSP1 Promoter Mapping

EMT and AP-1 family member transcription factors enriched in the HOMER motif analysis were mapped to the DUSP1 promoter (annotated in Ensembl GRCh38.p130) using TFmotifView (8). Coordinates relative to the TSS were obtained from the UCSC Genome Browser (GRCh38).

ChIP-seq Analysis

FASTQ files were aligned using Bowtie 2.5.0 (9) to the reference genomes (hg38, Genome Reference Consortium Human Build 38). Peak calling was performed using Model-based Analysis of ChIP-seq (MACS) 1.4.2 (10) with ‘--nomodel’ and ‘--nolambda’ options. For signal-based analyses, duplicate reads were filtered using Picard 3.0.0 [<https://broadinstitute.github.io/picard/>]. In order to address the potential effects of global histone modification alteration (Figure 1, A and B), a scale factor (SF) was determined for each experimental sample using ChIPseqSpikeInFree (11). Subsequently, the read density for each genomic region was normalized by scaling it with the factor of $1e7 / (\text{mapped read counts} * \text{SF})$ using the 'genomcov' command available in the BEDTools suite (12). This normalization process helped to account for variations stemming from global histone modification changes, ensuring more accurate data analysis and interpretation. Unique and overlapping peaks were defined using ‘intertect’ command in BEDTools suite. Motif occurrence analysis was performed using ‘findMotifs.pl’ command in hypergeometric optimization of motif enrichment (HOMER) suite version 4.4 (13). For ChIP-seq result correlation with RNA-seq data, expression level of each transcript isoform was quantified using Salmon version 1.9.0 (2). Peaks were visualized in the Integrative Genomics Viewer (IGV) (14). Figures show data from one representative biological replicate.

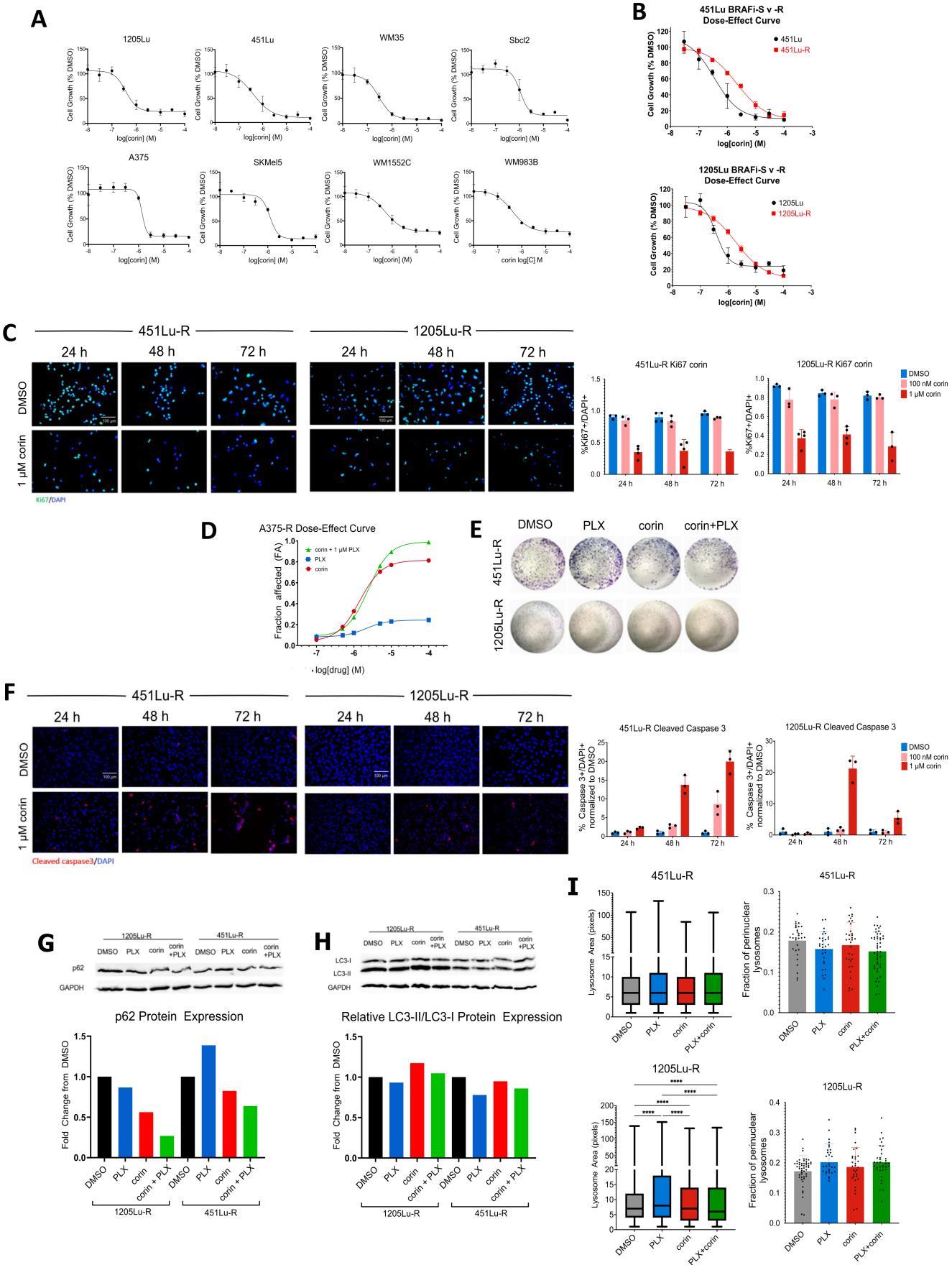
Open Source Database Analyses

DUSP1 expression in normal skin, benign nevi, and malignant melanoma was obtained from publicly available microarray data (15). Kaplan-Meier curves were generated by GEPIA (<http://gepia.cancer-pku.cn>) using the TCGA database (<http://cancergenome.nih.gov/>). Comparison analysis was performed using publicly available whole-exome sequence (WES) datasets of serial tumor biopsies (baseline and acquired resistant tumors) from patients with advanced melanoma treated with MAPK inhibitor (MAPKi) regimens (16) by comparing the list of 'Loss of Function (LOF)' genes from the dataset with corin-regulated genes. RCOR1, HDAC1, HDAC2, and KDM1A expression in BRAFi-resistant tumors compared to baseline was analyzed with data from the validation microarray dataset (GSE50509 and GSE61992) used by Song et al (16).

Calculation of the combination index (CI)

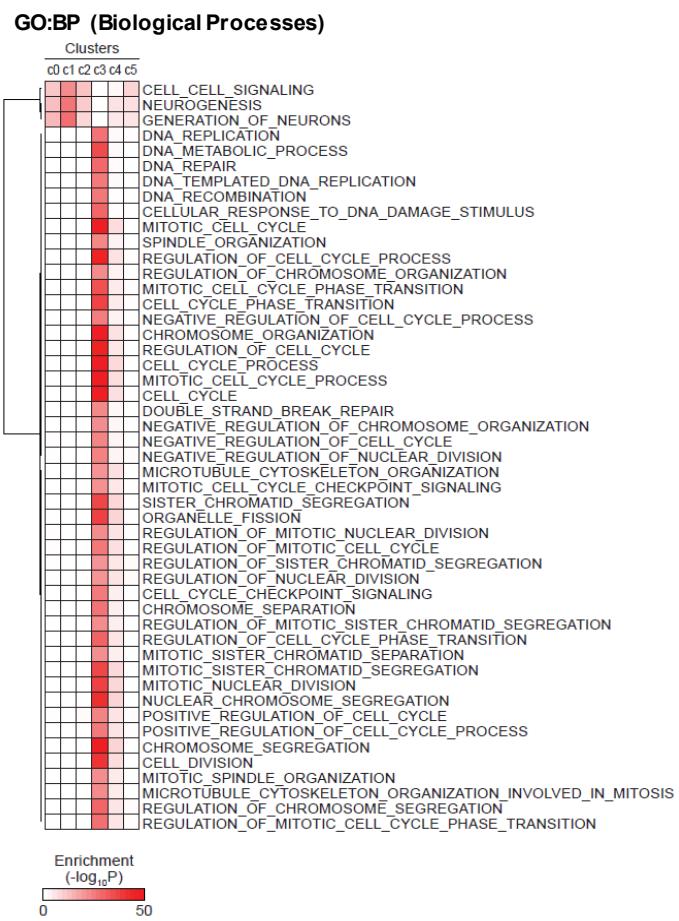
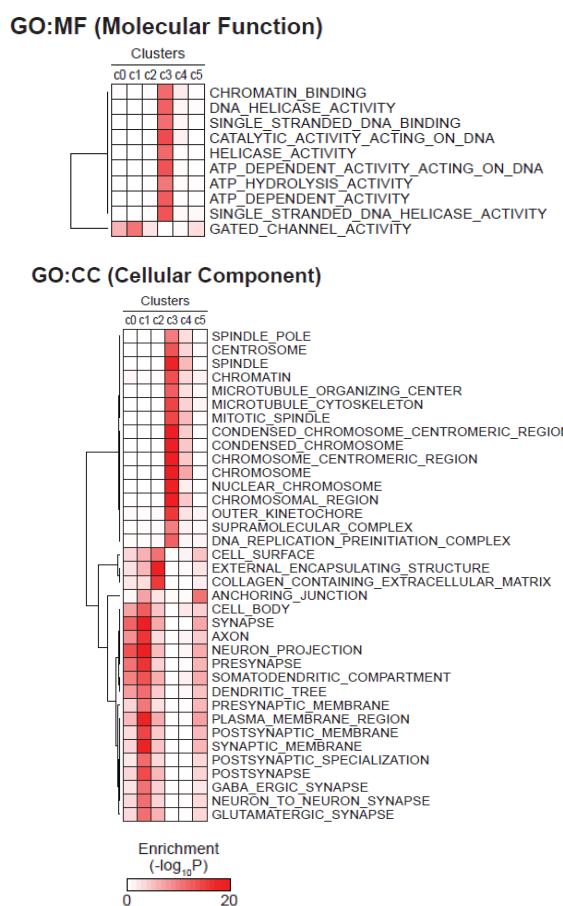
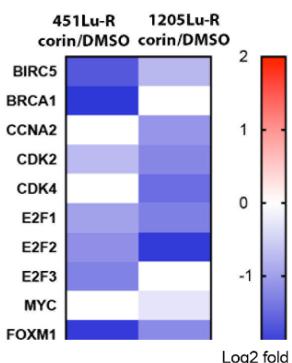
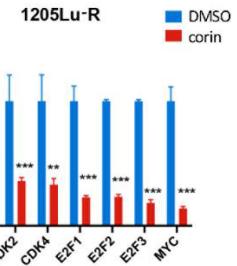
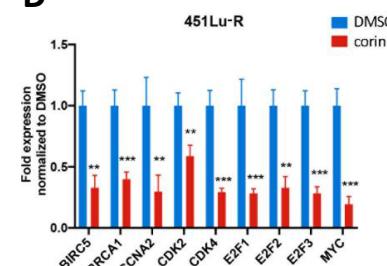
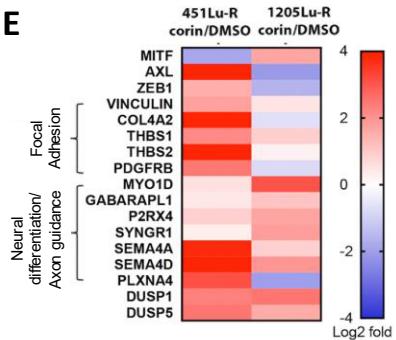
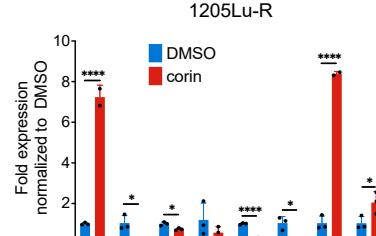
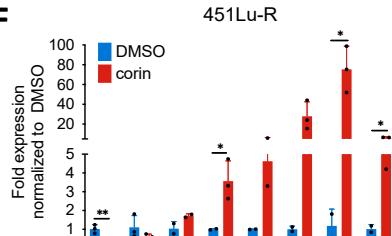
The combined activity of corin and PLX4032 was determined by calculating the CI for both compounds in 451Lu-R, 1205Lu-R and A375-R cells using Compusyn software (Biosoft). The concentrations used for PLX4032 alone are: 0.01 μ M, 0.1 μ M, 1 μ M, 5 μ M, 10 μ M; concentrations used for corin alone are: 0.01 μ M, 0.1 μ M, 1 μ M, 5 μ M, 10 μ M. Concentrations used for combination treatment are: 1 μ M PLX4032 + corin (0.01 μ M, 0.1 μ M, 1 μ M, 5 μ M, 10 μ M). The results are interpreted as: 0 < CI < 1 indicates synergism; CI = 1 indicates an additive effect; and CI > 1 indicates antagonism.

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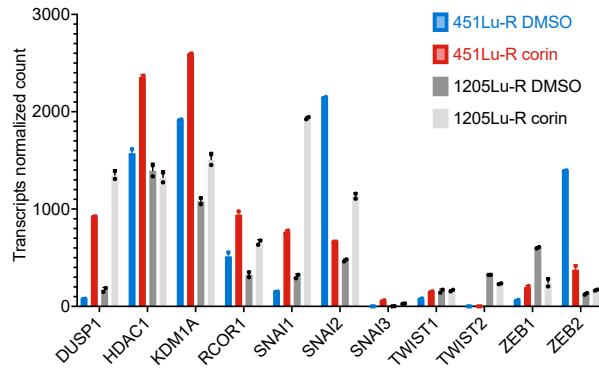


Supplemental Figure 1. CoREST inhibition promotes response to BRAFi-therapy in BRAFi-resistant melanoma cells. (A) Proliferation assays of BRAFi-sensitive cell lines (1205Lu, 451Lu, WM35, Sbcl2, A375, SKMel28, WM1552C and WM983B) treated with increasing doses of corin for 72 h. (B) Cell proliferation assay of 451Lu and 1205Lu BRAFi-sensitive (-S) and -resistant (-R) cell lines treated with increasing doses of corin for 72 h with the corin. (C) Immunofluorescent staining of Ki67 and quantification in 451Lu-R and 1205Lu-R BRAFi-resistant melanoma cell lines following 24 h, 48 h, 72 h treatment with DMSO, 100 nM corin, or 1 μ M corin. Representative images shown for 1 μ M corin, scale bar = 100 μ m. (D) Drug synergy graph for corin and PLX4032 in A375-R BRAFi-R melanoma lines. (E) Colony formation assays for 451Lu-R and 1205Lu-R melanoma cells treated with DMSO, 5 μ M PLX4032 alone, 2.5 μ M corin alone, and 2.5 μ M corin + 5 μ M PLX4032 for 10 days. (F) Immunofluorescence staining of cleaved caspase-3 and quantification in 451Lu-R and 1205Lu-R BRAFi-resistant melanoma cell lines following 24 h, 48 h, 72 h treatment with DMSO, 100 nM corin, or 1 μ M corin. Representative images shown for 1 μ M corin, scale bar = 100 μ m. (G-I) Autophagy analysis in 1205Lu-R and 451Lu-R melanoma cells following 24 h treatment with DMSO, 5 μ M PLX4032 alone, 2.5 μ M corin alone, or 5 μ M PLX4032 + 2.5 μ M corin using p62 Western blot protein expression and quantification (G), LC3-II/LC3-I Western blot protein expression and quantification (H), and (I), Lysotracker staining of lysosomes in 451Lu-R (top) and 1205Lu-R (bottom). Representative images shown, scale bar = 100 μ m. Significance asterisks indicate 1-way ANOVA with Turkey's test. All data represents mean + SE.

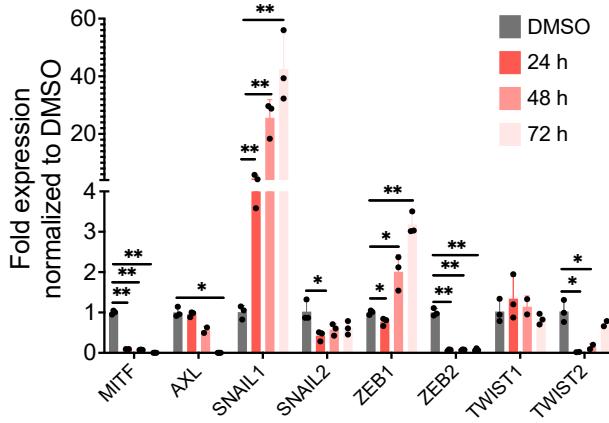
*** $P < 0.001$, ** $P < 0.0001$.

A**B****C****D****E****F**

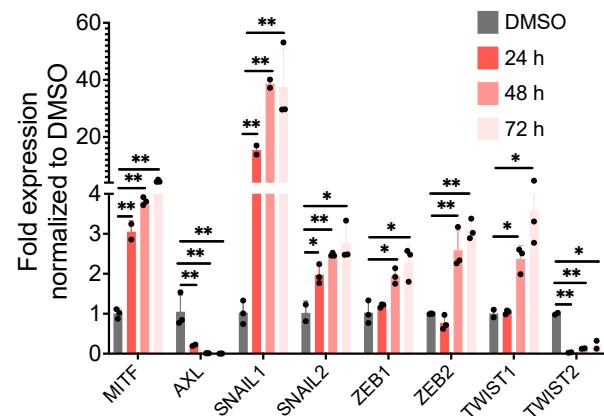
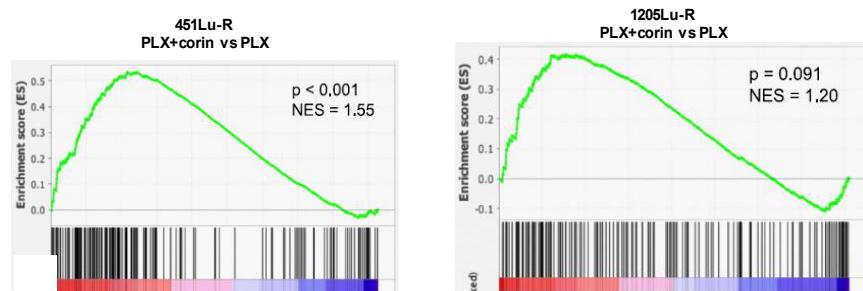
Supplemental Figure 2. Corin +/- PLX4032 differentially regulates signaling pathways in BRAFi-R melanomas with specific changes in cell cycle, focal adhesion, and neural differentiation gene sets. (A-B) Gene ontology analysis of k-means clusters shown in main Figure 3B. Significantly enriched ontologies with hypergeometric $P < 1e-20$ for biological processes (BP) (A) or $P < 1e-10$ for molecular function (MF) and cellular component (CC) (B) are shown. (C) heatmap of RNA-seq data illustrating gene expression patterns associated with cell cycle/DNA repair in 451Lu-R vs 1205Lu-R melanoma cells following corin treatment (2.5 μ M, 24 h). (D) qPCR validated expression of differentially expressed genes from RNA-seq data associated with cell cycle/DNA repair in 451Lu-R and 1205Lu-R melanoma cells following corin treatment (2.5 μ M, 24 h). (E) heatmap of RNA-seq data illustrating gene expression patterns associated with focal adhesions and neural differentiation/axon guidance in 451Lu-R and 1205Lu-R melanoma cells following corin treatment (2.5 μ M, 24 h). (F) qPCR validated expression of differentially expressed genes from RNA-seq data associated with focal adhesions and neural differentiation/axon guidance in 451Lu-R and 1205Lu-R following corin treatment (2.5 μ M, 24h). Significance asterisks indicate unpaired, two-tailed student's t test. All data represents mean + SE. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$.

A

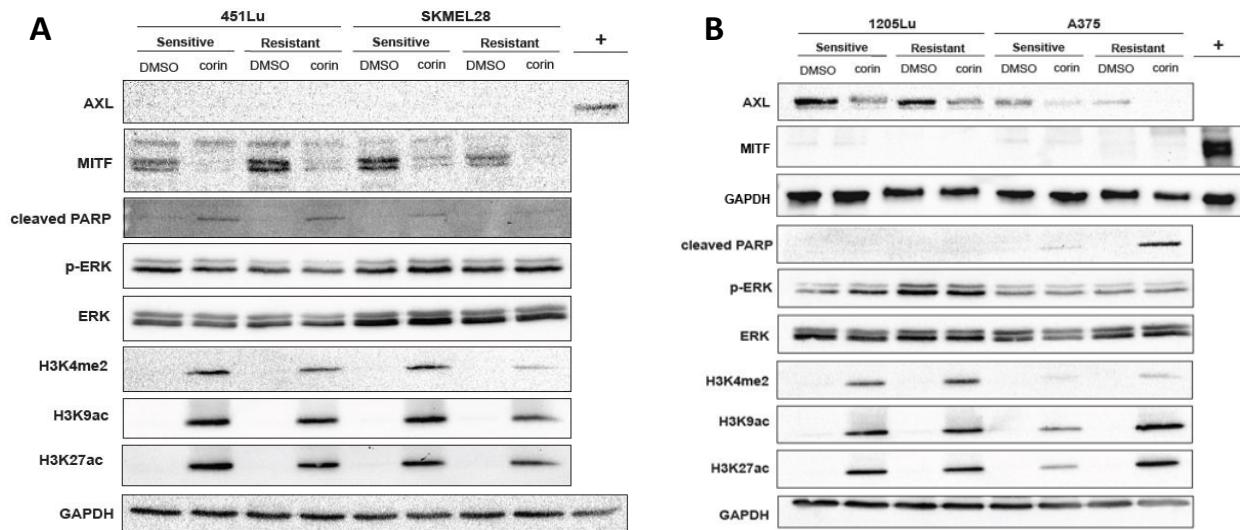
451Lu-R

B

1205Lu-R

C**Wouters et al. Intermediate Signature****D**

Supplemental Figure 3. Corin impacts the expression of EMT-related genes and induces an intermediate phenotype gene expression signature. **(A)** Raw RNA-seq transcript counts of EMT transcription factors in 451Lu-R and 1205Lu-R melanoma cells following corin treatment (2.5 μ M, 24 h). **(B, C)** qPCR validation of differentially expressed genes from RNA-seq data associated with the phenotype switch and EMT in 451Lu-R (**B**) and 1205Lu-R (**C**) melanoma cells following corin treatment (2.5 μ M; 24h, 48h, and 72h). **(D)** Corin-induced intermediate phenotype gene expression signature, defined in Wouters et al. (17), in 451Lu-R (left) and 1205Lu-R (right) melanoma cells treated with 5 μ M PLX4032 versus 2.5 μ M corin + 5 μ M PLX4032 for 24 h. Significance asterisks indicate 1-way ANOVA with Turkey's test. All data represents mean + SD. * $P < 0.05$, ** $P < 0.01$.



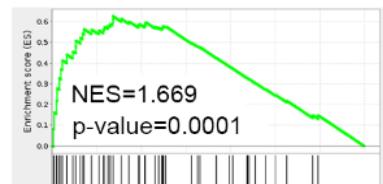
Supplemental Figure 4. Corin mediates phenotype switching in both BRAFi-sensitive and -resistant melanoma cell pairs. (A, B) Western blot analysis of MAPK pathway activity in BRAFi-sensitive and -resistant melanoma cell lines treated with 2.5 μ M corin for 24 h (A), in 451Lu and SKMel28 MITF^{high}/AXL^{low} melanoma cell lines with 1205Lu-R lysates run as a positive control for AXL (Western blots were run contemporaneously) and (B), 1205Lu and A375 MITF^{low}/AXL^{high} melanoma cell lines with 451Lu-R cell lysates run as a positive control for MITF. The 8-lane Western blots with same loading control were run contemporaneously.

A

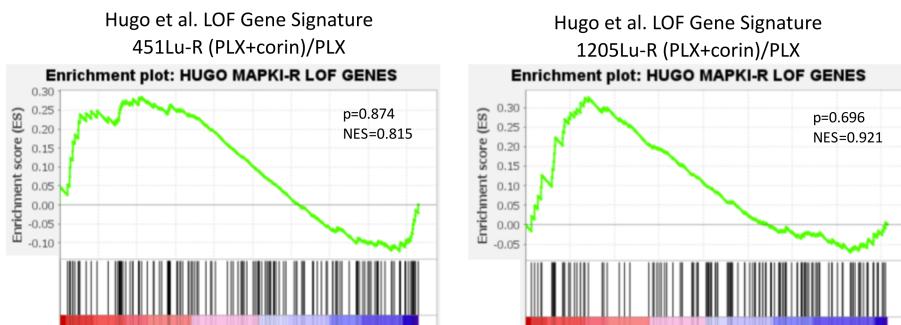
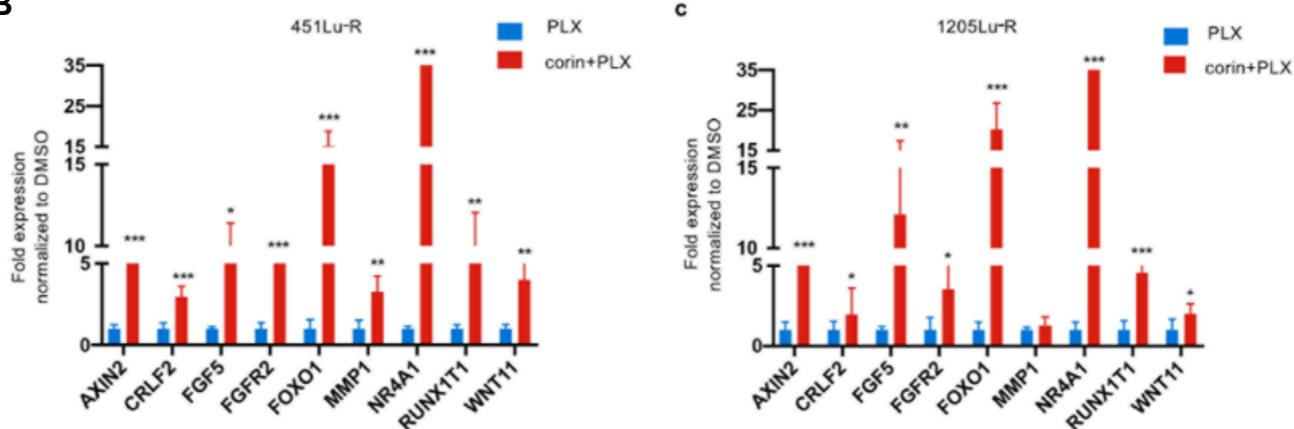
451Lu-R corin vs DMSO
Reactome Interferon gamma signaling

**B**

1205Lu-R corin vs DMSO
Reactome Interferon gamma signaling

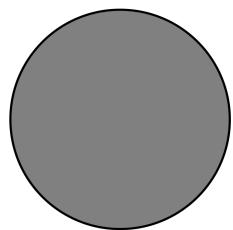


Supplemental Figure 5. Corin treatment of melanoma cells promotes interferon gamma expression of gene signatures in 451Lu-R and 1205Lu-R cells. (A, B) GSEA analysis in corin (2.5 μ M, 24 h) versus DMSO-treated melanoma cells illustrating enriched Interferon gamma signaling in 451Lu-R (**A**) and 1205Lu-R (**B**) melanoma cells. NES (Normalized Enrichment Score), (FDR < 0.05).

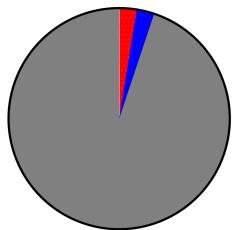
A**B****C**

Gene Expression in BRAFi-naive vs -resistant patient tumors

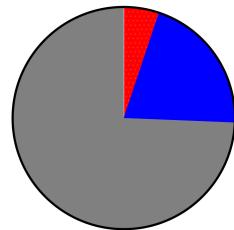
RCOR1



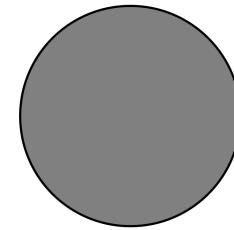
HDAC1



HDAC2



KDM1A



■ Upregulated (≥ 2 fold)
■ Downregulated (≤ -2 fold)
■ Unchanged

Total=39

Total=39

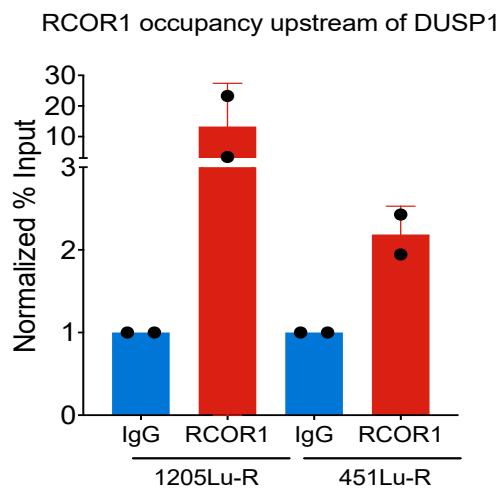
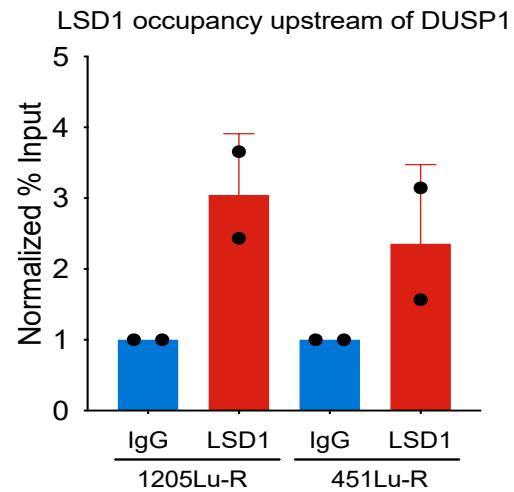
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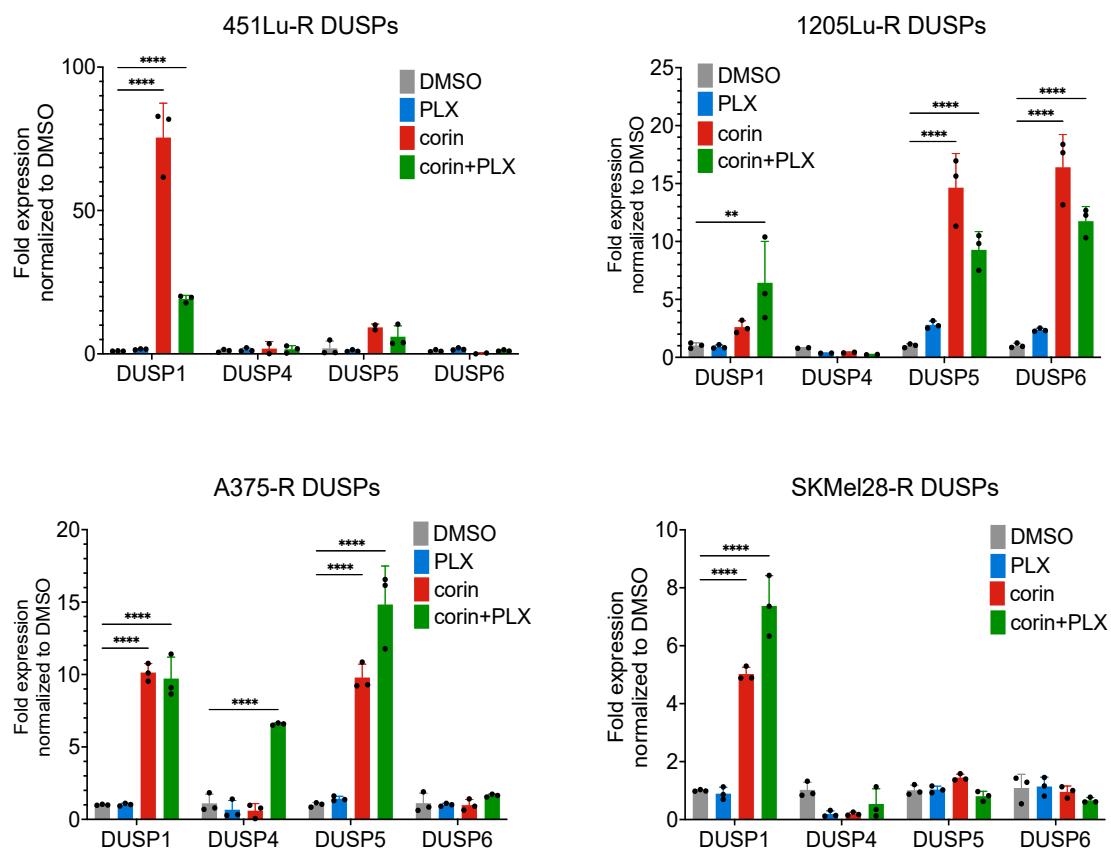
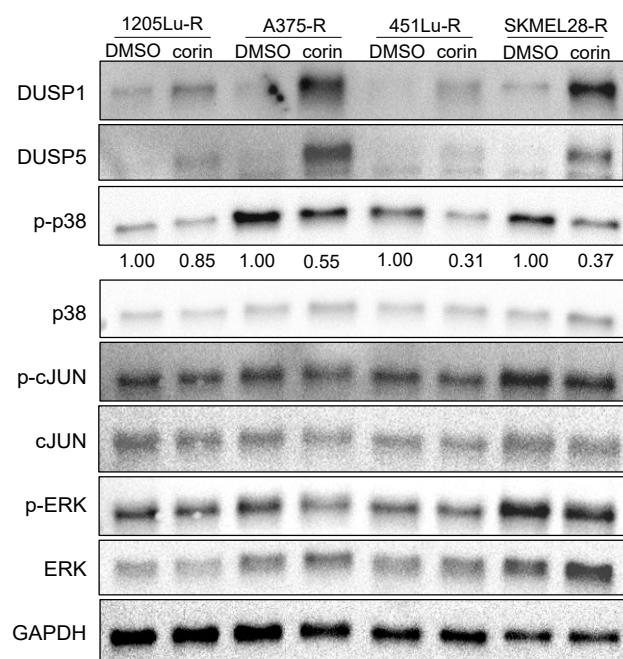
Supplemental Figure 6. Genes downregulated during the acquisition of MAPKi-resistance in melanoma patients are induced following corin treatment. (A) Corin-induced acquired MAPKi resistance LOF gene signature, defined in Hugo et al. (28), in 451Lu-R (left) and 1205Lu-R (right) melanoma cells treated with 2.5 μ M corin + 5 μ M PLX4032 versus 5 μ M PLX4032 for 24 h. (B) qPCR validation of genes downregulated during acquired MAPKi-resistance in human melanomas which are induced following 24 h treatment with corin + PLX4032 (5 μ M/2.5 μ M) vs. PLX4032 alone (5 μ M) in 451Lu-R (C) and 1205Lu-R (D) melanoma cells. (C) RCOR1, HDAC1, HDAC2, and KDM1A expression in BRAFi-resistant tumors compared to baseline analyzed with data from the validation microarray dataset (GSE50509 and GSE61992) used by Song et al.³⁶ Significance asterisks indicate unpaired, two-tailed student's *t* test. All data represents mean + SD. **P* < 0.05, ***P* < 0.01, ****P* < 0.001.

A

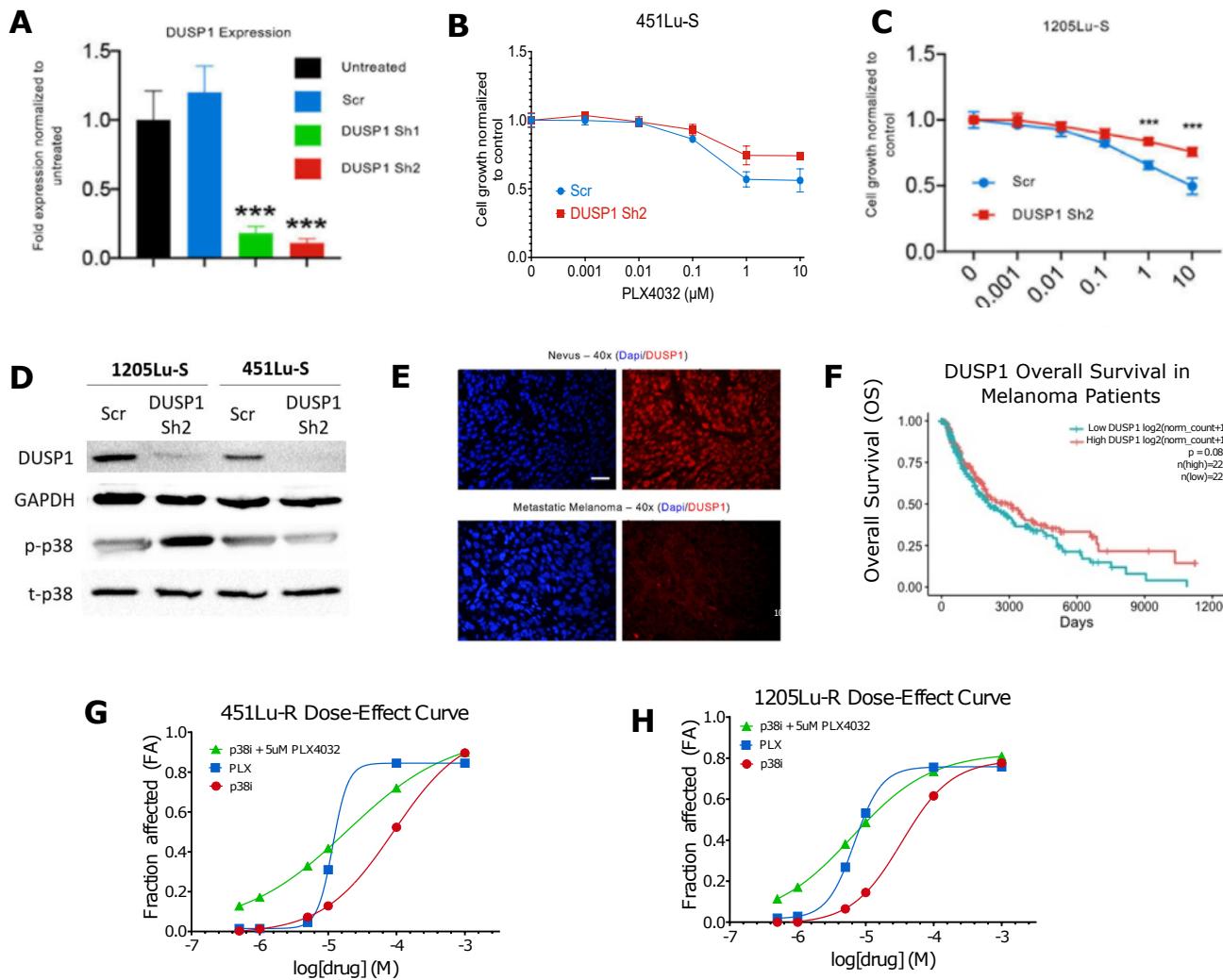
451Lu-R				1205Lu-R			
Rank	Motif	Best Match	P-value	Rank	Motif	Best Match	P-value
1		ZNF263	1e-16	1		NFYB	1e-11
2		MED1	1e-13	2		NEUROD1	1e-10
3		PRDM1	1e-12	3		MXI1	1e-10
4		HINFP	1e-10	4		DUX	1e-10
5		ZNF263	1e-10				
6		ZBTB3	1e-10				
7		CTCFL	1e-10				

B**C**

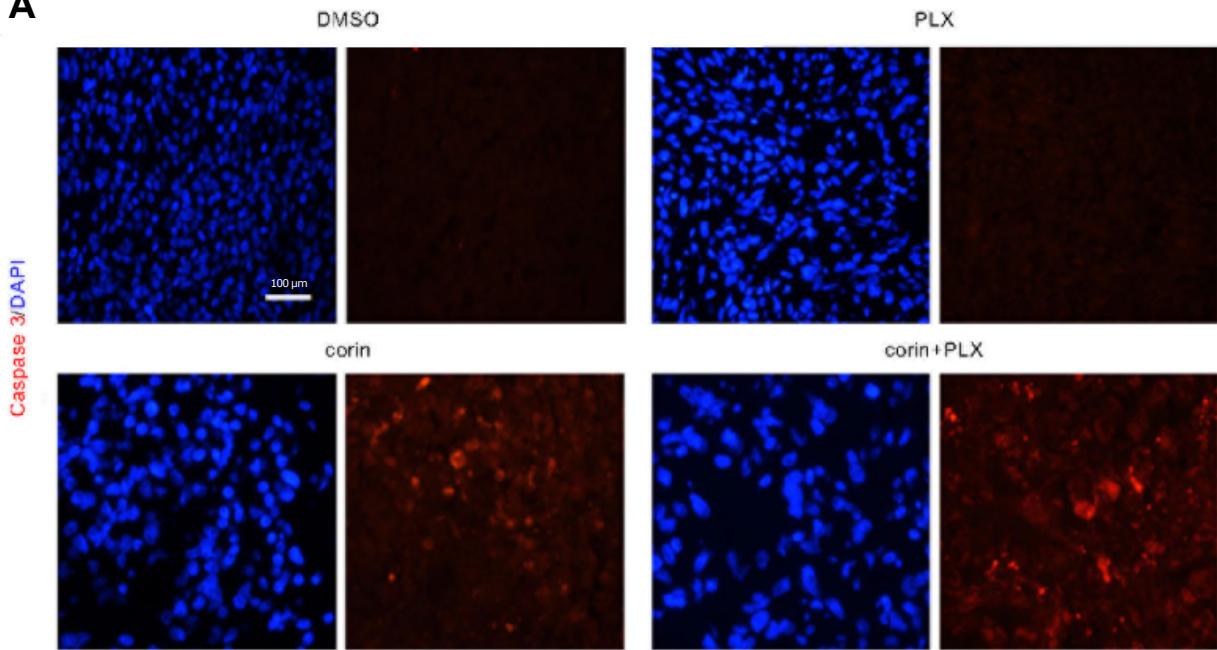
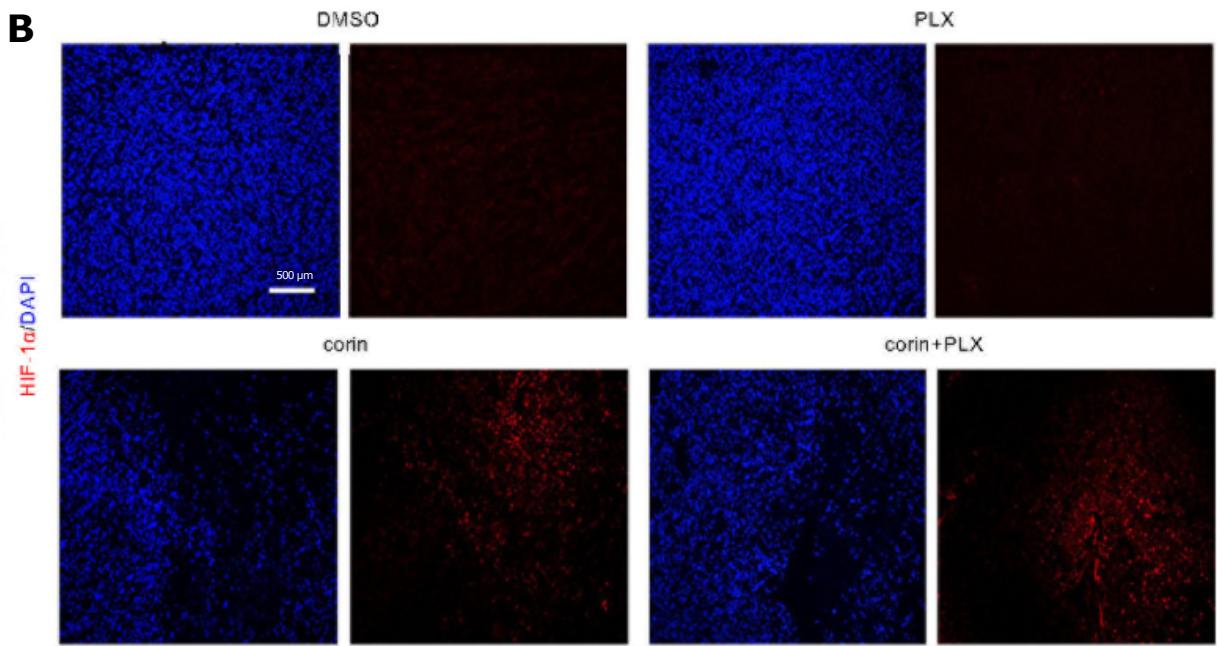
Supplemental Figure 7. The CoREST complex directly binds upstream of the DUSP1 promotor. (A) Top *de novo* motifs enriched in corin-upregulated genes in 451Lu-R (left) and 1205Lu-R (right) melanoma cells. **(B, C)** ChIP-qPCR analysis of RCOR1 **(B)** and LSD1 **(C)** occupancy upstream the DUSP1 promoter region in 1205Lu-R and 451Lu-R cells compared to IgG control (n=2).

A**B**

Supplemental Figure 8. Corin mediates re-sensitization to BRAFi therapy through DUSP1-associated inhibition of p38 MAPK in melanoma cells. **(A)** qPCR analysis of DUSP1, 4, 5, and 6 expression in 451Lu-R, 1205Lu-R, A375-R and SKMel28-R melanoma cells following 24 h treatment with 5 μ M PLX4032, +/- 2.5 μ M corin. **(B)** Western blot analysis of DUSP expression and MAPK-associated proteins in 1205Lu-R, A375-R, 451Lu-R and SKMel28-R melanoma cells treated with 2.5 μ M corin for 48 hours. Quantification of relative expression of p-p38 (active) versus p38 (total) protein expression relative to the DMSO control is shown below each p-p38 band. Western blots were run contemporaneously. Significance asterisks indicate 1-way ANOVA with Turkey's test. All data represents mean + SD. ** $P < 0.01$, *** $P < 0.0001$.

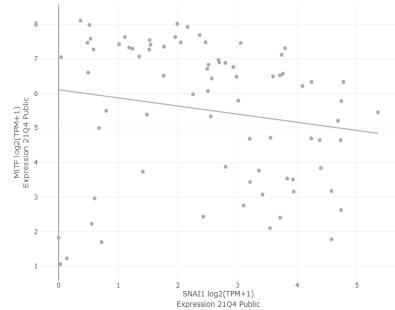


Supplemental Figure 9. DUSP1 mediates melanoma cell growth, response to BRAF inhibitor therapy and p38 MAPK activity and is downregulated in human melanomas versus benign nevi. (A) DUSP1 knockdown by shRNA lentivirus in 1205Lu cells, confirmed by qRT-PCR. (B, C) Cell growth assays of 451Lu and 1205Lu melanoma cells treated with PLX4032 (72 h) following knockdown of DUSP1 versus scramble control, (n=3). (D) Western blot analysis of DUSP1 and pp38/p38 expression in 1205Lu-S and 451Lu-S melanoma cells following knockdown of DUSP1 versus scramble control. (E) Quantification of DUSP1 protein following DUSP1 knockdown vs scramble control. (F) Immunostain for DUSP1 expression in human melanoma specimens and benign nevi. Representative images shown, scale bar = 100 μ m. (G) Kaplan-Meier curves illustrating the relationship between the expression of DUSP1 and overall survival in patient tumor specimens split at the median. Data obtained from the TCGA melanoma database (<https://portal.gdc.cancer.gov>). (H, I) Drug synergy graphs and Combination Index (CI) measurements for p38i and PLX4032 in 451Lu-R (H) and 1205Lu-R (I) BRAFi-R melanoma cells. Significance asterisks indicate 1-way ANOVA with Turkey's test. All data represents mean + SD. *** $P < 0.0001$.

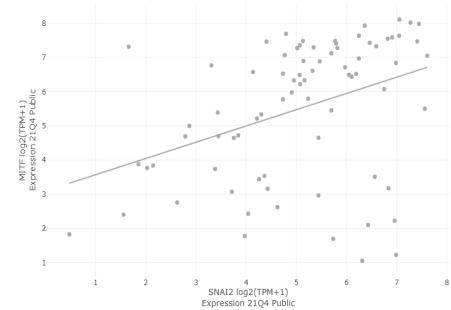
A**B**

Supplemental Figure 10. Corin promotes apoptosis and hypoxia in BRAFi-resistant melanoma xenografts. (A, B) Immunofluorescence stain of cleaved caspase 3 (A) and HIF-1 α (B) in 1205Lu-R melanoma xenografts treated with DMSO, PLX4032 alone, corin alone, or corin + PLX4032. Representative images shown, scale bar = 100 μ m (A) or 500 μ m (B).

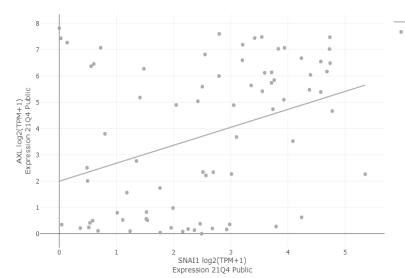
A MITF vs SNAI1 Expression in Melanoma Cells



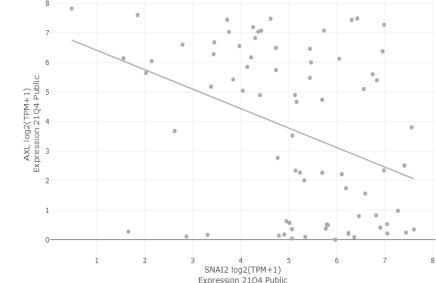
B MITF vs SNAI2 Expression in Melanoma Cells



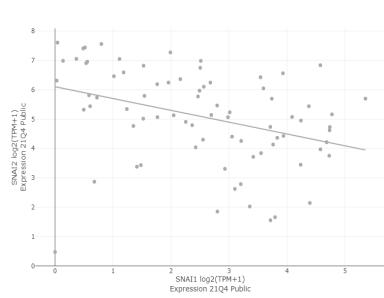
C AXL vs SNAI1 Expression in Melanoma Cells



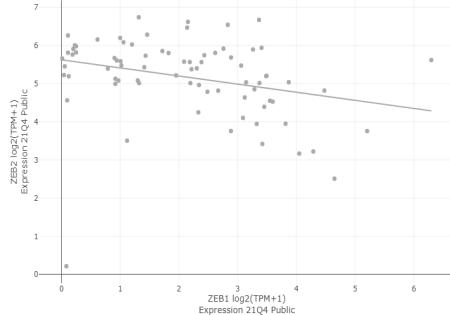
D AXL vs SNAI2 Expression in Melanoma Cells



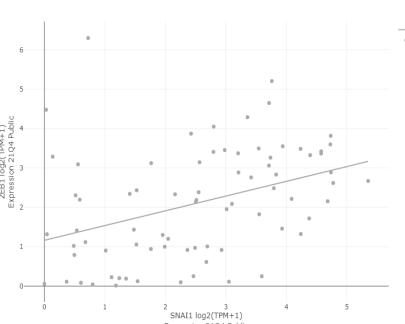
E SNAI2 vs SNAI1 Expression in Melanoma Cells



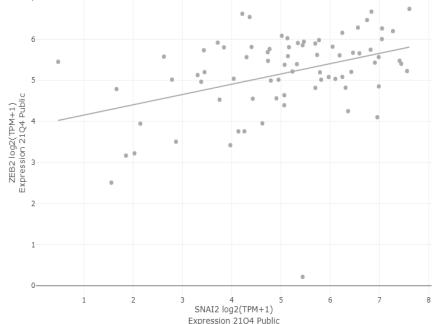
F ZEB2 vs ZEB1 Expression in Melanoma Cells



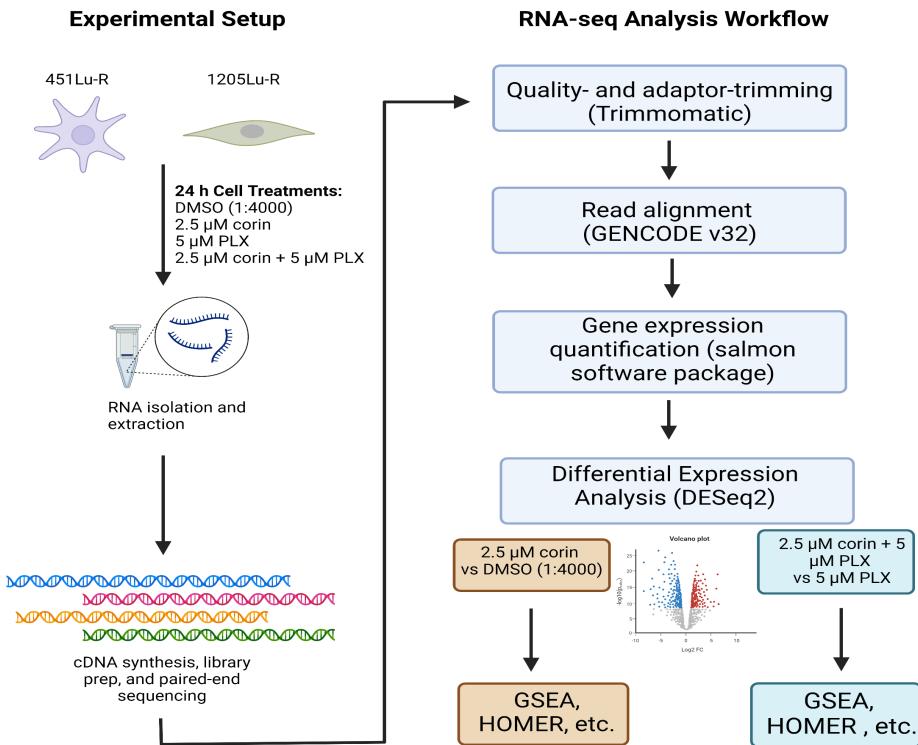
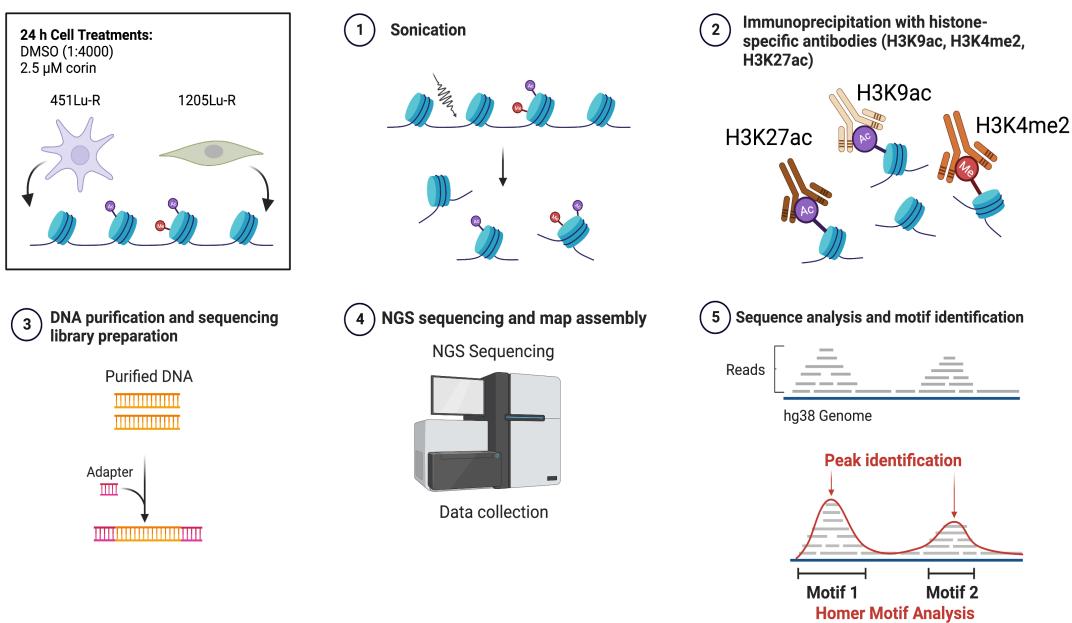
G ZEB1 vs SNAI1 Expression in Melanoma Cells



H ZEB2 vs SNAI2 Expression in Melanoma Cells



Supplemental Figure 11. Depmap analysis of EMT-associated transcription factors and their relevance to melanoma phenotypes in human melanoma cell lines. Survey of melanoma cell lines using the Broad Institute Depmap portal (<https://depmap.org/portal/>) (A-H) illustrating statistically significant inverse dependencies of expression of MITF with SNAI1 (A) and AXL with SNAI2 (D) as well as significant inverse dependencies of SNAI1 with SNAI2 (E) and ZEB1 with ZEB2 (F). The Depmap survey also found significant direct relationships between MITF and SNAI2 (B), AXL and SNAI1 (C) ZEB1 and SNAI1 (G) as well as ZEB2 and SNAI2 (H) in the melanoma cell lines evaluated.

A**B****ChIP-seq Workflow**

Supplemental Figure 12. Experimental Workflows for RNA-seq (A) and ChIP-seq (B)

studies. (A) Workflow schematic depicting RNA-sequencing experimental design. **(B)**

Workflow schematic depicting chromatin immunoprecipitation-sequencing (ChIP-seq)

experimental design. Created with BioRender.

Supplemental Table 1. IC50 values of BRAFi -sensitive vs -resistant 451Lu and 1205Lu cells.

Cell Line	LogIC50	IC50 (nM)
451Lu	-6.457	350
451Lu-R	-5.662	2177
1205Lu	-6.417	382
1205Lu-R	-5.711	1943

Supplemental Table 2. Combinations Index (CI) for A375-R cells treated with PLX4032 and corin. CIs indicate the degree of drug synergy where CI >1 indicates drug antagonism, CI=1 indicates drug additivity, and CI <1 indicates drug synergy for cell proliferation across a range of PLX and corin concentrations following 72 h treatment.

PLX (μ M)	corin (μ M)	A375-R Combination Index (CI)
1.0	0.1	40.52
1.0	0.5	4.12
1.0	1.0	1.72
1.0	5.0	0.06*
1.0	10	0.01*
1.0	100	7.43E-04*

Supplemental Table 3. Top leading edge genes of Reactome Interferon gamma signaling in 451Lu-R cells.

Gene	451Lu-R corin vs DMSO Log2foldChange	451Lu-R corin vs DMSO padj
<i>HLA-DRA</i>	9.622	3.30E-10
<i>CAMK2B</i>	7.493	6.72E-06
<i>OASL</i>	7.044	4.06E-05
<i>HLA-DRB1</i>	6.251	5.14E-08
<i>TRIM29</i>	5.935	2.31E-03
<i>GBP5</i>	4.834	9.49E-03
<i>HLA-DQA2</i>	4.798	4.82E-02
<i>HLA-DRB5</i>	4.778	5.05E-02
<i>HLA-DPB1</i>	4.483	3.99E-05
<i>HLA-DQA1</i>	4.116	4.64E-02

Supplemental Table 4. Top leading edge genes of Reactome Interferon gamma signaling in 1205Lu-R cells.

Gene	1205Lu-R corin vs DMSO Log2foldChange	1205Lu-R corin vs DMSO padj
<i>HLA-DRB1</i>	6.1749	7.46E-04
<i>HLA-DQB1</i>	6.0516	2.23E-04
<i>GBP5</i>	4.7505	1.89E-04
<i>TRIM48</i>	4.7095	5.10E-02
<i>TRIM10</i>	4.2114	1.18E-01
<i>GBP2</i>	4.0186	1.05E-41
<i>HLA-DRA</i>	3.5233	4.28E-17
<i>HLA-DPA1</i>	3.3493	3.66E-02
<i>HLA-DPB1</i>	3.2818	1.65E-01
<i>IRF5</i>	3.1357	1.06E-02

Supplementary Table 5. GSEA analysis of type I and type II IFN pathways for 451Lu-R and 1205Lu-R cells treated with corin vs DMSO.

	451Lu-R corin vs DMSO_pval	451Lu-R corin vs DMSO_NES	1205Lu-R corin vs DMSO_pval	1205Lu-R corin vs DMSO_NES
Reactome				
Interferon gamma signaling (type II)	0.0001	1.6653	0.0001	1.6694
Reactome				
Interferon gamma signaling (type I)	0.2715	1.1184	0.7778	0.8674

Supplementary Table 6. GSEA analysis of type I and type II IFN pathways for 451Lu-R and 1205Lu-R cells treated with PLX+corin vs PLX.

	451Lu-R PLX+corin vs PLX_pval	451Lu-R PLX+corin vs PLX_NES	1205Lu-R PLX+corin vs PLX_pval	1205Lu-R PLX+corin vs PLX_NES
Reactome				
Interferon gamma signaling (type II)	1.0036E-05	1.7631	1.0E-05	1.5643
Reactome				
Interferon gamma signaling (type I)	0.0272	1.3393	0.9723	0.6593

Supplementary Table 7. Transcript changes of RISC components and repetitive elements induced by corin.

	451Lu-R corin vs DMSO log2FoldChange	451Lu-R corin vs DMSO pvalue	1205Lu-R corin vs DMSO log2FoldChange	1205Lu-R corin vs DMSO pvalue
<i>DICER1</i>	-0.254	5.38E-02	-0.234	9.94E-02
<i>AGO2</i>	-0.674	2.59E-10	-0.816	3.19E-30
<i>ERV1</i>	0.336	1.28E-47	0.751	0
<i>ERVL</i>	0.286	3.74E-26	0.437	4.12E-64

Supplementary Table 8. Expression of genes downregulated in patient melanomas during acquired MAPKi resistance. Genes repressed in MAPKi-resistant melanoma specimens, reported by Hugo et al. (28), and associated level of induced expression following 24h treatment with corin + PLX4032 (2.5 µM/5 µM) versus PLX4032 alone (5 µM) in 451Lu-R and 1205Lu-R melanoma cells.

Gene	451Lu-R (PLX+corin)/PLX Log2FoldChange	1205Lu-R (PLX+corin)/PLX Log2FoldChange
<i>AXIN2</i>	3.02	3.22
<i>CRLF2</i>	7.52	4.13
<i>CTNNA2</i>	6.53	3.87
<i>FGF12</i>	5.93	2.29
<i>FGF5</i>	7.11	6.46
<i>FGFR2</i>	2.97	5.63
<i>FOXO1</i>	2.63	2.90
<i>MMP1</i>	13.73	4.02
<i>NR4A1</i>	3.30	4.19
<i>PLD1</i>	2.06	2.00
<i>RUNX1T1</i>	6.42	2.82
<i>SIRT4</i>	2.32	3.32
<i>WNT11</i>	7.47	6.98

Supplementary Table 9. MOTIFs enriched with corin treatment in 451Lu-R H3K9ac ChIPseq.

Motif name	Consensus	Log P-value	% of Target Sequences with Motif
Mef2c(MADS)/GM12878-Mef2c-ChIP-Seq(GSE32465)/Homer	DCYAAAAATAGM	-1640	0.6639
Mef2a(MADS)/HL1-Mef2a.biotin-ChIP-Seq(GSE21529)/Homer	CYAAAAATAG	-1495	0.6287
IDD5(C2H2)/colamp-IDD5-DAP-Seq(GSE60143)/Homer	TTTTGTCTTTTBTK	-1467	0.7511
REM19(REM)/colamp-REM19-DAP-Seq(GSE60143)/Homer	AAAAAAA	-1385	0.7387
IDD4(C2H2)/col-IDD4-DAP-Seq(GSE60143)/Homer	TTTGTCTTWB	-1257	0.7691
GSC(Homeobox)/FrogEmbryos-GSC-ChIP-Seq(DRA000576)/Homer	RGGATTAR	-1135	0.8286
VRN1(ABI3VP1)/col-VRN1-DAP-Seq(GSE60143)/Homer	TTTTTTTTTT	-1105	0.5166
RLR1?/SacCer-Promoters/Homer	WTTTTCYYTTT	-1077	0.6953
Mef2b(MADS)/HEK293-Mef2b.V5-ChIP-Seq(GSE67450)/Homer	GCTATTTTGGM	-953.8	0.6848
GLIS3(Zf)/Thyroid-Glis3.GFP-ChIP-Seq(GSE103297)/Homer	CTCCCTGGGAGGCCN	-704.8	0.825
Bapx1(Homeobox)/VertebralCol-Bapx1-ChIP-Seq(GSE36672)/Homer	TTRAGTGSYK	-583.7	0.8914
LXRE(NR),DR4/RRAW-LXRb.biotin-ChIP-Seq(GSE21512)/Homer	RGGTTACTANAGGTCA	-579.4	0.2808
Hoxd12(Homeobox)/ChickenMSG-Hoxd12.Flag-ChIP-Seq(GSE86088)/Homer	HDGYAATGAAAN	-500.1	0.7851
EAR2(NR)/K562-NR2F6-ChIP-Seq(Encode)/Homer	NRBCARRGGTCA	-417.5	0.701
Atf1(bZIP)/K562-ATF1-ChIP-Seq(GSE31477)/Homer	GATGACGTCA	-370.6	0.4496
COUP-TFII(NR)/K562-NR2F1-ChIP-Seq(Encode)/Homer	GKBCARAGGTCA	-356.1	0.7104
Gfi1b(Zf)/HPC7-Gfi1b-ChIP-Seq(GSE22178)/Homer	MAATCACTGC	-343	0.4783
CRX(Homeobox)/Retina-Crx-ChIP-Seq(GSE20012)/Homer	GCTAATCC	-341.5	0.8691
Snail1(Zf)/LS174T-SNAIL1.HA-ChIP-Seq(GSE127183)/Homer	TRCACCTGCY	-326.9	0.5628
NGA4(ABI3VP1)/col-NGA4-DAP-Seq(GSE60143)/Homer	TKNTCAGGTG	-307.2	0.8968
Atf7(bZIP)/3T3L1-Atf7-ChIP-Seq(GSE56872)/Homer	NGRTGACGTCA	-301.6	0.3556
THRa(NR)/C17.2-THRa-ChIP-Seq(GSE38347)/Homer	GGTCANYTGAGGWCA	-290.8	0.4406
Nkx2.2(Homeobox)/NPC-Nkx2.2-ChIP-Seq(GSE61673)/Homer	BTBRAGTGSN	-274.9	0.8148
FEA4(bZIP)/Corn-FEA4-ChIP-Seq(GSE61954)/Homer	TGACGTACCS	-263	0.5483
COUP-TFII(NR)/Artia-Nr2f2-ChIP-Seq(GSE46497)/Homer	AGRGGTCA	-257.2	0.7303
SUT1?/SacCer-Promoters/Homer	CCCCGCGC	-205.7	0.9507

RARa(NR)/K562-RARa-ChIP-Seq(Encode)/Homer	TTGAMCTTG	-198.2	0.9008
THRB(NR)/HepG2-THRB.Flag-ChIP-Seq(Encode)/Homer	GGTCACCTGAGGTCA	-159.4	0.4446
RIN(MADS)/Tomato-RIN-ChIP-Seq(GSE116581)/Homer	CYAAAAWWGG	-153.6	0.6319
Nkx2.1(Homeobox)/LungAC-Nkx2.1-ChIP-Seq(GSE43252)/Homer	RSCACTYRAG	-150.3	0.8709
Slug(Zf)/Mesoderm-Snai2-ChIP-Seq(GSE61475)/Homer	SNGCACCTGCHS	-146.3	0.382
Tbx5(T-box)/HL1-Tbx5.biotin-ChIP-Seq(GSE21529)/Homer	AGGTGTCA	-140.5	0.8724
ZEB2(Zf)/SNU398-ZEB2-ChIP-Seq(GSE103048)/Homer	GNMCAGGTGTGC	-129.2	0.4972
GAGA-repeat/SacCer-Promoters/Homer	CTYTCTYTCTCTCTC	-122.8	0.9163
E2A(bHLH),near _PU.1/Bcell-PU.1-ChIP-Seq(GSE21512)/Homer	NVCACCTGBN	-119.9	0.6404
Nkx3.1(Homeobox)/LNCaP-Nkx3.1-ChIP-Seq(GSE28264)/Homer	AAGCACTTAA	-119.2	0.8235
ZEB1(Zf)/PDAC-ZEB1-ChIP-Seq(GSE64557)/Homer	VCAGGTRDRY	-117.4	0.6786
E2A(bHLH)/proBcell-E2A-ChIP-Seq(GSE21978)/Homer	DNRCAGCTGY	-117.2	0.6339
GAGA-repeat/Arabidopsis-Promoters/Homer	CTCTCTCTCY	-113.9	0.533
Srebp2(bHLH)/HepG2-Srebp2-ChIP-Seq(GSE31477)/Homer	CGGTCACSCCAC	-111	0.1624
ZNF416(Zf)/HEK293-ZNF416.GFP-ChIP-Seq(GSE58341)/Homer	WDNCTGGCA	-104.2	0.6069
Nkx2.5(Homeobox)/HL1-Nkx2.5.biotin-ChIP-Seq(GSE21529)/Homer	RRSCACTYAA	-91.15	0.7889
Bcl11a(Zf)/HSPC-BCL11A-ChIP-Seq(GSE104676)/Homer	TYTGACCASWRG	-90.9	0.4169
Npas4(bHLH)/Neuron-Npas4-ChIP-Seq(GSE127793)/Homer	NHRTCACGACDN	-90.1	0.4462
At1g14580(C2H2)/colamp-At1g14580-DAP-Seq(GSE60143)/Homer	CASAAAAMGACAAAA	-83.4	0.1947
Srebp1a(bHLH)/HepG2-Srebp1a-ChIP-Seq(GSE31477)/Homer	RTCACSCCAY	-78.4	0.1822
HEB(bHLH)/mES-Heb-ChIP-Seq(GSE53233)/Homer	VCAGCTGBNN	-60.12	0.6795
ASHR1(ND)/col-ASHR1-DAP-Seq(GSE60143)/Homer	NTGGTGAN	-57.48	0.5859

Supplementary Table 10. MOTIFs enriched with corin treatment in 451Lu-R H3K4me2 ChIPseq.

Motif Name	Consensus	Log P-value	% of Target Sequences with Motif
Usf2(bHLH)/C2C12-Usf2-ChIP-Seq(GSE36030)/Homer	GTCACGTGGT	- 85.47	0.2711
Mef2a(MADS)/HL1-Mef2a.biotin-ChIP-Seq(GSE21529)/Homer	CYAAAAATAG	- 65.04	0.4921
Mef2c(MADS)/GM12878-Mef2c-ChIP-Seq(GSE32465)/Homer	DCYAAAAATAGM	- 64.62	0.5213
IDD5(C2H2)/colamp-IDD5-DAP-Seq(GSE60143)/Homer	TTTTGTCTTTTBTK	- 54.38	0.6152
GLIS3(Zf)/Thyroid-Glis3.GFP-ChIP-Seq(GSE103297)/Homer	CTCCCTGGGAGGCCN	- 41.06	0.7061
EAR2(NR)/K562-NR2F6-ChIP-Seq(Encode)/Homer	NRBCARRGGTCA	- 19.29	0.6261
VRN1(ABI3VP1)/col-VRN1-DAP-Seq(GSE60143)/Homer	TTTTTTTTTT	- 19.17	0.4213
Reverb(NR),DR2/Raw-Reverba.biotin-ChIP-Seq(GSE45914)/Homer	GTRGGTCASTGGGTCA	- 18.15	0.1083
p53(p53)/mES-cMyc-ChIP-Seq(GSE11431)/Homer	ACATGCCCGGGCAT	- 15.05	0.0162
IDD4(C2H2)/col-IDD4-DAP-Seq(GSE60143)/Homer	TTTGTCCTTWBT	- 14.83	0.6456
GSC(Homeobox)/FrogEmbryos-GSC-ChIP-Seq(DRA000576)/Homer	RGGATTAR	- 7.876	0.7121
COUP-TFII(NR)/K562-NR2F1-ChIP-Seq(Encode)/Homer	GKBCARAGGTCA	- 5.949	0.6393
SUT1?/SacCer-Promoters/Homer	CCCCGC	- 5.016	0.8883
REST-NRSF(Zf)/Jurkat-NRSF-ChIP-Seq/Homer	GGMGCTGTCCATGGTGCTGA	- 4.619	0.0043

Supplementary Table 11. Combinations Index (CI) for 451Lu-R cells treated with PLX4032 and p38i. CI data indicate the degree of drug synergy where CI >1 indicates drug antagonism, CI=1 indicates drug additivity, and CI <1 indicates drug synergy for cell proliferation inhibition across a range of PLX and p38i concentrations (72 h treatment).

PLX (μM)	p38i (μM)	Combination Index (CI)
5.0	0.5	1.82
5.0	1.0	0.93
5.0	5.0	0.27
5.0	10.0	0.27
5.0	100	1.18
5.0	1000	7.35

Supplementary Table 12. Combinations Index (CI) for 1205Lu-R cells treated with PLX4032 and p38i. CI data indicate the degree of drug synergy where CI >1 indicates drug antagonism, CI=1 indicates drug additivity, and CI <1 indicates drug synergy for cell proliferation inhibition across a range of PLX and p38i concentrations (72 h treatment).

PLX (μ M)	p38i (μ M)	Combination Index (CI)
5.0	0.5	4.72
5.0	1.0	1.76
5.0	5.0	0.20
5.0	10.0	0.10
5.0	100	0.22
5.0	1000	1.83

Supplementary Table 13. Antibody Information.

Antibody	Company	Catalog Number	Application	Dilution
H3K9ac	Abcam	ab32129	WB, ChIP	1:1000
H3K4me2	Abcam	ab32356	WB, ChIP, IF	1:5000
H3K27ac	Abcam	ab4729	WB, ChIP, IF	1:1000
Phospho-MEK1/2	Cell Signaling	9121	WB	1:1000
MEK1/2	Cell Signaling	9122	WB	1:1000
Total H3	Abcam	ab1791	WB	1:10000
DUSP1	Millipore	07-535	WB	1:1000
DUSP5	Abcam	ab200708	WB	1:1000
Phospho-c-Jun (Ser63)	Cell Signaling	9261s	WB	1:1000
c-Jun	Cell Signaling	9165s	WB	1:1000
Phospho-Erk	Cell Signaling	9101s	WB	1:1000
p44/42 MAPK (Erk1/2)	Cell Signaling	9102s	WB	1:1000
MITF	Cell Signaling	12590s	WB	1:1000
AXL	Cell Signaling	8661s	WB	1:1000
Cleaved PARP	Cell Signaling	9541s	WB	1:1000
Phospho-JNK	Cell Signaling	4668s	WB	1:1000
JNK	Cell Signaling	9252s	WB	1:1000
Phospho-p38 MAPK	Cell Signaling	4631s	WB	1:1000
p38 MAPK	Cell Signaling	9212s	WB	1:1000
GAPDH	Cell Signaling	2118L	WB	1:2000
p62	Cell signaling	88588s	WB	1:1000
LC3B	Cell signaling	2775S	WB	1:1000
Ki67	Abcam	ab15580	IF, IHC	1:500
Cleaved Caspase-3	Cell Signaling	9661s	IF, IHC	1:500
HIF1 alpha	Abcam	ab16066	IHC	1:500
Vinculin	Millipore	MAB3574	IF	1:300
HDAC1	Abcam	ab19845	ChIP	
LSD1	Abcam	ab129195	ChIP	
RCOR1	Millipore	07-455	ChIP	

Supplementary Table 14. List of primers and oligonucleotide sequences

RT-qPCR Primer Sequences		
Target	Forward Primer (5'->3')	Reverse Primer (5'->3')
<i>AXIN2</i>	GGATCACTGGCTCCCGCA	AGTCCTCTCAGCAATCGGC
<i>AXL</i>	CCAGGACACCCCAGAGGTGCTAAT	TGGTGGACTGGCTGTGCTTGC
<i>BIRC5</i>	CCACTGAGAACGAGCCAGACTTG	AGAAAGGAAAGCGCAACCGG
<i>BRCA1</i>	GAATTTATCGAGTGGCCAAAC	TCAAAGACTGACTGTTGTGG
<i>CCNA2</i>	GCATGTCACCCTCCTCCTT	GGGCATCTTCACGCTCTATT
<i>CDK2</i>	GCTTTGGAGTCCTGTTCG	GGTCCCAGAGTCCGAAAGA
<i>COL4A2</i>	CCTGAAGGCACAGCTAACCA	TGCTGTTGTCTCGTCTGTCC
<i>CRLF2</i>	ACTCCTGTTCAGGCATGGG	AGTCAGGTTGGCCTGGAGT
<i>DUSP1</i>	AGGACAACCACAAGGCAGAC	CAGTGGACAAACACCCTCC
<i>DUSP4</i>	CAAAGGCGGCTATGAG	GGTTATCTTCCACTGGG
<i>DUSP5</i>	GCCCCGGGTCTACTTCCTC	CTCGGAGGTCCGTCGGGAGA
<i>DUSP6</i>	CGAGACCCAATAGTGC	AATGGCCTCAGGGAAA
<i>E2F1</i>	AGGGGTGTGGGGTTGATACC	TCAGACACTGCAGGAGGGAC
<i>E2F2</i>	CCTTGAGGCTACTGACAGC	CCACAGGTAGTCGTCTGGT
<i>FGF5</i>	CTCTCTTCTCCCTCTCCCC	TAGGGCTGATTCTGGGCTCT
<i>FGFR2</i>	CCTGCGGAGACAGGTAACAG	GGTGTCTGCCGTGAAGAGA
<i>FOXO1</i>	GATCCCGTAAGTCGGGCGG	GCTGCTGCCTGTTGAATGTG
<i>GAPDH</i>	GGCTCTCCAGAACATCATCCCTGC	GGGTGTCGCTGTTGAAGTCAGAGG
<i>MITF</i>	GGAAATCTGGGCTTGATGGA	CCCGAGACAGGCAACGTATT
<i>MMP1</i>	AAGGCCAGTATGCACAGCTT	GGGCCACTATTCTCCGCTT
<i>MYC</i>	TCACCAAGACAACACTACGCG	CAGGATGTAGGCAGGTGGCTT
<i>NR4A1</i>	GCTACGAAACTTGGGGAGT	ATGTGGCTTGACCTGTTCT
<i>PDGFRB</i>	CAAGGACACCAGCGGCTTC	AGCAGGTAGAACGAAGGTG
<i>PLXNA4</i>	AATTCCCGTCTCACGGAGG	AGTCACTGTTGCTCTGTGGG
<i>RUNX1T1</i>	GTCAAGAACGAGACCGGGAA	CTAGTGCAACTGGGCTGGG
<i>SEMA4A</i>	CGGGATGGGGTTGAGAATGG	ATCATGCCAAGGCCAGCACT
<i>SEMA4D</i>	CTGGGCTCATCTCTAGCAC	CTCTCACCACCGCAATGTCA
<i>VCL</i>	GCTGCTGTTAACGCCATCCAA	CCAACCGAGTCACCTCATCT
<i>WNT11</i>	GTGAAGGACTCGGAACTCGT	GTCGCTCCGTTGGATGTCT
CHIP-qPCR Primer Sequences		
Target	Forward Primer (5'->3')	Reverse Primer (5'->3')
Upstream <i>DUSP1</i>	TGAGAGGTGGGCCTCAGTTA	GAGGTTGCAGTGAGCTGAGA
shRNA Sequences		
Target	Target Sequence (5'->3')	
RCOR1 Sh1	GATGGTCCAATAGAACCATAT	
RCOR1 Sh2	CAAACGACAGATCCAGAACATAT	
DUSP1 Sh1	GCCGCTCCTTCTCGCTTCA	
DUSP1 Sh2	ACGAGGCCATTGACTTCATAG	
DUSP1 Sh3	GAGGGTCACTACCAGTACAAG	
DUSP1 Sh4	GCTGATTATTATGACCTGAA	
DUSP1 Sh5	GCTCTGTCAACGTGCGCTTCA	