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965 Graphical Abstract

966 Supplemental Figure



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968 Supplementary Figure1: Effect of CTRP4 on T cell subsets

969 (A) Flow cytometry analyses of Th2 effector T cells (CD4⁺CD44⁺ IL-4⁺ cells) in the spleen of *Ctrp4^{-/-}*970 and WT mice.

971 **(B)** Flow cytometry analysis of Treg cells ($CD4^+CD25^+Foxp3^+$) in the spleen of $Ctrp4^{-/-}$ and WT mice.

972 (C) Gene expression levels of *Gata3, Foxp3* or *Tbx21* in CD4⁺ T cells were analyzed by quantitative 973 real-time PCR.

974 (D) Gene expression of *Ctrp4* was shown as summary bar graph. Naive CD4⁺CD62L^{high}CD44^{low}CD25⁻

975 T cells were sorted from WT mice were stimulated with anti-CD3 and anti-CD28 antibodies to obtain

976 effector CD4⁺ T cells, or differentiated towards Th subsets under Th0/1/2/17 or Treg differentiation

977 condition. Real-time RT-PCR of *Ctrp4* mRNA expression was analyzed and normalized against *Gapdh*.

- 978 (E) The productions of CTRP4 of WT CD4⁺T cells under Th0 or Th17 differentiation conditions were
- 979 quantitated by ELISA, respectively. Data were shown as mean \pm SEM and were from one of three

980 independent experiments with similar results. one-way ANOVA with Tukey's post-test was used for **D**. 981 Statistical significance was determined using unpaired Student t test or Mann-Whitney U test for **A-C** 982 and **E**; **p < 0.01, ***p < 0.001, ns not significance.

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Supplementary Figure2: *Ctrp4* deficiency exacerbates EAE progression with increased
 infiltration of CD4⁺IL-17A⁺T cells in the peripheral.

987 (A) Representative immunohistochemistry images of CD4 expression in the spinal cord of indicated EAE
 988 animals at the peak of disease were performed. Scale bar,100 μm.

989 **(B)** The summary bar graph showed the absolute number of $CD4^+$ in spleens and draining lymph nodes 990 of *Ctrp4^{-/-}* and WT mice at the peak of disease.

991 (C-D) Flow-cytometric analysis of the frequencies (C) or the absolute cell numbers (D) or of CD4⁺IL-992 17^+ , CD4⁺IL- 17^+ IFN γ^+ , CD4⁺IFN γ^+ cells in the spleens harvested from WT and *Ctrp4^{-/-}* mice at day18

993 postimmunization.

994 (E-F) Flow-cytometric analysis of the frequencies (E) or the absolute cell numbers (F) of CD4⁺IL-17⁺, 995 CD4⁺IL-17⁺IFN γ^+ , CD4⁺IFN γ^+ cells in the draining lymph nodes isolated from WT and *Ctrp4^{-/-}* mice at 996 day18 postimmunization. Data were shown as mean ± SEM and were from one of three independent 997 experiments with similar results. Statistical significance was determined using unpaired Student t test or 998 Mann-Whitney U test; *p < 0.05; **p < 0.01, ns not significance. 999

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1001 Supplementary Figure3: The deletion efficacy of T cell condition CTRP4 KO mice

- 1002 (A) Genotyping by PCR analysis of CTRP4^{flox/flox}, CTRP4^{+/+} and heterozygous mice or PCR analysis of
- 1003 CD4 Cre transgene mice with primers designed for indicated sites.
- 1004 **(B)** CD4⁺T cells from CTRP4^{flox/flox}, CTRP4^{+/flox}, CD4creCTRP4^{flox/flox}, or CD4creCTRP4^{+/+} mice were
- 1005 purified and lysates were subjected to western blot analysis for CTRP4 protein expression.
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Supplementary Figure 4: CTRP4 deletion did not affect the ability of Th17 cells to activate ormigrate into CNS

1010 **(A)** Representative flow cytometric analysis and quantification of CCR6/CCR2 expression in CD4⁺T 1011 cells from $Ctrp4^{-/-}$ and WT in the dLNs, spleens or CNS on day18 post EAE induction.

1012 (B) Representative flow cytometric analysis and quantification of CD49d and CD29 expression in CD4⁺

1013 T cells from spleens, lymph nodes and CNS at peak stage of disease.

1014 **(C)** Quantitative PCR analysis to determine the expression levels of indicated genes encoding multiple 1015 chemokines in spinal cord of MOG₃₅₋₅₅-immunized *Ctrp4*^{-/-} and WT at the peak of disease.

1016 **(D)** Comparable activation status of CD4⁺ T cells between $Ctrp4^{-/-}$ and WT mice on day18 post EAE 1017 induction were analyzed by flow cytometry. Data were shown as mean \pm SEM and were from one of 1018 three independent experiments with similar results. Statistical significance was determined using 1019 unpaired Student t-test or Mann-Whitney U test.



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Supplementary Figure 5: The in vitro differentiation ability of naïve CD4⁺ T cells into Treg cells was not impaired in CTRP4 KO mice

(A) Naïve CD4⁺CD62L^{high}CD44^{low}CD25⁻ T cells were sorted from WT and Ctrp4^{-/-} mice, and
 differentiated with 5 ng/ml TGFβ1 and 5 ng/ml IL-2 for 5 days. Numbers adjacent to outlined areas
 indicated the percentage of CD4⁺CD25⁺Foxp3⁺ cells.

1026(B) CFSE-labeled effector CD4+ T cells were cocultured with WT and $Ctrp4^{-/-}$ Treg cells to conduct the1027Treg suppression assay. The suppression capacity was determined through CFSE dilution when effector1028T cells cultured at a 4:1 ratio with Treg cells. Data were from one of three independent experiments with

1029 similar results.

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1032 Supplementary Figure 6: Ctrp4 deficiency did not alter the expression level of IL-6R

(A) The representative histograms showed the expression level of IL-6R on purified wild-type and *Ctrp4⁻* 1034 ^{/-} naïve CD4⁺T cells stimulated with anti-CD3 and anti-CD28 for indicated time.

(B)Flow cytometric analysis of the mean fluorescence intensity (MEI) of IL-6R on purified wild-type
 and *Ctrp4^{-/-}* naïve CD4⁺T cells stimulated with anti-CD3 and anti-CD28 for indicated time. Isotype
 means isotype-matched control antibody.

1038 (C) Flow cytometric analysis of the mean fluorescence intensity (MEI) of gp130 or IL-6R on CD4⁺ T 1039 cells isolated from CNS and peripheral lymphoid organs of $Ctrp4^{-/-}$ and WT mice on day18 post EAE 1040 induction. Data were shown as mean \pm SEM and were from one of three independent experiments with 1041 similar results. Statistical significance was determined using unpaired Student t-test or Mann-Whitney 1042 U test.

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Supplementary Figure 7: CTRP4 responded to IL-6 rather than other gp130 family cytokines
(A) Purified CD4⁺T cells from *Ctrp4^{-/-}* and WT mice were stimulated with IL-27 (50 ng/mL) for indicated
time. Lysates were subjected to western blot analysis for phosphorylated JAK2, p-JAK2, p-STAT3,
STAT3 and GAPDH (as a control). The samples derived from the same experiment and that gels/blots
were processed in parallel.

(B) The expression levels of gp130 and IL-6R in Ba/F3-gp130-IL-6R cells lysates were analyzed
by indicated antibodies (up). The expression level of gp130 in Ba/F3-gp130 cells lysates was
analyzed by anti-GFP antibodies (bottom).

- 1053 (C) Flow cytometric analysis of the expression of gp130 in Ba/F3-gp130 cells. Data were from one
- 1054 of three independent experiments with similar results.
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Gene	Forward Primer	Reverse Primer
Il17a	CTCCAGAAGGCCCTCAGACTC	GGGTCTTCATTGCGGTGG
Ifng	TCGAATCGCACCTGATCACTA	GGGTTGTTCACCTCGAACTTG
Rorc	CCGCTGAGAGGGCTTCAC	TGCAGGAGTAGGCCACATTAC
1117f	CCCCATGGGATTACAACATCC	CATTGATGCAGCCTGAGTGTT
II23r	AACATGACATGCACCTGGAA	TCCATGCCTAGGGAATTGAC
Foxp3	CCCATCCCCAGGAGTCTTG	ACCATGACTAGGGGGCACTGTA
Tbx21	GCCAGGGAACCGGTTATATG	GACGATCATCTGGGTCACAT
Gata3	AAGCTCAGTATCCGCTGACG	GTTTCCGTAGTAGGACGGGAC
116ra	CATTGCCATTGTTCTGAGGTTC	AGTAGTCTGTATTGCTGATGTC
Gapdh	GACTTCAACAGCAACTCCCAC	TCCACCACCCTGTTGCTGTA
Ccl2	CCGGCTGGAGCATCCACGTGT	TGGGGTCAGCACAGACCTCTCTCT
Ccl20	CGACTGTTGCCTCTCGTACA	GAGGAGGTTCACAGCCCTTT
Cxcl1	CACAGGGGGCGC CTATCGCCAA	CAAGGCAAGCCTCGCGACCAT
Cxcl2	ACCCCACTGCGCCCAGACAGAA	AGCAGCCCAGGCTCCTCCTTTCC
Ccl3	TGTACCATGACACTCTGCAAC	CAACGATGAATTGGCGTGGAA
Cx3cl1	ACGAAATGCGAAATCATG TGC	CTGTGTCGTCTCCAGGACAA
Ccl9	CCCTCTCCTTCCTCATTCTTACA	AGTCTTGAAAGCCCATGTGAAA
Cxcl9	TCCTTTTGGGCATCATCTTCC	TTTGTAGTGGATCGTGCCTCG
Cxcl11	GGCTTCCTTATGTTCAAACAGGG	GCCGTTACTCGGGTAAATTACA

Supplementary Table1: Primer sequence used for qPCR assays.