

Figure S1: Validation of PLK1 detection and examination of *Il1b* expression and IL1 β production under PLK1 inhibition.

(A) *Rosa^{CreErt2/wt}Plk1^{fl/fl}* BMDMs were treated with 4-OH Tamoxifen (0.002mg/mL 4-OH Tam, 24 hours), and cells treated without 4-OH Tam were used as controls. Cells were then stained with PLK1 for flow cytometry analysis. Isotype antibody control (Ctrl) and secondary antibody control were used. n = 4, 4, 3, 3 and 3, in order from the left column to the right. (B) Primed (100ng/mL LPS) NLRP3 knock-out (KO) BMDMs were activated (5mM ATP, 30 minutes), with or without selective PLK1 inhibitors (Cyclapolin9 (C9) 3 μ M; SBE13 10nM; Ro3280 50nM; BI6727 0.8nM) at the activation stage. The supernatants were collected to measure IL1 β concentration (n=5/group), cell death (n=6/group) and TNF α concentration (n=5/group). Treated samples from wide-type (WT) BMDMs without selective PLK1 inhibitors were used as control. (C) *Rosa^{CreErt2/wt}Plk1^{fl/fl}* BMDMs were treated with 0.002mg/mL 4-OH Tam for 24 hours before and during priming (100ng/mL LPS, 5 hours), and then cells were activated (5mM ATP, 30 minutes) with or without selective PLK1 inhibitors (3 μ M Cyclapolin9 (C9); 10nM SBE13; 50nM Ro3280; 0.8nM BI6727). The supernatants were collected to measure IL1 β concentration, cell death and TNF α concentration (n=4/group). Cells without 4-OH Tam treatment were used as control. (D) Murine BMDMs primed with low (L, 100ng/mL) or high (H, 500ng/mL) dose of LPS, were co-treated with the PLK1 inhibitor BI6727 at low (L, 0.4nM) or high (H, 0.8nM) dose. *Il1b* relative expression was quantified by qPCR (n = 2, 3, 3, 3, 3, 3, 3 and 3, in order from the left column to the right). Results are representative of 2 (A-C) or 3 (D) independent experiments. One-way ANOVA with Tukey post hoc test (A-C) and two-way ANOVA with Sidak post hoc test (D) were used for statistical analysis. All data are mean \pm SEM.

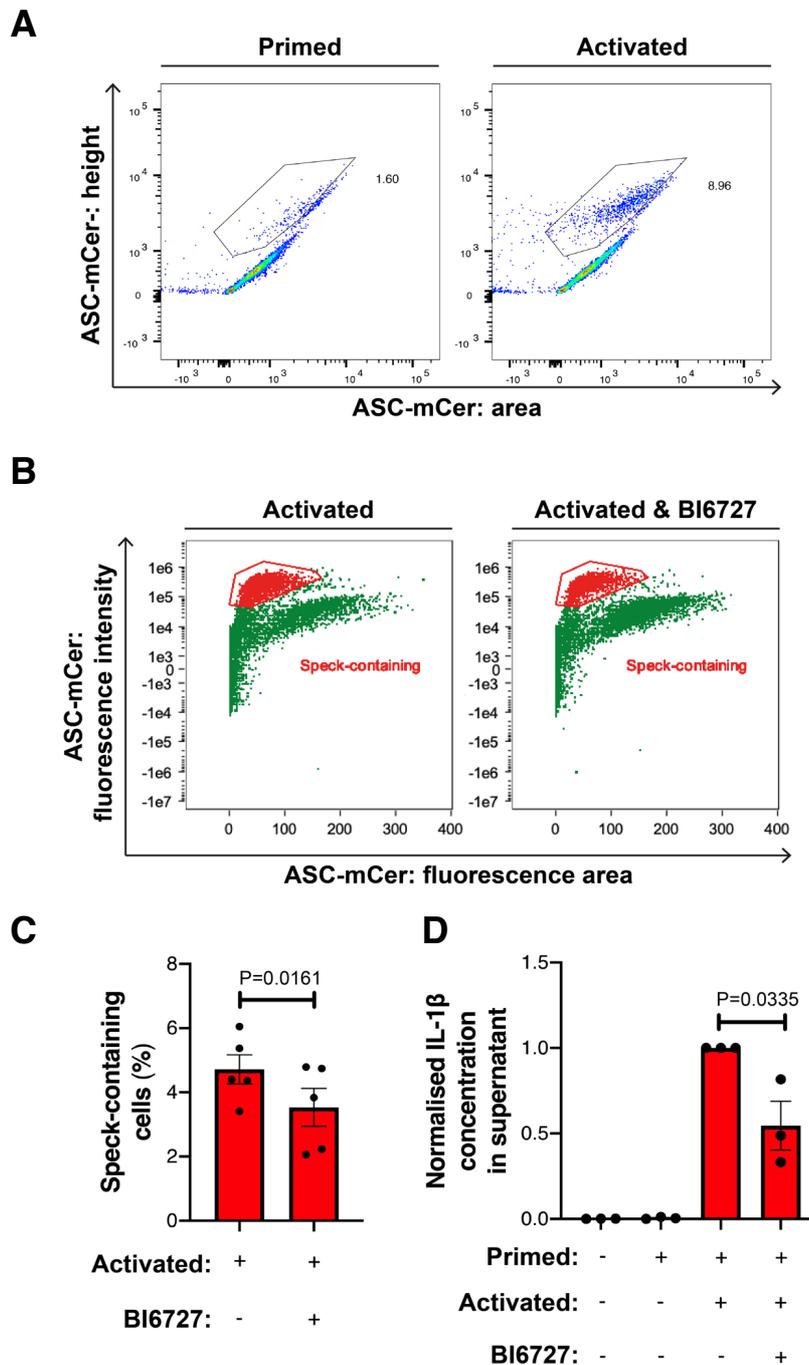


Figure S2: Validation of PLK1 effect on speck formation.

(A) mCerulean fluorescence area and height were measured on a flow cytometer to confirm speck detection, in immortalized BMDMs (iBMDMs) expressing ASC-mCerulean. These cells were primed (100ng/mL LPS, 5 hours) and activated (5 μ M nigericin, 2 hours). Cells without activation (primed) were used as controls. (B, C, D) iBMDMs expressing ASC-mCerulean were primed (100ng/mL LPS, 5 hours), and then activated (2.5 μ M nigericin, 30 minutes) under PLK1 inhibition (0.8nM BI6727). ASC specks were identified and quantified by imaging flow cytometry. Representative scatter plots (B) and quantification (C). Supernatants were collected for IL1 β quantification by ELISA (D). Cells without PLK1 inhibition were used as control. Results are representative of 3 independent replicates (A, B), or represent pooled data from 5 independent replicates (C). In (D), results represent pooled data from 3 independent biological replicates (each replicate comprises all 4 experimental conditions), and normalized IL1 β concentration was used to account for differences in plated cell numbers across different biological replicates. Paired t-test was used for statistical analysis (C, D). All data are mean \pm SEM.

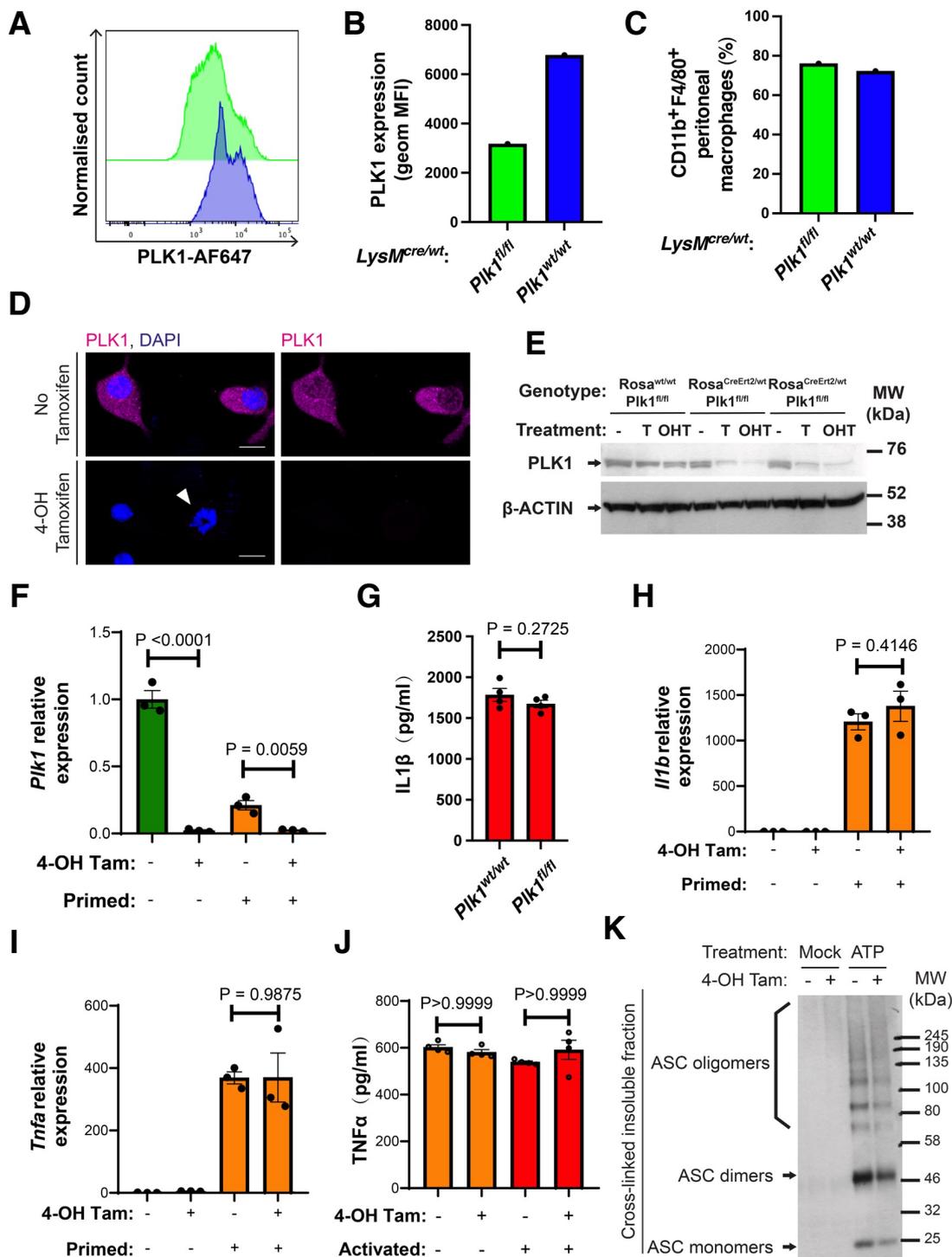


Figure S3: Validation of PLK1 genetic depletion and its effect on NLRP3 inflammasome activation.

(A-C) Peritoneal macrophages were elicited with 1mL of 3% thioglycolate (i.p.) for 4 days and isolated by AutoMACS separation. Cells were then analyzed on a flow cytometer to quantify PLK1 levels (A, B), and to evaluate purity (C). Control, *LysM^{cre/wt} Plk1^{wt/wt}*; deletion, *LysM^{cre/wt} Plk1^{fl/fl}*. (D) *Rosa^{CreErt2/wt} Plk1^{fl/fl}* BMDMs were treated with 4-OH Tamoxifen (0.002mg/mL 4-OH Tam, 24 hours), and then stained for PLK1 to assess protein levels. Arrowhead indicates a monopolar spindle of an arresting cell in mitosis, a phenotype of effective PLK1 inhibition. Scale bar = 10µm. (E) *Rosa^{CreErt2/wt} Plk1^{fl/fl}* and *Rosa^{wt/wt} Plk1^{fl/fl}* BMDMs were treated with 0.002mg/mL tamoxifen (T) or 0.002mg/mL 4-OH tamoxifen (OHT) for 24 hours, and cell lysates were then analyzed by western blot. (F, H, I) *Rosa^{CreErt2/wt} Plk1^{fl/fl}* BMDMs were treated with 0.002mg/mL 4-OH Tam for 24 hours before and during priming (100ng/mL LPS, 5 hours) to deplete PLK1. *Plk1* (F), *Il1b* (H) and *Tnfa* (I) mRNA relative expression was quantified by qPCR. n=3/group. (G) BMDMs derived from *Plk1^{fl/fl}*, and *Plk1^{wt/wt}* mice primed with LPS (100ng/mL, 5 hours) and activated with ATP (5mM, 30 minutes). Supernatants were collected for IL1β quantification by ELISA. n=4/group. (J) Supernatants from *Rosa^{CreErt2/wt} Plk1^{fl/fl}* BMDMs treated as in (F) were collected for TNFα quantification by ELISA. n=4/group. (K) *Rosa^{CreErt2/wt} Plk1^{fl/fl}* BMDMs were treated with 0.002mg/mL 4-OH Tam for 24 hours before and during priming (100ng/mL LPS, 5 hours) and then activated (5mM ATP, 30 minutes). Cross-linked insoluble protein fractions from those cells were analyzed by western blot to detect ASC oligomer formation. Results are representative of 3 (A, B, C) or 2 (F, H, I) independent experiments. Two-way ANOVA with Sidak post hoc test (F, H, I, J, K), and unpaired t test (G) were used for statistical analysis. All data are mean ± SEM.

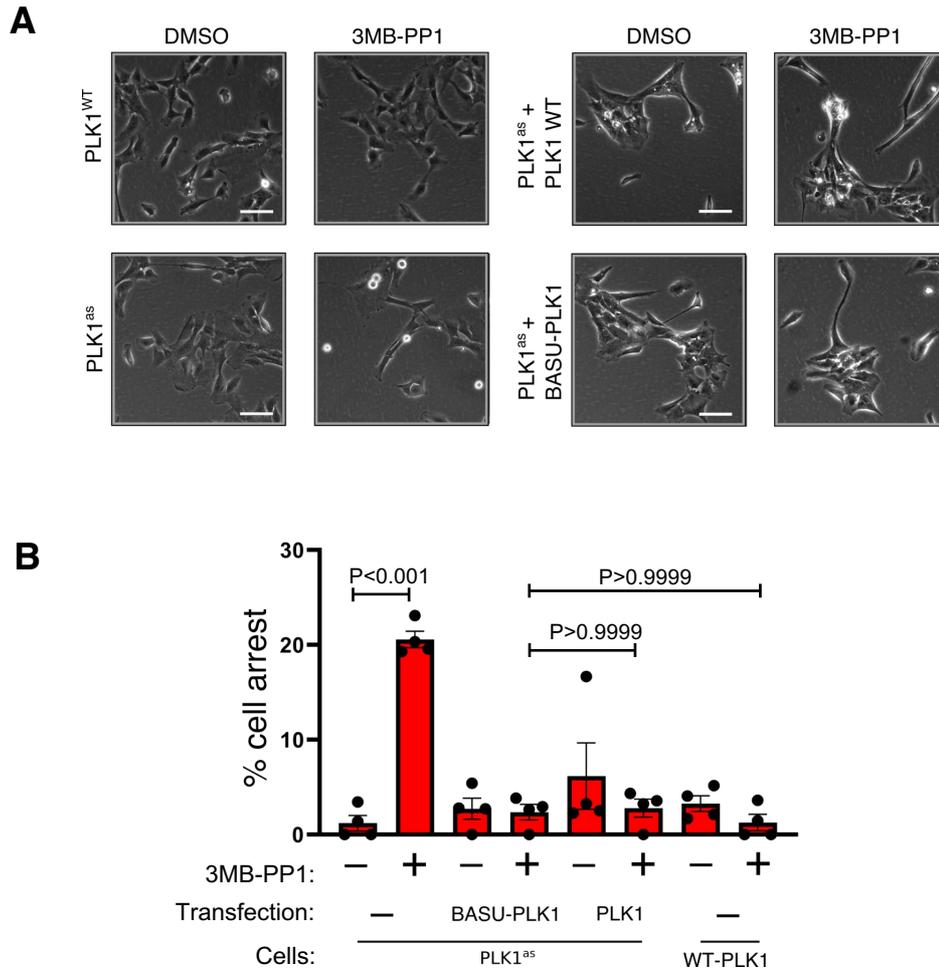


Figure S4. Validation of BASU-tagged PLK1.

tRPE PLK1^{as} cells were transfected with wild-type PLK1 or BASU-tagged PLK1 as indicated. tRPE PLK1^{as} cells were incubated with 3MB-PP1 as indicated to inhibit PLK1, and wild-type tRPE cells were used as controls. (A) Representative views used to count round cells in cell cycle arrest. Scale bar= 50 μ m. (B) Quantitative data of cells in cell cycle arrest (%). n=4/group. Two-way ANOVA with Sidak post hoc test was used. All data are mean \pm SEM.

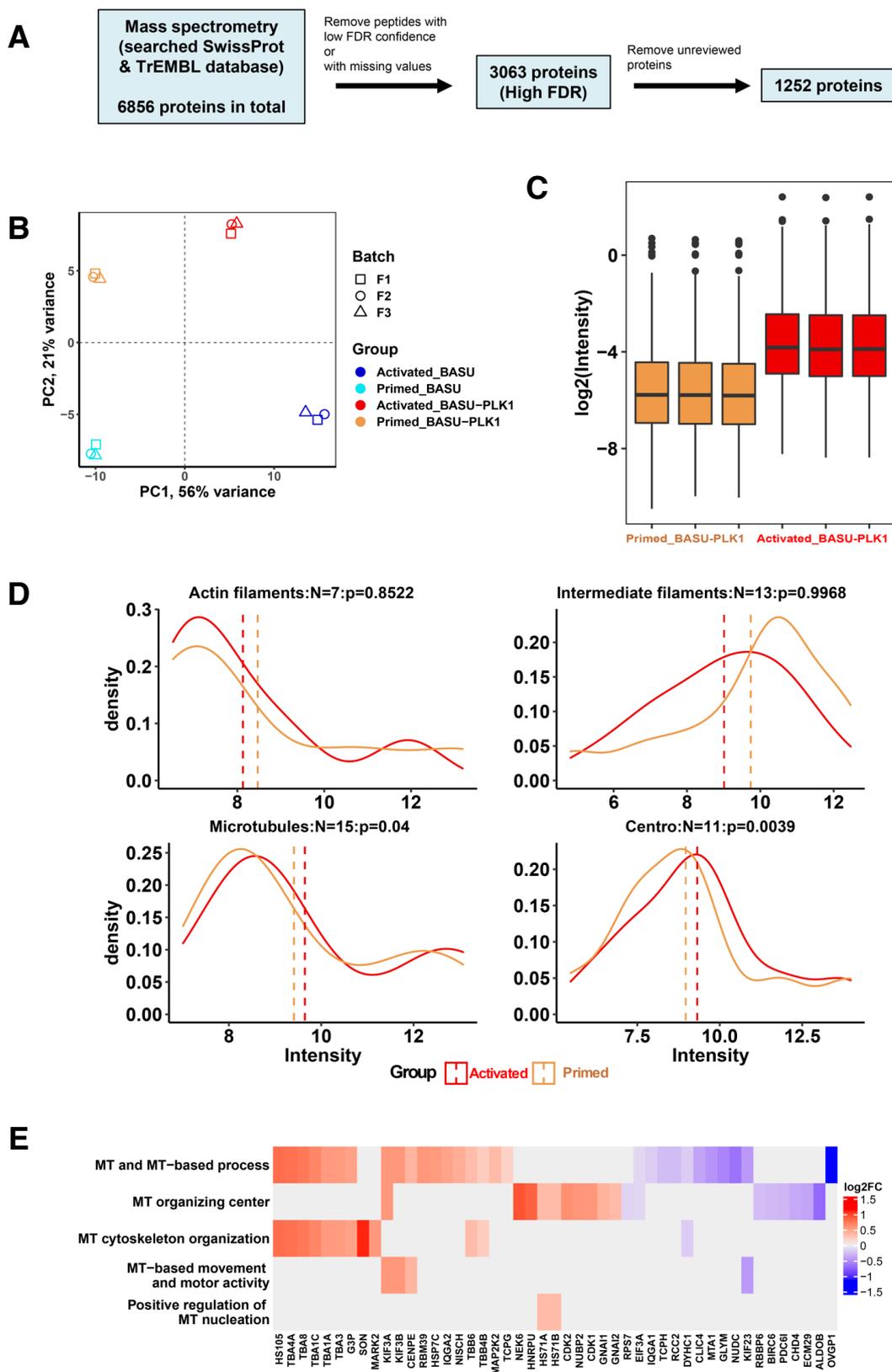


Figure S5. Bio-ID identified increased interactome with PLK1 after NLRP3 inflammasome activation.

(A) The analyzed interacting proteins which have passed the quality control are shown in the flow chart. (B) The interactome was analyzed under primed condition (primed), or after NLRP3 inflammasome activation (activated). PCA (Principal Components Analysis) shows the grouped replicates under each condition (BASU only under Primed or Activated condition; BASU-PLK1 under Primed or Activated condition). (C) Boxplot of normalized intensities of interacting proteins with PLK1 under primed condition or activated condition upon normalization using central median then PLK1 in each batch. (D) Density plot shows intensity shift of interacting proteins with PLK1 under primed or activated conditions at four subcellular cytoskeletal localization. Paired t-test for each location group is performed. (E) Heatmap plot shows the up (red shades) /down (blue shades)-regulated interacting proteins with PLK1 in microtubule-related pathways after NLRP3 inflammasome activation.

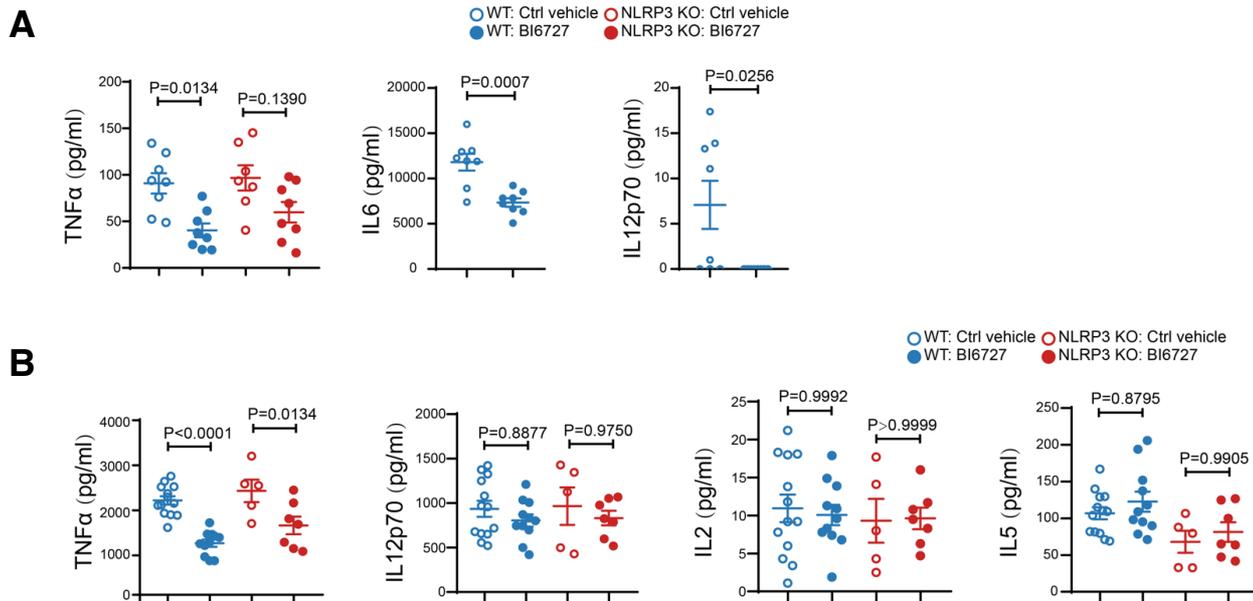


Figure S6. The effect of PLK1 inhibition on cytokines in LPS-induced endotoxemia model.

C57BL/6 wild-type (WT) and NLRP3 knock-out (KO) mice were treated with BI6727 (5mg/kg, i.p.) or control vehicle, followed by LPS administration (20mg/kg, i.p.). Samples for cytokine measurement were collected at 3 hours after LPS administration. **(A)** Peritoneal lavage fluid samples were collected, and multiple cytokine levels were measured. IL6 and IL12 were undetectable in NLRP3 KO samples WT: Ctrl vehicle, n=8; WT: BI6727, n=8; NLRP3 KO: Ctrl vehicle, n=7; NLRP3 KO: BI6727, n=8. **(B)** Serum samples were collected, and multiple cytokine levels were measured. WT: Ctrl vehicle, n= 13; WT: BI6727, n=11; NLRP3 KO: Ctrl vehicle, n=5; NLRP3 KO: BI6727 group, n=7. Unpaired t-test was used to compare two groups, and two-way ANOVA with Sidak post hoc test was used to compare multiple groups for statistical analysis. Non-parametric test was used to analyze the data of IL12p70 level in **(A)**. All data are mean \pm SEM.

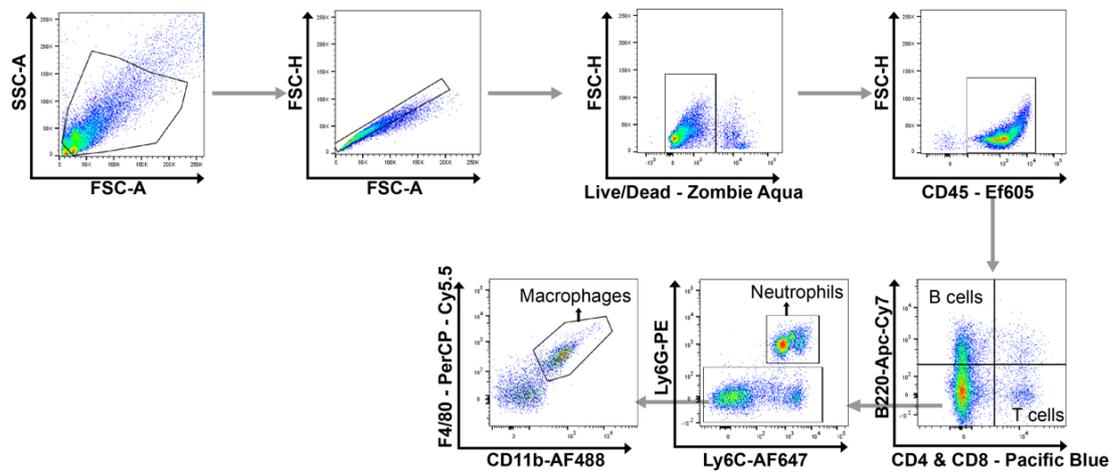


Figure S7. The gating strategy when examining the effect of PLK1 inhibition on immune cells in MSU-induced peritonitis model.

C57BL/6 wild-type (WT) and NLRP3 knock-out (KO) mice were treated with BI6727 (1mg/kg, i.v.) or control vehicle, followed by MSU (0.5mg/mouse, i.p.) for 6 hours. The cells were collected from the peritoneal cavity and analyzed on a flow cytometer. The gating strategy was shown.

Full unedited gel for all Figures

Figure 2D

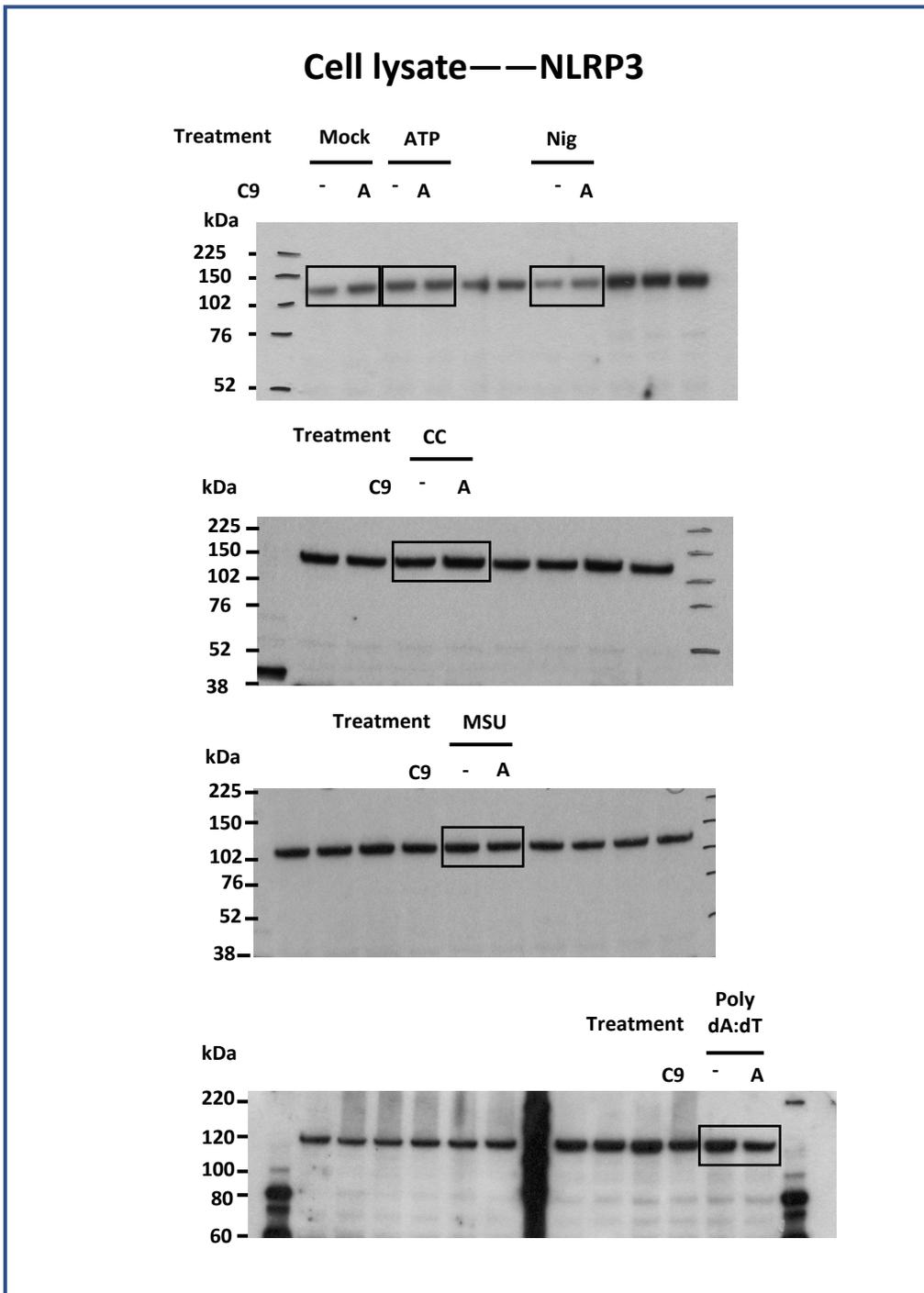


Figure 2D

Cell lysate—Pro-Casp1

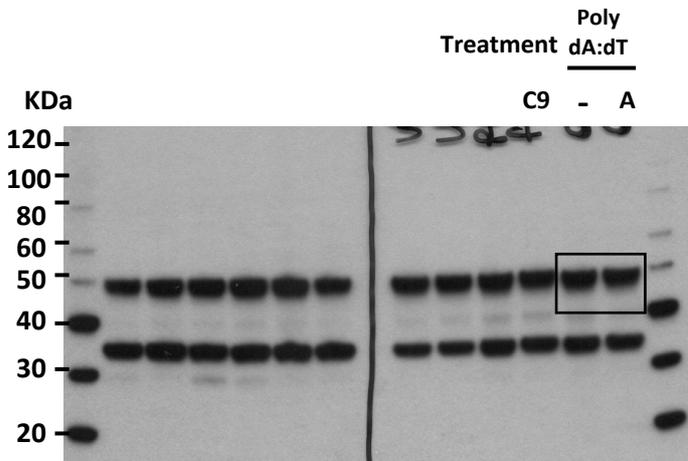
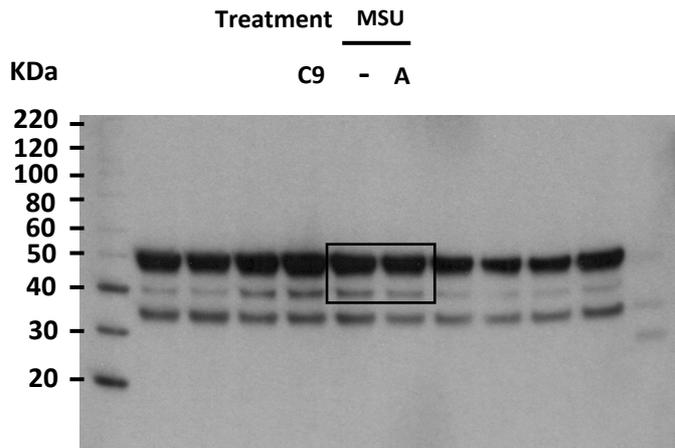
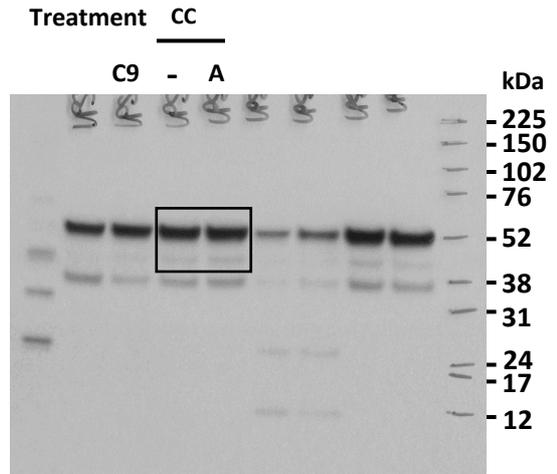
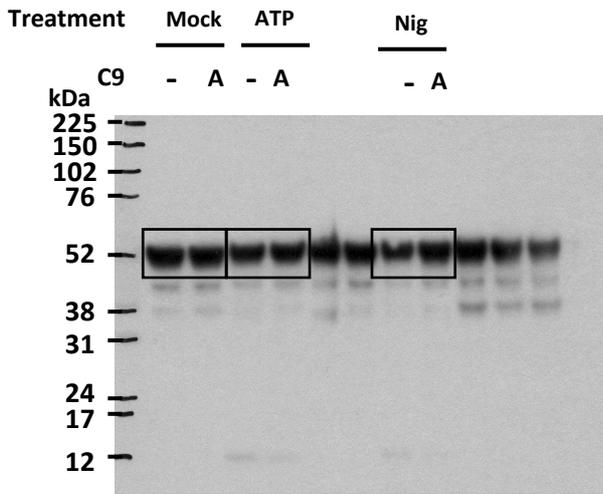


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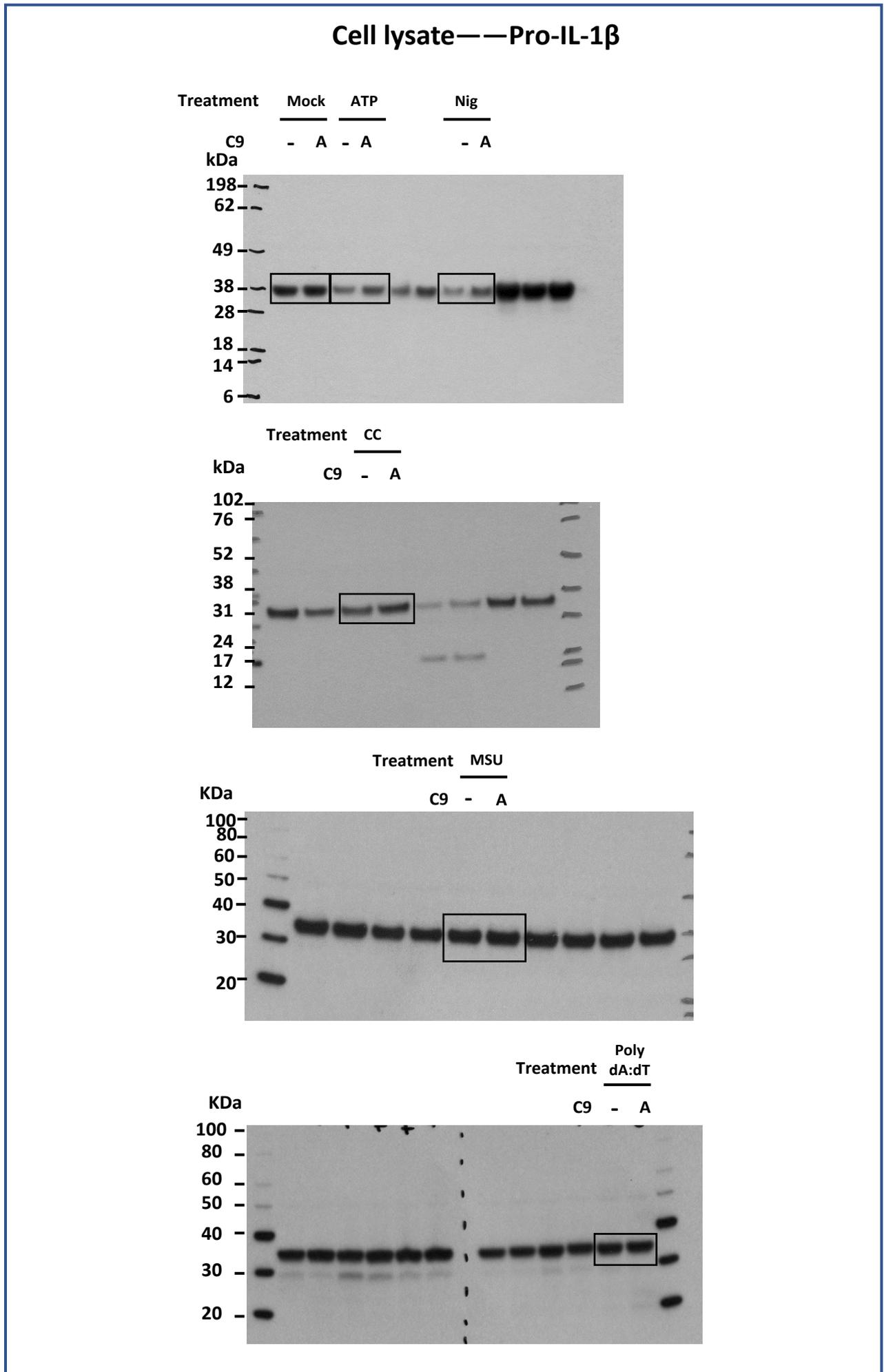


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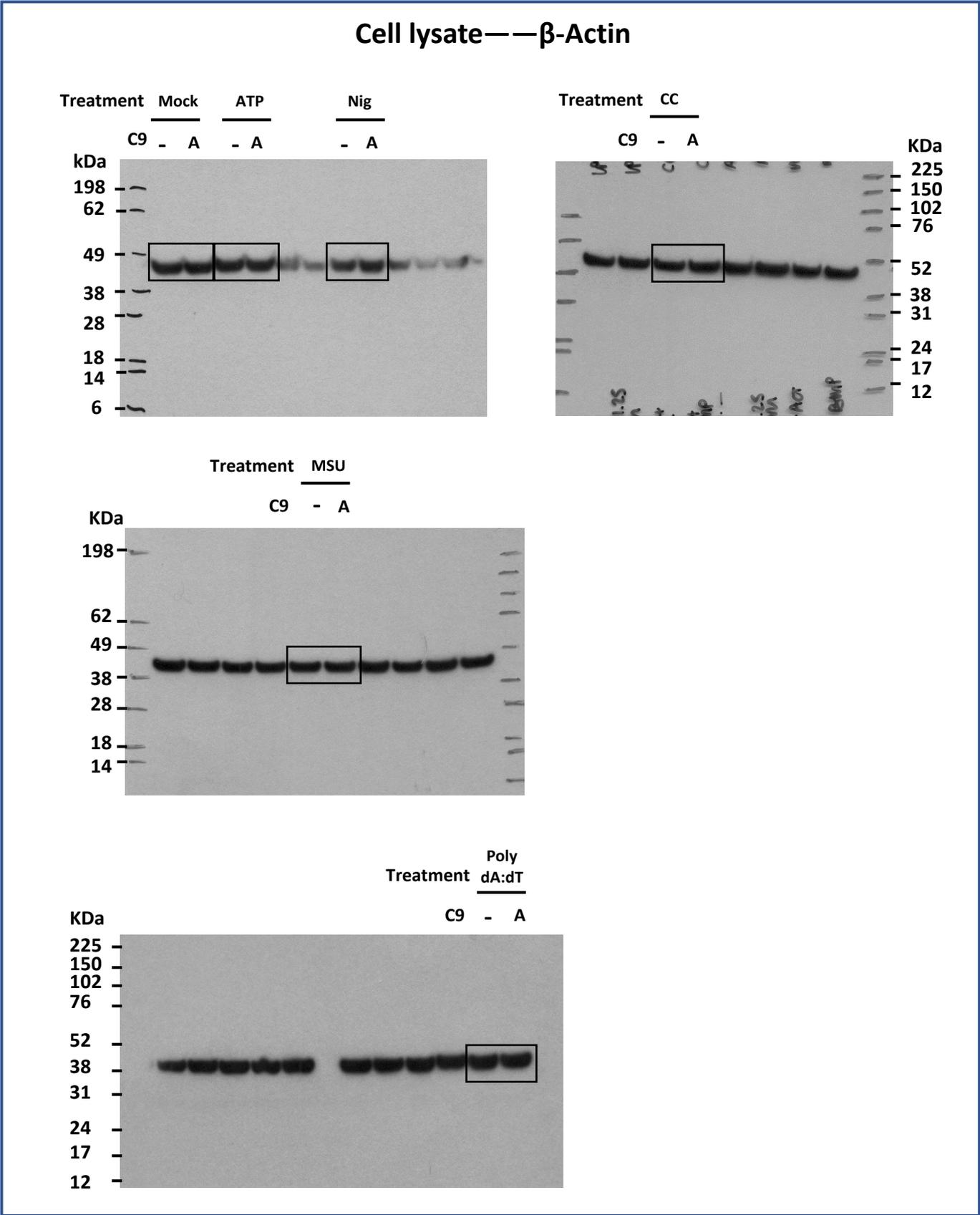


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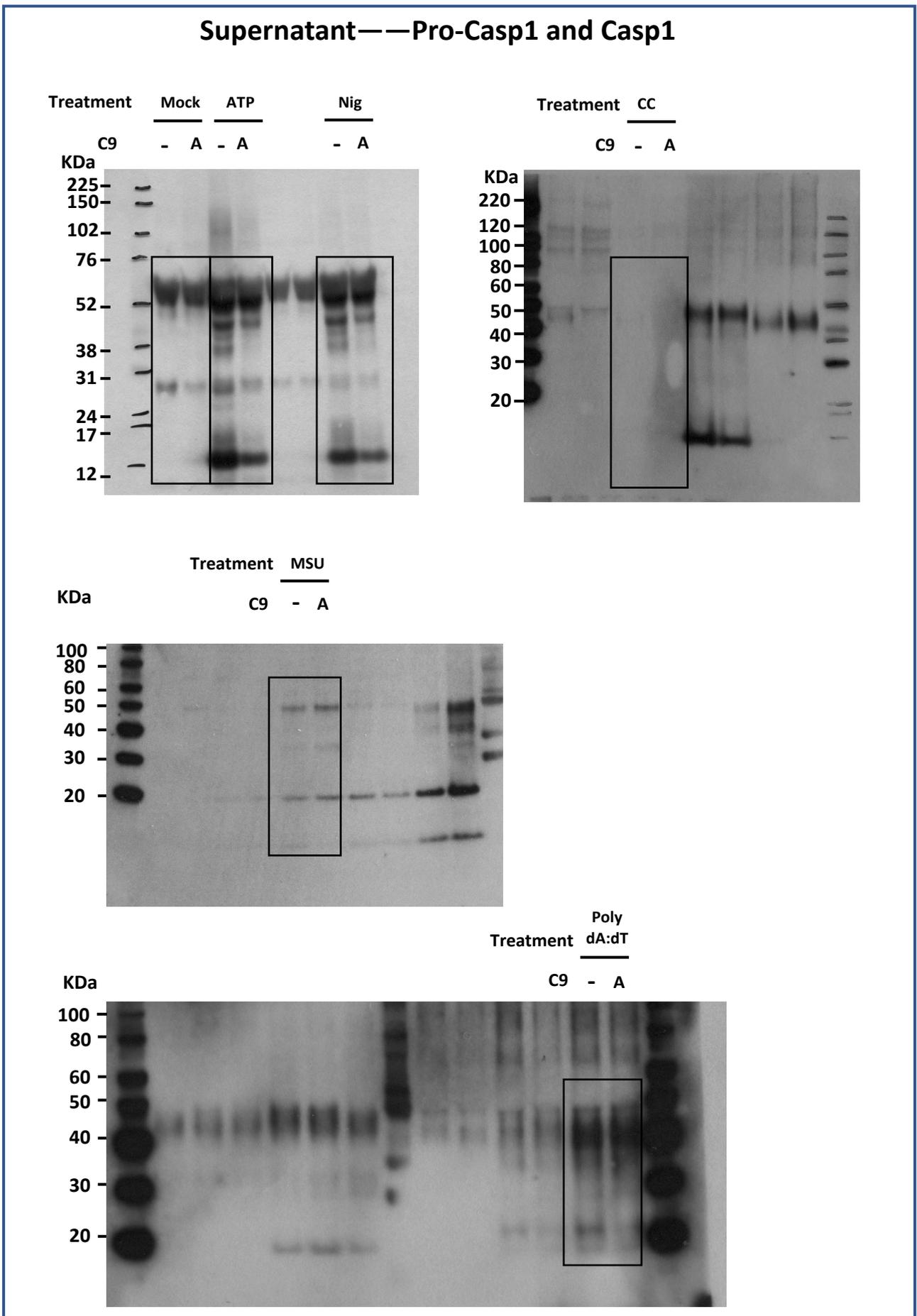


Figure 2D

Supernatant—Pro-IL-1 β and IL-1 β

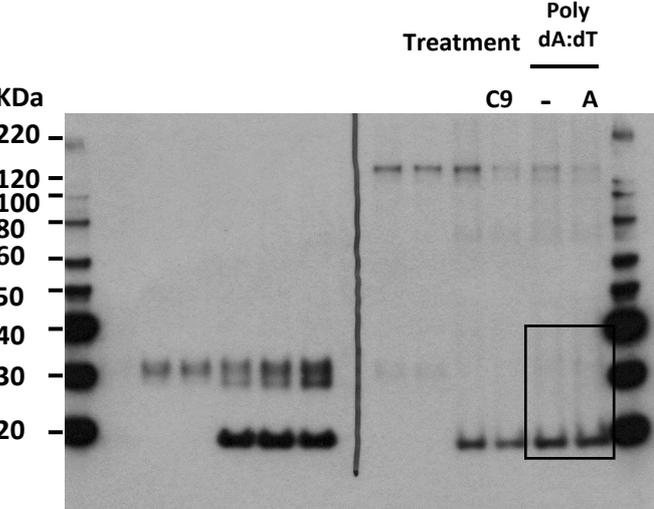
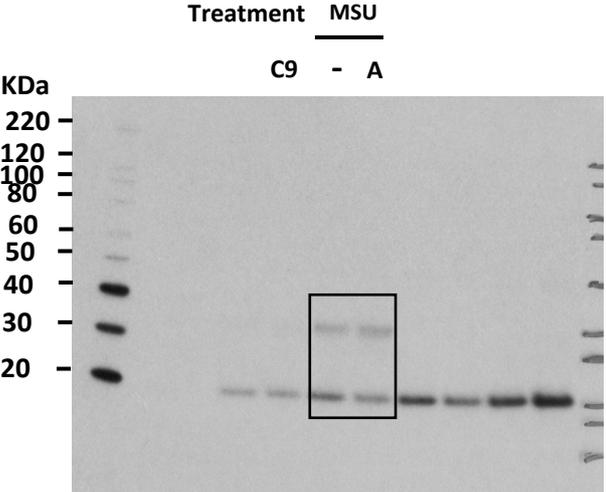
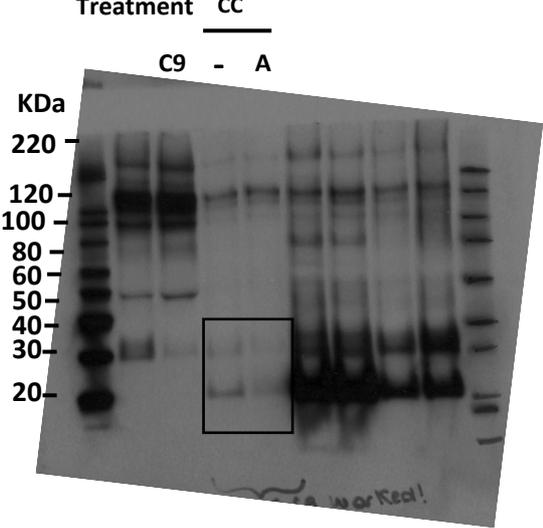
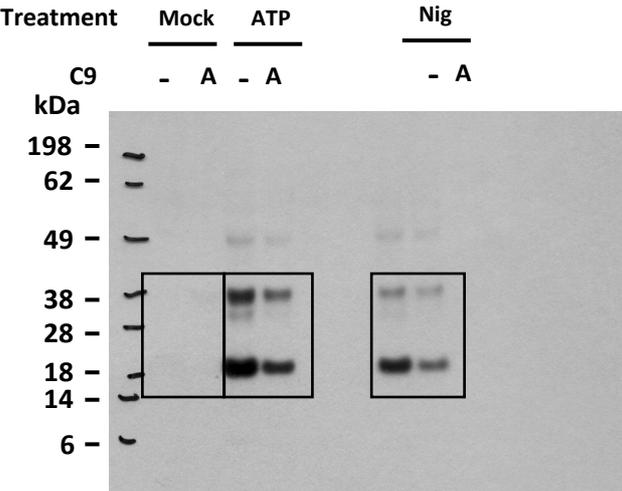


Figure 2G

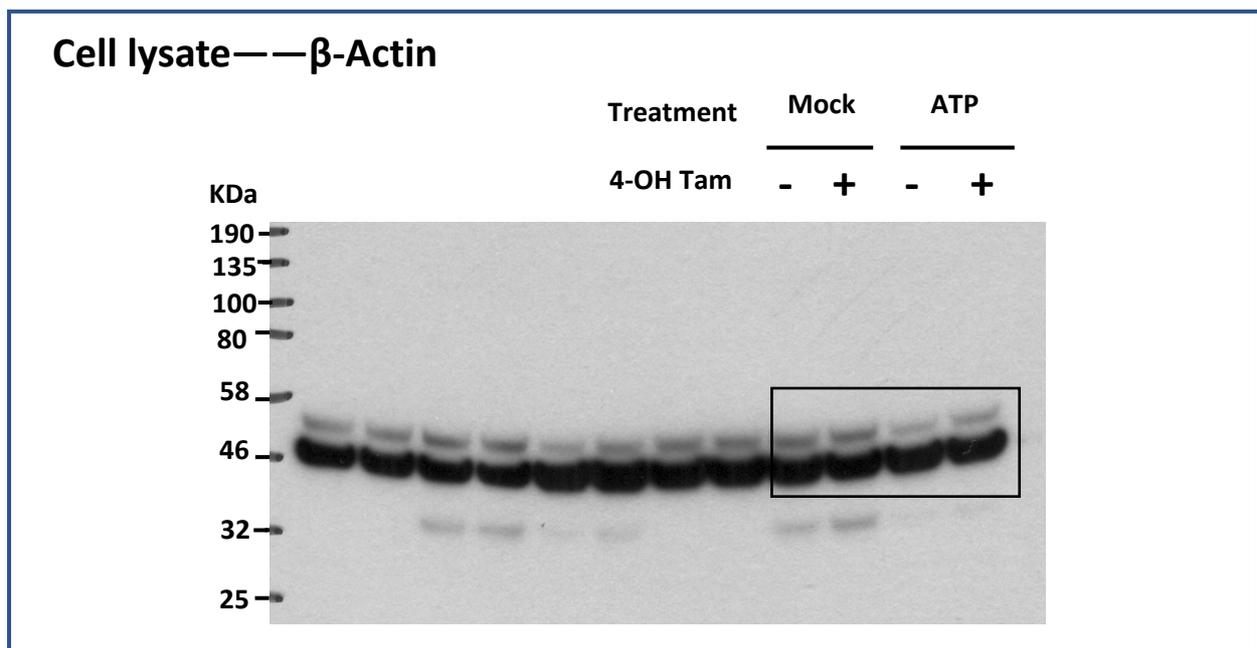
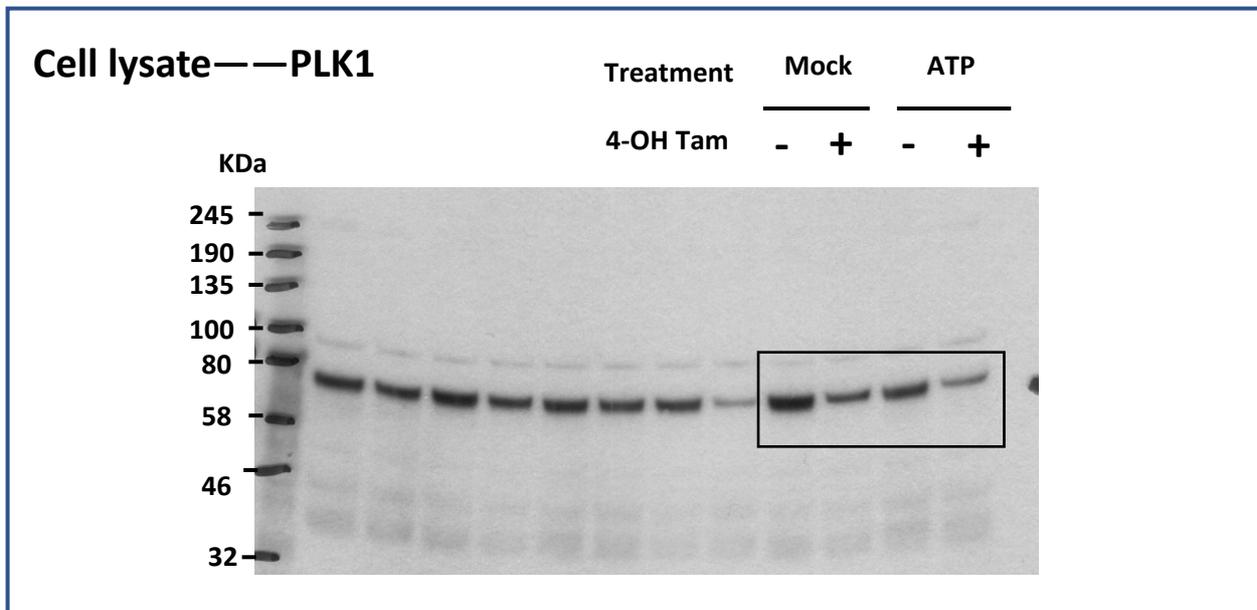
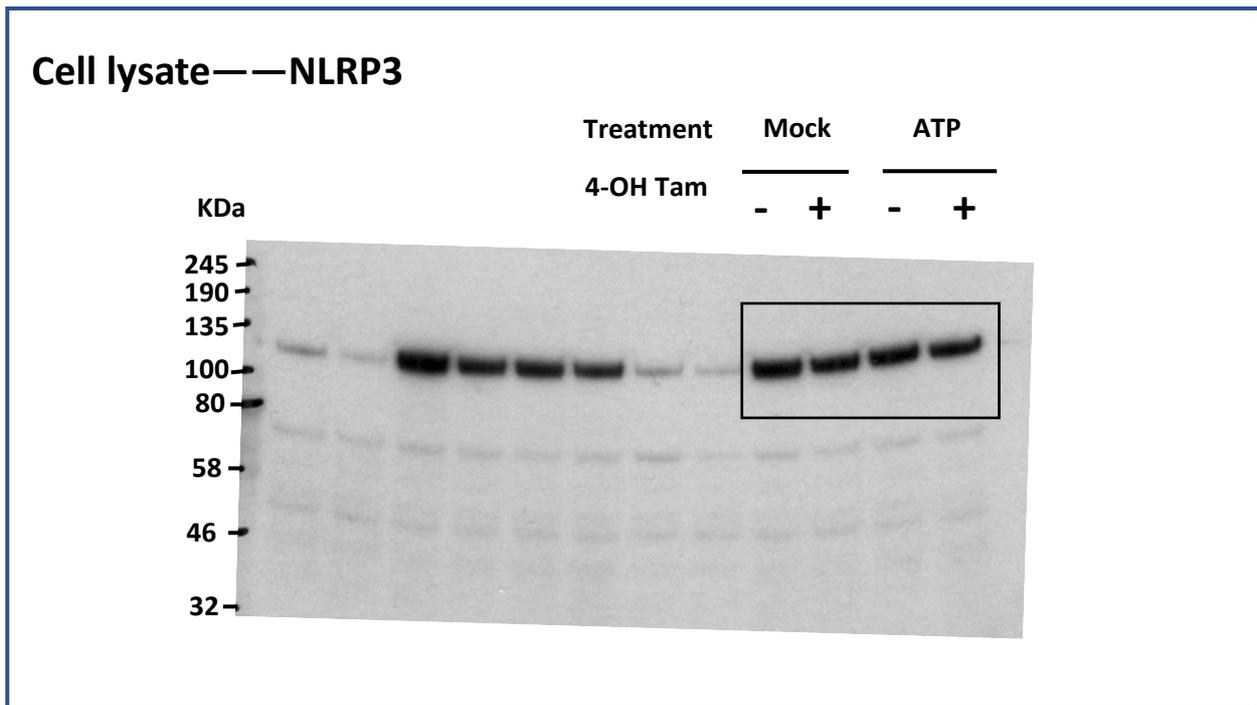


Figure 2G

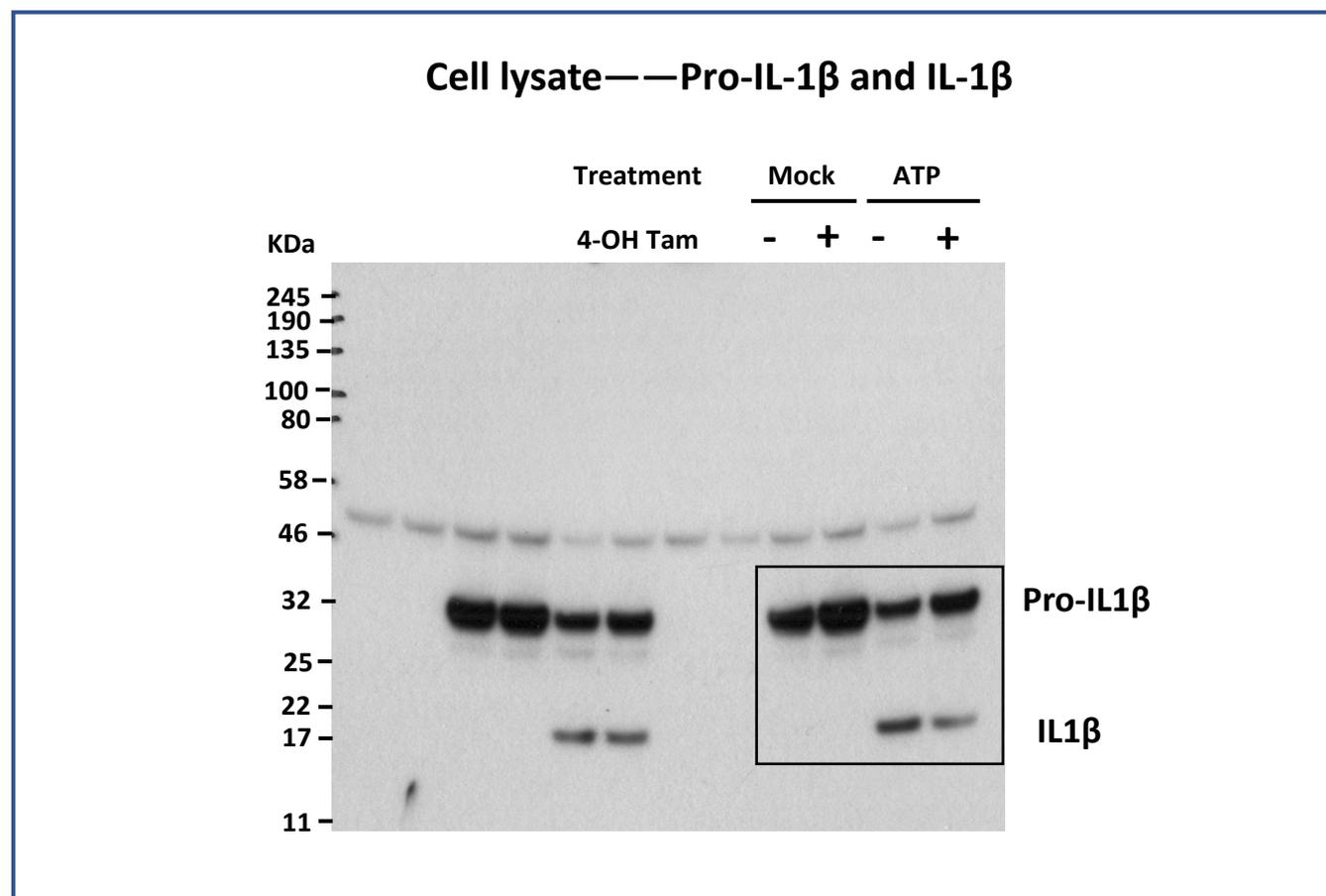
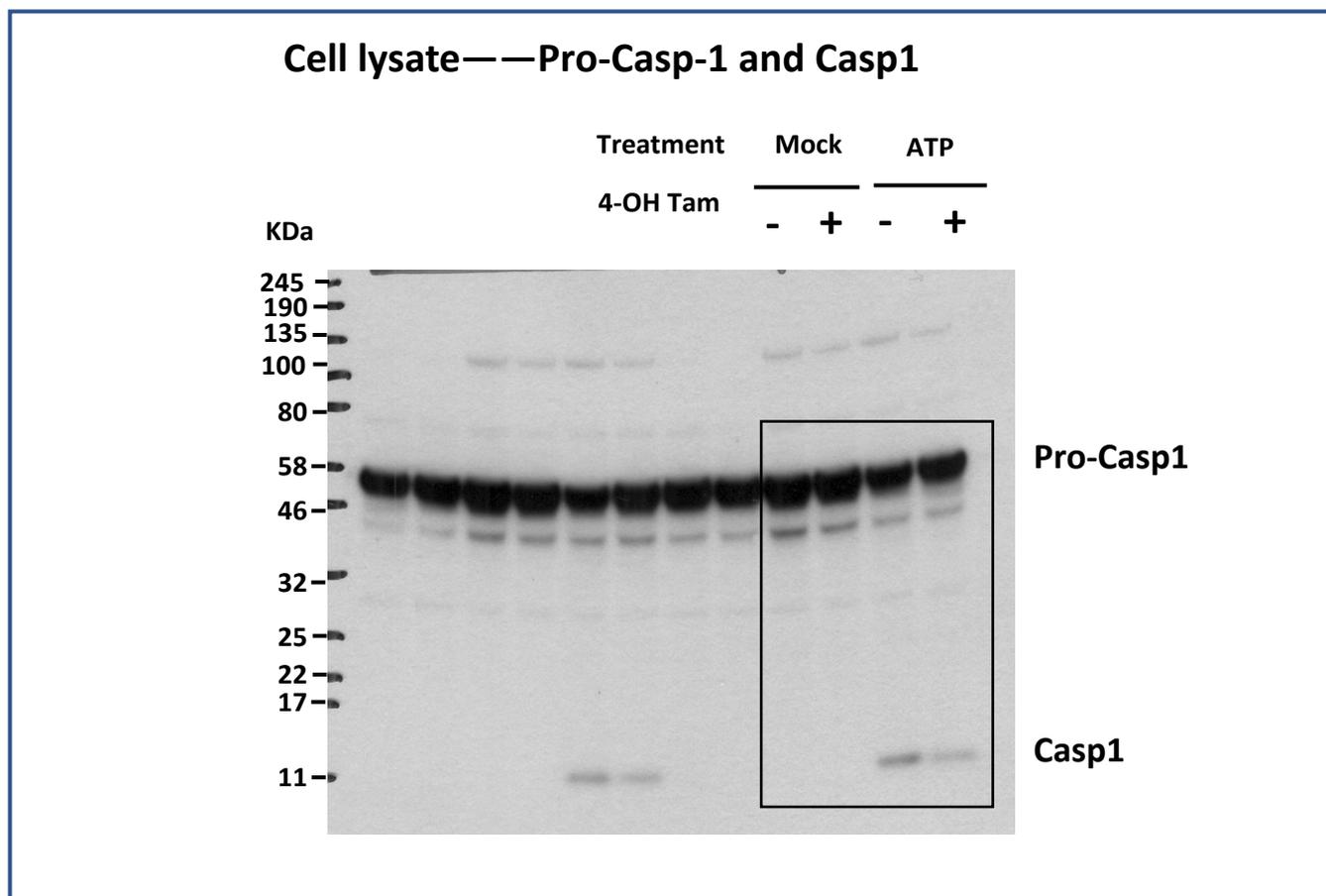


Figure 2G

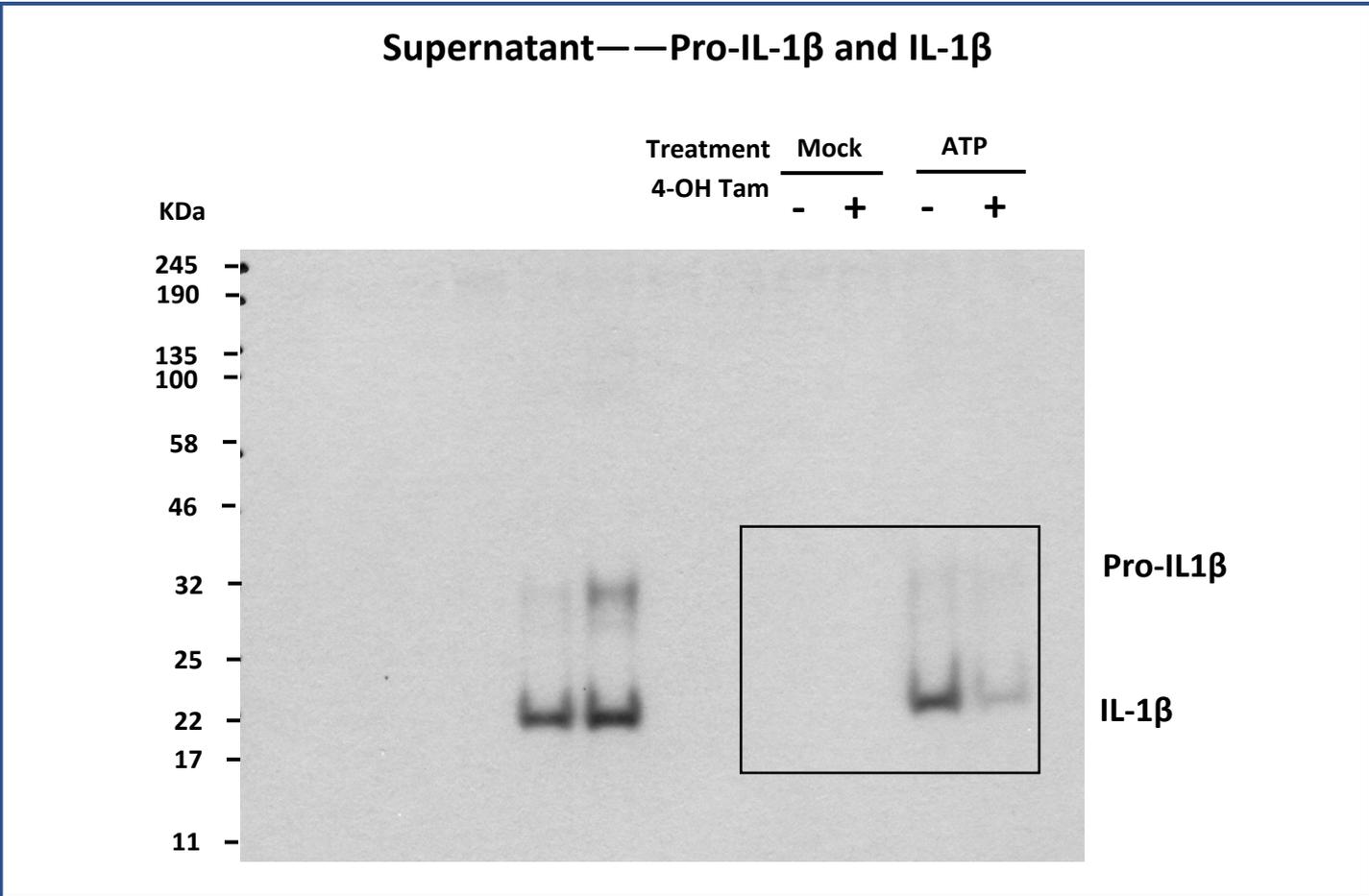
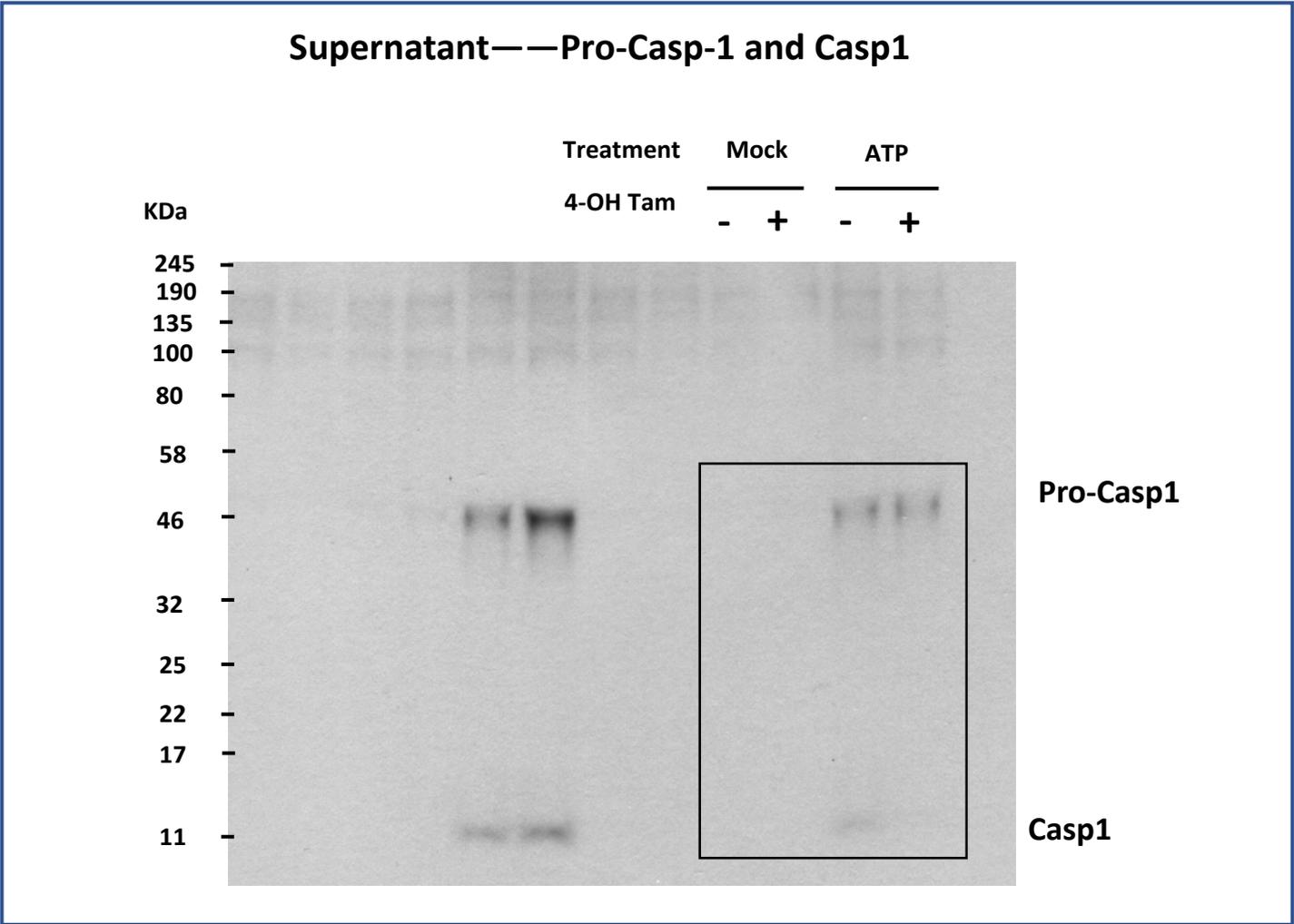


Figure 3C

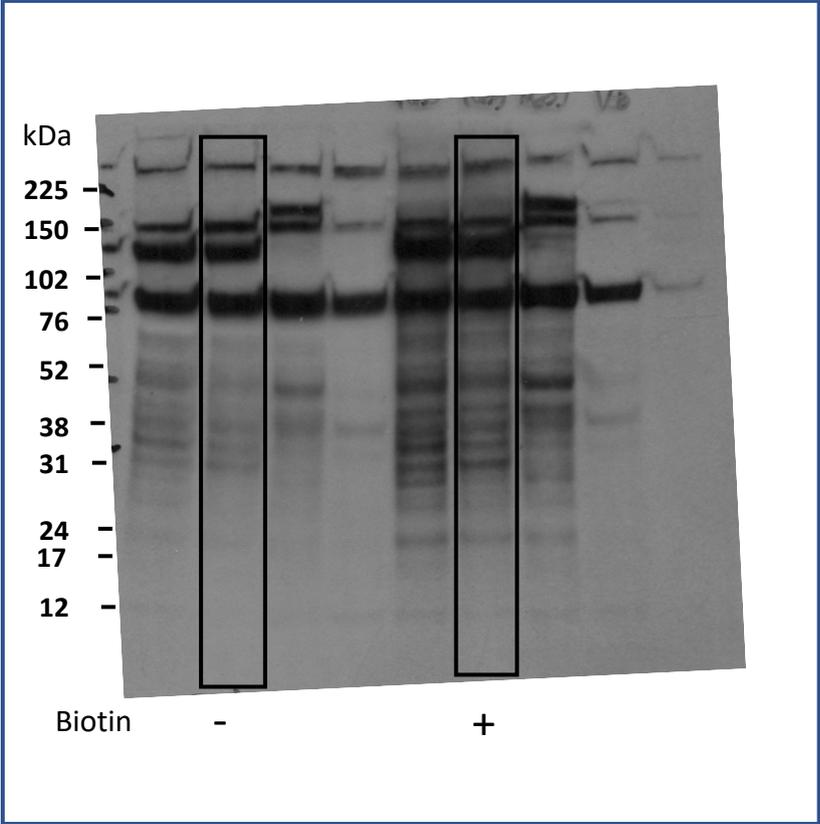


Figure 4A

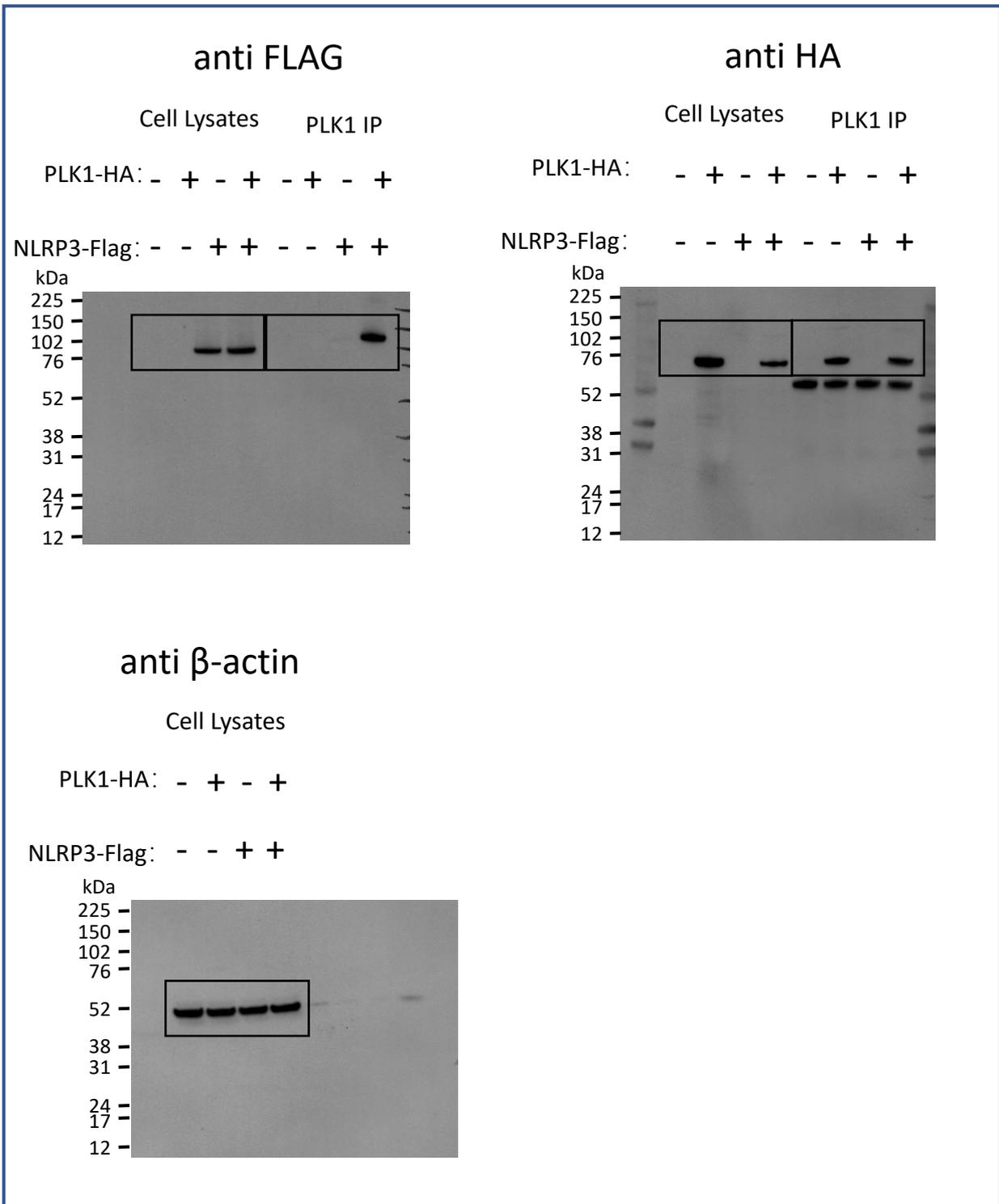
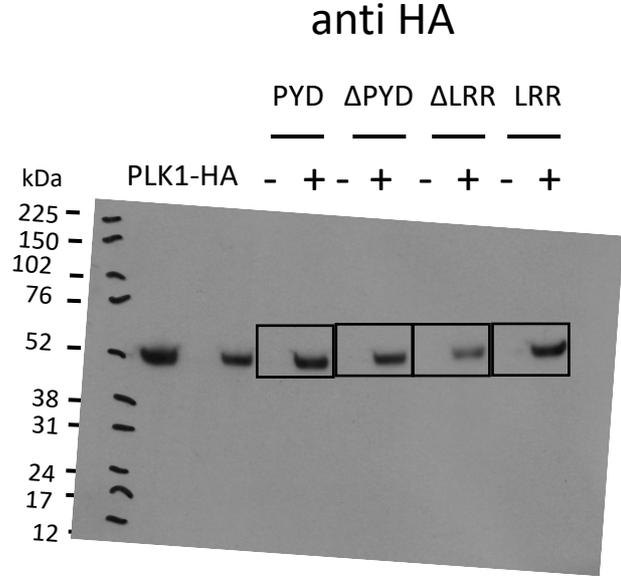
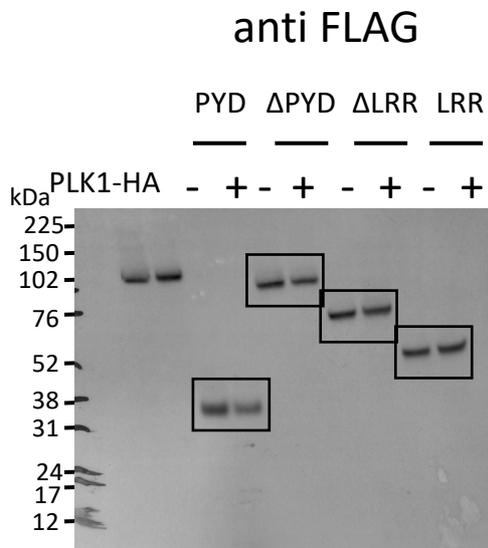
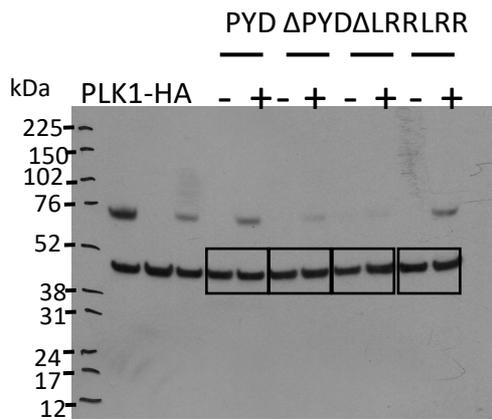


Figure 4B

Cell Lysates



anti β -ACTIN



PLK1 IP

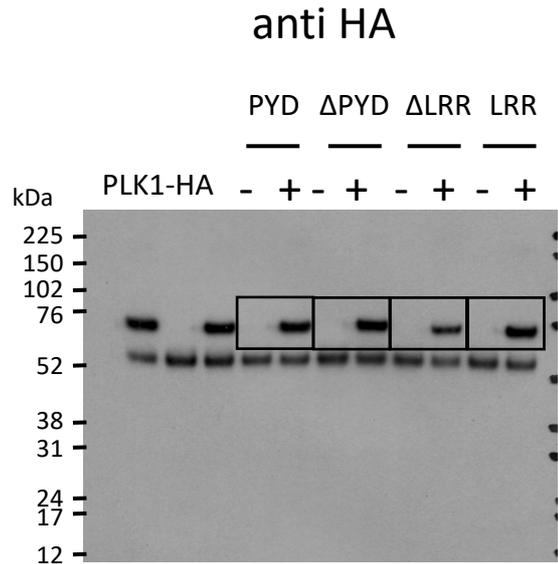
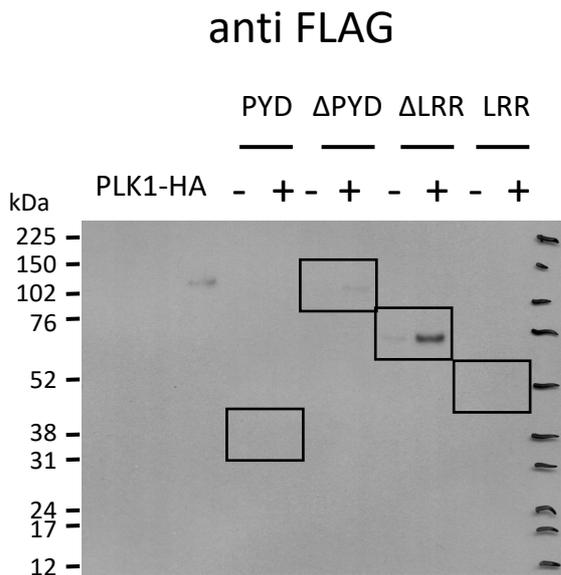


Figure 4C

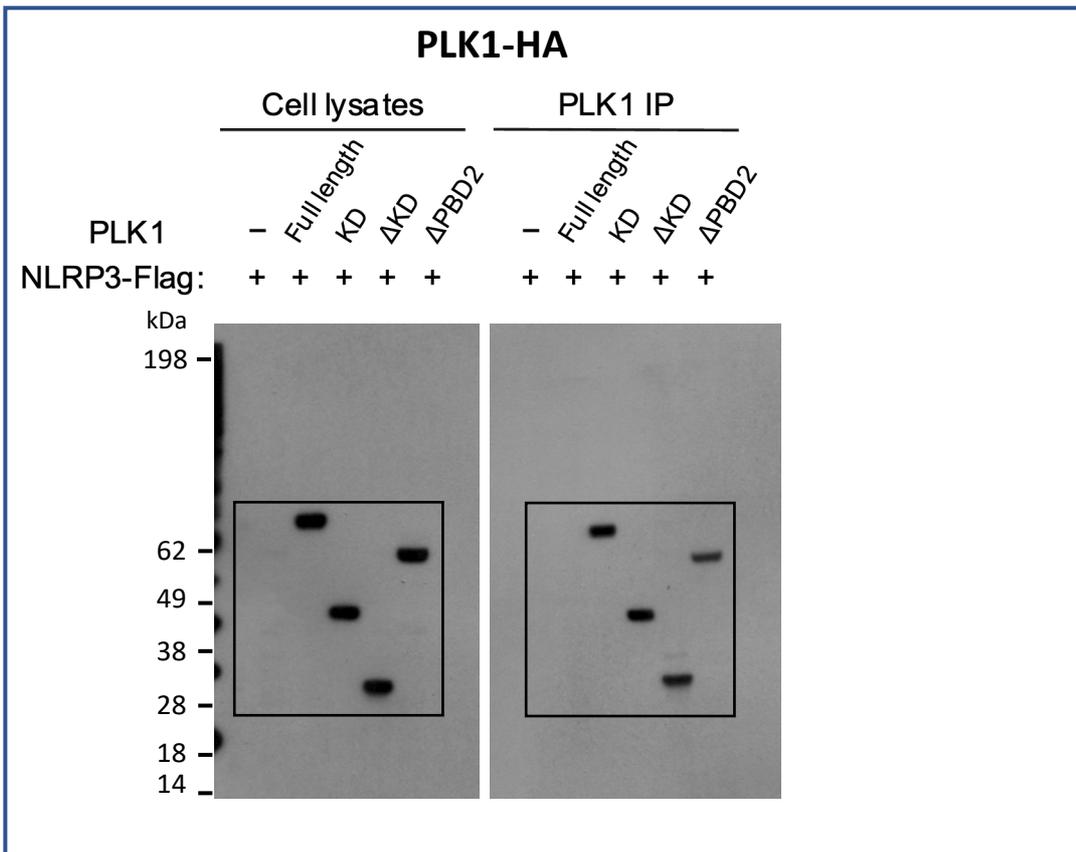
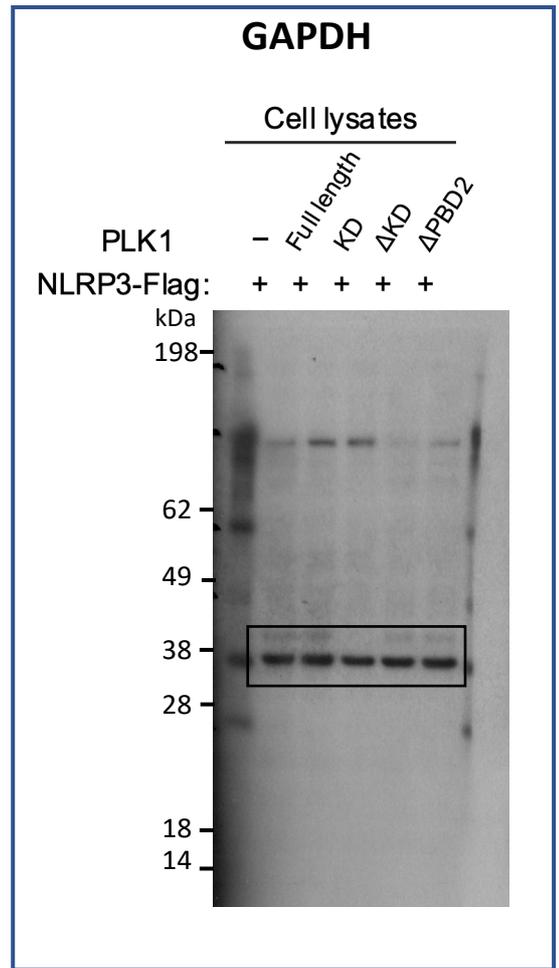
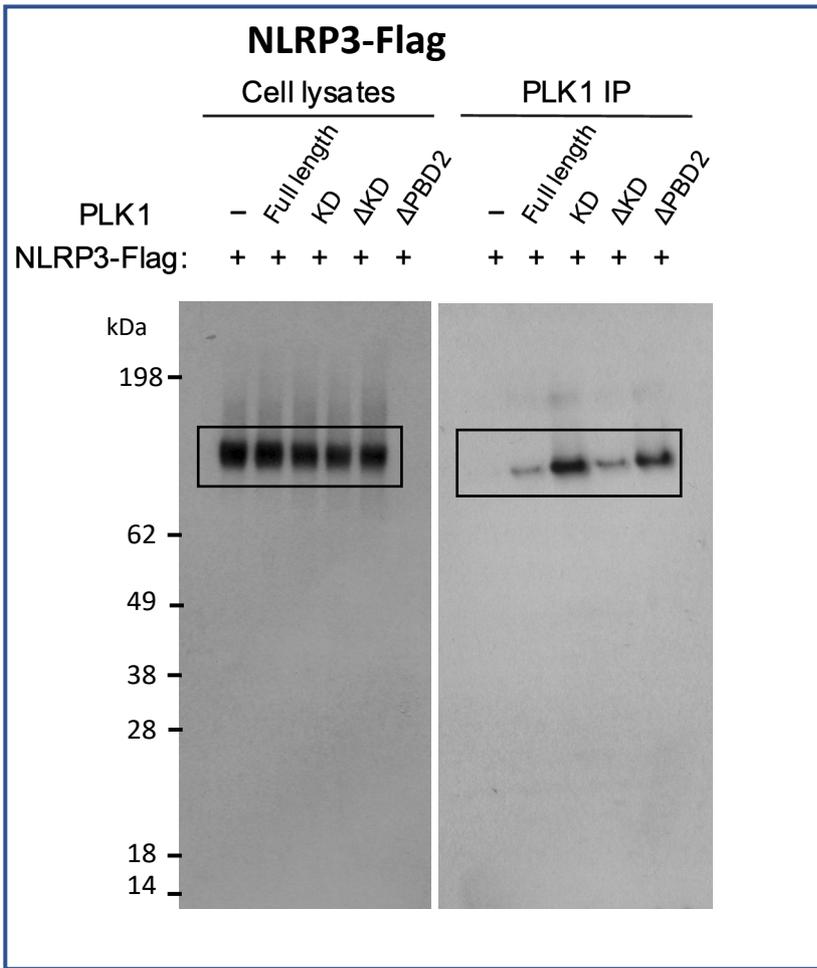


Figure 5F

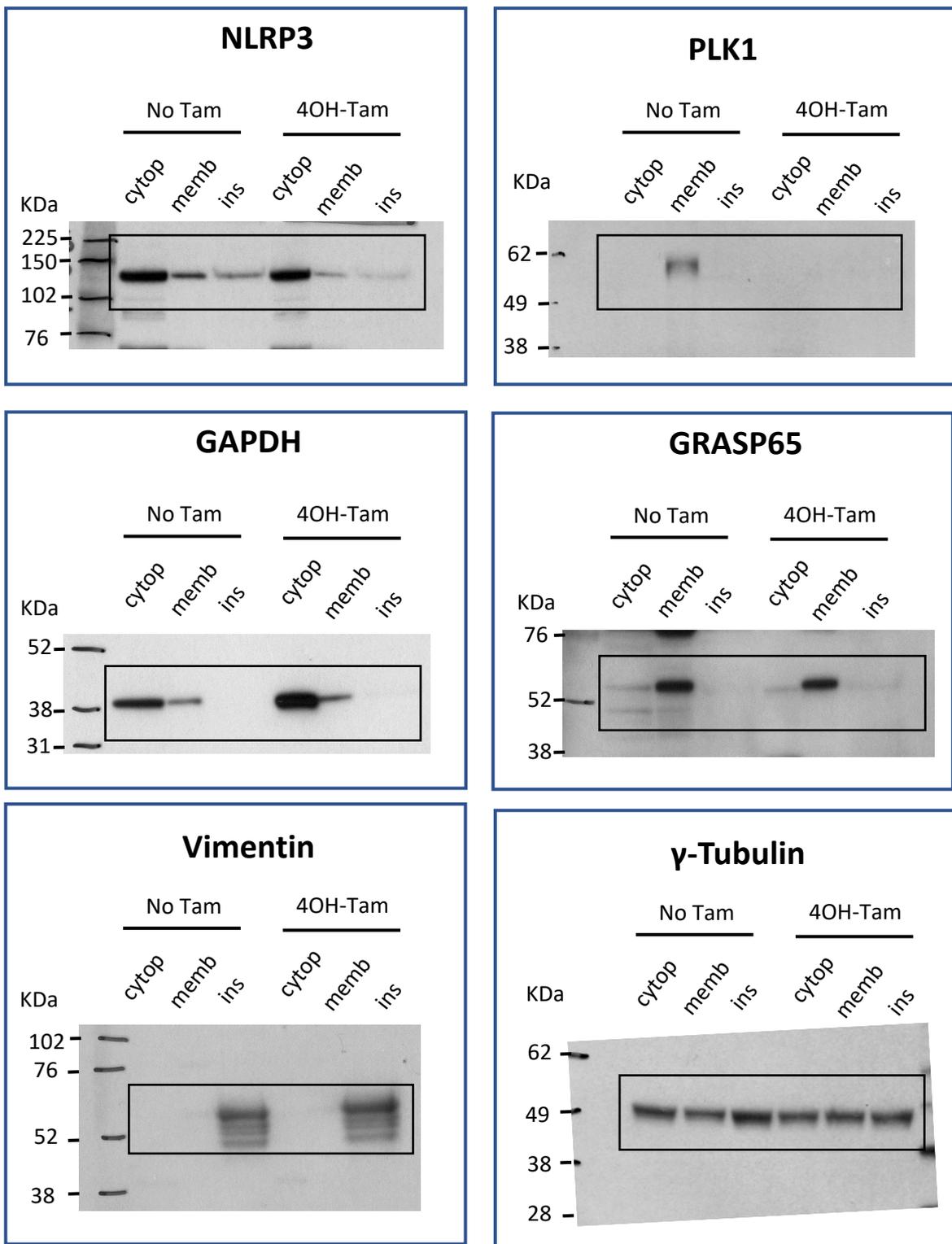


Figure S3E

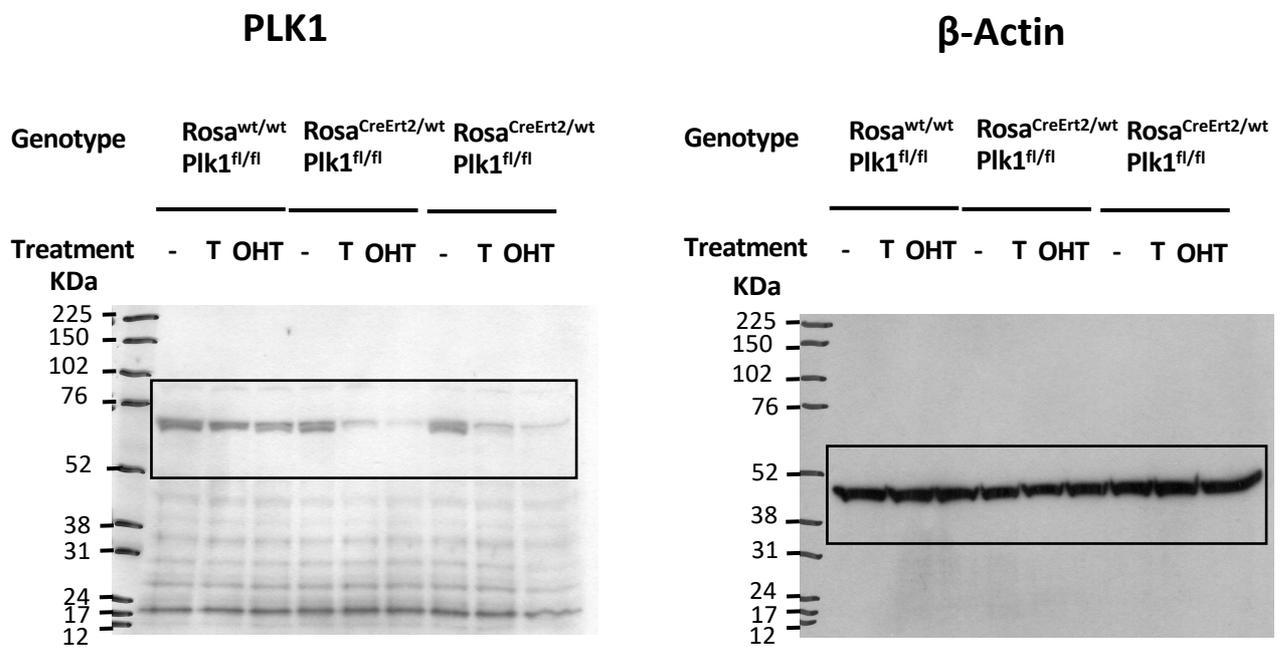


Figure S3K

