SUPPLEMENTAL MATERIALS

Breast Cancer Chemotherapy Induces Vascular Dysfunction and Hypertension through NOX4 dependent mechanism.

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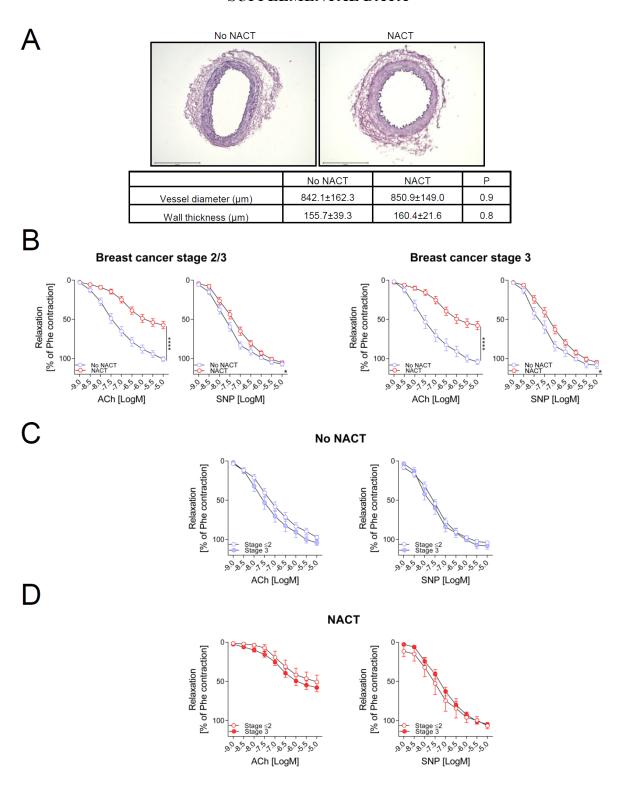
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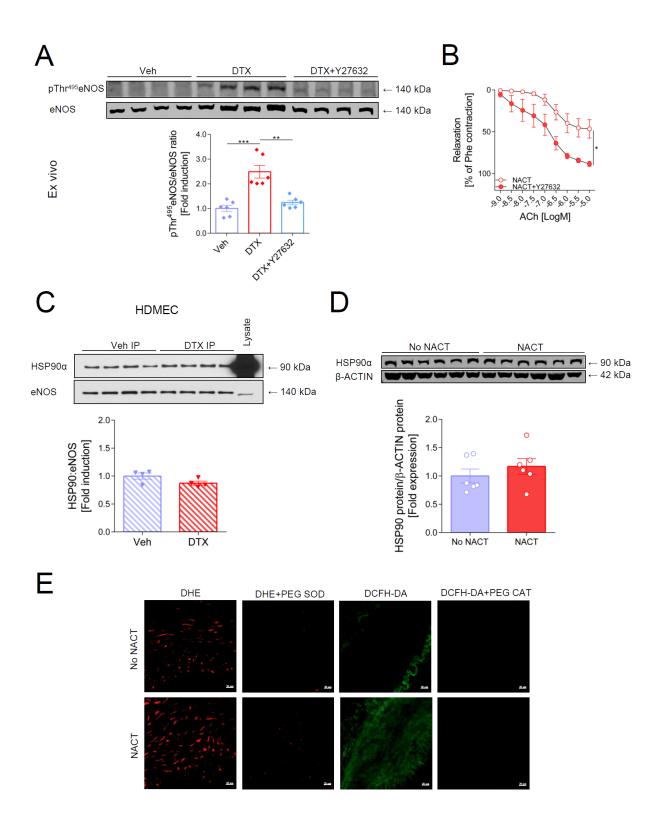
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SUPPLEMENTAL DATA



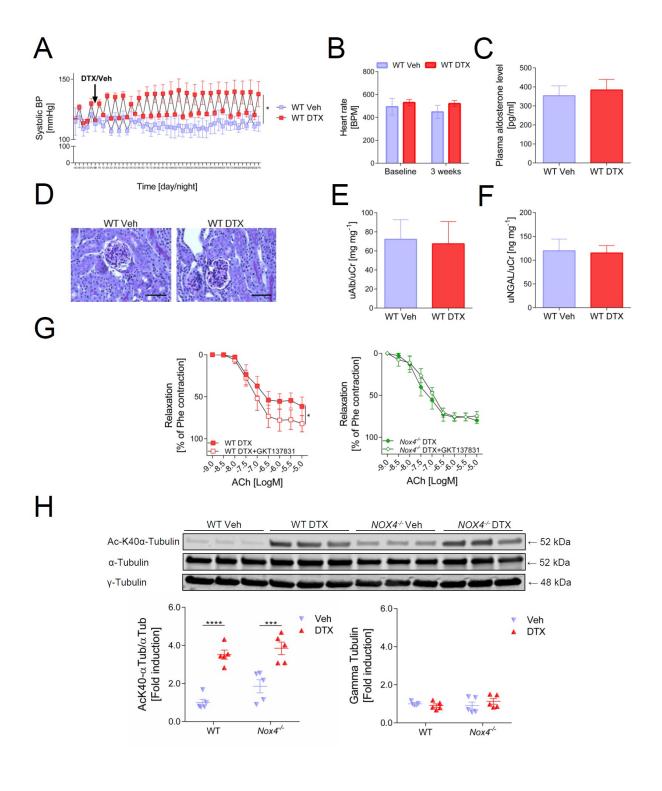
Supplemental Figure 1: Comparison of blood vessel morphology in NACT and No NACT and effects of the stage of breast cancer on endothelial dysfunction. (A) Vascular

morphology of studied vessels shown in representative H&E histograms (top) and quantification of vascular morphology (bottom) (n=5 per group). Scale bar, 500 μ m. Data are expressed as mean \pm s.e.m.; p-value - two-tailed unpaired t-test. (**B**) Average endothelium-dependent vasorelaxation curves to ACh (1 nM to 10 μ M) and endothelium-independent vasorelaxations in response to SNP (1 nM to 10 μ M) in vessels from patients without NACT (No NACT; n=30) and from patients who underwent NACT (NACT; n=39) with stages 2 and 3 of breast cancer (left) and in vessels from patients without NACT (No NACT; n=17) and from patients who underwent NACT (NACT; n=33) with stage 3 of breast cancer (right). Data are expressed as mean \pm s.e.m. ****P<0.0001, *P<0.05 vs. No NACT. Data were analyzed with repeated-measures ANOVA. (**C-D**) Comparison of endothelium-dependent vasorelaxations curves to ACh (1 nM to 10 μ M) (left) and to SNP (1 nM to 10 μ M) (right) breast cancer stage \leq 2 with breast cancer stage 3 in vessels (**C**) from patients without NACT (No NACT; left; n=17 vs 38) and (**D**) from patients who underwent NACT (NACT; right; n=7 vs 33). Data are expressed as mean \pm s.e.m. Data were analyzed with repeated measures ANOVA.



Supplemental Figure 2: Effects of breast cancer neoadjuvant chemotherapy on the regulation of endothelial nitric oxide synthase and the role of oxidative stress in NACT-induced endothelial dysfunction. (A) Effects of DTX (100 nM) on eNOS at Thr495

phosphorylation in NACT-naive vessels after 24-hour organ culture and the modulating effect of Y27632 (5 µM) (n=6 per group). Densitometric analysis of proteins normalized to the expression of total eNOS. Immunoblots are shown from one experiment (upper). Data are derived from two independent experiments and expressed as mean \pm s.e.m. ***P<0.001 vs. Veh, **P<0.01 vs. DTX. Data were analyzed with one-way ANOVA and Tukey's test. (B) Effect of Y27632 (5 μM) on endothelium-dependent vasorelaxation to acetylcholine (ACh) in vessels from patients with prior neoadjuvant chemotherapy (NACT) (n=7/group). Data shown as mean \pm s.e.m; *P<0.05 vs NACT by repeated measures ANOVA. (C) Effects of DTX (100 nM) on association between eNOS and HSP90α in Human Dermal Microvascular Endothelial Cells (HDMEC) (n=4/group). Data are expressed as mean \pm s.e.m. Two-tailed unpaired t-test. (**D**) Determination of HSP90α in vessels from patients with and without prior neoadjuvant chemotherapy (NACT) with densitometric analysis (n=6/group). Data are expressed as mean ± s.e.m. Two-tailed unpaired t-test. (E) High-power microphotographs of fluorescence for detection of superoxide (dihydroethidium; DHE; 10 µM- magnification of images in Figure 3F) and H₂O₂ production (2',7'-dichlorodihydrofluorescein diacetate; DCFH-DA; 10 μM) in vessel sections from patients with/without prior NACT. PEG-superoxide dismutase (PEG SOD; 500 U/ml) and PEG-catalase (PEG CAT; 500 U/ml) were used to show signal specificity for superoxide and H₂O₂ respectively (representative of n=5/group). Scale bar, 20 μm.



Supplemental Figure 3. Effects of docetaxel on blood pressure, renal function, microtubules and determination of the role of Nox4 in endothelial dysfunction induced by docetaxel. (A) Systolic blood pressure (SBP) measured using telemetry in wild-type (WT)

mice treated with docetaxel (DTX) or placebo (n=4 per group). Data are expressed as mean \pm s.e.m. *P<0.05 vs WT. Data were analyzed with repeated measures ANOVA. (B) Heart rate measured using telemetry in WT mice treated with DTX or placebo (n=4 per group). Data are expressed as mean \pm s.e.m. Data were analyzed with two-way ANOVA with Tukey's test. (C) Aldosterone levels in WT mice with DTX or placebo (n=7/group). Data are expressed as mean ± s.e.m. Data were analyzed with a two-tailed unpaired t-test. (D) Representative images of kidney staining with periodic acid - Schiff (PAS) in WT mice treated with DTX or placebo (n=6-7/group). Scale bar- 60 μm. (E) Urine albumin (uAlb) level in WT mice with DTX or placebo (n=6/group). Data are expressed as mean \pm s.e.m. Data were normalized with urine creatinine (uCr) level. Data were analyzed with a two-tailed unpaired t-test. (F) Urine NGAL level in WT mice with DTX or placebo (n=6/group). Data are expressed as mean \pm s.e.m. Data were normalized with urine creatinine (uCr) level. Data were analyzed with two-tailed unpaired t-test. (G) Endothelium dependent vasorelaxation in response to acetylcholine (ACh; 1 nM to 10 μM) after preincubation with GKT137831 (10 μM) in WT DTX mice (n=7 per group; left) and $Nox4^{-/-}$ DTX mice (n=7 per group; right). Data are expressed as mean \pm s.e.m. *P<0.05 vs. WT. Data were analyzed with repeated measures ANOVA. (H) Effect of DTX on Ack40-α-Tubulin and γ -Tubulin in $Nox4^{-1/2}$ and WT mice treated with DTX or placebo (n=5/group). Densitometric analysis of Ack40- α -Tubulin normalized to the expression of total α -Tubulin. Immunoblots are shown from one experiment (upper). Data are derived from two independent experiments and expressed as mean \pm s.e.m. ****P<0.0001 vs. WT DTX; ***P<0.001 vs. *Nox4*-/-DTX. Data were analyzed with two-way ANOVA and Bonferroni test.

Supplemental Table 1. Overall P values for two-way ANOVA.

Overall P values for two-way ANOVA				
Figure 1F for ACh	Presponse < 0.0001	Pgroup=0.0475	Presponse x group=0.4781	
Figure 1F for SNP	Presponse < 0.0001	Pgroup=0.8787	Presponse x group=0.9955	
Figure 2A for ACh	Presponse < 0.0001	Pgroup=0.0003	Presponse x group<0.0001	
Figure 6C for SBP	P ^{time} =0.0001	Pgroup=0.2436	P ^{time x group} =0.0096	
Figure 6D for ACh	Presponse<0.0001	Pgroup=0.0005	Presponse x group<0.0001	
Figure 6D for SNP	Presponse < 0.0001	Pgroup=0.3692	Presponse x group=0.9819	
Figure 6E for H ₂ O ₂	Pgenotype=0.0015	P ^{DTX} =0.0109	Pgenotype x DTX=0.1143	
Figure 6F for LGCL	Pgenotype=0.1755	P ^{DTX} =0.0012	Pgenotype x DTX=0.1289	
Figure 6H for <i>Nox4</i>	Pgenotype < 0.0001	P ^{DTX} =0.0198	Pgenotype x DTX=0.0198	
Figure 6I for Thr 495	Pgenotype=0.1133	P ^{DTX} =0.0071	Pgenotype x DTX=0.0007	
Figure 7B for SBP	P ^{time} =0.0261	Pgroup=0.0146	P ^{time x group} =0.0198	
Figure 7C for ACh	Presponse < 0.0001	Pgroup=0.0045	Presponse x group=0.0062	
Figure 7C for SNP	Presponse < 0.0001	Pgroup=0.1409	Presponse x group=0.0653	
Figure 7E for SBP	P ^{time} =0.3375	Pgroup=0.0120	P ^{time x group} =0.0281	
Figure 7F for ACh	Presponse < 0.0001	Pgroup=0.0300	Presponse x group=0.0139	
Figure 7F for SNP	Presponse < 0.0001	Pgroup=0.5048	Presponse x group=0.1105	
Supplemental Figure 3B for HR	P ^{time} =0.0554	Pgroup=0.4657	P ^{time x group} =0.1749	
Supplemental Figure 3H for Ack40-Tub	Pgenotype=0.0505	P ^{DTX} <0.0001	Pgenotype x DTX=0.3678	
Supplemental Figure 3H for gamma-Tub	Pgenotype=0.6167	P ^{DTX} =0.6263	Pgenotype x DTX=0.2384	

Supplemental Table 2. Antibodies.

Target antigen Source		Catalog number
β-ΑCTIN	Abcam	ab8226
phospho-eNOS (Thr495)	BD Biosciences	612707
phospho-eNOS (Ser1177)	BD Biosciences	612393
eNOS	BD Biosciences	610297
NOX1	Sigma-Aldrich	HPA035299
NOX2	Abcam	ab129068
NOX4	Abcam	ab109225
NOX5	Sigma-Aldrich	SAB2501641
α-Tubulin	Cell Signaling	2125S
acetyl-α-Tubulin	Cell Signaling	53358
gamma-Tubulin	Sigma-Aldrich	T6557
ΗЅΡ90α	BD Biosciences	610419
CD31	Abcam	ab24590
α-SMA	Sigma-Aldrich	A5228

Supplemental Table 3. Cell lines.

Cell line	Source
Human Dermal Microvascular Endothelial Cells (HDMEC)	PromoCell
Human Aortic Smooth Muscle Cells (HASMC)	Thermo Fisher
HEK 293	ATCC