Increased insulin and leptin sensitivity in mice lacking acyl CoA:diacylglycerol acyltransferase 1

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Acyl coenzyme A:diacylglycerol acyltransferase 1 (DGAT1) is one of two known DGAT enzymes that catalyze the final step in mammalian triglyceride synthesis. DGAT1-deficient mice are resistant to diet-induced obesity through a mechanism involving increased energy expenditure. Here we show that these mice have decreased levels of tissue triglycerides, as well as increased sensitivity to insulin and to leptin. Importantly, DGAT1 deficiency protects against insulin resistance and obesity in agouti yellow mice, a model of severe leptin resistance. In contrast, DGAT1 deficiency did not affect energy and glucose metabolism in leptin-deficient (ob/ob) mice, possibly due in part to a compensatory upregulation of DGAT2 expression in the absence of leptin. Our results suggest that inhibition of DGAT1 may be useful in treating insulin resistance and leptin resistance in human obesity.

stimulating hormone in the hypothalamus (11). They are also severely leptin resistant (12). Genotyping for Dgat1 and ob was performed as described (4, 13). AY/a mice were identified by their yellow fur. Mice were housed in a pathogen-free barrier facility (12-hour light/12-hour dark cycle) and fed rodent chow (Ralston Purina Co., St. Louis, Missouri, USA). For high-fat diet experiments, mice were fed a Western-type diet containing 21% fat by weight (Harlan Teklad Laboratory, Madison, Wisconsin, USA) for 4 weeks unless stated otherwise. All experiments were approved by the Committee on Animal Research of the University of California, San Francisco.

Tissue lipid analysis. Tissue lipids were extracted and separated by TLC as described (4). Triglycerides and diacylglycerol were scraped from the TLC plate, and heptadecanoic acid (~10% of total fatty acids) was added as an internal standard. Methyl esters were synthesized with a 10-ft glass column (10% SP-2330 on 100/120 Chromosorb WAW; Supelco, St. Louis, Missouri, USA) at 175°C for 5 minutes and increasing to 210°C at a rate of 2.5°C/min. The weight of triglycerides and diacylglycerol in the samples was calculated with reference to the internal standard.

Adipocyte size determination. Adipose tissue was obtained from the reproductive fat pads of 14-week-old male mice. The samples were fixed in paraformaldehyde, embedded in paraffin, cut into 5-μm sections, and stained with hematoxylin and eosin. Images of the histology sections were analyzed with Adobe Photoshop 5.01 (Adobe Systems Inc., San Jose, California, USA) and Image Process Tool Kit (Reindeer Games, Gainesville, Florida, USA) as described (14).

Glucose metabolism studies. Glucose (1 g/kg body wt) or bovine insulin (1 U/kg body wt; Sigma Chemical Co., St. Louis, Missouri, USA) was injected intraperitoneally, and glucose concentrations were measured with a glucometer (Accu-Chek; Roche Diagnostics Corp., Indianapolis, Indiana, USA). The hyperinsulinemic-euglycemic clamp studies were performed as described (15) with slight modifications. Weight-matched 10- to 14-week-old male mice were fasted for 4 hours before the clamp studies. Insulin was infused at 20 mU/kg/min, and plasma glucose concentration was clamped at 120 mg/dl. For ob/ob and AY/a mice, nonfasting blood samples for glucose and insulin measurements were obtained at noon. Insulin was measured with a rat insulin RIA kit (Linco Research Inc., St. Charles, Missouri, USA).

Leptin infusion studies. Mice were infused with recombinant human leptin (gift from F. Chehab, University of California, San Francisco) with a microosmotic pump (Alza model 1002; DURECT Corp., Cupertino, California, USA) as described (14). Plasmatrope levels were measured by AniLytics Inc. (Gaithersburg, Maryland, USA).

Northern blots. White adipose tissue (WAT) was obtained from the reproductive fat pads and brown adipose tissue (BAT) from the interscapular region of 10- to 14-week-old male mice. Total RNA was isolated, and

### Table 1
Real-time PCR primer and probe sequences

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer pair or probe</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Actin</td>
<td>5′ ‑CATCTTAGGCTCCTACGTGCA-3′</td>
<td>3′ ‑CGCCGGTGAGCTAGCTGAC-3′</td>
</tr>
<tr>
<td>Probe</td>
<td>5′ ‑GCCTTACCAGACGTCCTGAC-3′</td>
<td>3′ ‑GGCGGGGCTCATGCTGAC-3′</td>
</tr>
<tr>
<td>Acyl CoA:diacylglycerol acyltransferase 2</td>
<td>5′ ‑AGTGAGCTTACGTGAC-3′</td>
<td>3′ ‑GGCGGGGCTCATGCTGAC-3′</td>
</tr>
<tr>
<td>Probe</td>
<td>5′ ‑GCCTTACCAGACGTCCTGAC-3′</td>
<td>3′ ‑GGCGGGGCTCATGCTGAC-3′</td>
</tr>
<tr>
<td>Leptin</td>
<td>5′ ‑CTCCGGAGGATTACCCTCCTGTC-3′</td>
<td>3′ ‑CTCCGGAGGATTACCCTCCTGTC-3′</td>
</tr>
<tr>
<td>Probe</td>
<td>5′ ‑CTCCGGAGGATTACCCTCCTGTC-3′</td>
<td>3′ ‑CTCCGGAGGATTACCCTCCTGTC-3′</td>
</tr>
<tr>
<td>PPARα</td>
<td>5′ ‑AGAGGGCGCCTCCCTGCTGTT-3′</td>
<td>3′ ‑AGAGGGCGCCTCCCTGCTGTT-3′</td>
</tr>
<tr>
<td>Probe</td>
<td>5′ ‑AGAGGGCGCCTCCCTGCTGTT-3′</td>
<td>3′ ‑AGAGGGCGCCTCCCTGCTGTT-3′</td>
</tr>
</tbody>
</table>

### Table 2
Lipid composition of tissues in Dgat1−/− and Dgat1+/+ mice

<table>
<thead>
<tr>
<th></th>
<th>Triglycerides (% of tissue weight)</th>
<th>Diacylglycerol (% of tissue weight)</th>
<th>Fatty acid composition of triglycerides (% of total fatty acids)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>16:0</td>
<td>16:1</td>
<td>18:0</td>
</tr>
<tr>
<td>White adipose tissue</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>+/+</td>
<td>32.4 ± 7.9</td>
<td>0.06 ± 0.02</td>
<td>20.0 ± 0.6</td>
</tr>
<tr>
<td>−/−</td>
<td>20.3 ± 3.0a</td>
<td>0.04 ± 0.01</td>
<td>26.4 ± 1.5a</td>
</tr>
<tr>
<td>Skeletal muscle</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>+/+</td>
<td>4.3 ± 1.9</td>
<td>0.008 ± 0.002</td>
<td>21.0 ± 2.0</td>
</tr>
<tr>
<td>−/−</td>
<td>2.4 ± 0.5a</td>
<td>0.006 ± 0.003</td>
<td>27.3 ± 0.7a</td>
</tr>
<tr>
<td>Liver</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>+/+</td>
<td>0.2 ± 0.1</td>
<td>0.013 ± 0.003</td>
<td>ND</td>
</tr>
<tr>
<td>−/−</td>
<td>0.1 ± 0.0</td>
<td>0.003 ± 0.001</td>
<td>ND</td>
</tr>
</tbody>
</table>

Twelve- to sixteen-week-old male mice fed a chow diet were used. ND, not determined. aP < 0.05, bP < 0.01 versus Dgat1+/+, n = 3 per genotype.
were fed a high-fat diet for 10 weeks. Adipocytes were measured per mouse. For high-fat experiments, mice mean adipocyte surface area of one female mouse. More than 100 Decreased adipocyte size in Figure 1 in a 20- RNA was extracted. RNA (1 µl reaction containing oligo(dT)12-18 primer and Superscript II enzyme (Invitrogen Corp., Carlsbad, California, USA). Primer and probe sequences (Table 1) Superscript II enzyme (Invitrogen Corp., Carlsbad, California, USA). Primer and probe sequences (Table 1) of cDNA probes. Blots were rehybridized with a β-actin probe (Ambion Inc, Austin, Texas, USA) for loading normalization. Probes for uncoupling protein 2 (UCP-2) and UCP-3 were generated by PCR with WAT cDNA and the following primers: 5′-GTCGATTCCGCCCTCGGTG-3′ and 5′-GAGGGAAAGTGATGAGATCT-3′ (UCP-2); 5′-GTCGGAGCACAGCCTTCTGC-3′, 5′-ACCTTGAGCCCGAGCCCGG-3′ (UCP-3). The remaining probes were gifts from M. Reitman, NIH, Bethesda, Maryland, USA (UCP-1); B. Staels, Institut National de la Santé et de la Recherche Médicale (Lille, France) (acyl CoA oxidase); B. Spiegelman, Dana-Farber Cancer Institutes, Boston, Massachusetts, USA (peroxisome proliferator-activated receptor γ); and I. Shimomura, M. Brown, and J. Goldstein, University of Texas Southwestern, Dallas, Texas, USA (fatty acid synthase). Signals were quantified with a PhosphorImager (Bio-Rad Laboratories, Hercules, California, USA).

Real-time PCR. Tissues were homogenized, and total RNA was extracted. RNA (1 µg) was reverse-transcribed in a 20-µl reaction containing oligo(dT)12-18 primer and Superscript II enzyme (Invitrogen Corp., Carlsbad, California, USA). Primer and probe sequences (Table 1) were selected with Primer Express (Perkin-Elmer Applied Biosystems, Foster City, California, USA). The PCR reaction (50 µl) contained 1 µl of cDNA, 1× Gold buffer, 4 mM MgCl₂, 500 µM dNTP, primers (200 nM), 100 nM probe (labeled with 6-carboxyfluorescein), and 1.25 U of AmpliTaq Gold DNA polymerase (Perkin-Elmer Applied Biosystems). Real-time PCR was performed with the ABI Prism 7700 System (Perkin-Elmer Applied Biosystems). Expression levels were calculated by the comparative cycle of threshold detection method, according to the manufacturer. Expression of β-actin was used for loading normalization.

Statistical methods. Data are shown as mean ± SD unless stated otherwise. Measurements were compared with the t test or Mann-Whitney rank-sum test. Differences in body weight or food intake were compared with ANOVA, followed by the Tukey-Kramer test.

Results
Altered lipid composition in tissues of Dgat1–/– mice. We hypothesized that DGAT1 deficiency results in decreased tissue triglyceride content. Indeed, Dgat1–/– mice had a 30–40% reduction of triglyceride levels in WAT and skeletal muscle (Table 2). Liver triglyceride levels trended lower in chow-fed Dgat1–/– mice, although the difference was not statistically significant. On a high-fat diet, however, Dgat1–/– mice had significantly lower liver triglyceride levels than Dgat1+/+ mice (28 ± 16 versus 157 ± 28 mg/g tissue weight, P < 0.05). Unexpectedly, levels of diacylglycerol, a substrate for the DGAT reaction, were not elevated, and in fact tended to be lower in Dgat1–/– tissues (Table 2). DGAT1 deficiency also altered the fatty acid composition of triglycerides in WAT and skeletal muscle, resulting in a relative decrease in monounsaturated (16:1 and 18:1) fatty acids and a relative increase in saturated (16:0 and 18:0) fatty acids (Table 2).

Decreased adipocyte size in Dgat1–/– mice. Concomitant with the decreased triglyceride levels in WAT, Dgat1–/– mice had smaller adipocytes than Dgat1+/+ mice on both chow and high-fat diets (Figure 1). Adipocytes from Dgat1–/– mice developed minimal hypertrophy in response to a high-fat diet, whereas those from Dgat1+/+ mice doubled in size. This protection from diet-induced adipocyte hypertrophy in Dgat1–/– mice mirrored their weight response to a high-fat diet (4). In addition to having smaller adipocytes, male Dgat1–/– mice had a lower mean DNA content in reproductive fat pads than Dgat1+/+ mice (169 ± 36 versus 273 ± 50 µg/fat pad, P < 0.05), suggesting that Dgat1–/– mice also had fewer adipocytes.

Increased insulin sensitivity in Dgat1–/– mice. Previous studies in Dgat1–/– mice suggested that they have enhanced glucose disposal (4). To address this further,
we performed glucose- and insulin-tolerance tests. Dgat1–/– mice had lower plasma glucose concentrations than Dgat1+/+ mice after a glucose load (Figure 2a) or an insulin injection (Figure 2b), suggesting that Dgat1–/– mice have increased insulin sensitivity. This was confirmed in hyperinsulinemic-euglycemic clamp studies, which showed that Dgat1–/– mice required an approximately 20% higher glucose infusion rate than Dgat1+/+ mice to maintain euglycemia (Figure 2c).

Increased weight loss in response to leptin infusion in Dgat1–/– mice. We hypothesized that the decreased tissue triglyceride content in Dgat1–/– mice resulted in an increased sensitivity to leptin. To test this hypothesis, we infused leptin subcutaneously into Dgat1+/+ mice and Dgat1–/– mice and measured their response in body weight and food intake. Leptin infusion achieved comparable levels of increase in plasma leptin levels in Dgat1+/+ and Dgat1–/– mice (0.7 ± 0.1 versus 0.5 ± 0.2 ng/ml for 6 µg/day, P > 0.05). In young (10- to 14-week-old) Dgat1+/+ mice, leptin administration suppressed the normal weight gain seen in control (PBS-infused) mice (Figure 3a). The same doses of leptin caused an additional 3% weight loss in age-matched Dgat1–/– mice (Figure 3b), indicating an enhanced response to leptin.

Dgat1–/– mice consistently ate more than Dgat1+/+ mice at baseline (25.3% ± 1.6% versus 19.5% ± 3.2% of body weight, P < 0.05). During leptin infusion, Dgat1–/– mice continued to eat more than Dgat1+/+ mice (19.8% ± 3% versus 15.8% ± 2.4% of body weight after 5 days of leptin (6 µg/day) infusion, P < 0.05). Expressed as percentage of change from baseline, reduction in food intake in response to leptin infusion was comparable in Dgat1+/+ and Dgat1–/– mice (Figure 3c and d). Similarly, the absolute reduction in food intake per day was similar in Dgat1+/+ and Dgat1–/– mice in response to leptin infusion in several different experiments (not shown). These results imply that the increased weight loss in leptin-treated Dgat1–/– mice resulted from increased energy expenditure rather than reduced food intake.

Expression of leptin-regulated genes in Dgat1–/– mice. The enhanced response to leptin in Dgat1–/– mice suggested that the leptin pathway was activated in these mice at baseline. To examine this possibility, we measured the expression of several leptin-regulated genes in BAT and WAT of Dgat1–/– mice (Figure 4). In Dgat1–/– BAT, UCP1 expression was increased by approximately 70% versus controls. In WAT, Dgat1–/– mice had increased levels of UCP3 expression, and UCP2 expression levels trended higher. The expression of genes involved in fatty acid oxidation (peroxisome proliferator-activated receptor α [PPARα] and acyl CoA oxidase) was also higher in Dgat1–/– WAT. In contrast, Dgat1–/– mice had decreased levels of expression for genes involved in adipogenesis (PPARγ) and lipid synthesis (fatty acid synthase) in WAT. Dgat1–/– mice also had an approximate 50% reduction in leptin (ob) mRNA expression. These results are consistent with...
those observed in rodents with leptin overexpression (16–18), suggesting that Dgat1−/− mice have increased leptin sensitivity at baseline.

DGAT1 deficiency protects against obesity and insulin resistance in AY/a mice but not in ob/ob mice. DGAT1 deficiency protects against obesity and insulin resistance induced by high-fat feeding (4). To determine whether DGAT1 deficiency had similar effects in genetic models of obesity and insulin resistance, we introduced DGAT1 deficiency into AY/a and ob/ob mice through breeding. DGAT1 deficiency protected against obesity in AY/a mice (Figure 5a), decreasing body weight (Figure 5b) and fat pad content (Figure 5c) by approximately 20% at 25 weeks. AY/a Dgat1−/− and AY/a Dgat1+/+ mice had similar plasma glucose levels (Figure 5d), but DGAT1 deficiency reduced plasma insulin levels by approximately 80% in AY/a mice (Figure 5e), most likely by increasing their insulin sensitivity. In contrast, DGAT1 deficiency had no apparent effects in ob/ob mice. In the setting of leptin deficiency, both Dgat1−/− and Dgat1+/+ mice became obese (Figure 5f) and diabetic, with similar growth curves (Figure 5g), fat pad content (Figure 5h), and plasma glucose (Figure 5i) and insulin (Figure 5j) levels. DGAT1 deficiency also had no apparent effect on the obesity and diabetes of mice lacking functional leptin receptors (db/db mice, not shown).

Increased DGAT2 mRNA expression in WAT of leptin-deficient Dgat1−/− mice. One possible explanation for the lack of effect of DGAT1 deficiency in ob/ob mice is that an alternative pathway for triglyceride synthesis is upregulated in leptin-deficient Dgat1−/− mice. To test this hypothesis, we measured DGAT2 mRNA expression in WAT of Dgat1−/− mice in different backgrounds and conditions (Figure 6). DGAT2 expression was not increased in Dgat1−/− mice at baseline (chow) or after 15 weeks of a high-fat diet. DGAT2 expression was also not increased in AY/a Dgat1−/− mice. In contrast, DGAT2 expression was elevated approximately threefold in leptin-deficient Dgat1−/− mice. This suggests that leptin normally downregulates DGAT2 expression in WAT and that the upregulation of DGAT2 expression may compensate for the loss of DGAT1 in leptin-deficient Dgat1−/− mice.

Discussion

This study demonstrates that mice lacking DGAT1, one of two known enzymes that catalyze the final step in mammalian triglyceride synthesis, have reduced levels of tissue triglycerides. More significantly, these reductions in tissue triglycerides were associated with increased sensitivity to insulin and to leptin. These mechanisms appear to underlie the increased energy consumption observed in rodents with leptin overexpression (16–18), suggesting that Dgat1−/− mice have increased leptin sensitivity at baseline.
expenditure and protection against diet-induced obesity and insulin resistance in \textit{Dgat1}^+/− mice (4).

One potential mechanism for the increased insulin sensitivity in \textit{Dgat1}^+/− mice is their decreased tissue levels of triglycerides. Recent studies have provided a strong correlation between tissue triglyceride content and insulin resistance (6, 14, 19, 20). Thus, the reduced triglyceride content in the skeletal muscle and liver of \textit{Dgat1}^+/− mice may enhance the insulin sensitivity of these tissues. Because we performed the hyperinsulinemic-euglycemic clamp studies with an insulin infusion rate that suppresses hepatic glucose production, further studies will be required to determine the relative contribution of the liver to the increased systemic insulin sensitivity in \textit{Dgat1}^+/− mice.

Several additional features of the \textit{Dgat1}^+/− phenotype have been associated with, and may contribute to, the increased insulin sensitivity in these mice. Decreased adipocyte size, as we observed in \textit{Dgat1}^+/− mice, is associated with increased insulin sensitivity (21). Because muscle contraction enhances the insulin sensitivity of skeletal muscles, the increased physical activity in \textit{Dgat1}^+/− mice (4) may contribute to their enhanced glucose disposal during high-fat feeding. Finally, leptin enhances insulin action in several murine models of diabetes (10, 22, 23), and \textit{DGAT1} deficiency appears to increase activation of the leptin pathway.

The marked reduction of plasma insulin levels in \textit{A^n/a} mice clearly demonstrates that \textit{DGAT1} deficiency enhances insulin sensitivity and may provide insight into the underlying molecular mechanism. The ratio of plasma insulin to glucose levels, a simple estimation of insulin resistance, is approximately 0.67 in \textit{A^n/a Dgat1}^+/− mice, whereas it is 0.13 in \textit{A^n/a Dgat1}^+/− mice. Thus, a relatively small change in body weight (∼20%) is associated with a remarkable reduction in insulin resistance (∼80%). This finding suggests that \textit{DGAT1} deficiency may directly enhance insulin signaling in target tissues. Interestingly, the levels of diacylglycerol, a molecule thought to antagonize insulin signaling by activating isoforms of typical protein kinase C (24), tended to be decreased in \textit{Dgat1}^+/− tissues. Thus, changes in diacylglycerol levels, or perhaps fatty acyl CoAs, may alter intracellular signaling pathways to enhance insulin signaling. It will be of interest to examine the activities of protein kinase C-θ (7) and IκB kinase-β (25), which may have important roles in the regulation of insulin signaling by intracellular lipids. Our findings do not exclude a hormonal effect of \textit{DGAT1} deficiency on insulin sensitivity. For example, adipocytes secrete several proteins that modulate glucose metabolism, for example, adiponectin (26, 27), TNF-α (28), and resistin (29). \textit{DGAT1} deficiency in \textit{WAT} could alter the secretion of one or more of these molecules, thereby affecting systemic glucose metabolism.

The findings of enhanced weight loss in response to leptin infusion and changes in the expression of leptin-regulated genes that are consistent with increased activation of the leptin pathway indicate that \textit{Dgat1}^+/− mice have increased leptin sensitivity. Although obesity is clearly associated with leptin resistance in both humans and animal models (9, 30), less evidence exists to indicate that decreasing adiposity and, specifically, decreasing tissue triglyceride content, can increase leptin sensitivity. To our knowledge, our findings are among the first to indicate that decreased tissue triglyceride content is associated with increased leptin sensitivity.

The increased leptin sensitivity in \textit{Dgat1}^+/− mice appears primarily to affect energy expenditure without a comparable effect on food intake. In response to leptin infusion, \textit{Dgat1}^+/− mice lost more weight but did not eat less than \textit{Dgat1}^+/− mice. One possible explanation for these findings is that leptin sensitivity in \textit{Dgat1}^+/− mice is increased predominantly in peripheral tissues. There is accumulating evidence that leptin has direct, peripheral effects on lipolysis (18), adipogenesis (17), and fatty acid oxidation (31). Because \textit{DGAT1} is highly expressed in peripheral tissues that have important roles in energy and lipid metabolism (e.g., skeletal muscle, liver, and \textit{WAT}) (2), \textit{DGAT1} deficiency may somehow enhance peripheral leptin sensitivity without affecting the hypothalamic leptin pathway. It would be of interest to determine whether other murine models of increased leanness and obesity resistance (e.g., mice lacking protein tyrosine phosphatase-1B) (32, 33, reviewed in ref. 34) have a similar response to leptin infusion (i.e., increased weight loss without an enhanced decrease in food intake). If not, it would suggest that \textit{DGAT1} deficiency might be unique in selectively enhancing the peripheral effects of leptin.

Although the increased physical activity in \textit{Dgat1}^+/− mice (4) may contribute to their enhanced insulin sensitivity on a high-fat diet, it is less clear whether this finding can help to explain their increased leptin sensitivity. To our knowledge, no studies have directly examined the effects of physical activity on leptin sensitivity. Recent findings, however, suggest that physical activity could enhance peripheral leptin sensitivity. Muscle contraction, for example, activates 5′-AMP-activated protein kinase, which mediates the effects of leptin on fatty acid oxidation (31). Thus, the increased physical activity in \textit{Dgat1}^+/− mice fed a high-fat diet may help to enhance leptin-mediated fatty acid oxidation.

\textit{DGAT1} deficiency protected against obesity and insulin resistance in mice fed a high-fat diet and in \textit{A^n/a} mice, but had no apparent effect in \textit{ob/ob} mice. The lack of phenotypic effects of \textit{DGAT1} deficiency in the setting of leptin deficiency is similar to what we observed for changes of sebaceous gland atrophy and hair loss in \textit{Dgat1}^−/− mice (35). Why is the phenotype of \textit{DGAT1} deficiency not apparent in the setting of leptin deficiency? One possibility is that the effects of \textit{DGAT1} deficiency are masked by the severe metabolic derangement of leptin deficiency. This seems unlikely, however, because several other murine models of decreased adiposity, for example, mice lacking perilipin (36) or high-mobility group protein I-C (37), can counter the obesity of \textit{ob/ob} mice. A more likely explanation is that the
effects of DGAT1 deficiency on energy and glucose metabolism require a functional leptin pathway. In the absence of leptin, for example, an alternative pathway of triglyceride synthesis may be upregulated to compensate for the loss of DGAT1, thereby eliminating the effects of DGAT1 deficiency on energy and glucose metabolism. Indeed, as we found previously in the skin of Dgat1−/− mice (35), DGAT2 mRNA expression was upregulated approximately threefold in WAT of leptin-deficient ob/ob mice. Importantly, this upregulation was not observed in high-fat feeding and in A1/a mice, conditions in which DGAT1 deficiency confers protection against insulin resistance and obesity. This increase in DGAT2 mRNA expression may adequately compensate for DGAT1 deficiency in the absence of leptin.

In conclusion, DGAT1 deficiency reduced tissue triglyceride levels, and these reductions were associated with increased insulin and leptin sensitivity. Moreover, DGAT1 deficiency protected against insulin resistance and obesity in agouti yellow mice, a model of severe leptin resistance. Because most human obesity is associated with insulin and leptin resistance, understanding the potential mechanisms of enhancing insulin and leptin sensitivity has significant medical relevance. Our findings suggest that pharmacological inhibition of DGAT1, by reversing insulin and leptin resistance, may represent an effective therapy for diabetes and obesity.

Acknowledgments

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