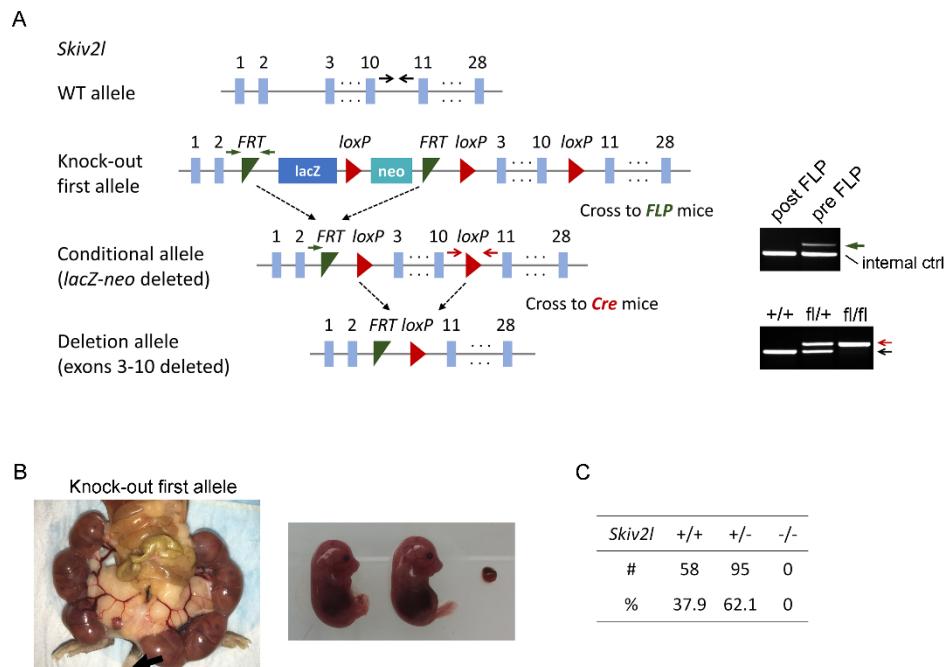


## Supplementary Figures



**Supplemental Figure 1. Generation of *Skiv2l* germline and conditional knockout mice**

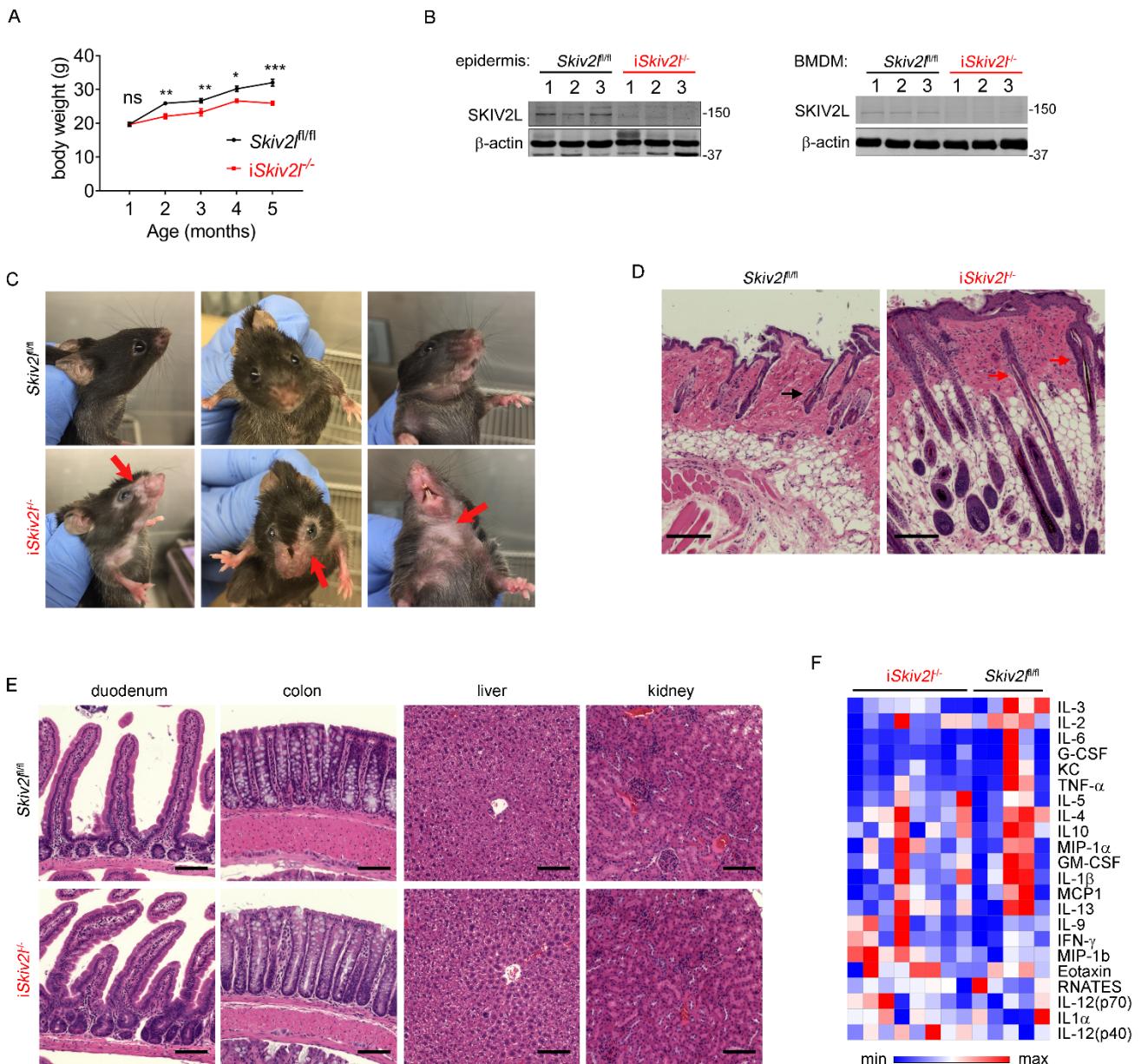
(A) The breeding scheme of *Skiv2l*<sup>f/f</sup> mice and genotyping results of knockout-first and conditional alleles (right graphs).

(B) Representative embryos from intercrossing knockout-first-allele heterozygous (*Skiv2l*<sup>+/+</sup>) female breeder mouse at embryonic day E13.5. Arrows indicated absorbed dead embryos. Morphologically normal embryos were either wild-type or heterozygotes.

(C) A summary of viable pups from *Skiv2l*<sup>+/+</sup> intercrossing.

C

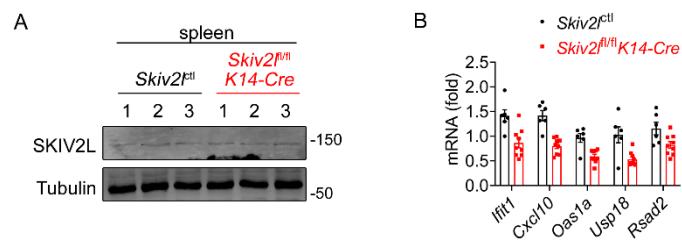
<i>Skiv2l</i>	+/+	+/-	-/-
#	58	95	0
%	37.9	62.1	0



### Supplemental Figure 2. Characterization of postnatal inducible whole-body *Skiv2l* knockout mice.

- (A) Body weight of *iSkiv2l<sup>-/-</sup>* male mice and *Skiv2l<sup>fl/fl</sup>* littermate controls. n=4-8 mice per genotype. Unpaired two-sided Student's *t*-test, \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001. ns, not significant.
- (B) Western blot analysis of SKIV2L in epidermis and BMDM from *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls. n=3 mice per genotype.
- (C) Skin lesion, alopecia of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls at 6 weeks of age.
- (D) H&E staining analysis of hair follicles (HFs) in dorsal skin of 3-month-old *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* littermate control. Black arrow, healthy HF without inflammation. Red arrow, dystrophic anagen HF with clustering inflammatory cells. Scale bar, 200 μm.
- (E) H&E staining of indicated organs of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls at 3 months of age. Scale bar, 100 μm.

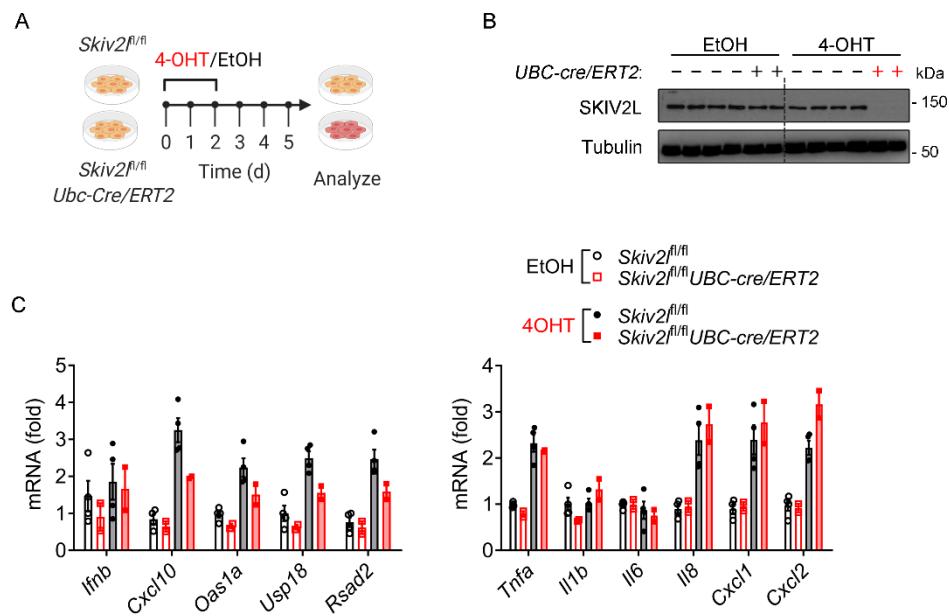
(F) Cytokine array analysis of serum of *iSkiv2l*<sup>-/-</sup> male mice (n=8) and sex-matched *Skiv2l*<sup>f/f</sup> littermates (n=5) at 2 months of age.



**Supplemental Figure 3. Germline keratinocyte-specific *Skiv2l* knockout mice do not show IFN signature in epidermis.**

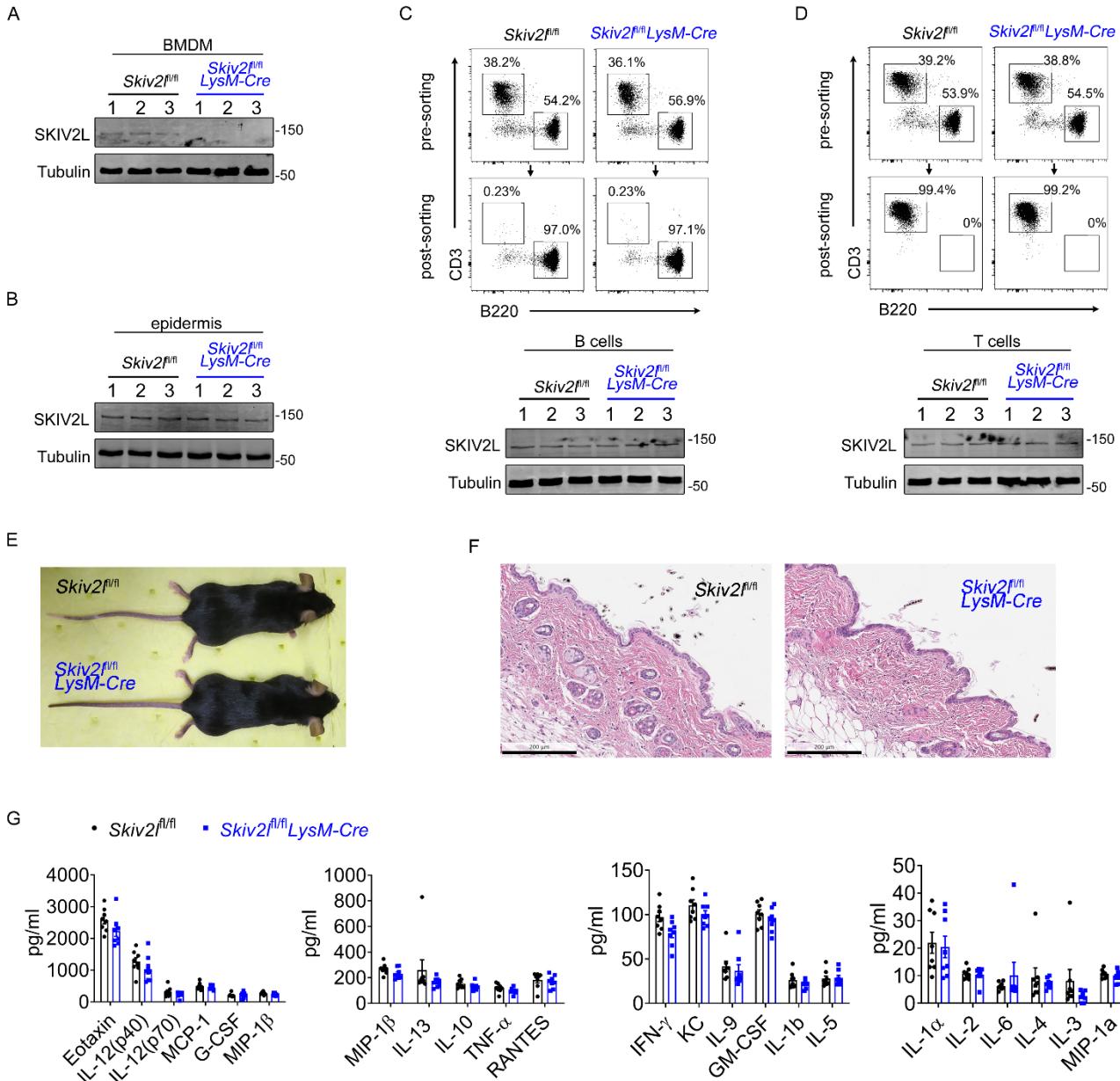
(A) Western blot analysis of SKIV2L in spleen from *Skiv2l*<sup>ctl</sup> and *Skiv2l*<sup>f/f</sup>*K14-Cre* P0 pup. n=3 mice per genotype.

(B) qPCR analysis of immune gene expression in P0 *Skiv2l*<sup>ctl</sup> (n=6) and *Skiv2l*<sup>f/f</sup>*K14-Cre* (n=9) pup epidermis.



**Supplemental Figure 4. Ex vivo deletion of *Skiv2l* in primary keratinocytes does not trigger IFN response.**

- (A) A schematic diagram showing experimental design for ex vitro deletion of *Skiv2l* in primary mouse neonatal keratinocytes. Primary mouse neonatal keratinocytes were isolated from *Skiv2l<sup>fl/fl</sup> UBC-Cre/ERT2* P0 pups and *Skiv2l<sup>fl/fl</sup>* littermates, and treated with 4-hydroxytamoxifen (4-OHT) or vehicle (ethanol, EtOH) for 48 h followed by 72-h culture.
- (B) Western blot analysis of SKIV2L in primary mouse neonatal keratinocytes from *Skiv2l<sup>fl/fl</sup> UBC-Cre/ERT2* P0 pups and *Skiv2l<sup>fl/fl</sup>* littermates after 4-OHT treatment as in (A).
- (C) qPCR analysis of IFNs, IFN-stimulated genes ISGs, inflammatory cytokine, and chemokine mRNA expression in neonatal keratinocytes of *Skiv2l<sup>fl/fl</sup> UBC-Cre/ERT2* mouse and littermate after 4-OHT treatment as in (A).



**Supplemental Figure 5. Germline myeloid-specific *Skiv2l* knockout does not cause skin disease or systemic inflammation.**

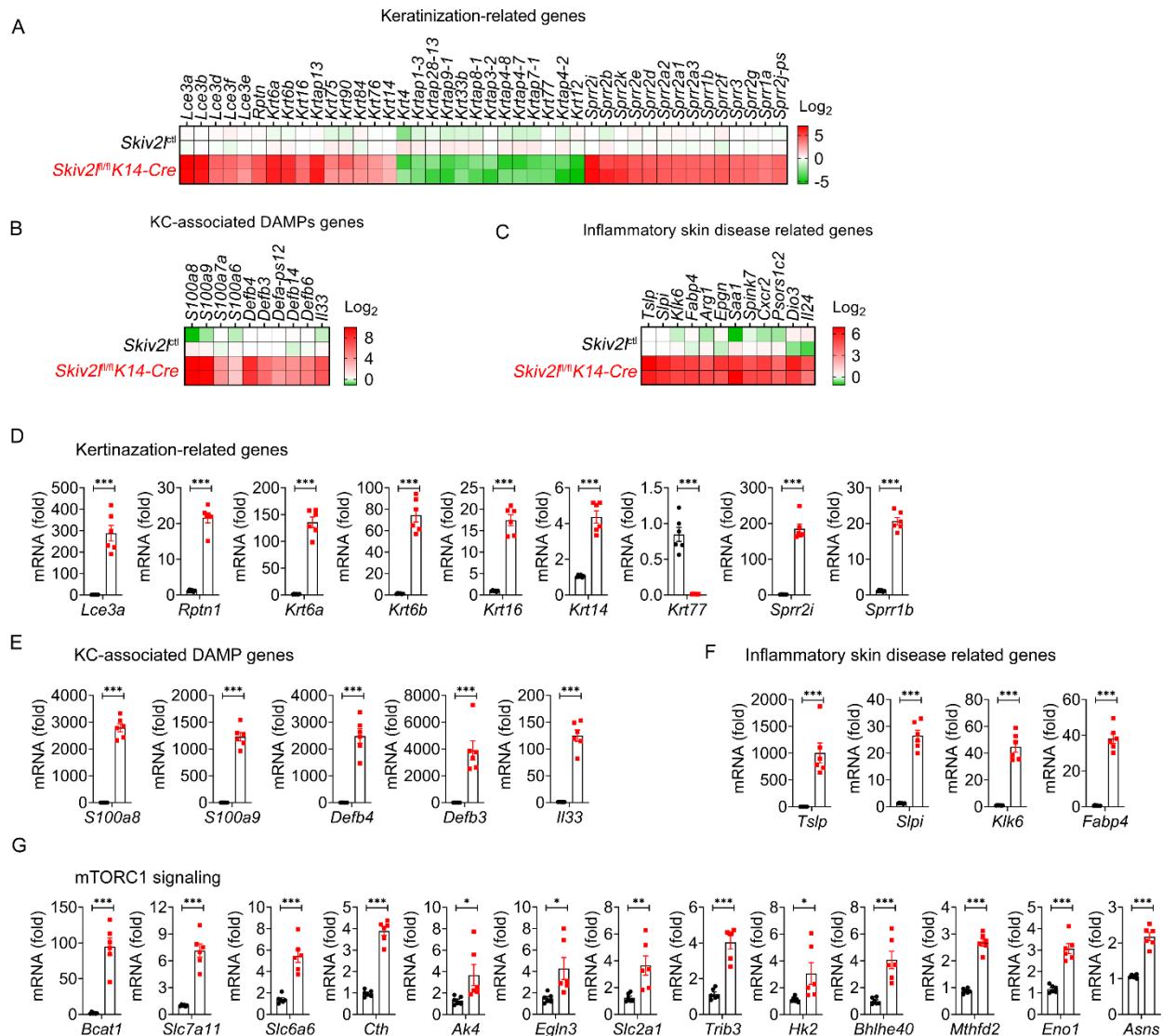
(A, B) Western blot analysis of SKIV2L in BMDM (A) or epidermis (B) of *Skiv2l<sup>fl/fl</sup>LysM-Cre* mice and *Skiv2l<sup>fl/fl</sup>* littermate control. n=3 mice per genotype.

(C, D) Western blot analysis of SKIV2L in sorted splenic B cells (C) or T cells (D) of *Skiv2l<sup>fl/fl</sup>LysM-Cre* mice and *Skiv2l<sup>fl/fl</sup>* littermate control. Flow cytometry analysis of sorting purity is shown in upper panels. n=3 mice per genotype.

(E) Skin and hair appearance of 7-month-old *Skiv2l<sup>fl/fl</sup>LysM-Cre* (germline myeloid-specific *Skiv2l* knockout) mice and *Skiv2l<sup>fl/fl</sup>* littermates.

(F) H&E staining of dorsal skin of 7-month-old *Skiv2l<sup>fl/fl</sup>LysM-Cre* mice and *Skiv2l<sup>fl/fl</sup>* littermates. Scale bar, 200 μm.

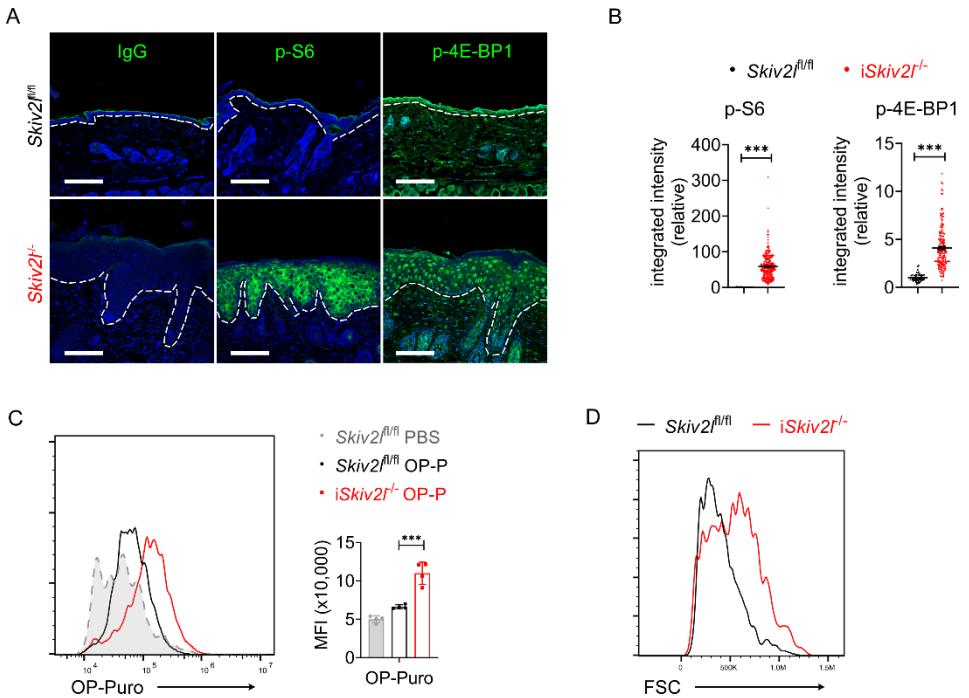
**(G)** Serum cytokine analysis of 7-month-old *Skiv2l<sup>f/f</sup>**LysM-Cre* female mice and *Skiv2l<sup>f/f</sup>* littermates. n=9 per genotype.



**Supplemental Figure 6. Dysregulated pathways in epidermis of germline keratinocyte-specific *Skiv2l* knockout mice.**

**(A-C)** A heatmap showing mRNA expression of keratinization-related genes, keratinocyte-associated DAMPs genes and inflammatory skin disease related genes in *Skiv2l<sup>ctl</sup>* and *Skiv2l<sup>fl/fl</sup>K14-Cre* (germline keratinocyte-specific *Skiv2l* knockout) P0 epidermis (n=2 mice per genotype). Relative mRNA level of each gene is shown after normalizing FPKM of each sample to that of the average value of *Skiv2l<sup>ctl</sup>* samples.

**(D-G)** qRT-PCR analysis of representative gene expression in *Skiv2l<sup>ctd</sup>* and *Skiv2l<sup>f/f</sup>K14-Cre* P0 epidermis. n=6 mice per genotype. Two-sided Student's *t*-test, \**P*<0.05, \*\**P*<0.01, \*\*\**P*<0.001.

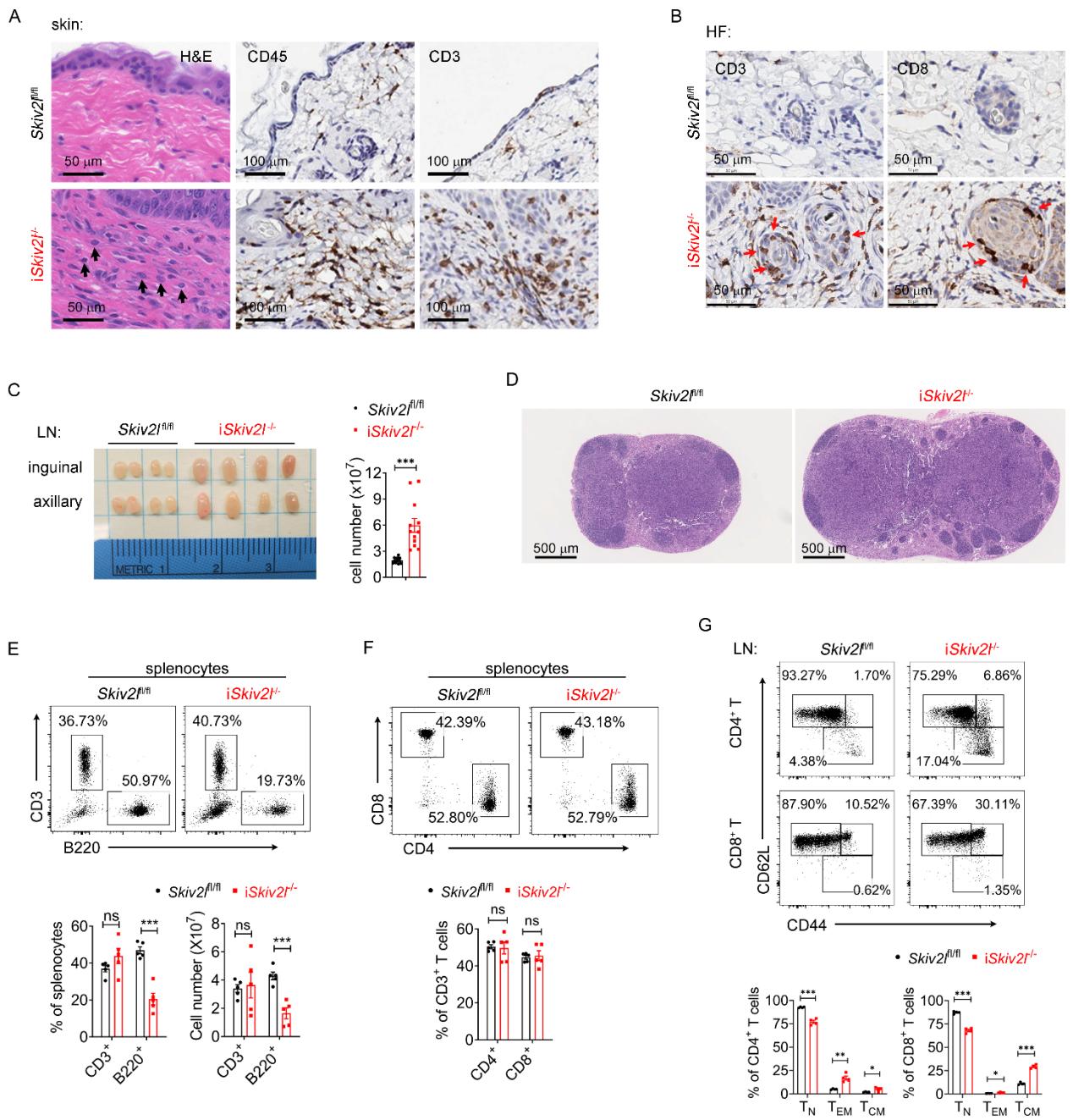


**Supplemental Figure 7. Aberrant activation of the mTORC1 pathway in epidermis of postnatal whole-body inducible *Skiv2l* knockout mice.**

(A, B) Fluorescent immunohistochemistry analysis of phospho-S6 ribosomal protein (S235/236) and phospho-4E-BP1 (T37/46) in dorsal skin of *iSkiv2l<sup>-/-</sup>* and *Skiv2l<sup>f/f</sup>* mice at 3 months of age (A). Dashed line, epidermal-dermal junction. Scale bar, 50  $\mu$ m. Quantification of pS6 or p4E-BP1 fluorescence intensity per cell (>60 cells each genotype) is shown in (B). Two-sided unpaired Student's *t*-test, \*\*\**P*<0.001.

(C) Global protein translation measured by OP-Puro incorporation in keratinocyte of *iSkiv2l<sup>-/-</sup>* and *Skiv2l<sup>f/f</sup>* mice in vivo. n=4 mice per group. *Skiv2l<sup>f/f</sup>* mice were injected with OP-Puro or PBS and analyzed 1 h after administration. One-way ANOVA with post hoc Tukey's multiple comparison, \*\*\**P*<0.001.

(D) Flow cytometry analysis of keratinocyte cell size (indicated by FSC). Keratinocytes were isolated from *iSkiv2l<sup>-/-</sup>* and *Skiv2l<sup>f/f</sup>* mice and analyzed by FACS.



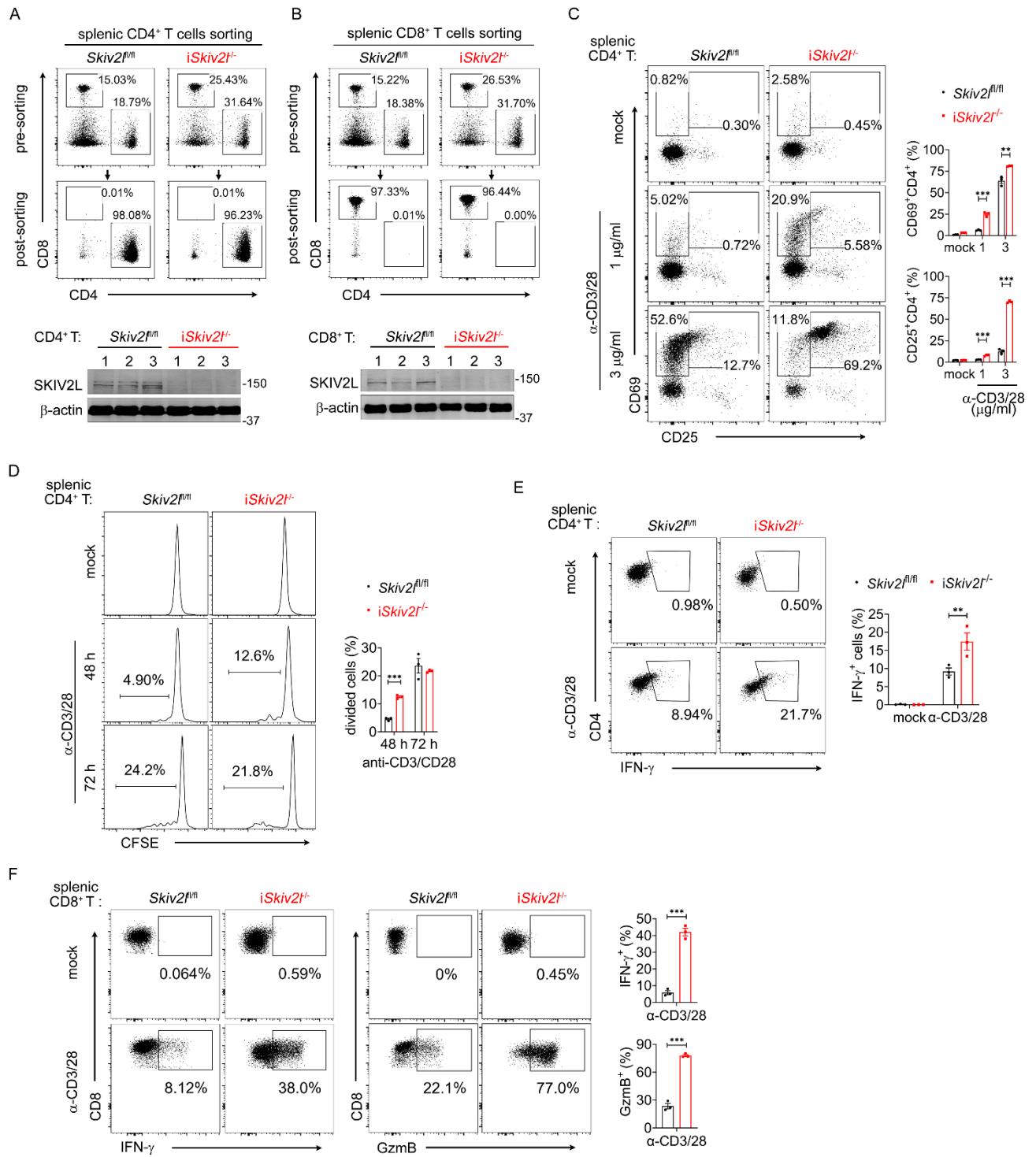
**Supplemental Figure 8. Disruption of immune homeostasis in postnatal whole-body inducible *Skiv2l* knockout mice.**

(A) H&E staining and immunohistochemistry analysis of leukocytes (CD45) and T cells (CD3) in dorsal skin of iSkiv2l<sup>-/-</sup> mice and Skiv2l<sup>fl/fl</sup> littermate control (3-month-old). Black arrows, immune infiltrates in dermis. The H&E staining images of Skiv2l<sup>fl/fl</sup> (top left) and iSkiv2l<sup>-/-</sup> (bottom left) are different crops of images shown in Figure 1C (top right and bottom right, respectively).

(B) Immunohistochemistry analysis of T cells (CD3, CD8) in hair follicles (HF) of 3-month-old iSkiv2l<sup>-/-</sup> mice and Skiv2l<sup>fl/fl</sup> littermate control. Red arrows, immune infiltrates. Scale bar, 50  $\mu$ m.

(C) Skin-draining inguinal and axillary lymph nodes and their cell numbers (lymph nodes from one side were counted). n=12 mice per genotype. Two-sided Student's t-test, \*\*\*P<0.001.

- (D)** H&E staining of inguinal lymph nodes of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* littermate controls (3-month-old).
- (E)** Flow cytometry analysis of CD3<sup>+</sup> T cells and B220<sup>+</sup> B cells in splenocytes of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls. n=5 mice per genotype. Two-sided Student's *t*-test, \*\*\*P<0.001, ns, not significant.
- (F)** Flow cytometry analysis of CD4<sup>+</sup> T cells and CD8<sup>+</sup> B cells in spelnocytes (gated on CD3<sup>+</sup>) of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls. n=5 mice per genotype. Two-sided Student's *t*-test; ns, not significant.
- (G)** Flow cytometry analysis of *iSkiv2l<sup>-/-</sup>* and *Skiv2l<sup>fl/fl</sup>* inguinal lymph node T cells. Numbers adjacent to each gate indicate the percentage of each population. n=3 mice per genotype. Two-sided Student's *t*-test, \*P<0.05, \*\*P<0.01, \*\*\*P<0.001.



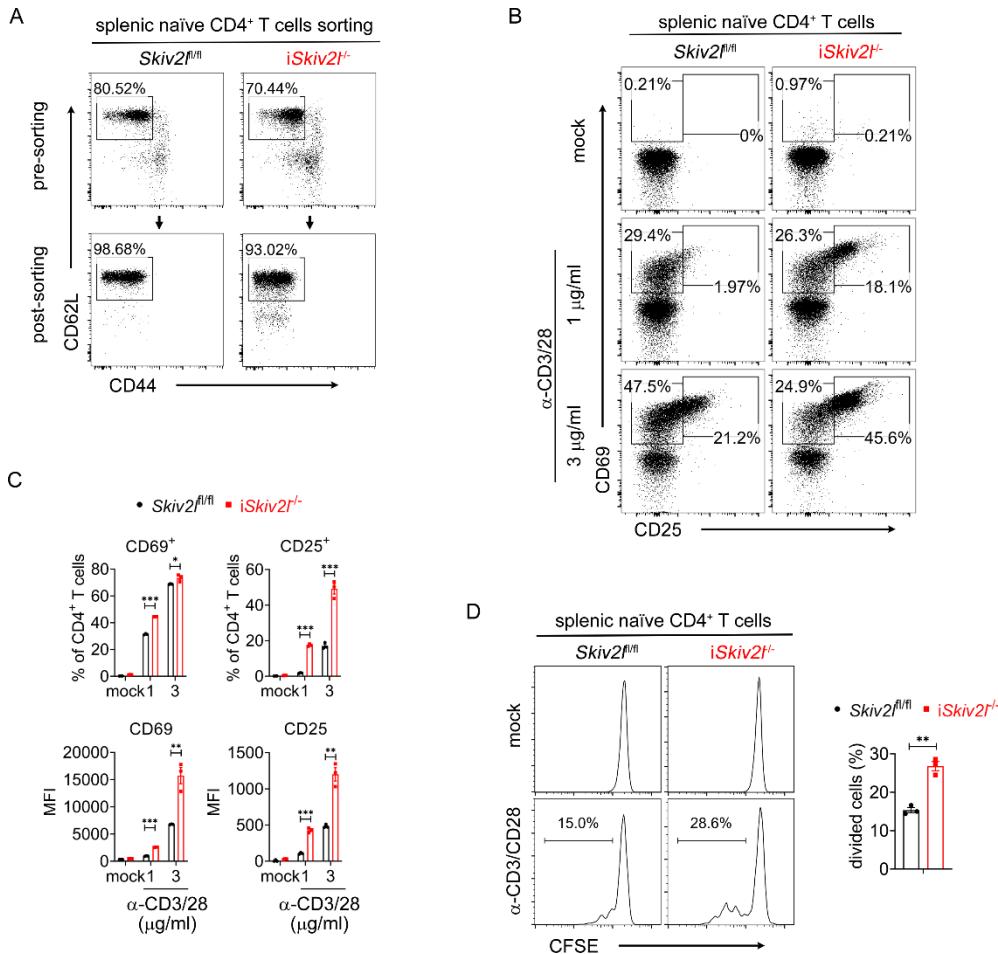
**Supplemental Figure 9. T-cell hyperactivation in postnatal inducible whole-body *Skiv2l* knockout mice.**

(A, B) Western blot analysis of SKIV2L in sorted splenic CD4<sup>+</sup> (A) or CD8<sup>+</sup> T cells (B) of *iSkiv2l-/-* mice and *Skiv2l<sup>fl/fl</sup>* littermate controls. Flow cytometry analysis of sorting purity is shown in upper panels. n=3 mice per genotype.

(C) Expression of activation markers of *iSkiv2l-/-* and *Skiv2l<sup>fl/fl</sup>* splenic CD4<sup>+</sup> T cells stimulated with indicated concentration of anti-CD3 and anti-CD28 antibodies for 16 h. n=3 per genotype. Two-sided Student's *t*-test, \*\*P<0.01, \*\*\*P<0.001.

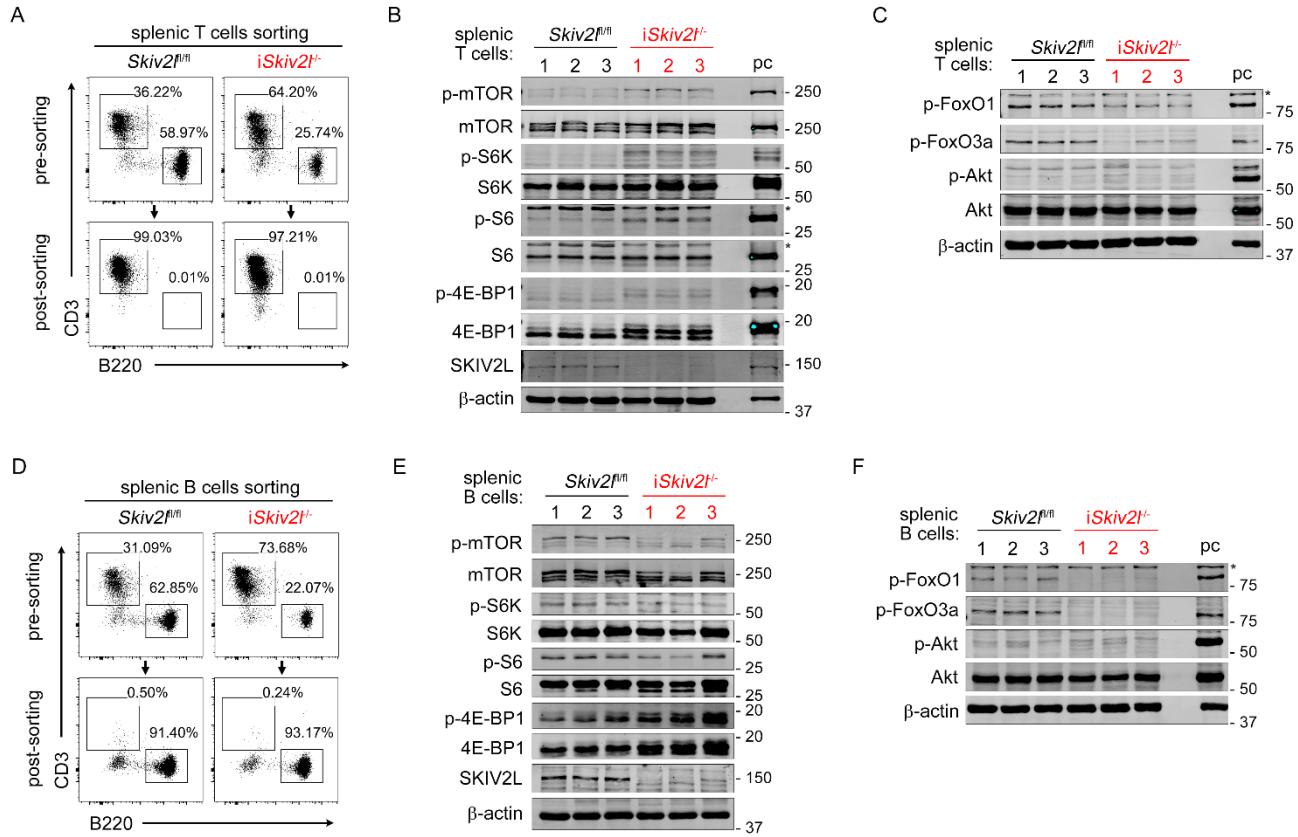
**(D)** T cell proliferation analysis by CFSE dilution assay. i*Skiv2l*<sup>-/-</sup> and *Skiv2l*<sup>f/f</sup> splenic CD4<sup>+</sup> T cells were stained with CFSE then stimulated with anti-CD3 and anti-CD28 (3 µg/ml) for indicated times. n=3 per genotype. Two-sided Student's *t*-test, \*\*\*P<0.001.

**(E, F)** Intracellular IFN-γ or granzyme B (GzmB) staining of i*Skiv2l*<sup>-/-</sup> and *Skiv2l*<sup>f/f</sup> splenic CD4<sup>+</sup> T (**E**) and CD8<sup>+</sup> T cells (**F**) after stimulation with anti-CD3 and anti-CD28 (3 µg/ml). Two-sided Student's *t*-test, \*\*P<0.01, \*\*\*P<0.001.



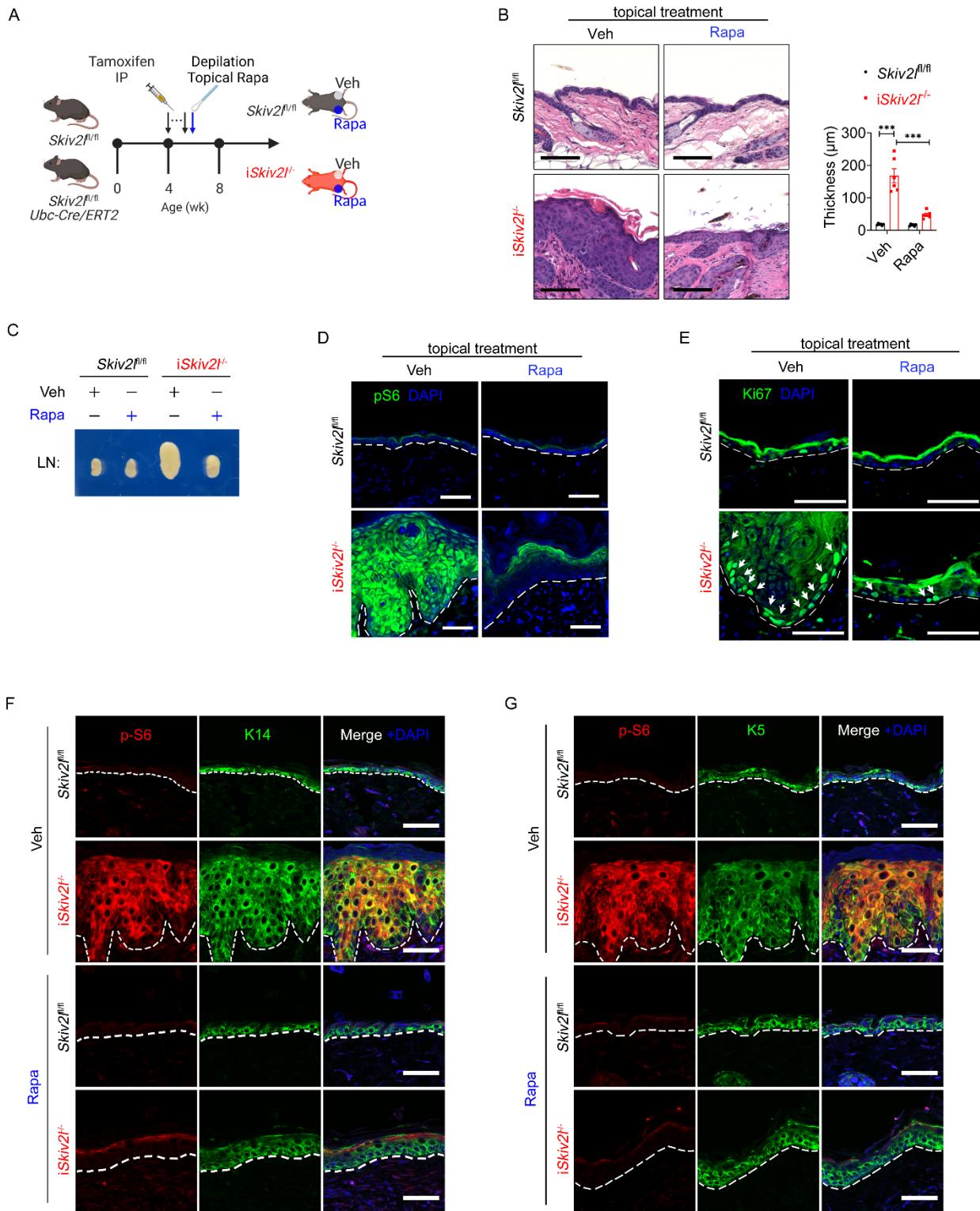
**Supplemental Figure 10. Hyperactive response of *Skiv2l*-deficient naïve CD4<sup>+</sup> T cells.**

- (A) Flow cytometry analysis of naïve CD4<sup>+</sup> T cells purity after sorting from *iSkiv2l*<sup>-/-</sup> and *Skiv2l*<sup>fl/fl</sup> splenocytes.
- (B, C) Expression of activation markers CD69 and CD25 of *iSkiv2l*<sup>-/-</sup> and *Skiv2l*<sup>fl/fl</sup> naïve CD4<sup>+</sup> T cells stimulated with indicated concentration of anti-CD3 and anti-CD28 antibodies for 16 h. n=3 per genotype. Two-sided Student's *t*-test, \*P<0.05, \*\*P<0.01, \*\*\*P<0.001.
- (D) T cell proliferation analysis by CFSE dilution assay. *iSkiv2l*<sup>-/-</sup> and *Skiv2l*<sup>fl/fl</sup> naïve CD4<sup>+</sup> T cells were stained with CFSE then stimulated with anti-CD3 and anti-CD28 (3 μg/ml) for 72 h. n=3 per genotype. Two-sided Student's *t*-test, \*\*P<0.01.



**Supplemental Figure 11. Activation of the mTORC1 pathway in T cells of postnatal inducible whole-body *Skiv2l* knockout mice.**

- (A) Flow cytometry analysis of T cells purity after sorting of *iSkiv2l<sup>-/-</sup>* and *Skiv2l<sup>fl/fl</sup>* splenocytes.
- (B, C) Western blot analysis of mTORC1 (B) and mTORC2 (C) pathway in sorted splenic T cells of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls. n=3 mice per genotype. Whole cell lysate of B16 melanoma cells was used as positive control (pc). \*, non-specific band.
- (D) Flow cytometry analysis of B cells purity after sorting of *iSkiv2l<sup>-/-</sup>* and *Skiv2l<sup>fl/fl</sup>* splenocytes.
- (E, F) Western blot analysis of mTORC1 (E) and mTORC2 (F) pathway in sorted splenic B cells of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls. n=3 mice per genotype. Whole cell lysate for B16 melanoma cells was used as positive control (pc). \*, non-specific band.



**Supplemental Figure 12. Topical rapamycin treatment ameliorates skin pathology of postnatal whole-body inducible *Skiv2l* knockout mice**

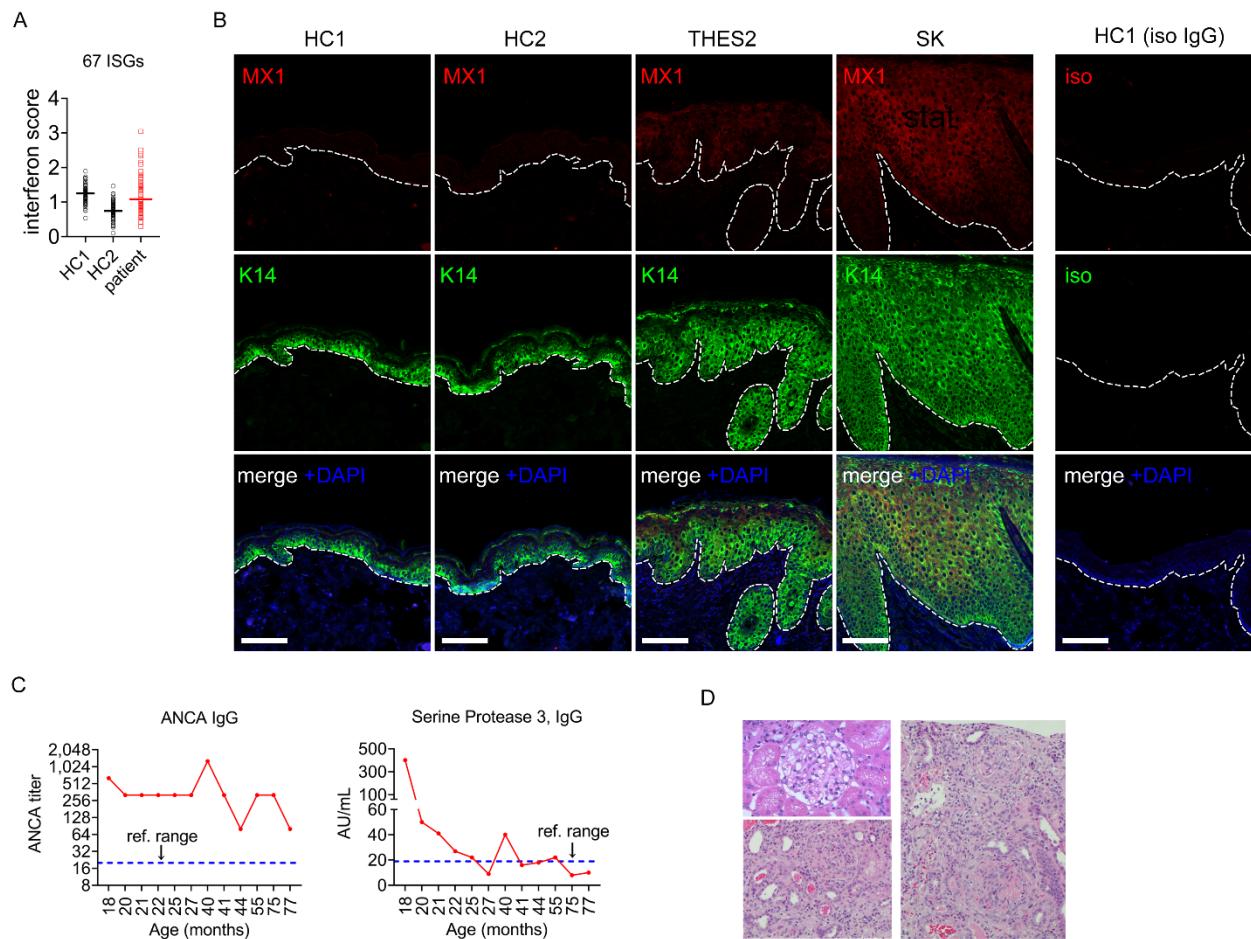
(A) A schematic diagram of experimental design for topical rapamycin treatment of *iSkiv2l<sup>-/-</sup>* mice and *Skiv2l<sup>fl/fl</sup>* controls after tamoxifen injection and depilation.

**(B)** H&E staining of iSkiv2l<sup>-/-</sup> and Skiv2l<sup>f/f</sup> mice skin topically treated with rapamycin or vehicle as above (A). Scale bar, 50  $\mu$ m. Quantification of epidermal thickness is shown on the right bar graph. n=6 mice per group. Two-way ANOVA with post hoc Tukey's multiple comparisons test, \*\*\*P<0.001.

**(C)** Skin-draining inguinal and axillary lymph nodes of iSkiv2l<sup>-/-</sup> and Skiv2l<sup>f/f</sup> mice skin topically treated with rapamycin or vehicle.

**(D, E)** Fluorescent immunohistochemistry analysis of phospho-S6 ribosomal protein (S235/236) **(D)** and proliferation marker Ki67 **(E)** in iSkiv2l<sup>-/-</sup> and Skiv2l<sup>f/f</sup> mouse skin treated with rapamycin or vehicle. White arrows in **(E)**, Ki67 positive cells. Dashed line, epidermal-dermal junction. Scale bar, 50  $\mu$ m.

**(F, G)** Fluorescent immunohistochemistry analysis of phospho-S6 ribosomal protein (S235/236) and keratinocyte marker K14 **(F)** or K5 **(G)** in iSkiv2l<sup>-/-</sup> and Skiv2l<sup>f/f</sup> mouse skin treated with rapamycin or vehicle. Dashed line, epidermal-dermal junction. Dashed line, epidermal-dermal junction. Scale bar, 50  $\mu$ m.



**Supplemental Figure 13. Immunological parameters of the THES2 patient.**

- (A) Interferon score from average expression of 67 ISGs in PBMCs from the THES2 patient and two unrelated healthy controls (HC).
- (B) Fluorescent immunohistochemistry analysis of MX1 and K14 in skin biopsies from healthy controls, THES2, and an unrelated case of seborrheic keratosis (SK). Dashed line, epidermal-dermal junction. Scale bar, 100  $\mu$ m. The skin biopsy of THES2 is from the same patient as presented in **Figure 7E**.
- (C) Titers of anti-neutrophil cytoplasmic antibody (ANCA) and serine protease 3 IgG antibodies in THES2 patient. Patient's titers are shown in solid red line and reference range are in dashed blue line.
- (D) H&E staining of kidney biopsy at 17-month old indicates pauci-immune necrotizing and crescentic glomerulonephritis (NCGN), necrotizing arteriolitis, acute tubulointerstitial nephritis, global glomerulosclerosis, interstitial fibrosis and tubular atrophy.

**Supplemental Table 1. qRT-PCR primer sequences**

<b>Gene</b>	<b>Primer</b>	<b>Sequence 5'-3'</b>
<i>Ifit1</i>	Fwd	TTCACATGGAAGCTGCTATTGAAA
	Rev	TGCTCAGCTGCTCGCTGGATCAA
<i>Cxcl10</i>	Fwd	GGGATCCCTCTCGCAAGGACGGTCC
	Rev	ACGCTTCATTAAATTCTTGATGGT
<i>Oas1a</i>	Fwd	GGATGGCATAGATTCTGGGA
	Rev	CTGCATCAGGAGGTGGAGTT
<i>Usp18</i>	Fwd	GTGTCCGTGATCTGGTCCTT
	Rev	CTGCAGAAATACAACGTGCC
<i>Rsad2</i>	Fwd	GGACGCTTCATGGTGTATTG
	Rev	TGATTGGTCGCCTGTTATCT
<i>Ifnb</i>	Fwd	CTGCGTTCCCTGCTGTGCTTCTCCA
	Rev	TTCTCCGTCATCTCCATAGGGATC
<i>Tnfa</i>	Fwd	CTACCTTGTGCTCCTCTTT
	Rev	GAGCAGAGGTTCACTGATGTAG
<i>Il1b</i>	Fwd	GGTACATCAGCACCTCACAA
	Rev	TTAGAAACAGTCCAGGCCATAC
<i>Il6</i>	Fwd	CACAAGTCCGGAGAGGGAGAC
	Rev	CAGAATTGCCATTGCACAAC
<i>Cxcl1</i>	Fwd	CGAACATAGCCACACTCAA
	Rev	GAGCAGTCTGTCTTCTTCTCC
<i>Cxcl2</i>	Fwd	TAAGCACCGAGGAGAGTAGAA
	Rev	GTCCAAGGGTTACTCACAAACA
<i>Lce3a</i>	Fwd	GTCTGGGCTCTGTGTTCTT
	Rev	CATGGTTGGACACAGGTGAT
<i>Rptn</i>	Fwd	GAAGGAACACGGAGGCCATAAA
	Rev	CCTTCAGACTGATTGTGGTGAG
<i>Krt6a</i>	Fwd	CAACATCATAACCCTCCCTGTC
	Rev	GAGGAAGCCAAGAGCATCAA
<i>Krt6b</i>	Fwd	GTCTCTGAGTTGCCCTGGTAAAG
	Rev	CCAGGCCATTGGAAACTAGAA
<i>Krt16</i>	Fwd	CTCCTCTGGACAGTCCTATTCT
	Rev	GTCCCTGGAACTCTGACTTTG
<i>Krt14</i>	Fwd	GAGCGGCAAGAGTGAGATT
	Rev	CTTGGTCTCCTCCAGGTTATT
<i>Krt77</i>	Fwd	GCTGCTTCATGGGCAAATC
	Rev	GGGACAGCTCCGTATCAAATAG
<i>Sprr2i</i>	Fwd	TCCACATAGCACCTCCTCTA
	Rev	ATTCTCTGCAGGCCCTTAC
<i>Sprr1b</i>	Fwd	CCATATACCAGGCTCATCCATC
	Rev	GGCTGTTCACTTGTGCTC

<i>S100a8</i>	Fwd	CTTGTCAGCTCCGTCTCA
	Rev	TGTAGAGGGCATGGTGATTTC
<i>S100a9</i>	Fwd	GAGGAGTGTATGATGCTGATGG
	Rev	GTCACATGGCTGACCTCTTAAT
<i>Defb4</i>	Fwd	CGAAGAACAGCAAGATGAATAAA
	Rev	CTAGAACTGGAGTTAGAGAAGGTAATC
<i>Il33</i>	Fwd	GGGTACCAAGCATGAAGAGAA
	Rev	GTCAACAGACGCAGCAAATG
<i>Tslp</i>	Fwd	TCATGACCTGACTGGAGATTG
	Rev	AGCCAGGGATAGGATTGAGA
<i>Slpi</i>	Fwd	CGCCTCCTGGTAAAGACATAAA
	Rev	GGAGCACCGTGAAAGGTAAA
<i>Klk6</i>	Fwd	AGAGCACAGAACACTGCTAAAT
	Rev	CATCTGCTAACACCACCCATAG
<i>Fabp4</i>	Fwd	GCTCCTCCTCGAAGGTTAC
	Rev	CCCACTCCCACCTCTTCAT
<i>Bcat1</i>	Fwd	GTGGTAGCATTGTGGTAATA
	Rev	TCCTAACCGCTGGATTAAAG
<i>Slc7a11</i>	Fwd	GGGAAGGTGATGGCTGTATT
	Rev	CACCACTCAGCATCTGTGTATC
<i>Slc6a6</i>	Fwd	CTGCCAAGTGACCCCTTAATC
	Rev	TAACACGAGAGCAACCAGAAATA
<i>Cth</i>	Fwd	GGTCGGATGGAGAACATT
	Rev	AGAGGGTAGCCCAGGATAAA
<i>Ak4</i>	Fwd	CCCTTCACAGGAACGAGTATAG
	Rev	CCTGTGGAAGGTGGATAAA
<i>Egln3</i>	Fwd	GCCCAGGACTGCTTCTTATT
	Rev	TGGCATCTGTCACCAACTTA
<i>Slc2a1</i>	Fwd	AAGACTGCTGCTCAGATCTATT
	Rev	GGAGATAGGAGAGTGGCTGATA
<i>Trib3</i>	Fwd	GTCTCAGCTTCTCCTGACTTT
	Rev	CACTAGGCACAGGAACGAATAA
<i>Hk2</i>	Fwd	CTCGGCCTTGTGAGTAGATAGA
	Rev	CGCGGGTGAATGAGGTATT
<i>Bhlhe40</i>	Fwd	CTGCTGCCTGCTTCTTTC
	Rev	CTGAATCTCTGTGGGTCTG
<i>Mthfd2</i>	Fwd	GTGCTTGGACCAGTACTCTATG
	Rev	CCAGCCACTACCACATTCTT
<i>Eno1</i>	Fwd	GATGGACGGCACAGAGAATAAA
	Rev	TCAGCAATGTGGCGGTAAA
<i>Asns</i>	Fwd	CAGTGTCTGAGTGCATGAA
	Rev	CAAGCGGTGAAAGCCAAAG

<i>Actb</i>	Fwd	GAGGTATCCTGACCCTGAAGTA
	Rev	CACACGCAGCTCATTGTAGA

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