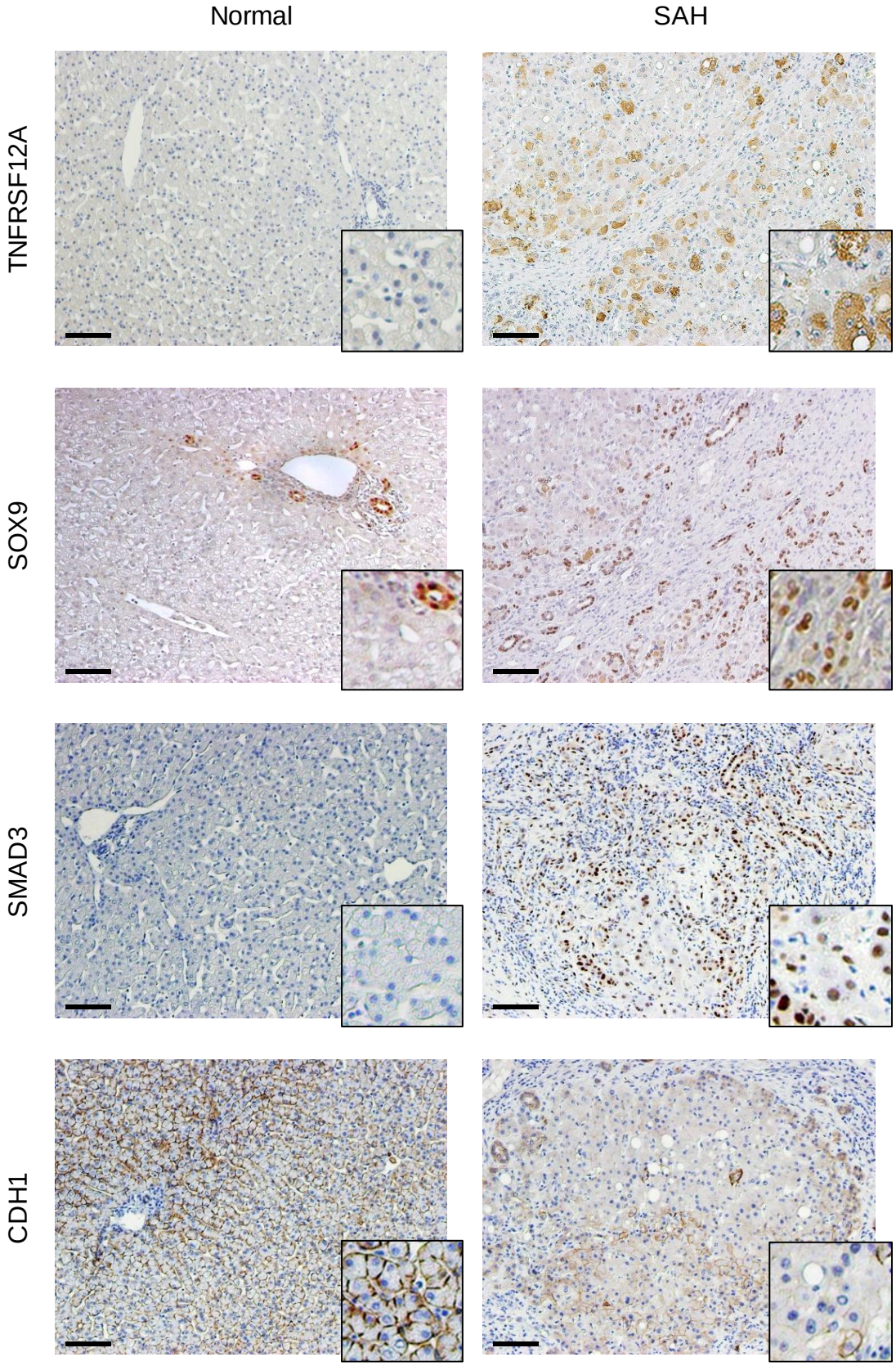
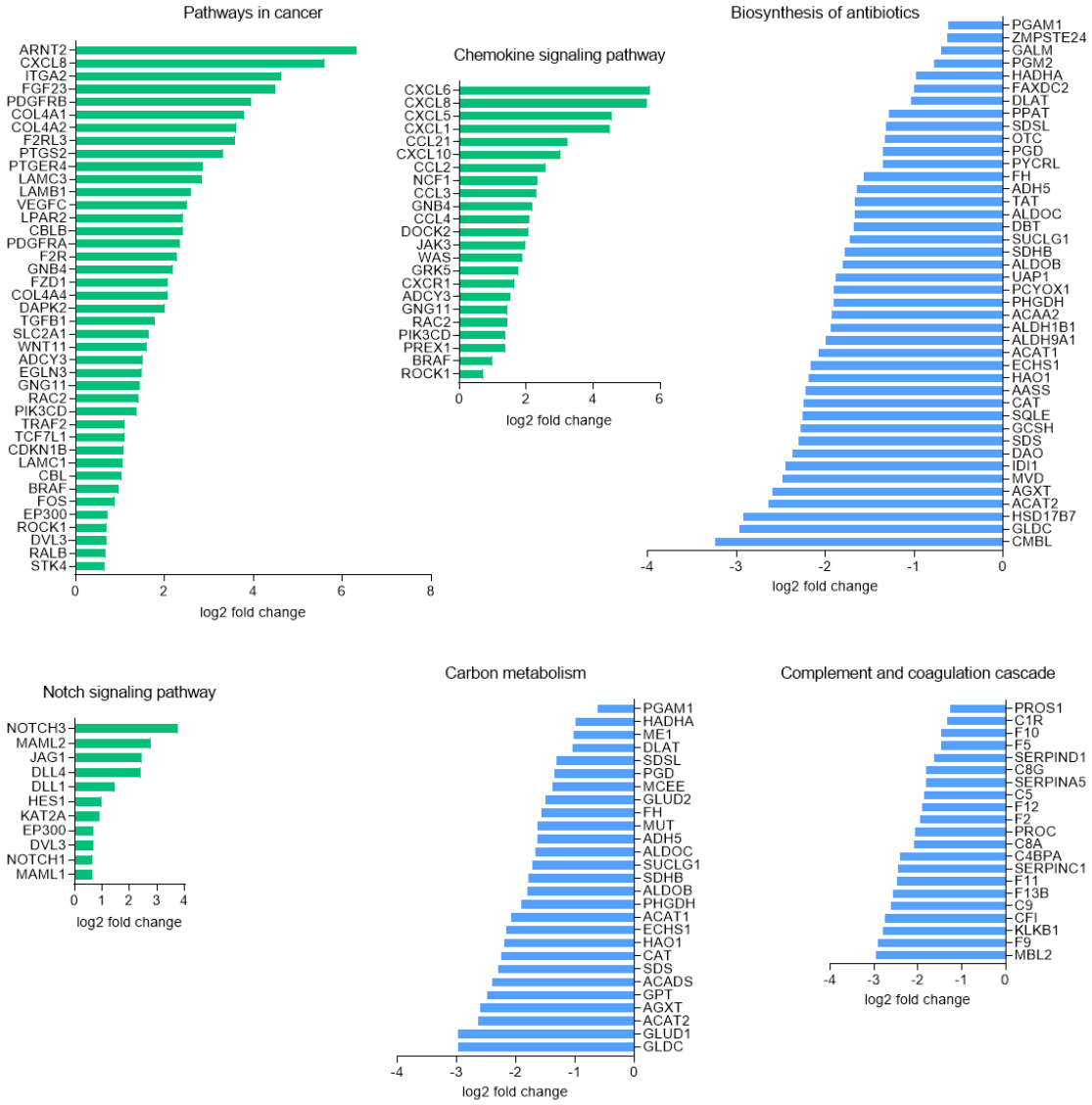
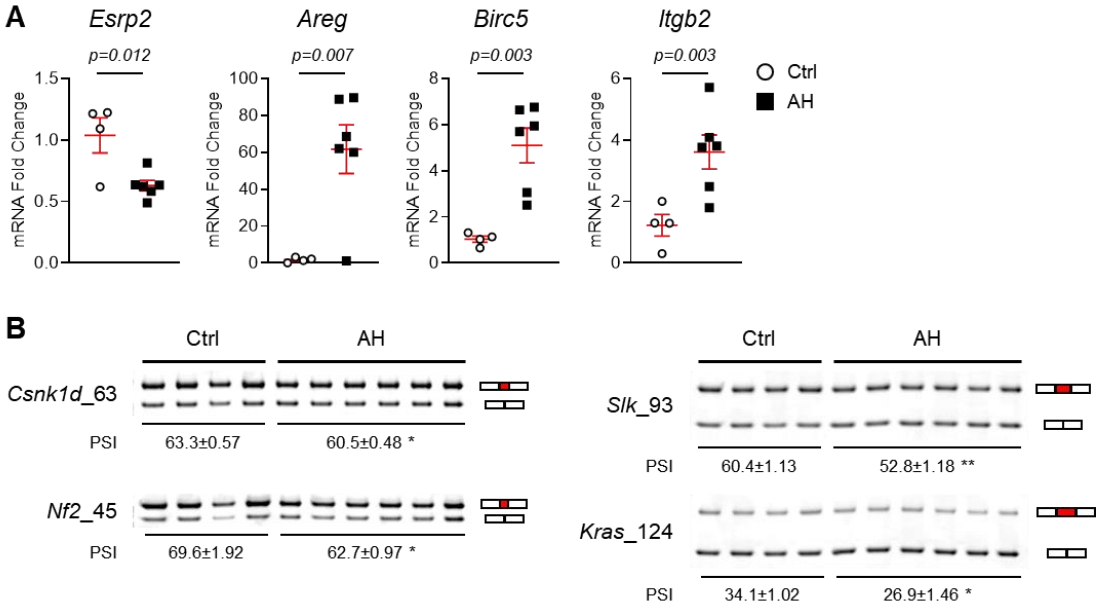

Normal
SAH
TNFRSF12A
SOX9
SMAD3
CDH1

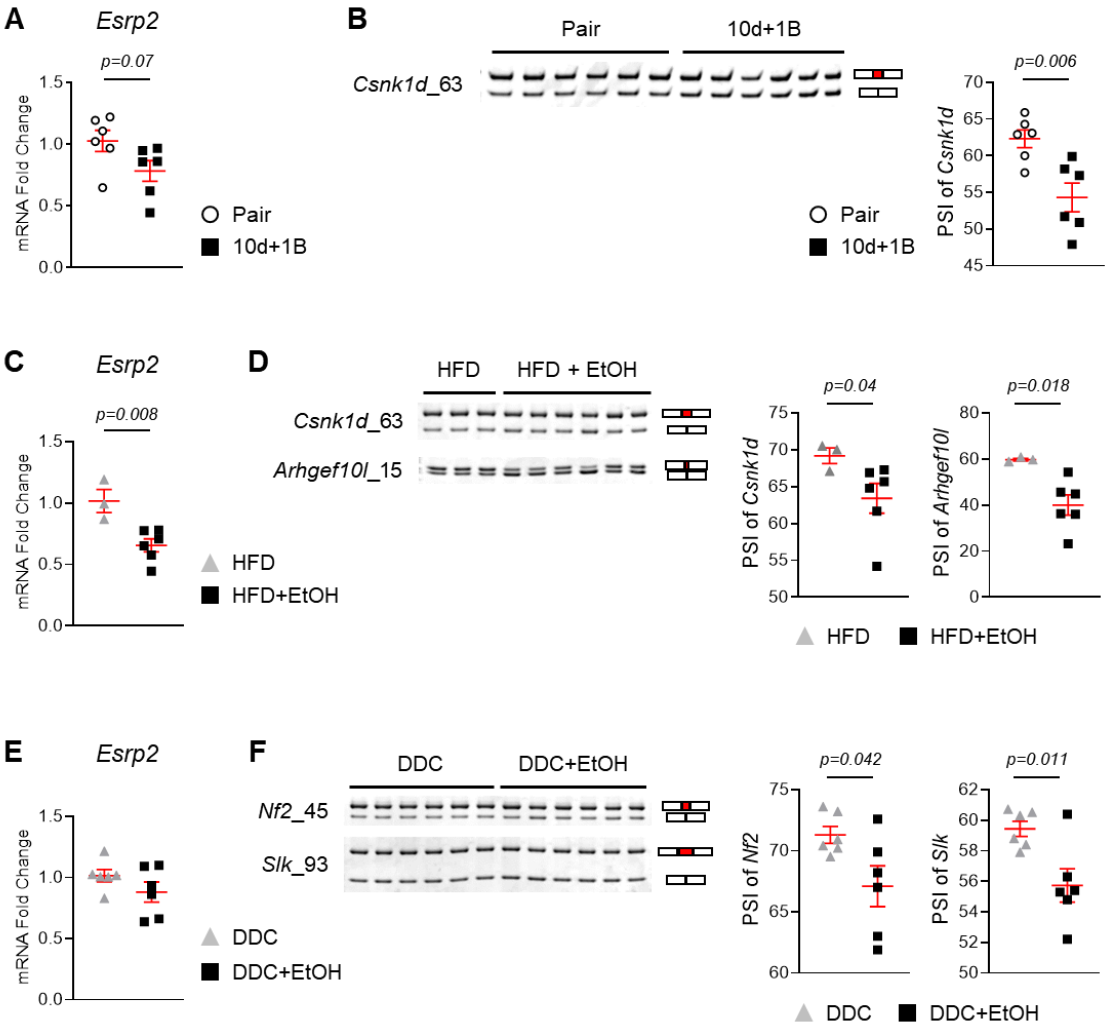
Supplemental Figure 1. Fetal-like liver cells accumulate in patients with severe alcoholic hepatitis. Immunohistochemistry (IHC) for TNFRSF12A, SOX9, SMAD3 and CDH1 in explanted liver tissues of patients with severe alcoholic hepatitis (SAH) and healthy human liver. Representative images are shown. Scale bar=100μm. Magnification=100×. Each higher magnification image (400×) is shown in right-bottom.



Supplemental Figure 2. Lists of genes that are involved in notable KEGG pathways.

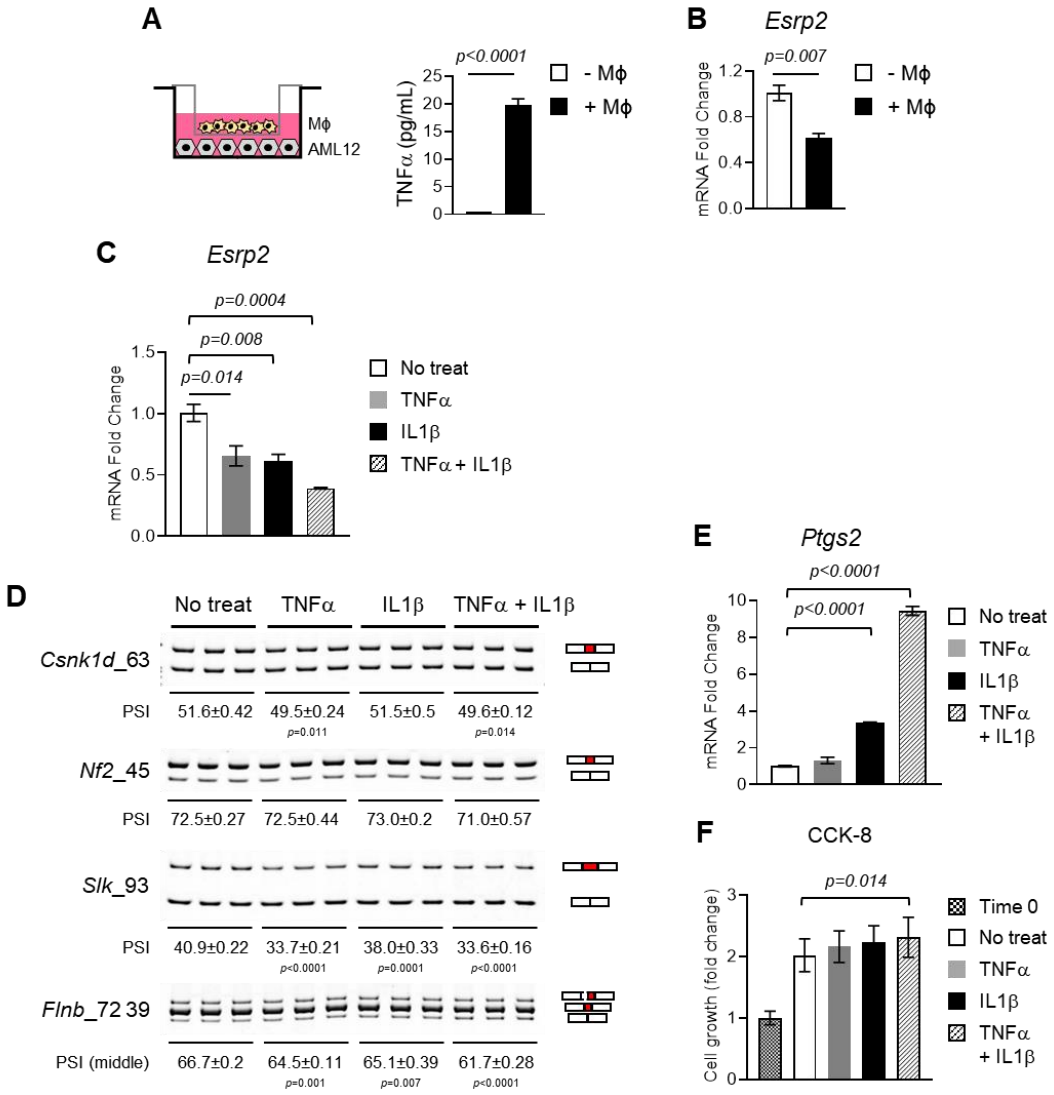


Supplemental Figure 3. Adult-to-fetal reprogramming is robust in Tsukamoto-French mouse model of alcoholic hepatitis. (A) qRT-PCR analysis for *Esrp2* and YAP/TAZ target genes (*Areg*, *Birc5*, *Itgb2*) in liver tissues of control (Ctrl) or alcoholic hepatitis (AH) mice of Tsukamoto-French model. Results are graphed as dot plots (AH: black square, Ctrl: white circle) with mean±s.e.m. (red bars). **(B)** Alternative splicing of ESRP2 target mRNAs including *Csnk1d*, *Nf2*, *Slk* and *Kras* in liver tissues of control or Tsukamoto-French AH mice. Results are shown as mean±s.e.m. below each image. Statistical analysis for both qRT-PCR and splicing assays was performed by using two-tailed student *t* test between two groups (n=4-6 mice/group, *p* values are specified otherwise, **p*<0.01, ***p*<0.005). Full unedited gel images are shown in Supplemental Material.



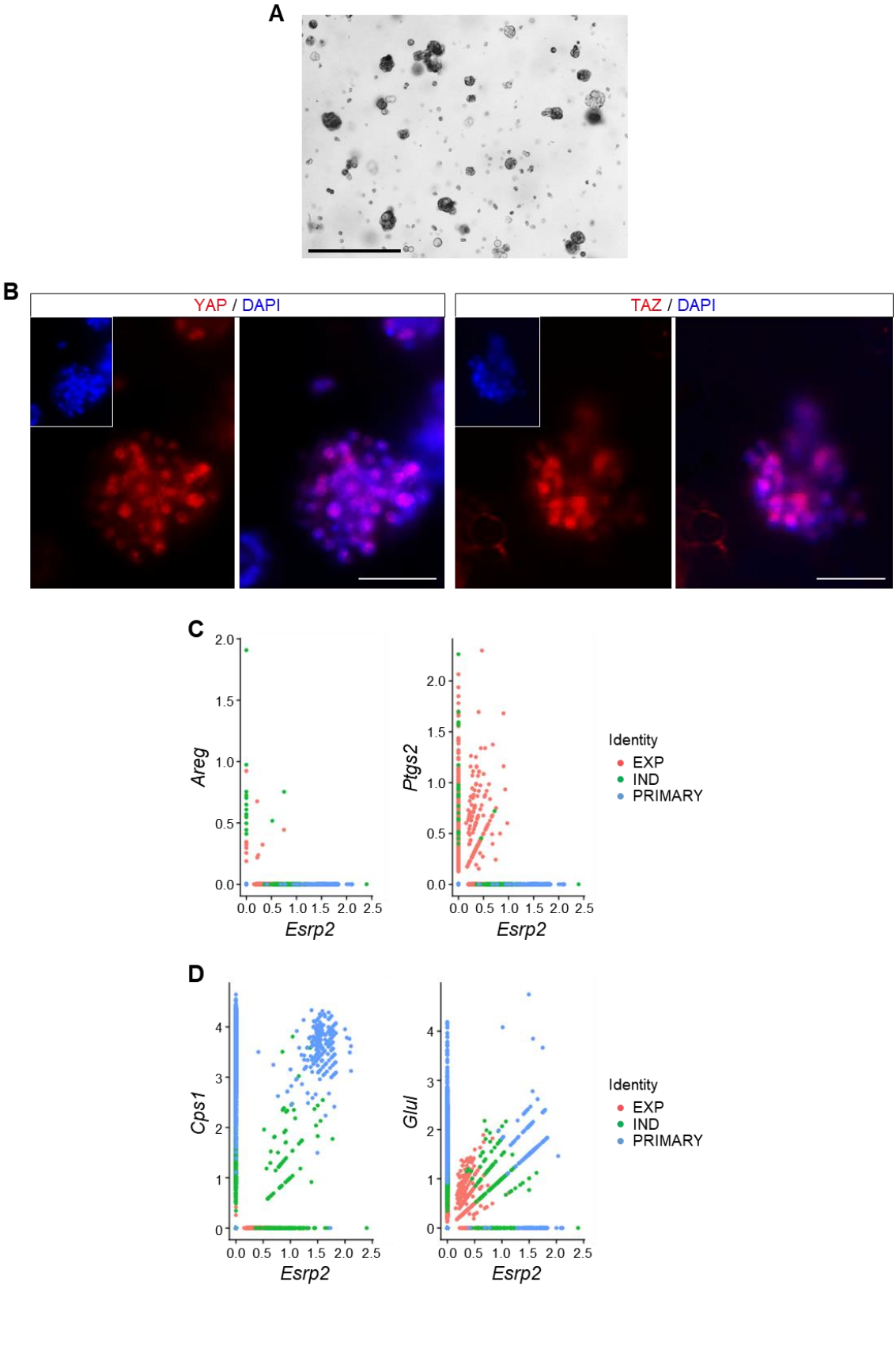
Supplemental Figure 4. Mouse models of alcohol-induced liver injury suppress ESRP2-mediated adult splicing program. (A and B) qRT-PCR analysis for *Esrp2* **(A)** and alternative splicing of *Csnk1d* in whole liver tissues of Gao Binge model (10d+10B) and pair-fed mice. 10d+1B, 10 days of ethanol feeding plus one binge. **(C and D)** qRT-PCR analysis for *Esrp2* **(C)** and alternative splicing of *Csnk1d* and *Arhgef10l* **(D)** in liver tissues of Lieber DeCarli high fat model (HFD+EtOH) and only HFD-fed mice. HFD, high fat diet; EtOH, ethanol. **(E and F)** qRT-PCR analysis for *Esrp2* **(E)** and alternative splicing of *Nf2* and *Slk* **(F)** in liver tissues of only DDC- or 'DDC+EtOH'-fed mice. All results are graphed as dot plots with mean±s.e.m. (n=3-6 mice/group). Statistical analysis was performed by using two-tailed student *t* test between two groups. The upper band includes the exon (red part), while the lower band skips

exon. The exon length is shown besides the gene name. PSI, percent spliced in; s.e.m., standard error of the mean. Full unedited gel images are shown in Supplemental Material.

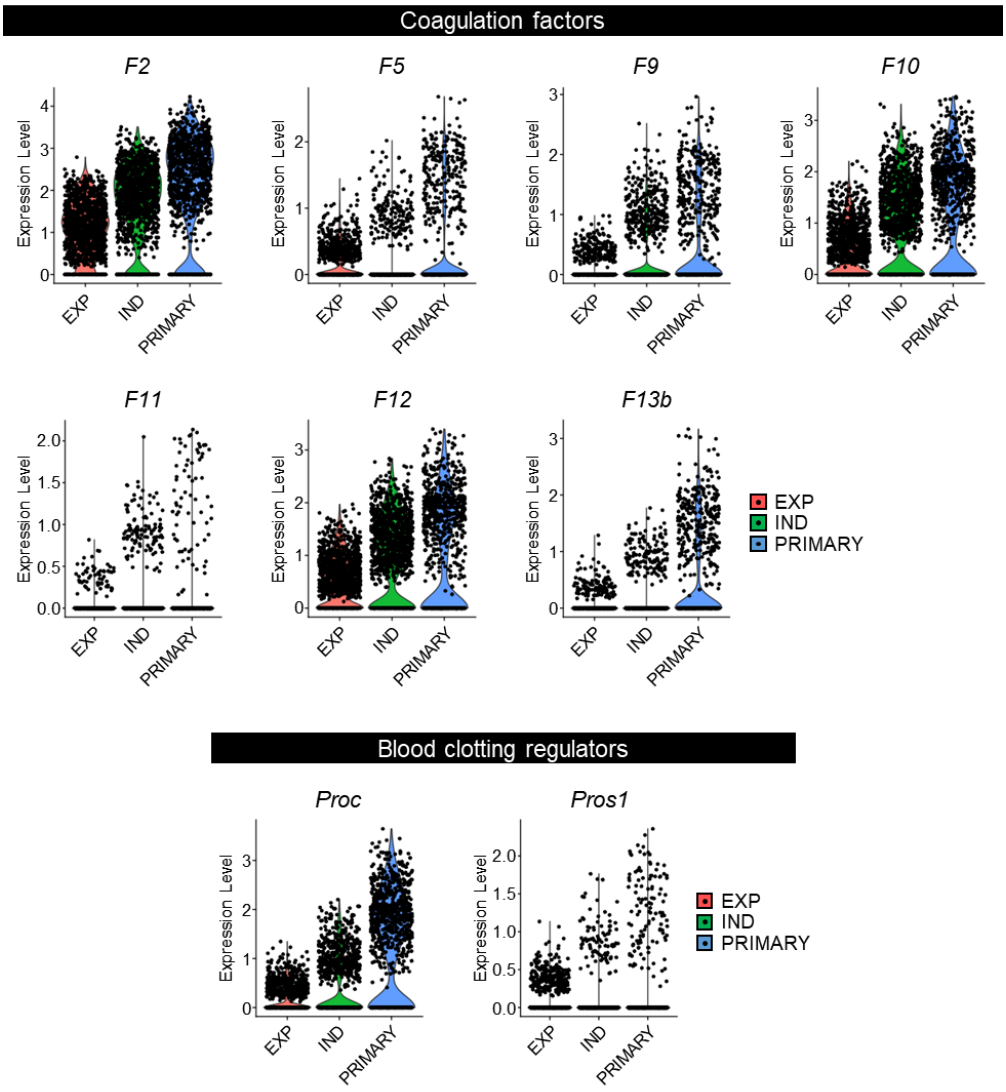


Supplemental Figure 5. Proinflammatory cytokines suppress ESRP2, reactivate fetal splicing program, and promote cell growth in hepatocytes. (A) AML12 cells (mouse hepatocyte cell line) were co-cultured with RAW264.7 cells (mouse macrophage cell line, Mϕ) for 24 hours in the Transwell system as depicted in the schematic image. ELISA assay was performed for the level of TNFα in conditioned medium of AML12 cells cultured alone (-Mϕ) or co-cultured with RAW264.7 cells (+Mϕ). **(B)** qRT-PCR analysis for *Esrp2* in AML12 cells cultured alone (-Mϕ) or co-cultured with RAW264.7 cells (+Mϕ). The mean±s.e.m. results are graphed, and the statistical analysis was performed by using two-tailed student *t* test between two groups. **(C)** qRT-PCR analysis for *Esrp2* in AML12 cells non-treated or treated with TNFα

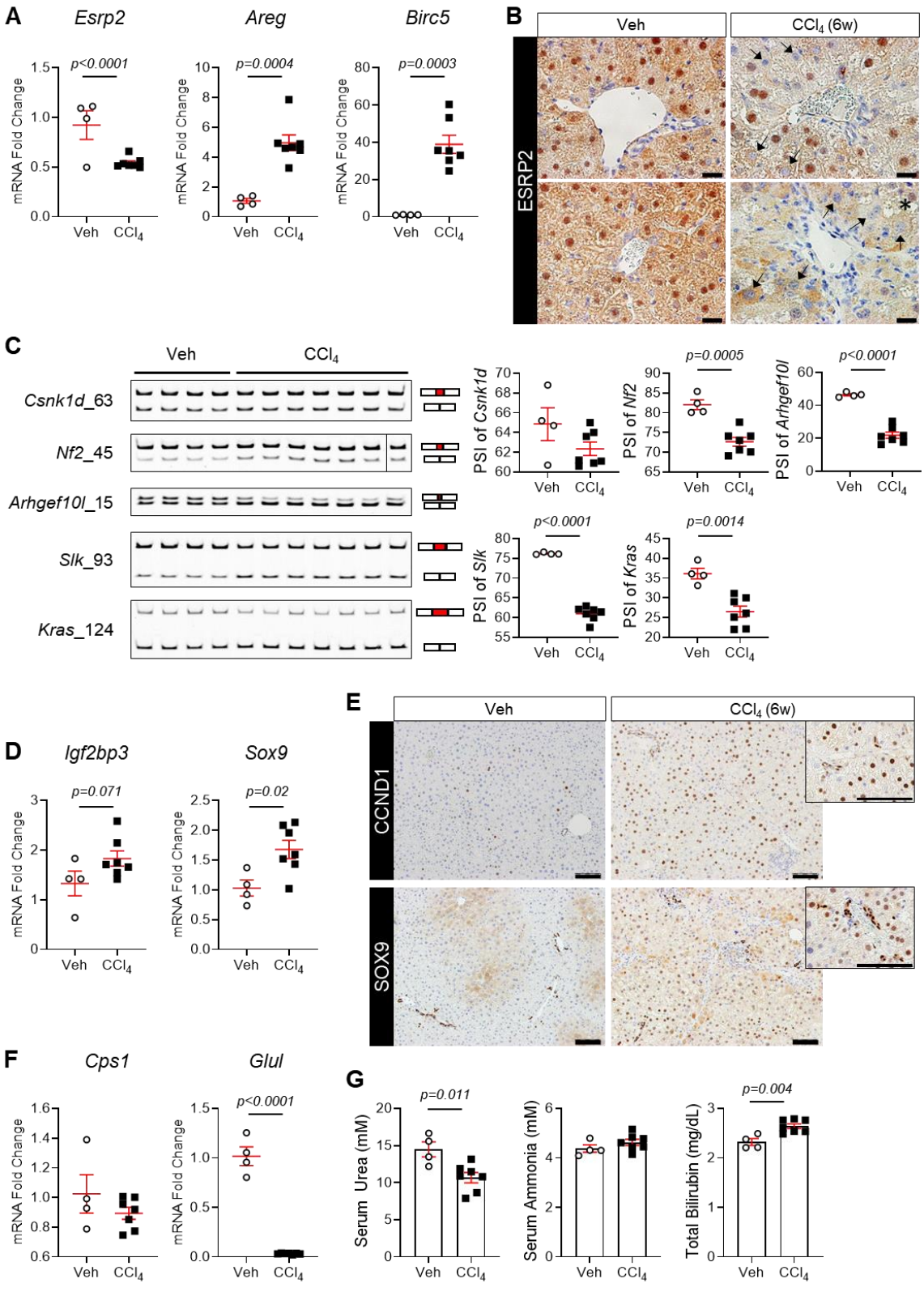
(100 ng/ml), IL1β (100 ng/ml), or TNFα + IL1β (50 ng/ml each) for 24 hours. **(D)** Alternative splicing of ESRP2 target mRNAs including *Csnk1d*, *Nf2*, *Slk* and *Flnb* in AML12 cells ± TNFα / IL1β / TNFα + IL1β treatment. Full unedited gel images are shown in Supplemental Material. **(E)** qRT-PCR analysis for a YAP/TAZ target gene, *Ptgs2*, in AML12 cells treated as designated. **(F)** Cell Counting Kit-8 (CCK-8) assay for the growth of AML12 cells at time 0 and 24 hours after treatment. The mean±s.e.m. results are graphed, and the statistical analysis was performed by one-way analysis of variation (ANOVA) with Tukey corrections (n=3 biological replicates/group).


A
B
YAP / DAPI
TAZ / DAPI
C
Areg
Ptgs2
Esrp2
Identity
EXP
IND
PRIMARY
D
Cps1
Glul
Esrp2

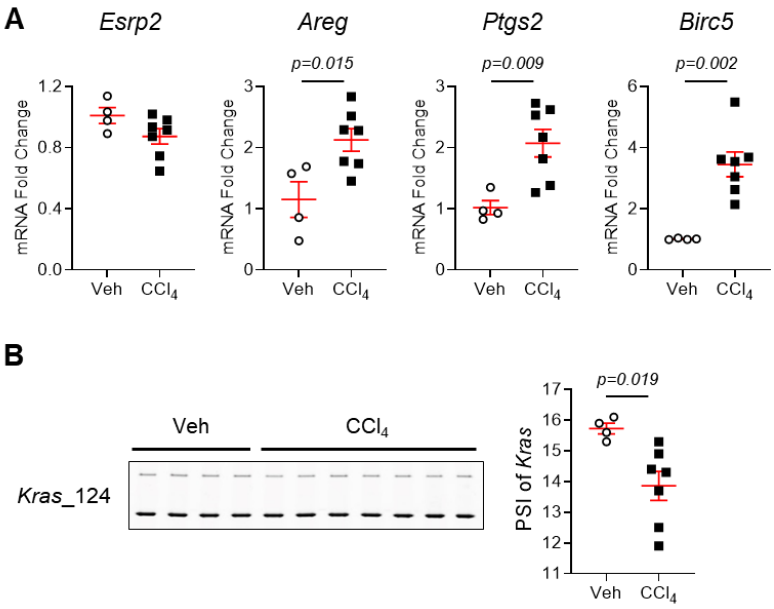
Supplemental Figure 6. As *Esrp2* decreases, YAP/TAZ target genes are upregulated and metabolic enzyme genes are downregulated in hepatocyte spheroids. (A) A representative brightfield image of primary hepatocytes under spheroid culture system for 18 days. Scale bar=10^5 µm, Magnification=50×. **(B)** Images of hepatocyte spheroids stained for YAP (left, red) and TAZ (right, red) visualized by whole mount immunofluorescence. Nuclear counterstaining was done by 4',6-diamidino-2-phenylindole (DAPI, blue) and merged images are shown. Scale bar=50µm. **(C and D)** Co-expression plots of *Esrp2* with mRNAs that encode YAP/TAZ targets (*Areg*, *Ptgs2*) **(C)** and metabolic enzymes (*Cps1*, *Glul*) **(D)** graphed from scRNA-seq data in primary hepatocytes ("PRIMARY") and hepatocyte spheroids under TNFα-mediated expansion medium ("EXP") or TNFα-withdrawn induction medium ("IND") as described in a previous publication [35].



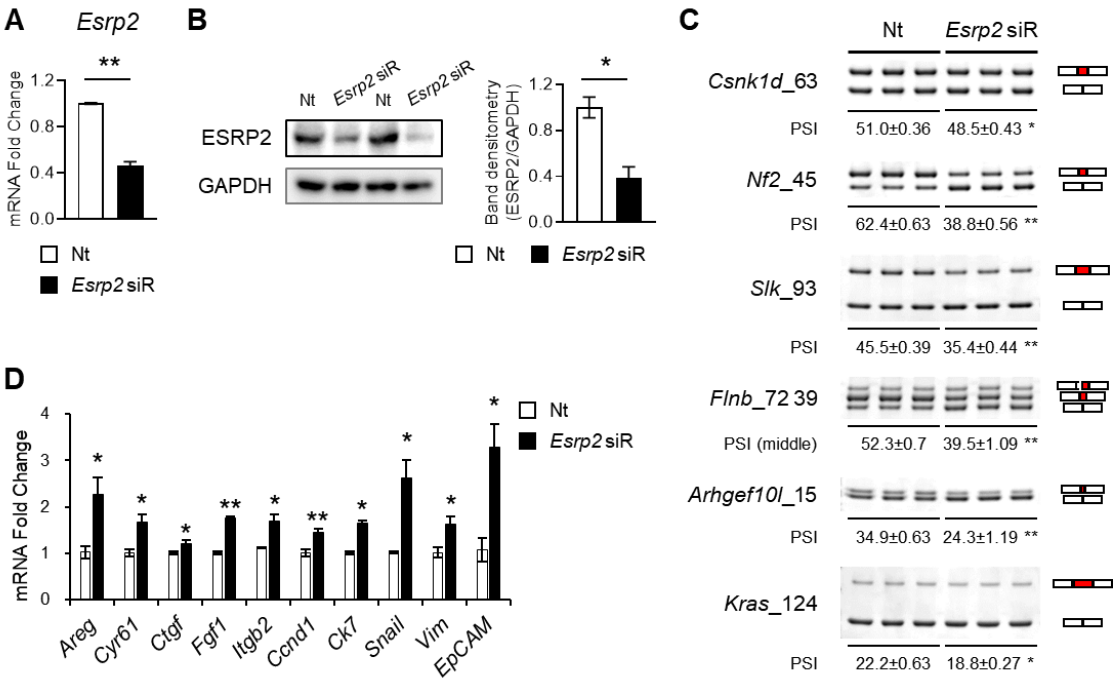
Supplemental Figure 7. The coagulation function is impaired in fetal-like hepatocytes. scRNA-seq analysis for coagulation factors (*F2, F5, F9, F10, F11, F12, F13b*) and blood clotting regulators (*Proc, Pros1*) in primary hepatocytes ("PRIMARY") and hepatocyte spheroids under TNFα-mediated expansion medium ("EXP") or TNFα-withdrawn induction medium ("IND") as described in a previous publication [35].


A
Esrp2
Areg
Birc5
p<0.0001
p=0.0004
p=0.0003
mRNA Fold Change
Veh
CCl4
B
Veh
CCl4 (6w)
ESRP2
C
Veh
CCl4
Csnk1d_63
Nf2_45
Arhgef10l_15
Slk_93
Kras_124
PSI of Csnk1d
PSI of Nf2
PSI of Arhgef10l
PSI of Slk
PSI of Kras
p=0.0005
p<0.0001
p<0.0001
p=0.0014
D
Igf2bp3
Sox9
p=0.071
p=0.02
E
Veh
CCl4 (6w)
CCND1
SOX9
F
Cps1
Glul
p<0.0001
G
Serum Urea (mM)
Serum Ammonia (mM)
Total Bilirubin (mg/dL)
p=0.011
p=0.004

Supplemental Figure 8. Adult-to-fetal reprogramming occurs in a mouse model of non-alcoholic liver injury. (A) qRT-PCR analysis for *Esrp2* and YAP/TAZ target genes (*Areg* and *Birc5*) in livers of mice with intraperitoneal injection of carbon tetrachloride (CCl_4) or corn oil as vehicle (Veh) for 6 weeks. **(B)** IHC for ESRP2 in liver sections from Veh- or CCl_4-treated mice. Representative images of periportal (upper panels) and pericentral (lower panels) areas are shown. Arrows indicate ESRP2-negative hepatocytes and asterisk indicates hepatocytes expressing nuclear ESRP2. Scale bars=20μm. **(C)** Alternative splicing for HK mRNAs and EMT-related genes (*Arhgef10l*, *Slk* and *Kras*) in Veh- and CCl_4-injected mouse liver tissues. The lanes of *Nf2* were run on the same gel but were noncontiguous. Full unedited gel images are shown in Supplemental Material. **(D)** qRT-PCR analysis for *Igf2bp3* and *Sox9* in liver tissues of these mice. **(E)** IHC for CCND1 and SOX9 in liver sections of mice with or without chronic CCl_4 treatment. Enlarged images are shown. Scale bars=100μm. **(F)** qRT-PCR analysis for hepatic *Cps1* and *Glul* in these mice. **(G)** The serum levels of urea, ammonia and total bilirubin in mice treated with Veh or CCl_4. All results are graphed as dot plots (Veh: white circle, CCl_4: black square) with mean±s.e.m. (red bars) (n=4-7 individuals/group). All statistical analyses were performed by using two-tailed student *t* test between two groups.



Supplemental Figure 9. Adult-to-fetal reprogramming occurs in kidneys of mice with CCl_4–induced liver injury. (A) qRT-PCR analysis for *Esrp2* and YAP/TAZ target genes (*Areg*, *Ptgs2* and *Birc5*) in kidney tissues of mice injected with CCl_4 or corn oil as vehicle (Veh) for 6 weeks. **(B)** Alternative splicing for an EMT-related gene (*Kras*) in Veh- and CCl_4-treated mouse kidney tissues. All results are graphed as dot plots (Veh: white circle, CCl_4: black square) with mean±s.e.m. (red bars) (n=4-7 individuals/group). All statistical analyses were performed by using two-tailed student *t* test between two groups. Full unedited gel image is shown in Supplemental Material.



Supplemental Figure 10. Knocking down *Esrp2* promotes adult-to-fetal reprogramming in mouse hepatocytes. (A and B) qRT-PCR analysis **(A)** and immunoblot **(B)** for ESRP2 in AML12 cells transfected with mouse *Esrp2*-specific siRNA (*Esrp2* siR) or non-targeting control oligo (Nt) for 48 hours. Band intensity of ESRP2 blot normalized to GAPDH was calculated. **(C)** Alternative splicing of ESRP2 target mRNAs including *Csnk1d*, *Nf2*, *Slk*, *Flnb*, *Arhgef10l and Kras* between *Esrp2* siR- and Nt-treated AML12 cells. **(D)** qRT-PCR analysis for direct YAP/TAZ target genes (*Areg*, *Cyr61*, *Ctgf*, *Fgf1*, *Itgb2*) and immature/mesenchymal cell markers (*Ck7*, *Snail*, *Vim*, *EpCAM*) in AML12 cells transfected with *Esrp2* siR or Nt. The mean±s.e.m. results are displayed, and statistical analysis was performed by using two-tailed student *t* test between two groups (n=3 biological replicates/group, **p*<0.05, ***p*<0.005). Full unedited blots and gel images are shown in Supplemental Material.

Supplemental Table 1. Clinical information of patients with severe alcoholic hepatitis

ID	AH1	AH2	AH3	AH4	AH5	AH6
Age	32	34	49	48	41	61
Gender	M	F	F	M	M	M
Race	White	White	White	White	White	White
PT (9.4-11.6)	18.4	17	16.8	41	34.7	23.6
INR (0.9-1.1)	1.9	1.7	1.7	4.2	3.5	2.4
WBC (K/ cu mm) (4.5-11.0)	23.65	5.7	16.98	8.65	14.01	7.86
Hemoglobin (g/ dL) (M: 13.5-17.5; F: 12.0-15.5)	8	7.4	7.6	8.1	7.2	9.3
Platelets (K/ cu mm) (150-350)	147	119	27	66	57	45
AST (U/L) (10-40)	141	144	127	121	95	71
ALT (U/L) (7-56)	76	49	52	41	44	36
AP (U/L) (20-140)	188	87	175	110	129	98
Creatinine (mg/dL) (M: 0.6-1.2; F: 0.5-1.1)	2.1	1.5	1	2.7	0.9	1.5
Albumin (g/dL) (3.4-5.4)	3.2	3.7	2.9	4	2.9	3.4
Total Bilirubin (mg/dL) (0.1-1.2)	38.8	27.7	14.2	48	33.4	35.6
BMI (18.5-25)	36.9	26.7	26.2	30.1	22.9	26.3

PT, prothrombin time; INR, international normalized ratio; WBC, white blood cell; AST, aspartate transaminase; ALT, alanine transaminase; AP, alkaline phosphatase; BMI, body mass index.

Supplemental Table 2. Primer sequences used in the study

Target	Type	Primer sense	Sequence (5' → 3')
hESRP2	qRT-PCR	F	TGCCACAGAGGATGACTTTG
		R	ATTGACTGCTGGGCTCTTTG
hAREG	qRT-PCR	F	TGTCGCTCTTGATACTCGGC
		R	ATGGTTCACGCTTCCCAGAG
hCYR61	qRT-PCR	F	GGGCTGGAATGCAACTTC
		R	TACAGTTGGGCTGGAAACT
hCTGF	qRT-PCR	F	CTCCACCCGGGTTACCAATG
		R	CTTCCAGGTCAGCTTCGCAA
hPTGS2	qRT-PCR	F	ATATGTTCTCCTGCCTACTGGAA
		R	GCCCTTCACGTTATTGCAGATG
hIGF2BP3	qRT-PCR	F	GGGAGGTGCTGGATAGTTTAC
		R	CTAGCTTGGTCCTTACTGGAATAG
hCPS1	qRT-PCR	F	ATTCCTTGGTGTGGCTGAAC
		R	ATGGAAGAGAGGCTGGGATT
hGLUL	qRT-PCR	F	CTCGCGGCCTAGCTTTACCC
		R	CCACTCAGGCAACTCTTCCACA
hS9	qRT-PCR	F	GACTCCGGAACAAACGTGAGGT
		R	CTTCATCTTGCCCTCGTCCA
hCSNK1D_64	RT-PCR	F	ATGGAGAGAGAGCGGAAAGTGA
		R	GGTAACAGAGTAGATCAGCCATGC
hNF2_45	RT-PCR	F	CATGGAAAAGAGCAAGCATCTG
		R	AAAGAAGGCCACTCGGGACT
hSLK_93	RT-PCR	F	AGACTATCGAACGCCTGGAA
		R	AACTCTGCCTTCTGCTGCTG
hFLNB_72 39	RT-PCR	F	CGGGAAGGGTAAAGTGACCT
		R	CCGTTCATGTCACTCACTGG
mEsrp2	qRT-PCR	F	TATAAAGCCACAGGGGAGGA
		R	TCTTCCCGTGATAGGAAACG
mAreg	qRT-PCR	F	CCATCATCCTCGCAGCTATT
		R	CTTGTCGAAGCCTCCTTCTT

mCyr61	qRT-PCR	F	GGGCTGGAATGCAACTTC
		R	TACAGTTGGGCTGGAAACT
mCtgf	qRT-PCR	F	TCAAGCTGCCTGGGAAATG
		R	TCTGGGCCAAATGTGTCTTC
mFgf1	qRT-PCR	F	AGGAAACGTCCACAGTCAGG
		R	CTCCTACGCCCACTCTTCAG
mItgb2	qRT-PCR	F	ATGTGGGCCCACACTCACTGC
		R	TTAACAAAAGGCAGCACCGT
mCk7	qRT-PCR	F	TAGAGTCCAGCATCGCAGAG
		R	CACAGGTCCCATTCCGTC
mCol1α1	qRT-PCR	F	GAGCGGAGAGTACTGGATCG
		R	GCTTCTTTTCCTTGGGGTTC
mSnail	qRT-PCR	F	GGAAAGGCCTTCTCTGGC
		R	TTGGAGCGGTCAGCAAA
mVim	qRT-PCR	F	CGTCACCTTCGTGAATACCA
		R	TCCAGCAGCTTCCTGTAGGT
mEpCAM	qRT-PCR	F	GACCGGAAGTGGCAGAAGAG
		R	GGTCTTCATCTTCCCCAGGTTT
mBirc5	qRT-PCR	F	GAACCCGATGACAACCCGAT
		R	TGGTCTCCTTTGCAATTTTGTTCT
mPtgs2	qRT-PCR	F	TACCCGGACTGGATTCTATG
		R	AGTGGGTCAGGATGTAGTG
mIgf2bp3	qRT-PCR	F	GGGAGGTGCTGGATAGTTTAC
		R	CTAGCTTGGTCCTTACTGGAATAG
mSox9	qRT-PCR	F	CGGCTCCAGCAAGAACAAG
		R	GCGCCCACAGGATGAAG
mCcnd1	qRT-PCR	F	TAGGCCCTCAGCCTCACTC
		R	CCACCCCTGGGATAAAGCAC
mCps1	qRT-PCR	F	CACCAATTTCCAGGTGACCA
		R	TACTGCTTTAGGCGGCCTTT
mGlul	qRT-PCR	F	ACTGCGCTGCAAGACCCGTA
		R	GTTGGTCTCTGAAGGTTTCCGG
	qRT-PCR	F	CGTCAGCCGATTTGCTATCT

mTnfα		R	CGGACTCCGCAAAGTCTAAG
mIl1β	qRT-PCR	F	CCACCTCAATGGACAGAATATCA
		R	CCCAAGGCCACAGGTATTT
mCd68	qRT-PCR	F	TGCGGCTCCCTGTGTGT
		R	TCTTCCTCTGTTCCTTGGGCTAT
mLy6G	qRT-PCR	F	TGCGTTGCTCTGGAGATAGA
		R	CAGAGTAGTGGGGCAGATGG
mPparγ	qRT-PCR	F	AACTGCAGGGTGAAACTCTGGGAGA
		R	GGATTCAGCAACCATTGG GTCAGCT
mS9	qRT-PCR	F	GGGCCTGAAGATTGAGGATT
		R	CGGGCATGGTGAATAGATTT
mCsnk1d_63	RT-PCR	F	GGAACGAGAACGGAAAGTGA
		R	GGGGGCGTGTCACTAGTAAAG
mNf2_45	RT-PCR	F	ACACAGCGAGAGCTCAGACA
		R	ACAAGCCAGCCCTCTACTGA
mSlk_93	RT-PCR	F	AAGGAGCTGTCCAAGTTCCA
		R	TATTGGCCAACTCTGCCTTC
mFlnb_72 39	RT-PCR	F	GGCGAGGAGGTGGGCTTTGTAG
		R	CCTGACGGCAAATGGAATCACCAA
mArhgef10l_15	RT-PCR	F	CTGCTCAATGACATGCTGGT
		R	TCCTGACCCACCTCTACCAC
mKras_124	RT-PCR	F	GATGTGCCTATGGTCCTGGT
		R	TCTTCTTCCCATCTTTGCTCA

Supplemental Methods

Immunohistochemistry

Liver specimens were fixed in 10% neutral buffered formalin, embedded in paraffin using standard methods and cut into 5 μm sections. For immunohistochemistry, liver sections were deparaffinized, hydrated and incubated in 3% hydrogen peroxide to block endogenous peroxidase. Antigen retrieval was performed by heating in 10 mM sodium citrate buffer (pH 6.0) for 10 min using a microwave. Specimens were blocked in Dako Protein Block solution (Agilent, Santa Clara, CA) for 30 min at room temperature followed by incubation with primary antibody at 4 °C overnight. Other sections were also incubated at 4 °C overnight in non-immune sera. Rabbit anti-C/EBPα (Santa Cruz Sc-61, Dallas, TX), rabbit anti-YAP (Cell Signaling 14074S, Danvers, MA), rabbit anti-TAZ (Abcam ab110239, Cambridge, UK), rabbit anti-ESRP2 (Abcam ab113486), rabbit anti-IGF2BP3 (Millipore 03-198, Burlington, MA), goat anti-TNFRSF12A (Biolegend 314002, San Diego, CA), rabbit anti-SOX9 (Millipore AB5535), rabbit anti-SMAD3 (Abcam ab28379), rabbit anti-CDH1 (BD Biosciences 610181, San Jose, CA) and rabbit anti-Cyclin D1 (Abcam ab134175) were used as primary antibodies and diluted in Antibody Diluent (Sigma-Aldrich, St. Louis, MO). Polymer-horseradish peroxidase (HRP) anti-rabbit (Dako) and polymer-HRP anti-goat (ThermoFisher Scientific, Waltham, MA) were used as secondary antibodies and 3,3′-diaminobenzidine (DAB) as brown color was used to visualize the protein. Sections were counterstained with hematoxylin. To quantify CCND1 and SOX9(+) cells, more than 10 randomly chosen 100× or 400× fields/section were evaluated by counting the total number of stained cells/field for each mouse.

Liver histology

Paraffin-embedded liver sections were stained with hematoxylin and eosin (H&E) for pathological evaluation, and optimal cutting temperature (OCT)-embedded liver sections were stained with Oil Red O for visualization of lipid droplets.

Immunoblot analysis

Total protein was extracted from whole liver tissues or cell lysates using RIPA buffer (Sigma-Aldrich) with protease inhibitor cocktail (Sigma-Aldrich). Protein concentration was measured by Pierce BCA Protein Assay Kit (ThermoFisher Scientific) and an equal amount of total protein lysates were separated by SDS-PAGE on 4-20% Mini-PROTEAN® TGX™ Precast Protein Gels (BIO-RAD, Hercules, CA, USA) and then transferred onto a polyvinylidene difluoride membrane (Invitrogen). Primary antibodies used in this study were as follows: rabbit anti-ESRP2 (Abcam ab113486), rabbit anti-IGF2BP3 (Millipore 03-198) and mouse anti-GAPDH (Santa Cruz sc-365062). HRP-conjugated Amersham ECL anti-rabbit or anti-mouse IgG (GE Healthcare, Little Chalfont, UK) were used as secondary antibodies. Protein bands were developed using an ECL Western Blotting Detection Reagent (ThermoFisher Scientific) and detected using the ChemiDoc XRS+ (BIO-RAD). Densitometric analyses for protein quantification were conducted using CS Analyzer software (Version 3.00.1011, ATTO & Rise Corporation). Band density of each target protein was normalized to the density of its respective loading control.

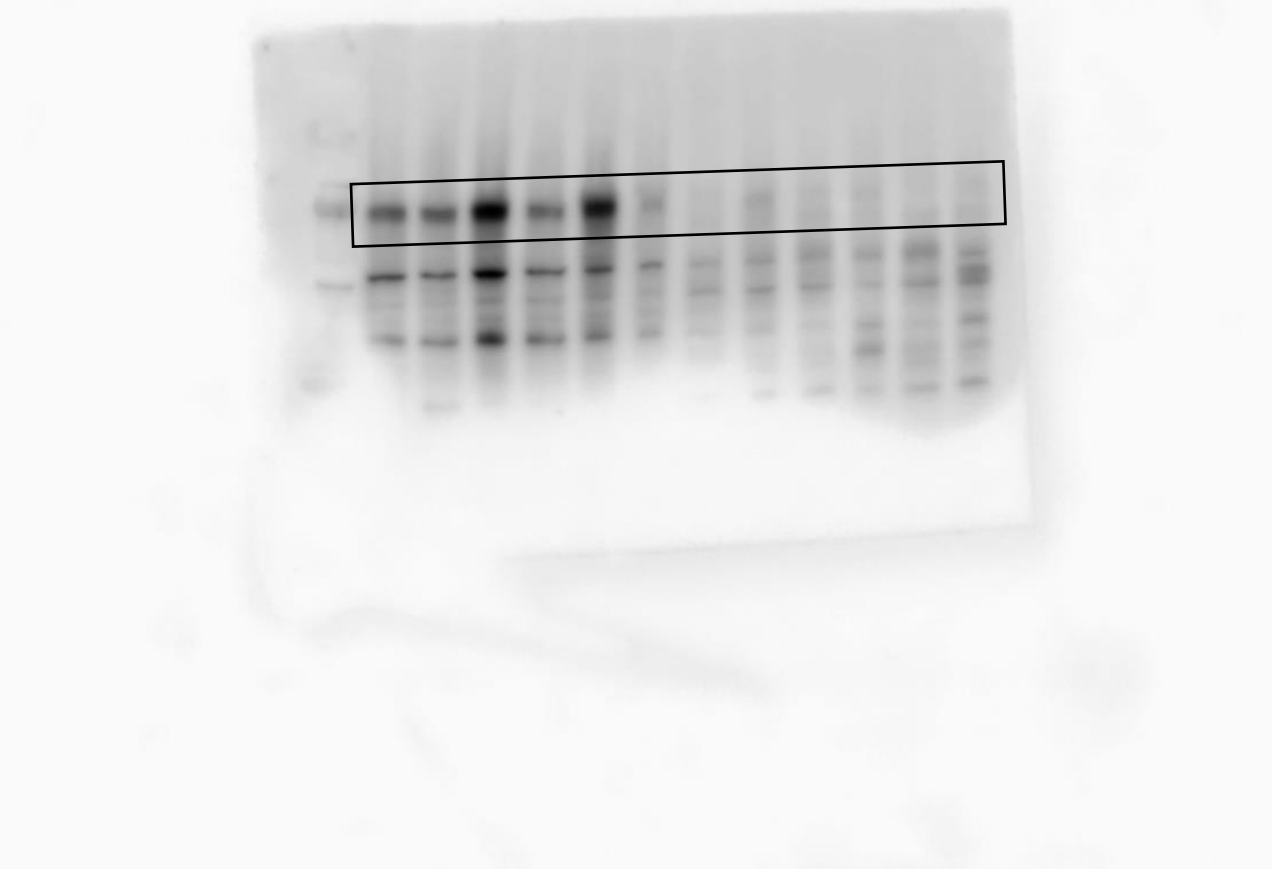
Measurements of glutamine, urea, ammonia, alanine transaminase (ALT), bilirubin, albumin (ALB) and triglyceride (TG)

The concentrations of glutamine, urea and ammonia were measured in conditioned media of primary hepatocytes at 2 days after media were changed, and in mouse serum using the Glutamine/Glutamate-Glo Assay kit (Promega, Madison, WI), Urea Colorimetric Assay Kit (BioVision, Inc., Milpitas, CA) and Ammonia Colorimetric Assay Kit (BioVision, Inc.), respectively, following manufacturer's instructions. Also, serum levels of albumin (ALB), bilirubin and alanine transaminase (ALT) and hepatic triglyceride (TG) contents of mouse models were quantified using the Mouse Albumin ELISA Kit (Abcam), Bilirubin Colorimetric Assay Kit (BioVision, Inc.), ALT (SOFT) Kinetic Kit (Biotron Diagnostic Inc., Hemet, CA) and

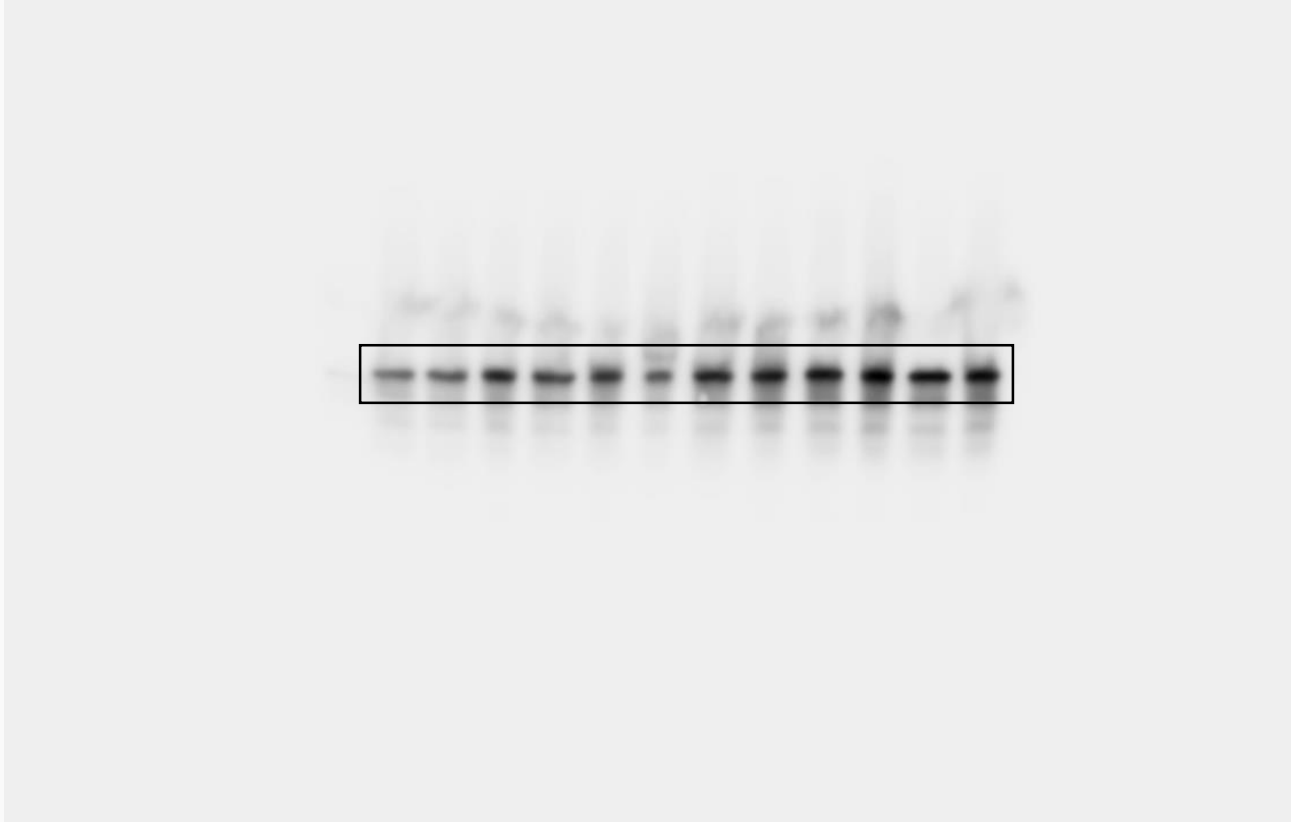
Triglyceride Colorimetric Assay Kit (Cayman Chemical, Ann Arbor, MI), respectively, per manufacturers' specifications. Luminescence was read on a FLUOstar OPTIMA (BMG LABTECH, Ortenberg, Germany), and absorbance was measured on an Infinite M200 PRO (TECAN, Männedorf, Switzerland).

Full unedited blot for Figure 2

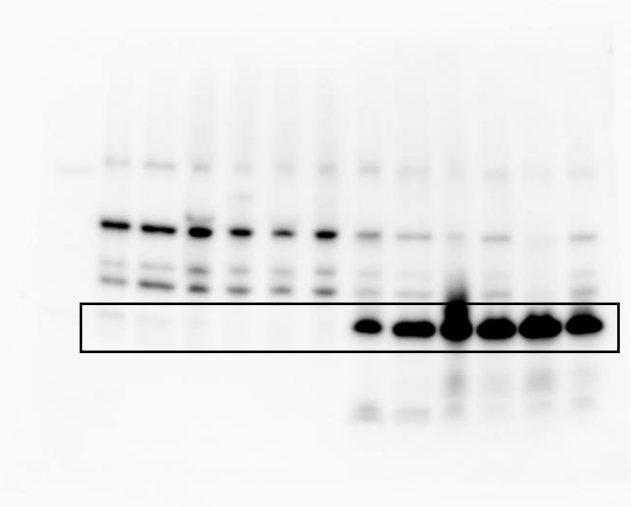
Immunoblot for ESRP2 (Fig 2C)
using rabbit anti-ESRP2 (Abcam ab113486)



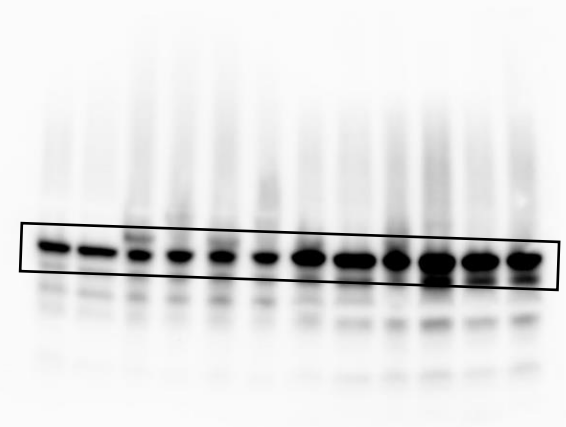
Immunoblot for GAPDH (Fig 2C)
using mouse anti-GAPDH (Santa Cruz sc-365062)



Immunoblot for IGF2BP3 (Fig 2G)
using rabbit anti-IGF2BP3 (Millipore 03-198)

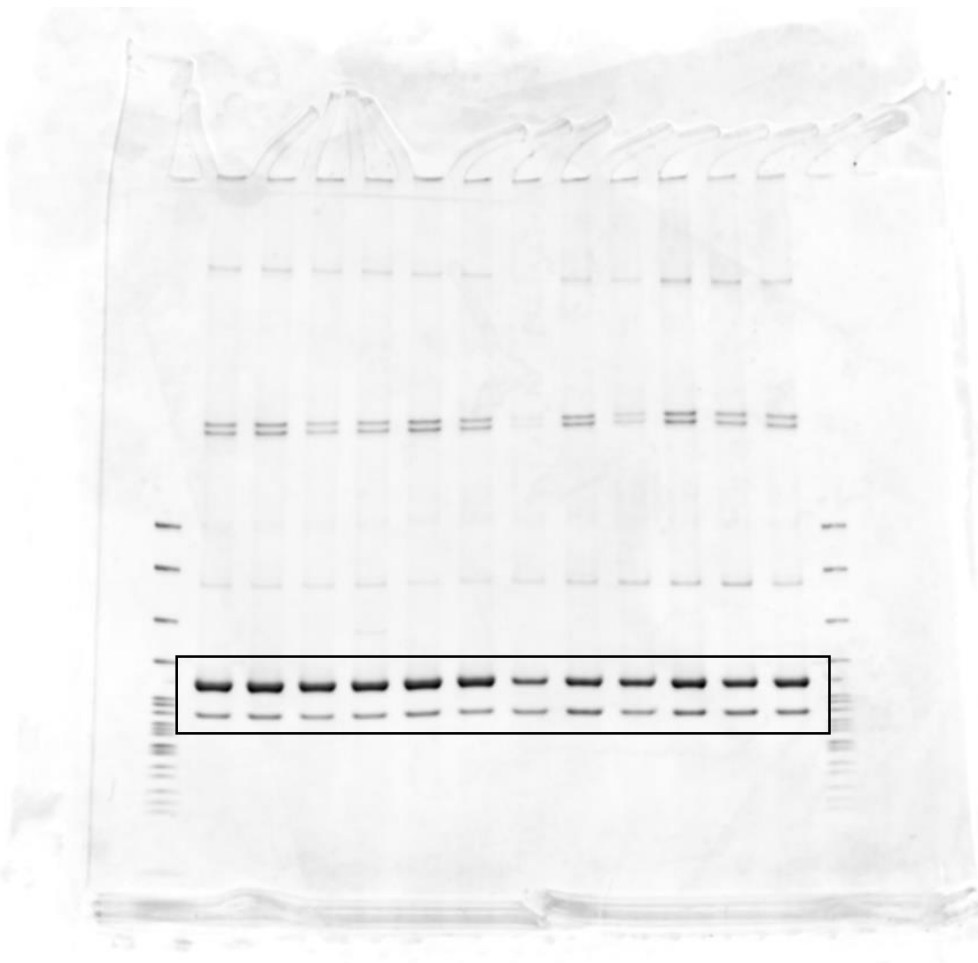


Immunoblot for GAPDH (Fig 2G)
using mouse anti-GAPDH (Santa Cruz sc-365062)

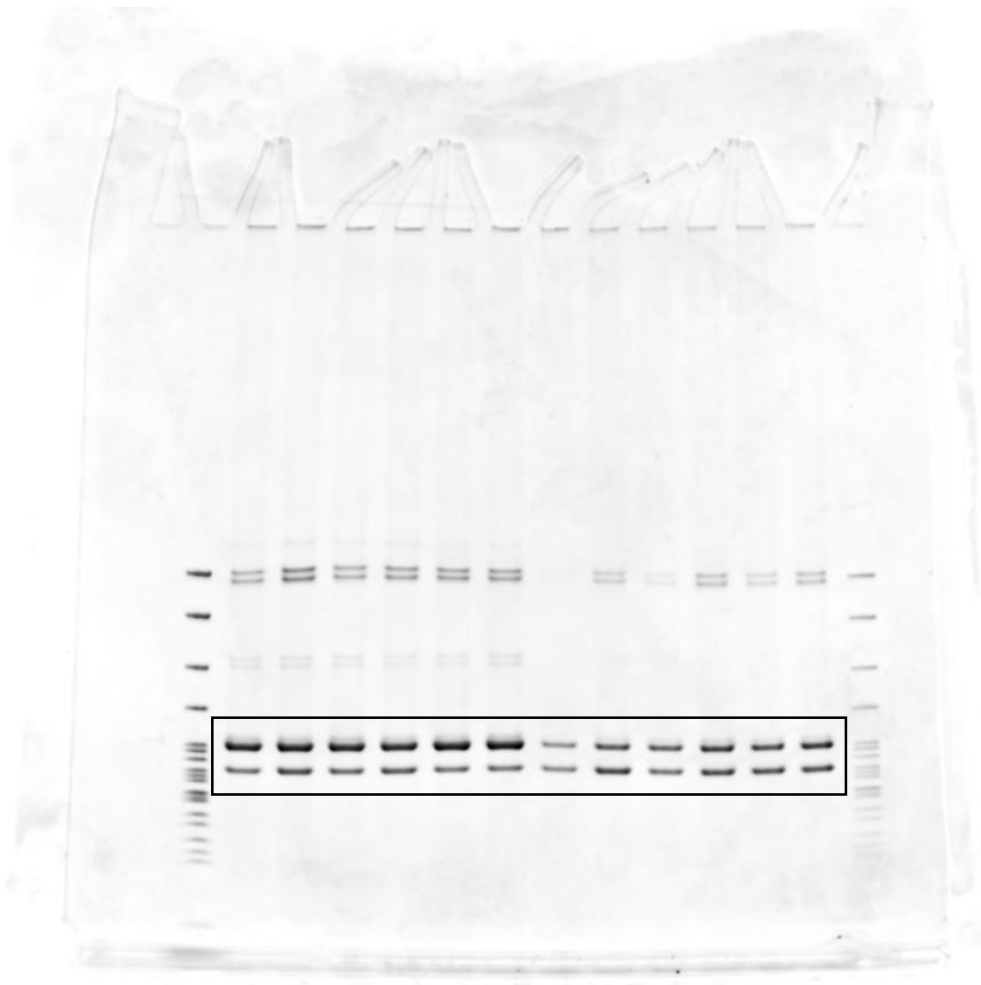


Full unedited gel for Figure 2

Splicing for *CSNK1D* (Fig 2E)

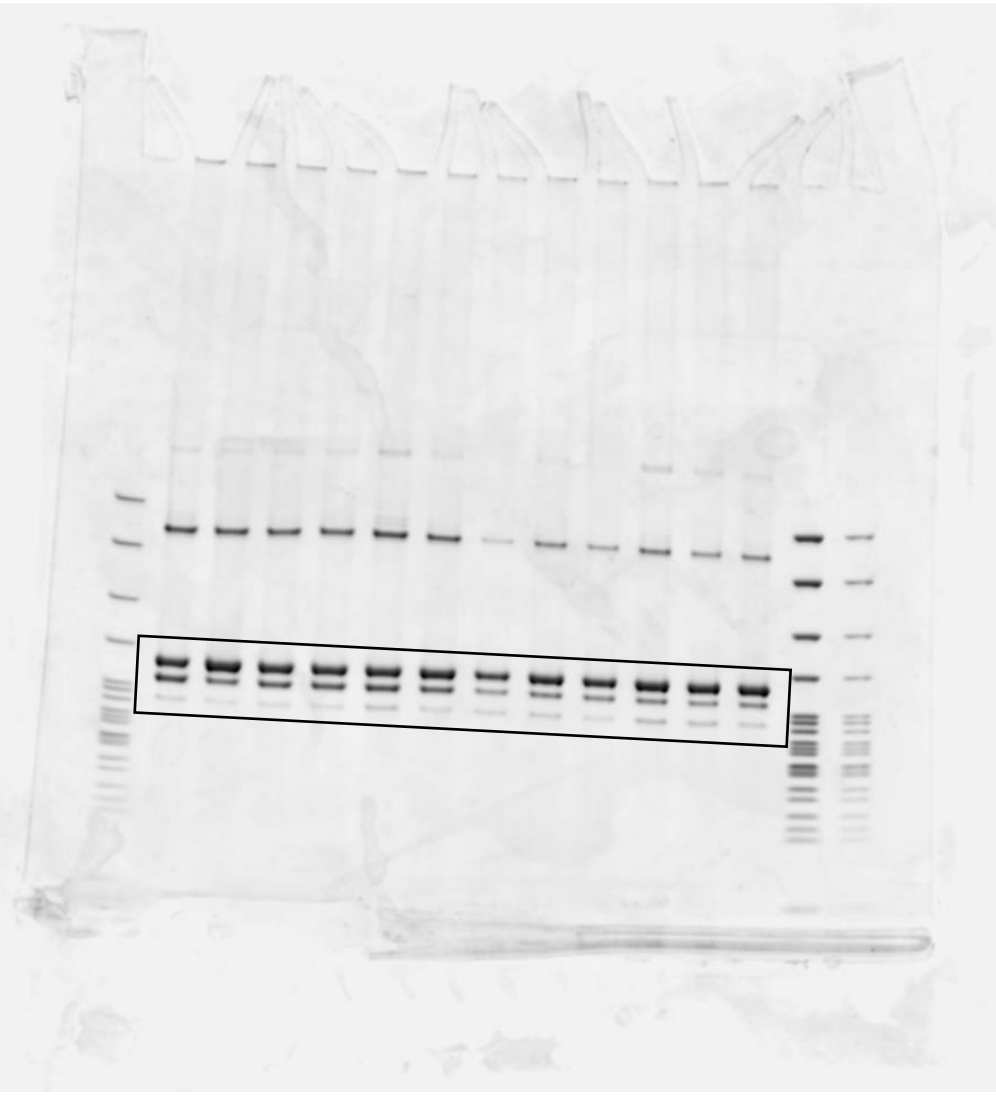


Splicing for *NF2* (Fig 2E)



Splicing for *SLK* (Fig 2E)

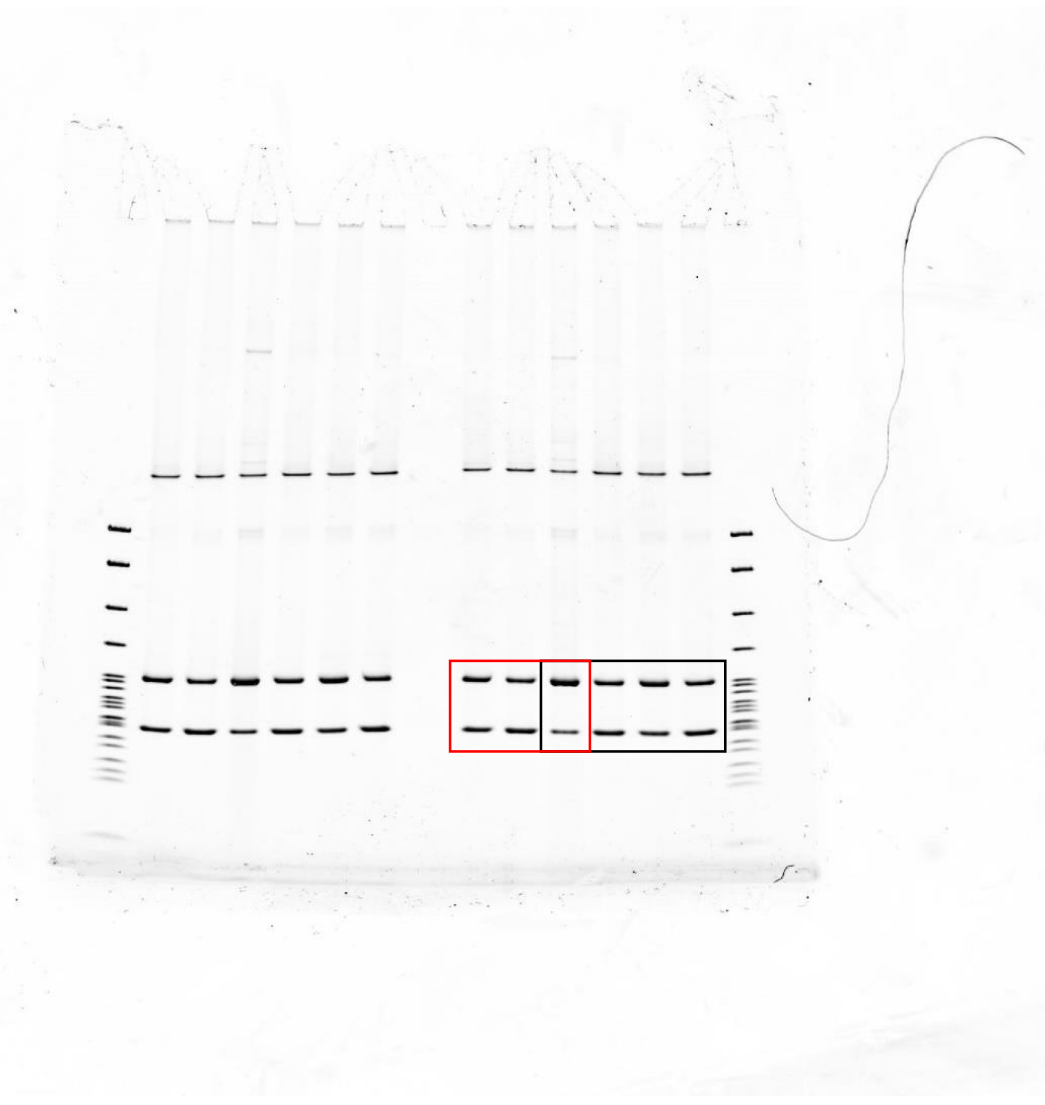
Splicing for *FLNB* (Fig 2E)



Full unedited gel for Figure 4 and 5

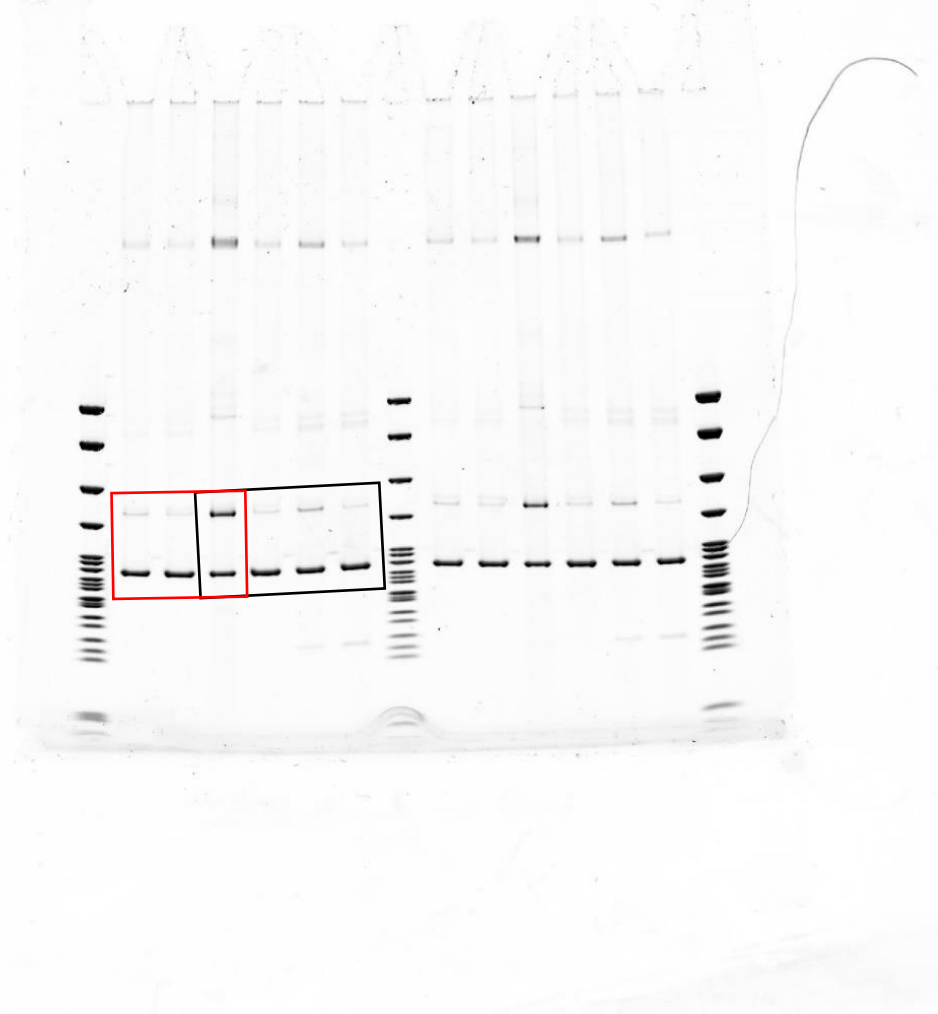
Splicing for *Nf2* (Fig 4B, Fig 5B left-right inverted)

Splicing for *Slk* (Fig 4B, Fig 5B left-right inverted)



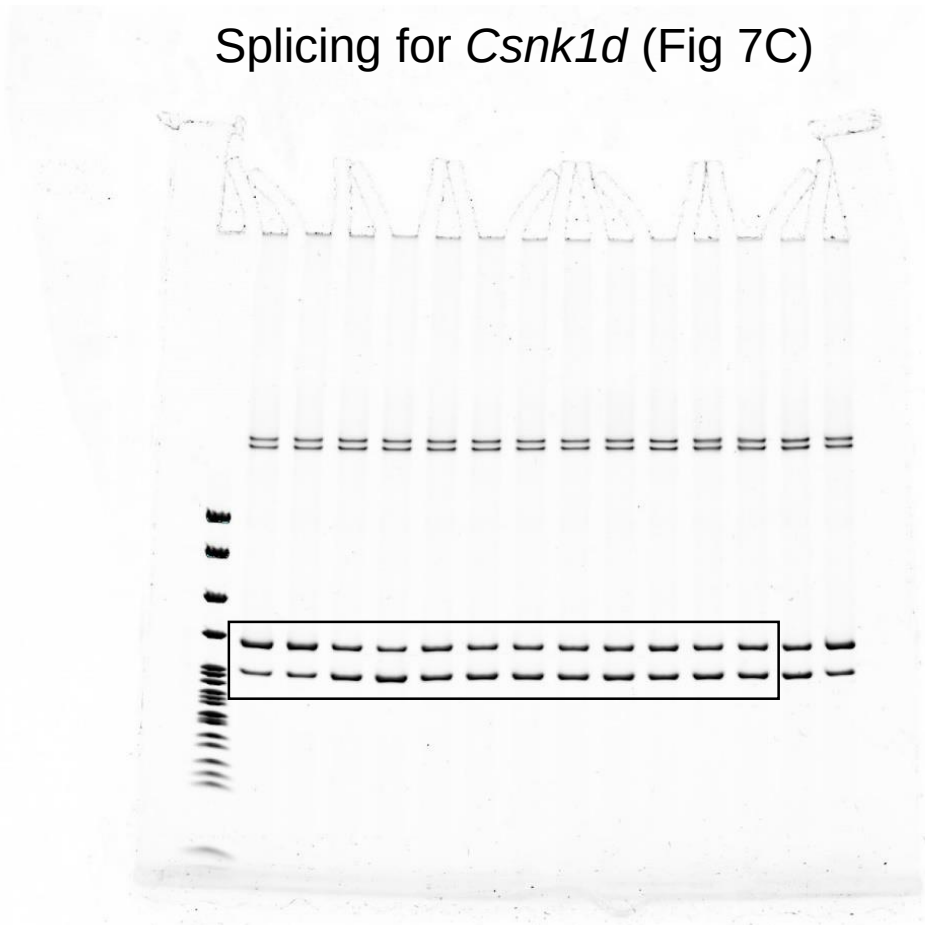
Splicing for *Flnb* (Fig 4B, Fig 5B left-right inverted)

Splicing for *Kras* (Fig 4B, Fig 5B left-right inverted)

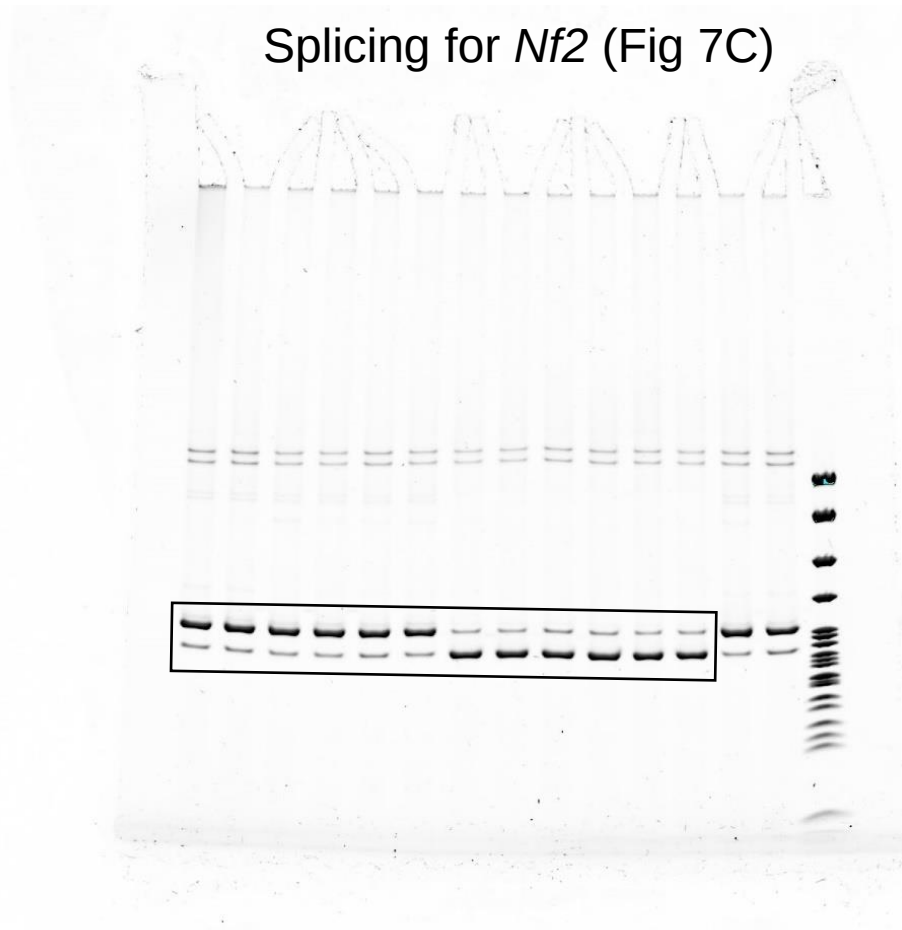


Full unedited gel for Figure 7

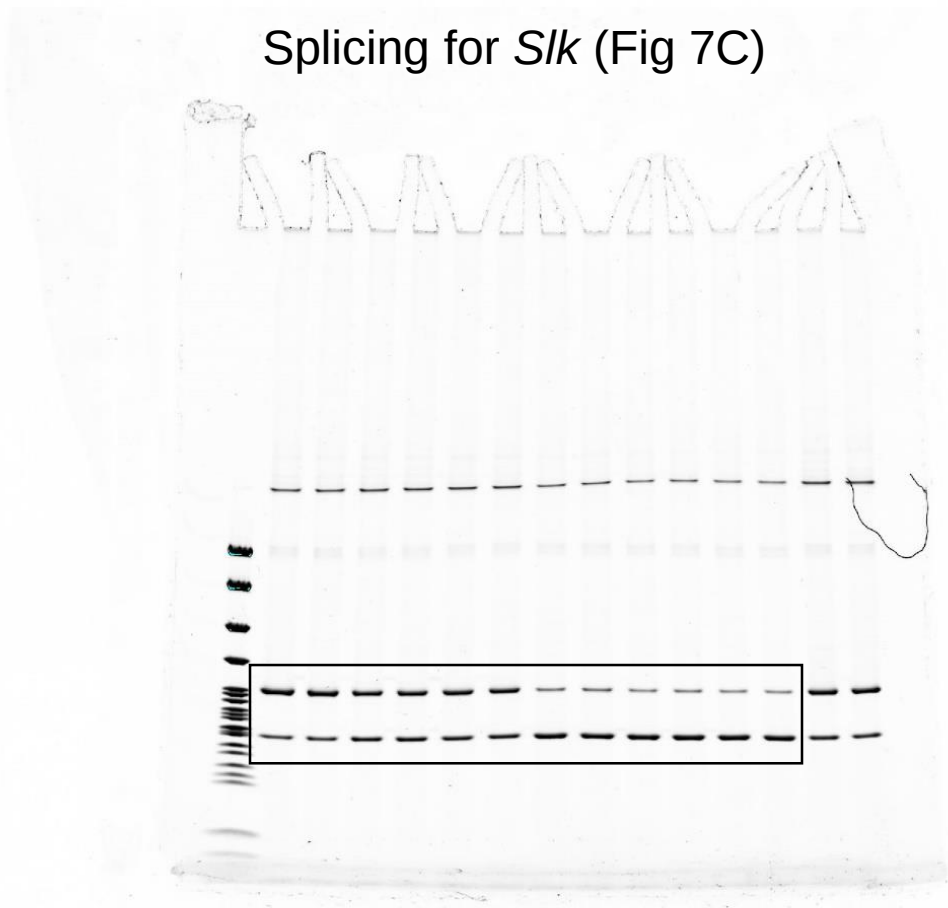
Splicing for *Csnk1d* (Fig 7C)



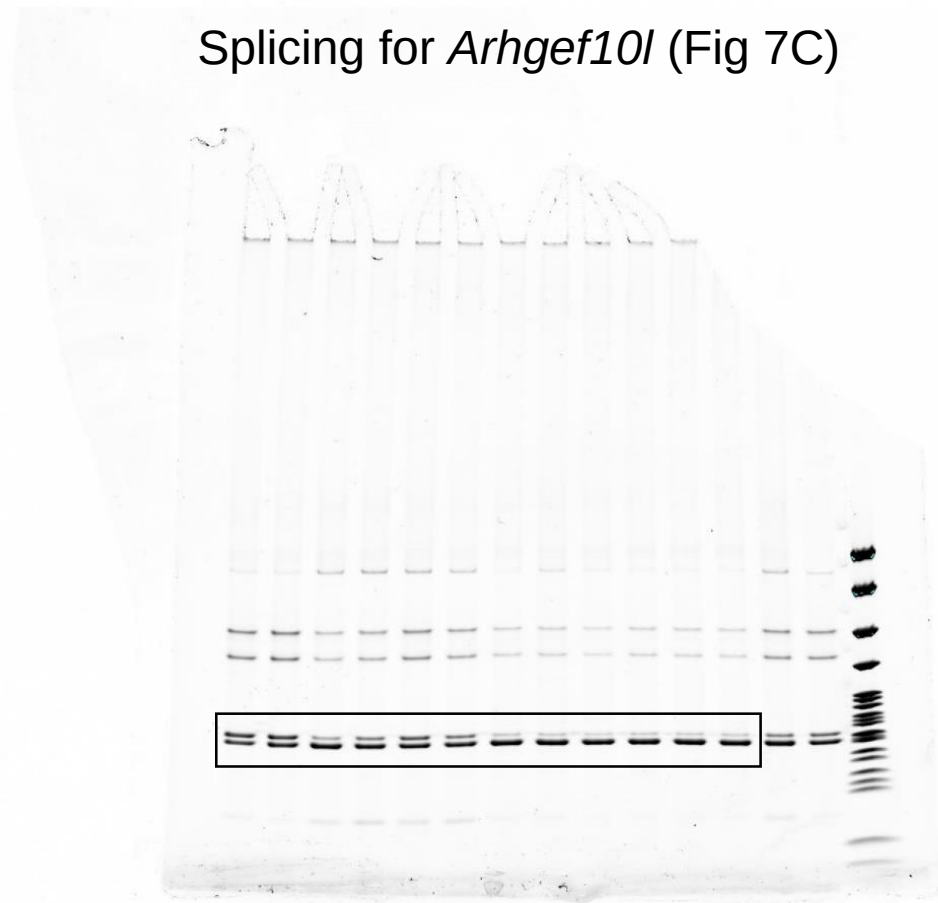
Splicing for *Nf2* (Fig 7C)



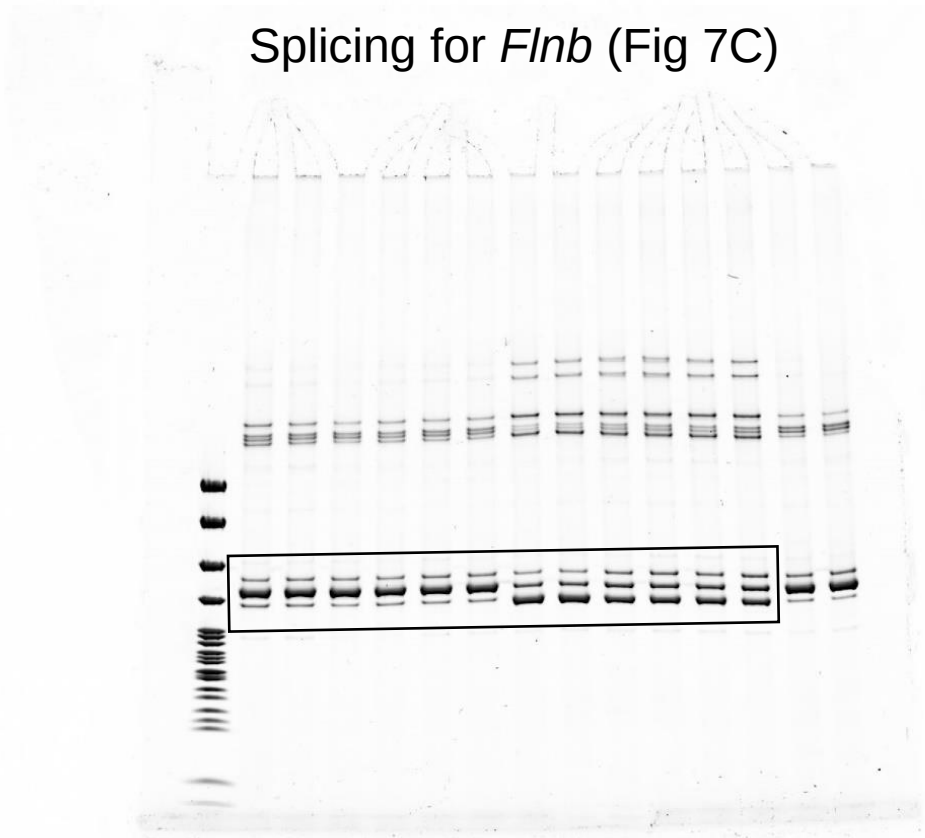
Splicing for *Slk* (Fig 7C)



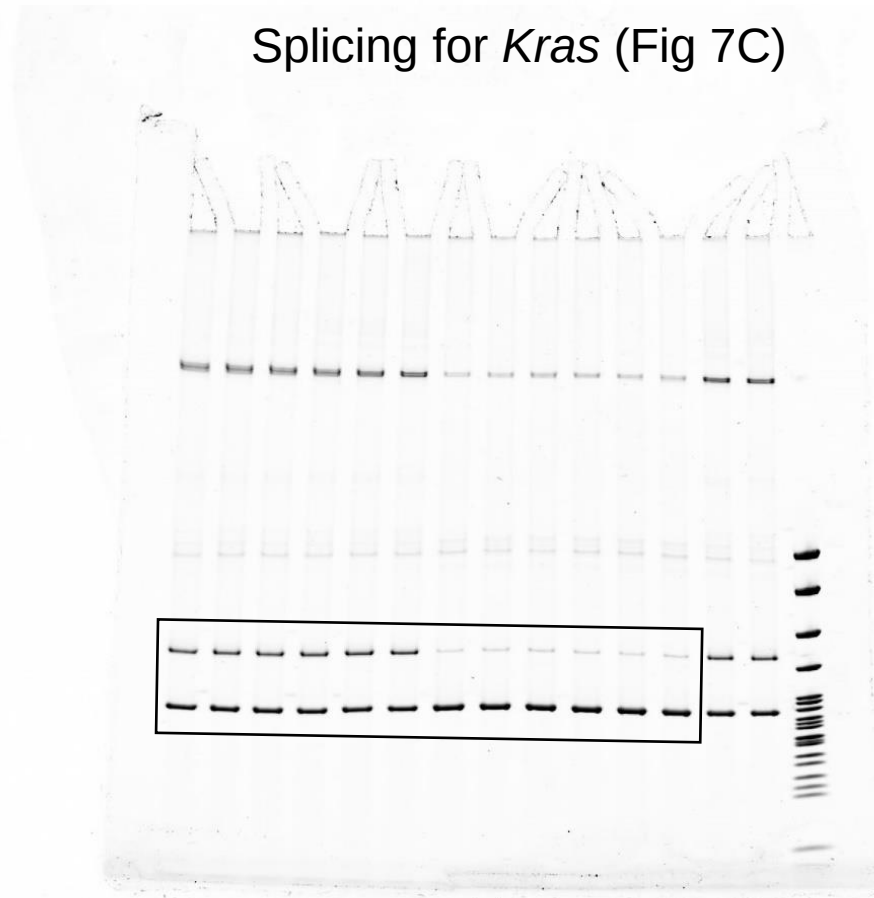
Splicing for *Arhgef10l* (Fig 7C)



Splicing for *Flnb* (Fig 7C)

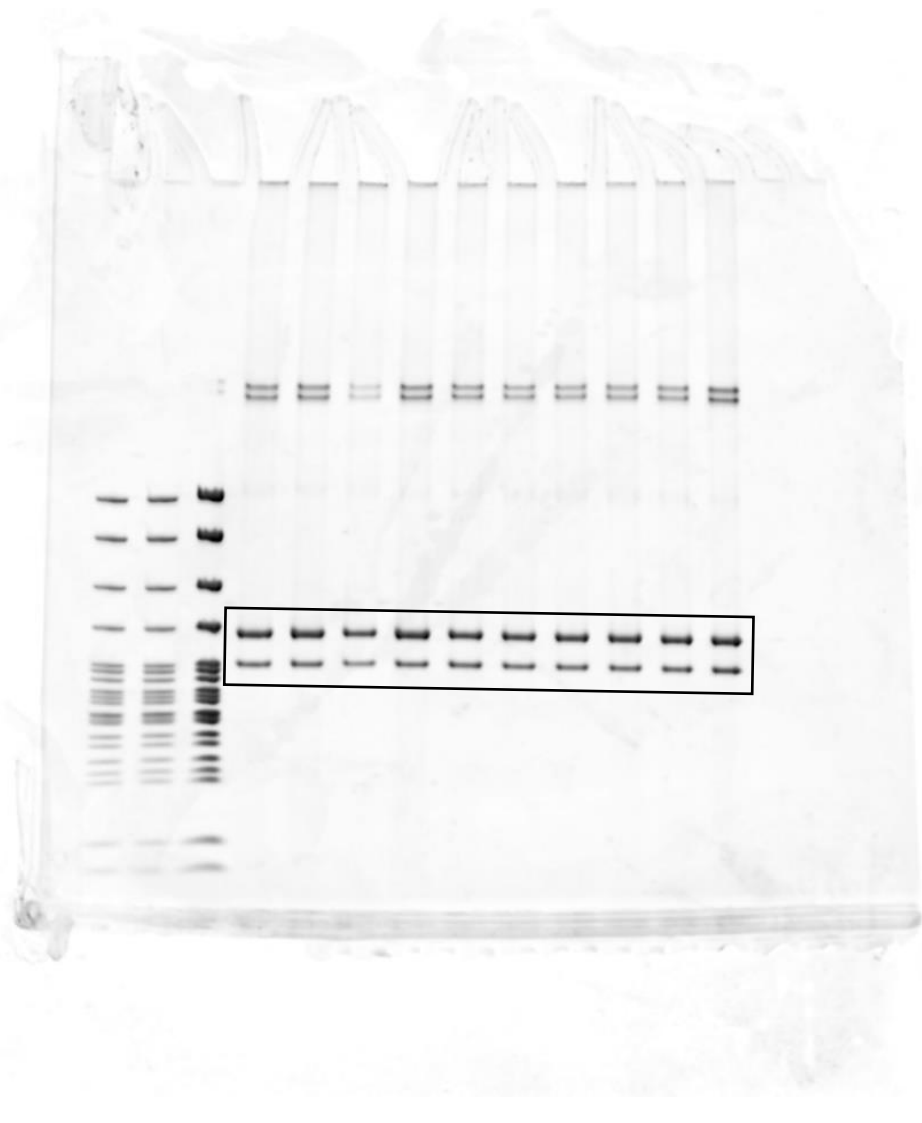


Splicing for *Kras* (Fig 7C)



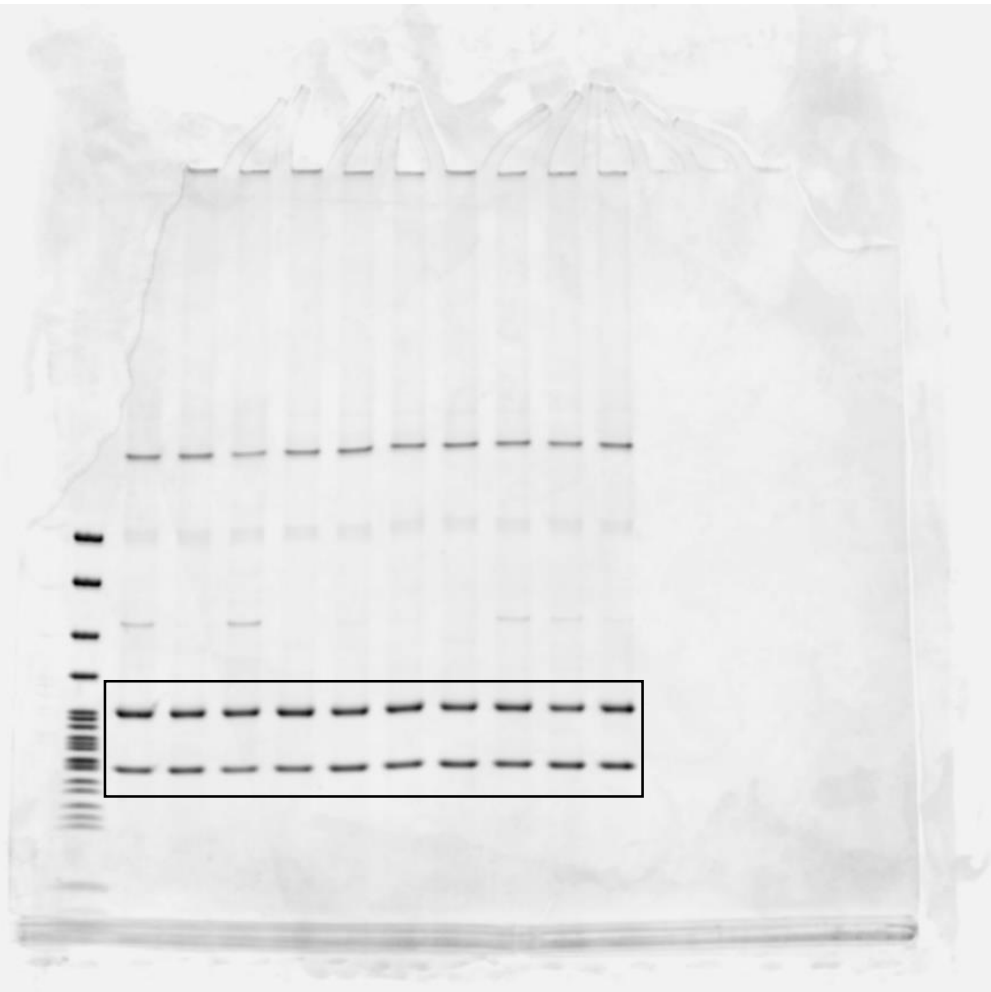
Full unedited gel for Figure S3

Splicing for *Csnk1d* (Fig 3B)

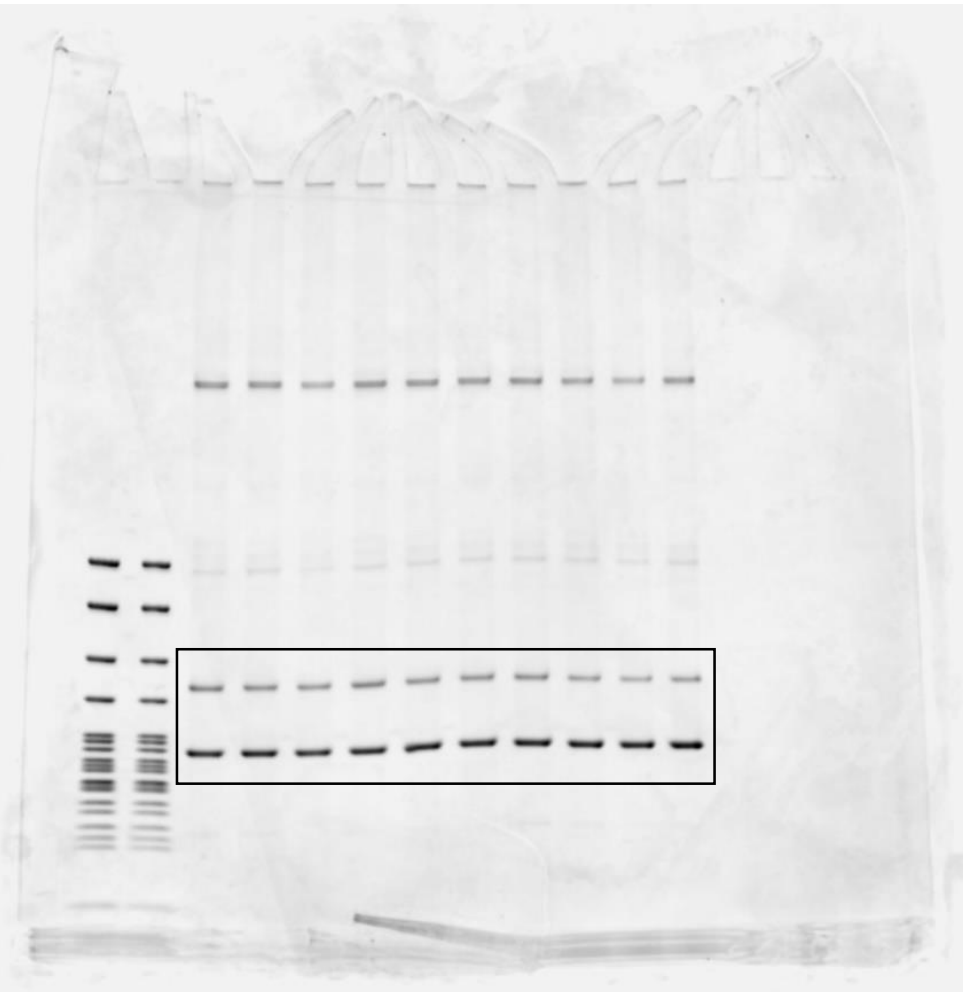


Splicing for *Nf2* (Fig 3B)

Splicing for *Slk* (Fig 3B)

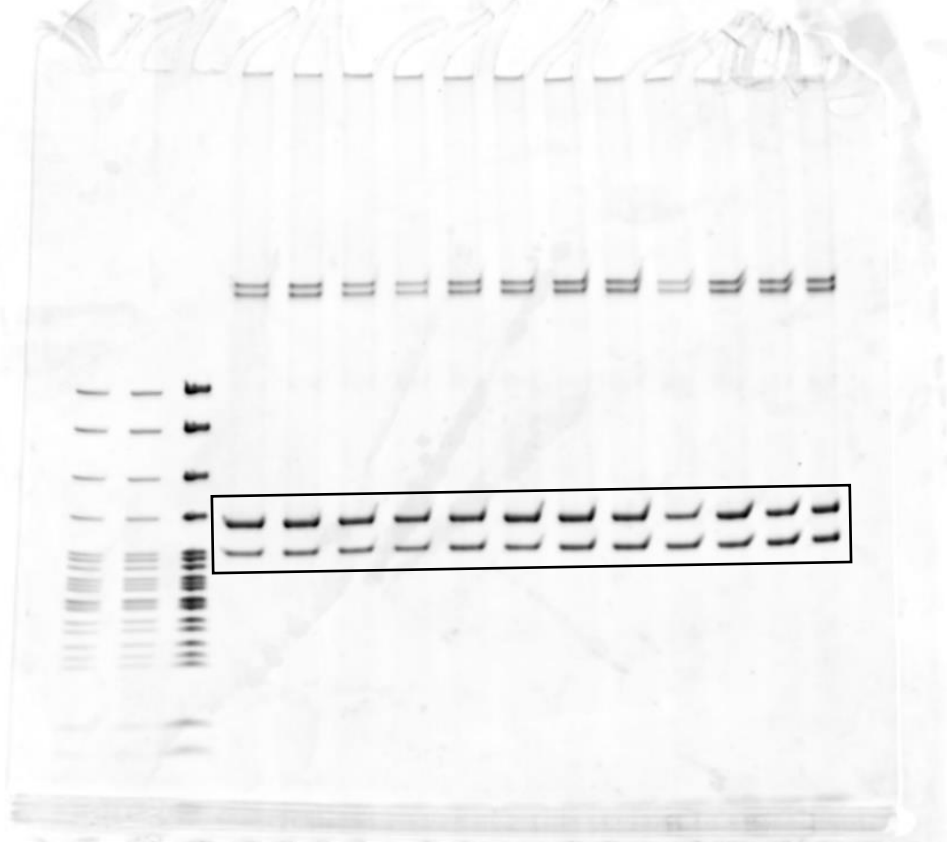


Splicing for *Kras* (Fig 3B)

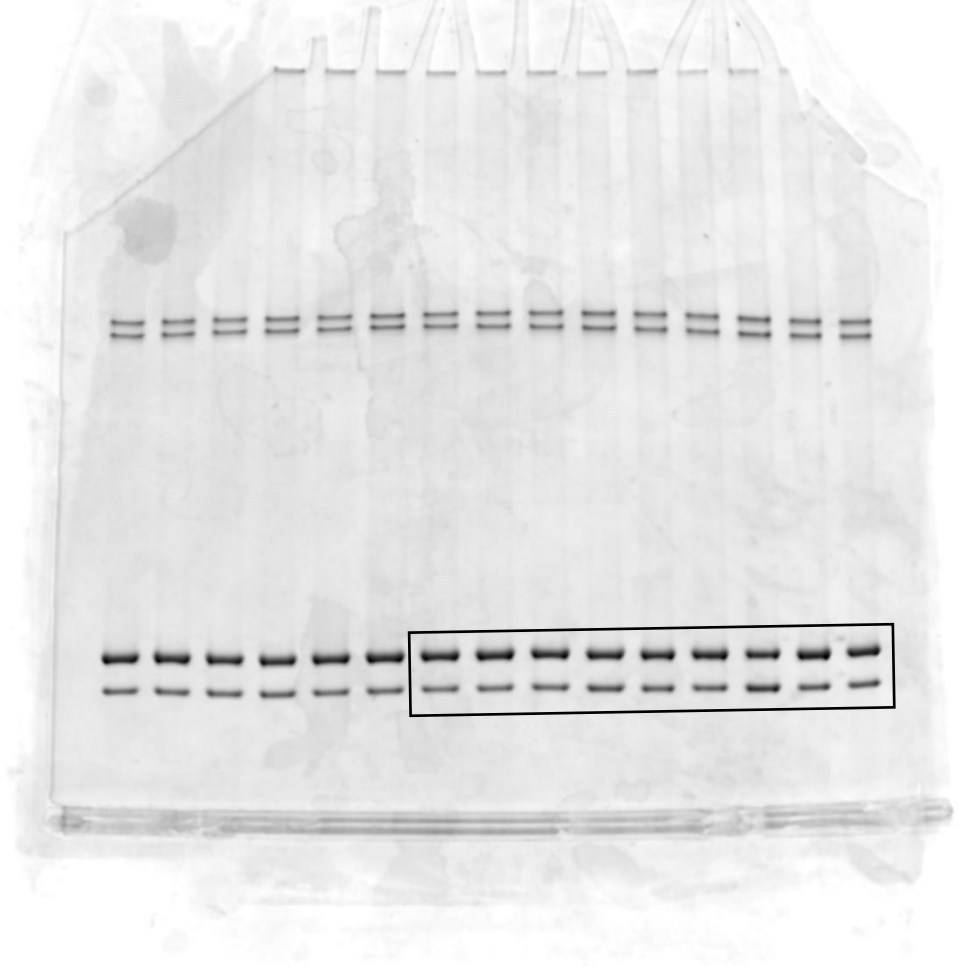


Full unedited gel for Figure S4

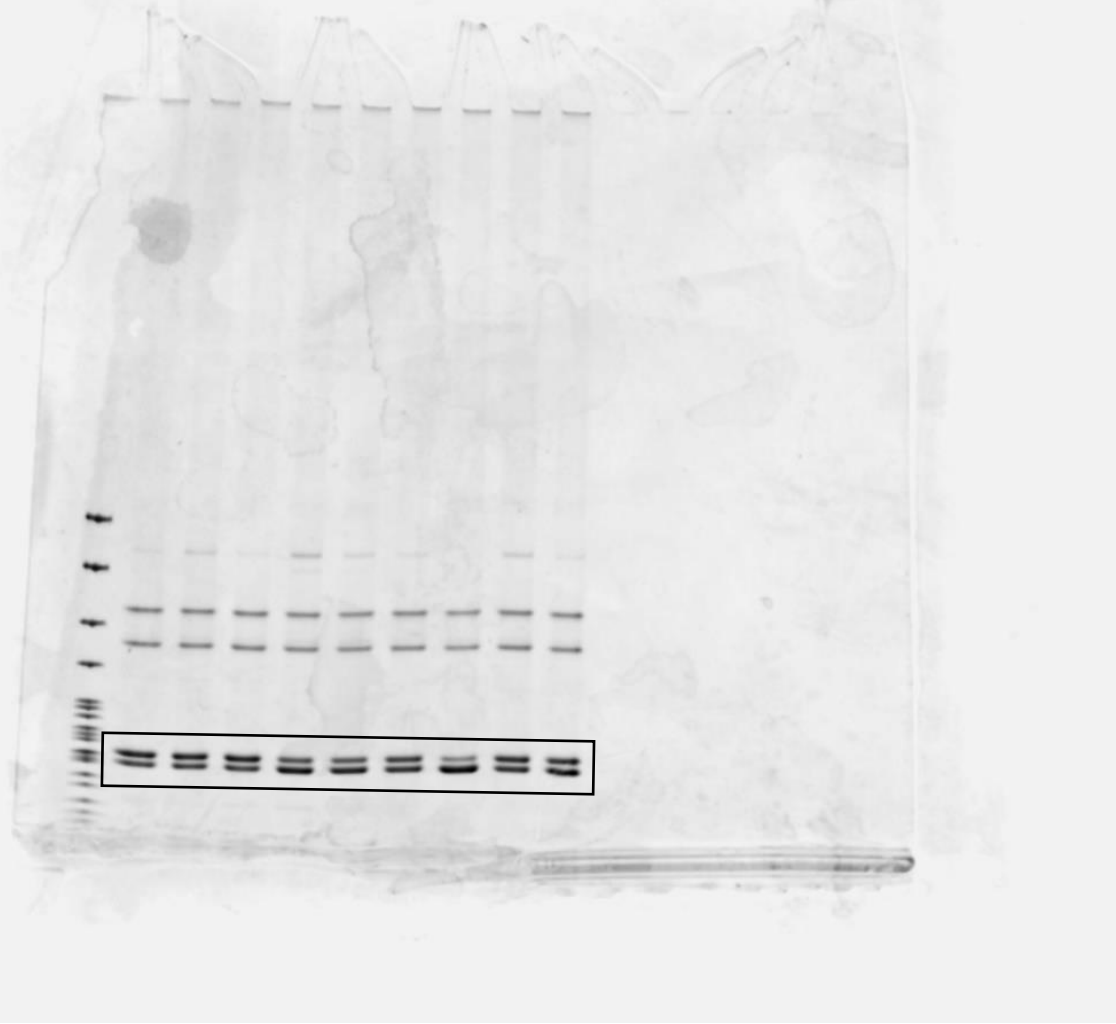
Splicing for *Csnk1d* (Fig S4B)



Splicing for *Csnk1d* (Fig S4D)

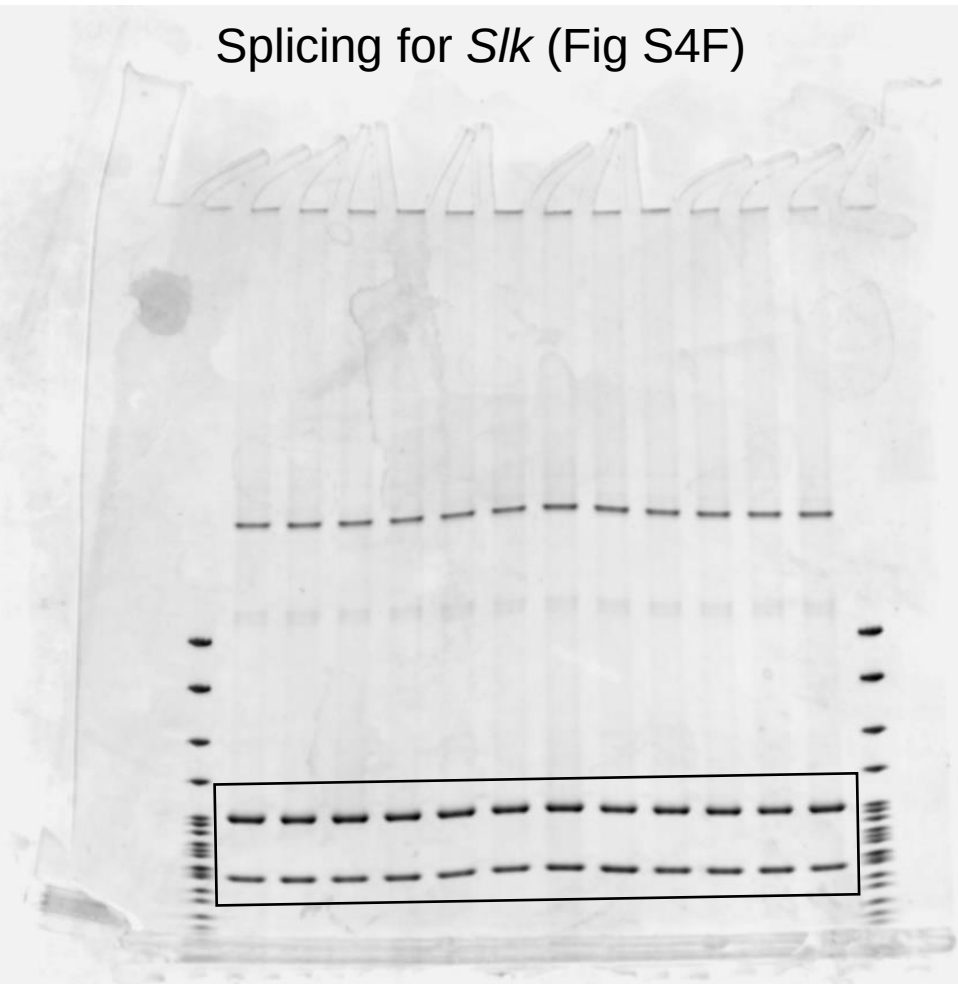


Splicing for *Arhge10l* (Fig S4D)



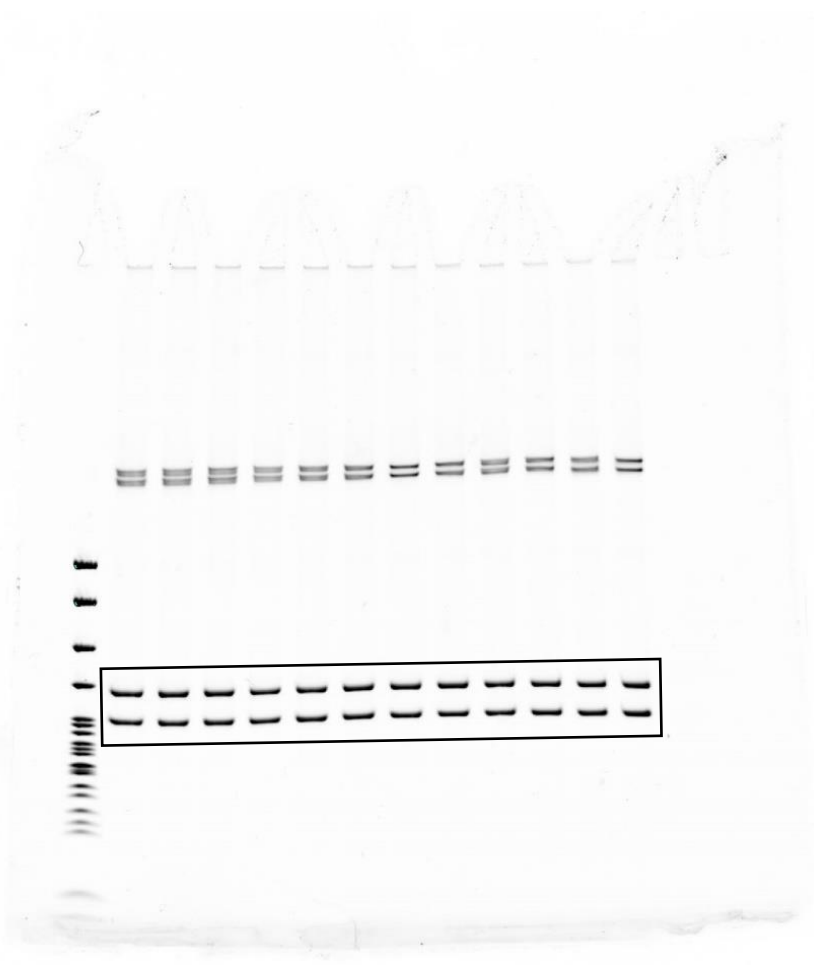
Splicing for *Nf2* (Fig S4F)

Splicing for *Slk* (Fig S4F)

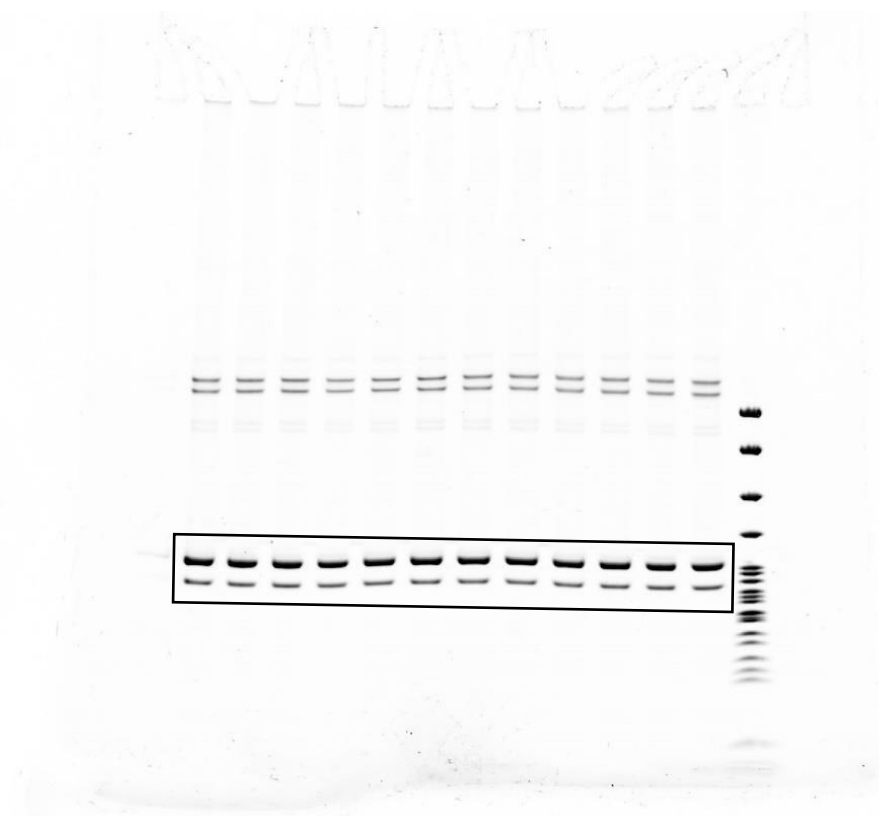


Full unedited gel for Figure S5

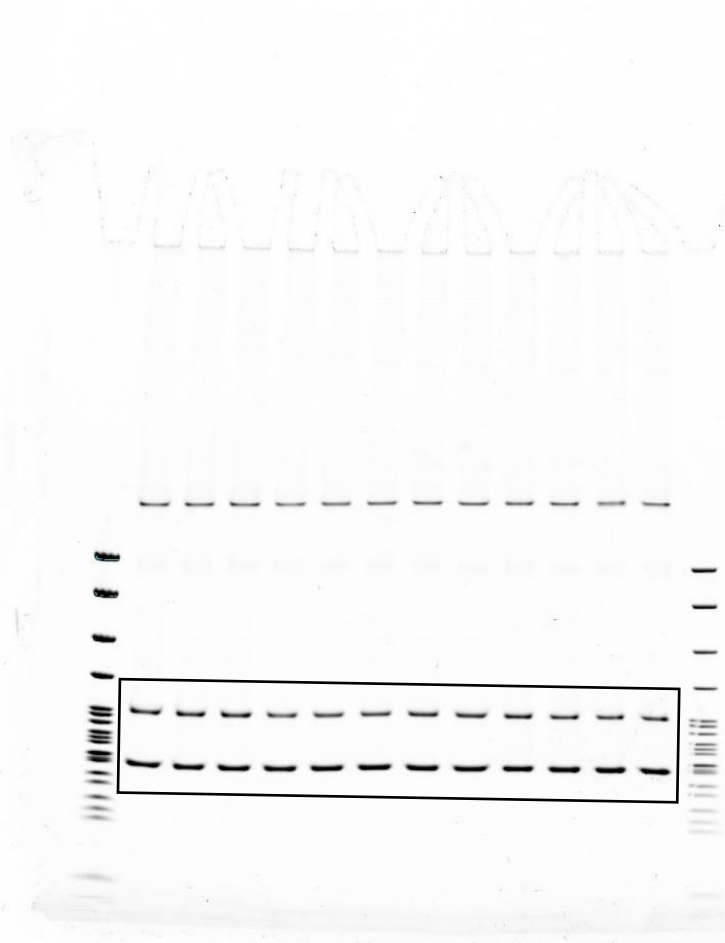
Splicing for *Csnk1d* (Fig 5D)



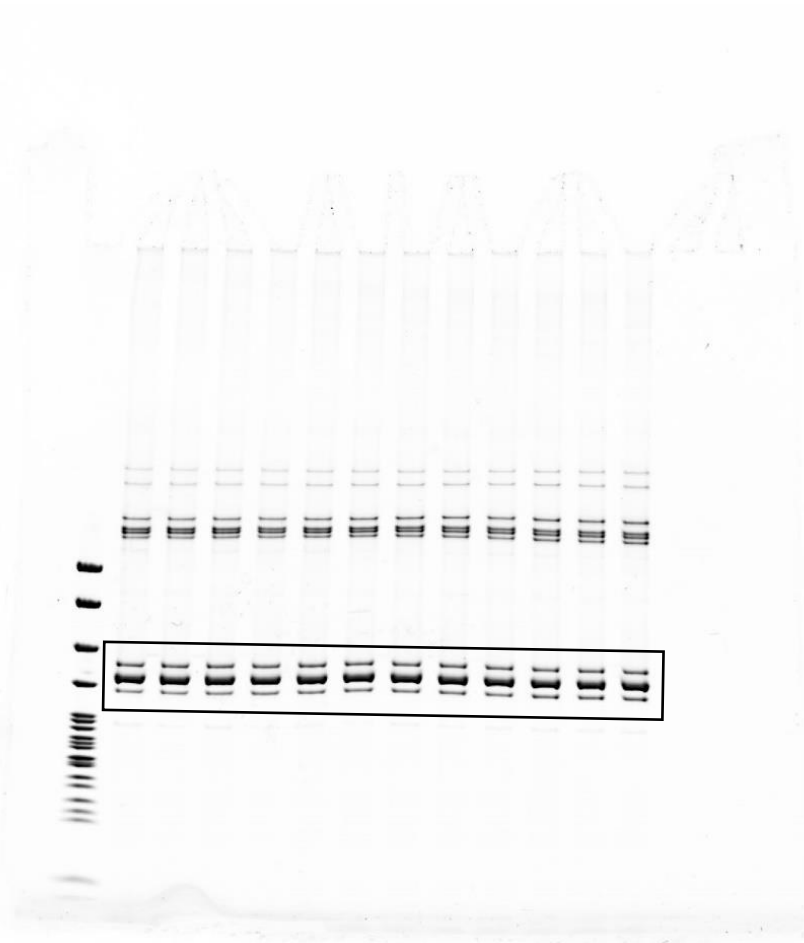
Splicing for *Nf2* (Fig 5D)



Splicing for *Slk* (Fig 5D)

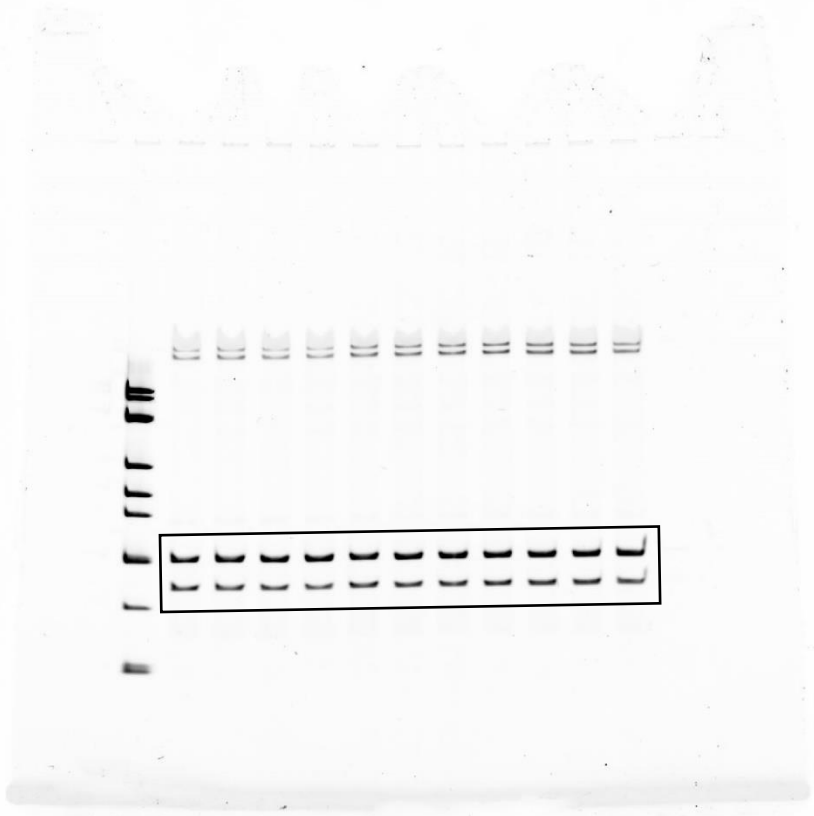


Splicing for *Flnb* (Fig 5D)

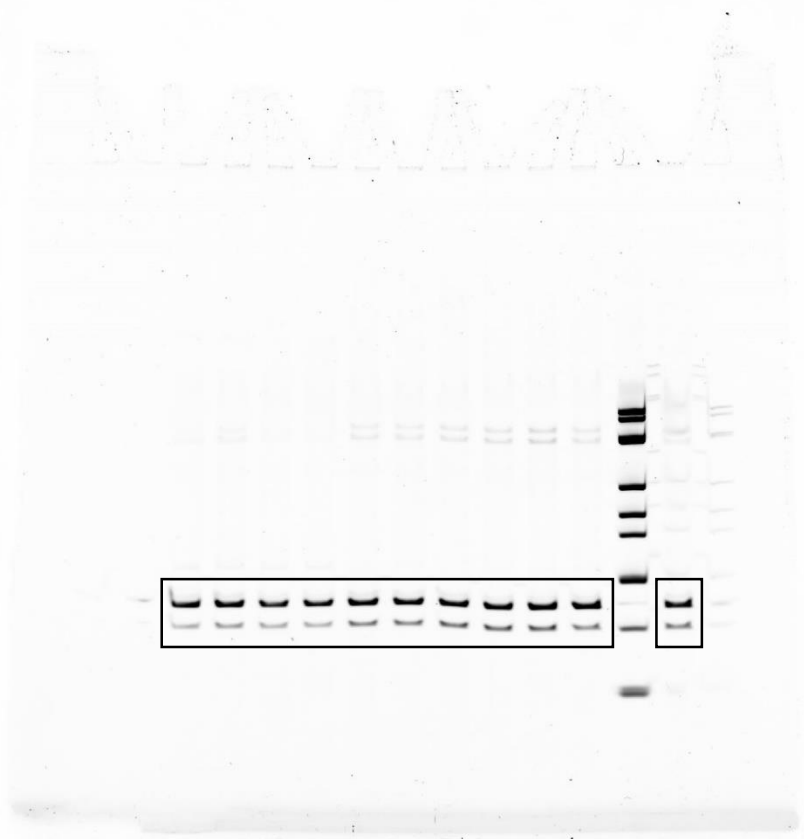


Full unedited gel for Figure S8

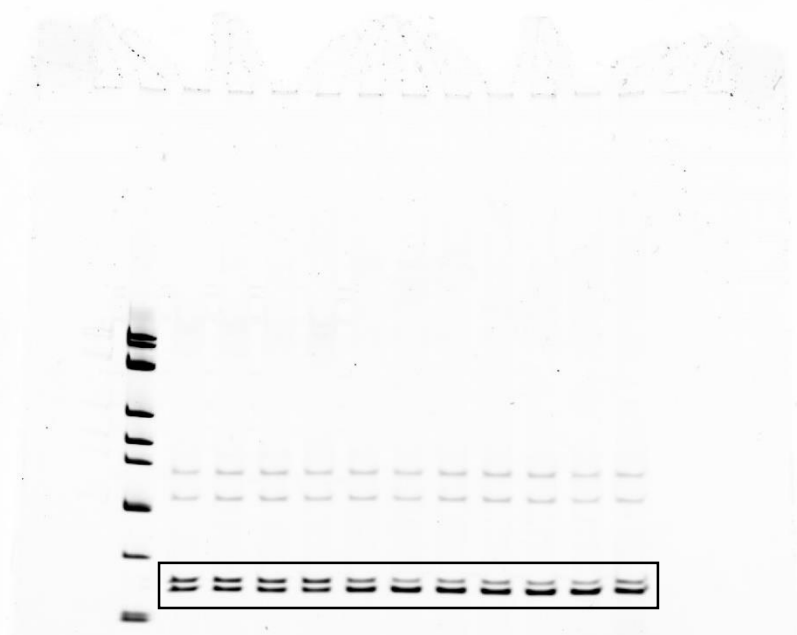
Splicing for *Csnk1d* (Fig S8C)



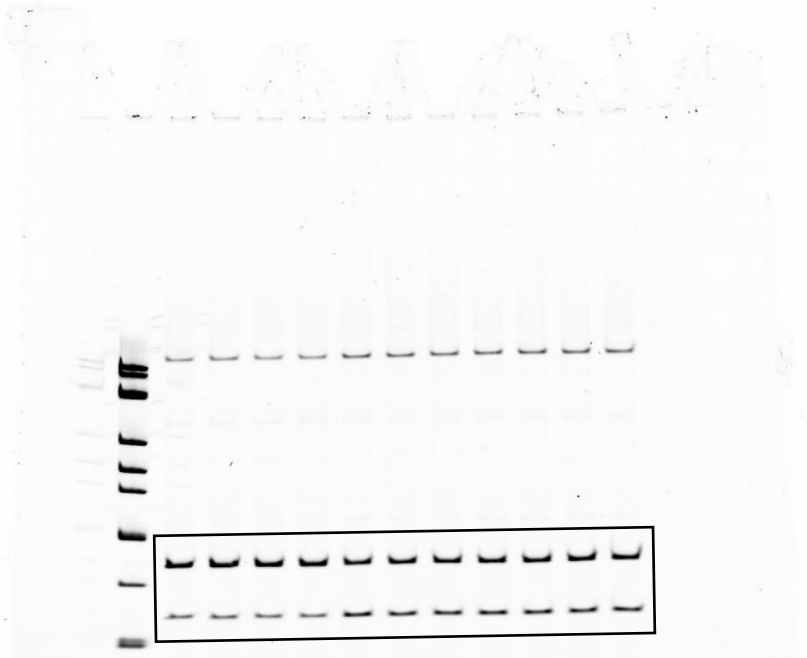
Splicing for *Nf2* (Fig S8C)



Splicing for *Arhgef10l* (Fig S8C)



Splicing for *Slk* (Fig S8C)



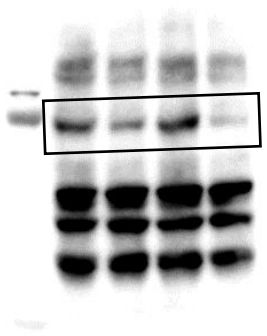
Splicing for *Kras* (Fig S8C)

Full unedited gel for Figure S9

Splicing for *Kras* (Fig S9B)

Full unedited blot for Figure S10

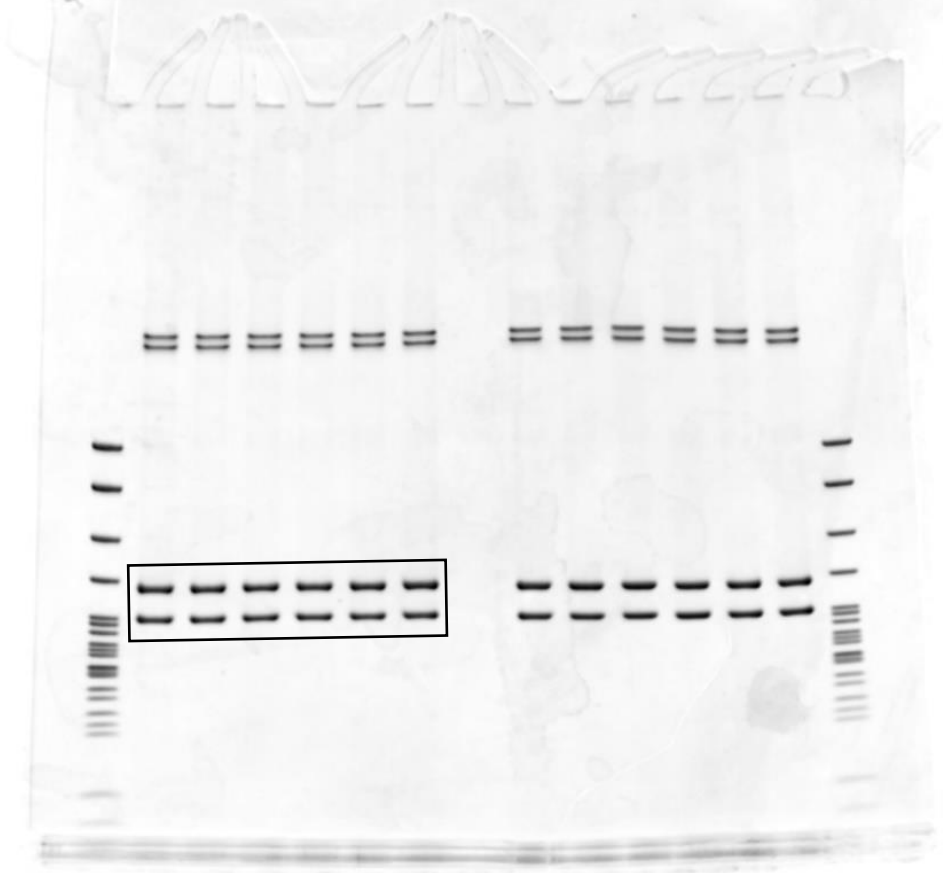
Immunoblot for ESRP2 (Fig S10B)
using rabbit anti-ESRP2 (Abcam ab113486)



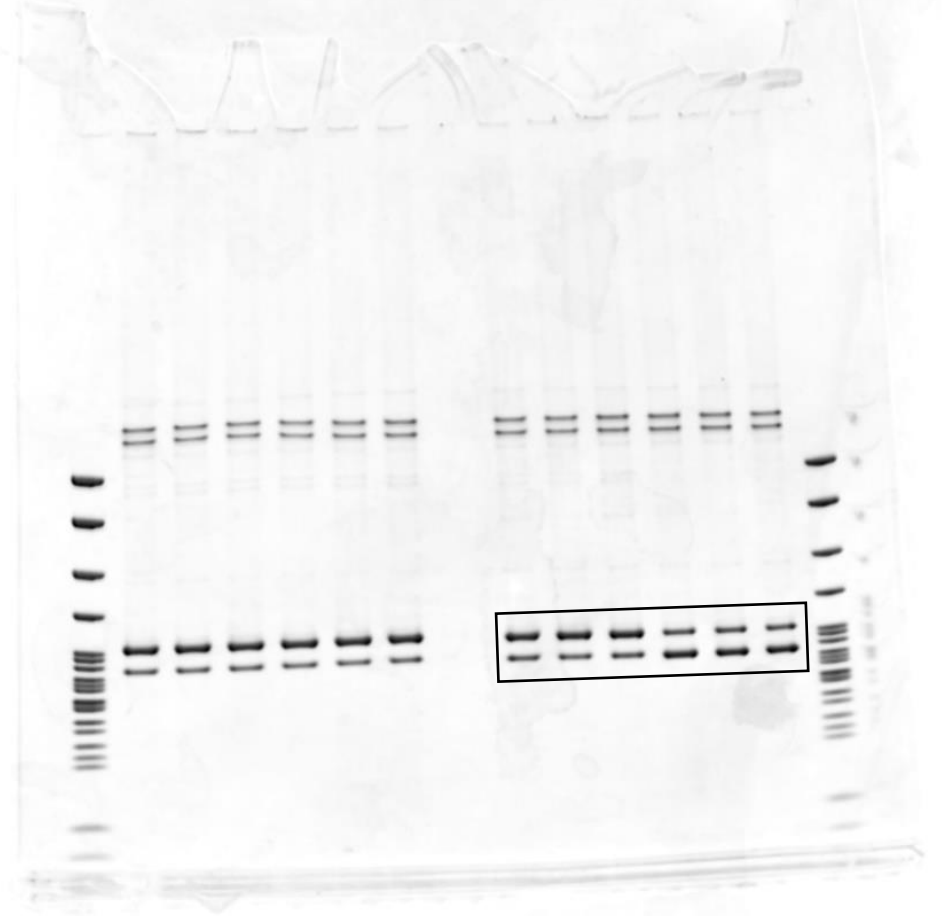
Immunoblot for GAPDH (Fig S10B)
using mouse anti-GAPDH (Santa Cruz sc-365062)

Full unedited gel for Figure S10

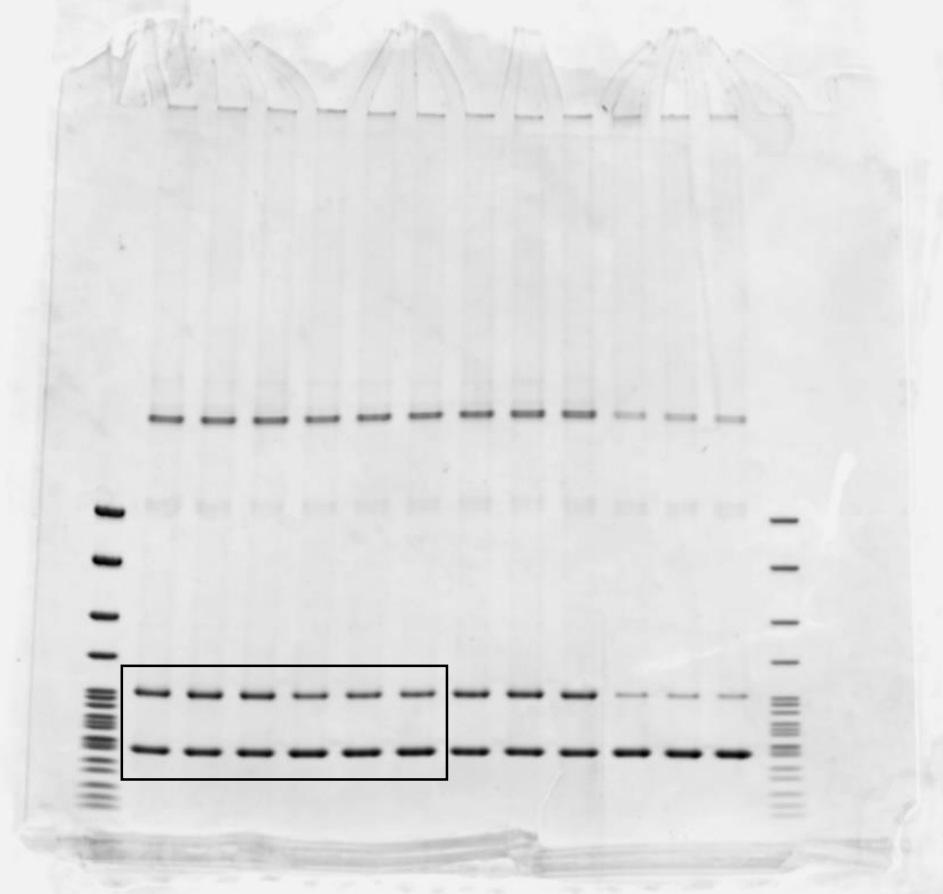
Splicing for *Csnk1d* (Fig S10C)



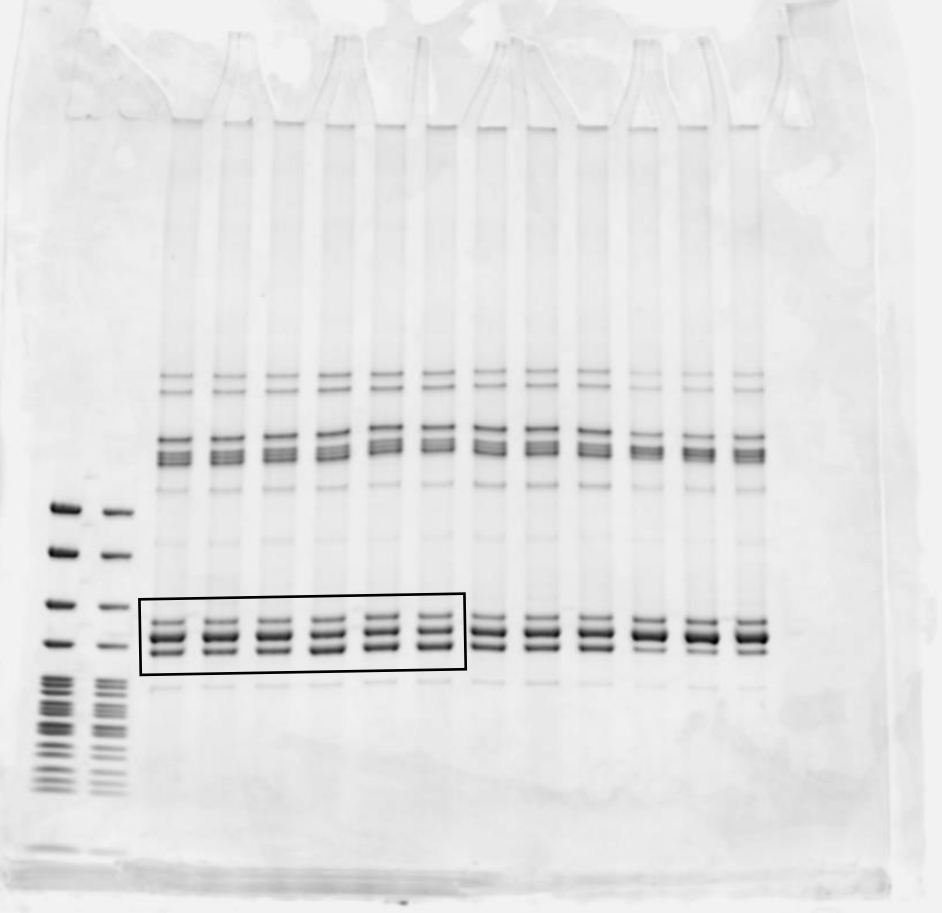
Splicing for *Nf2* (Fig S10C)



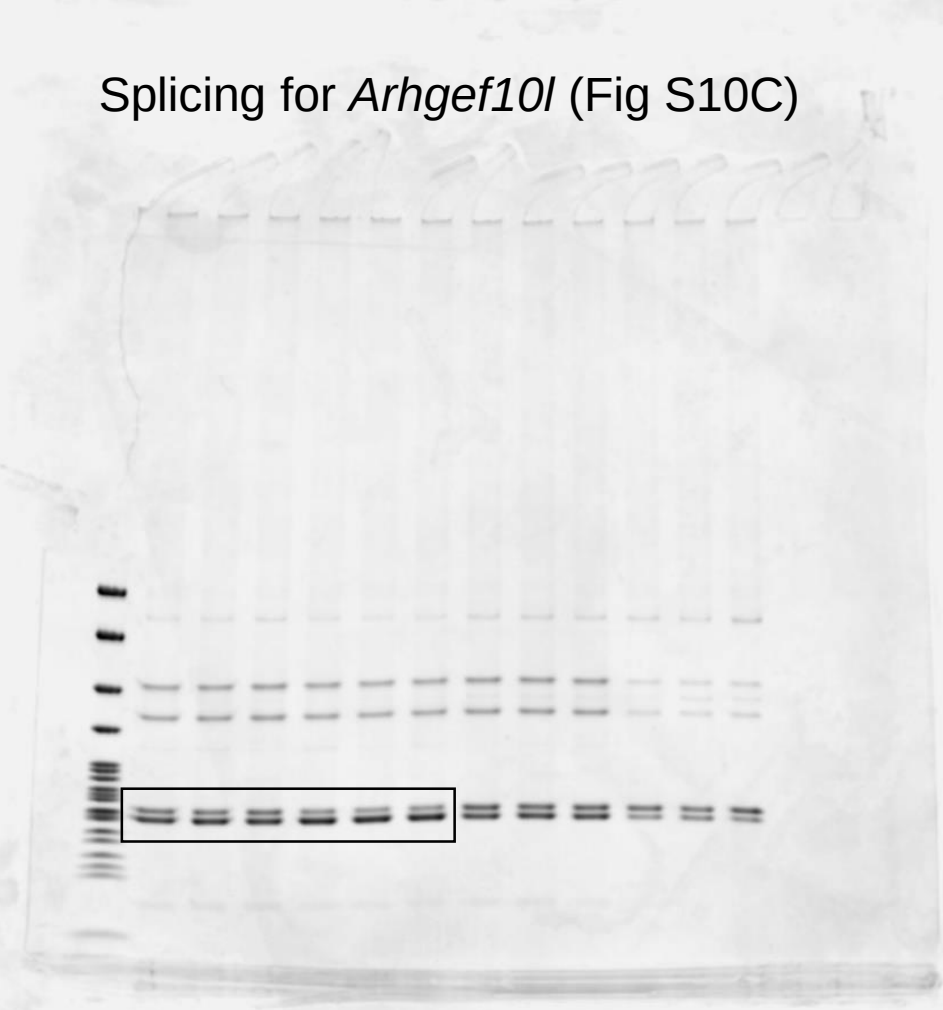
Splicing for *Slk* (Fig S10C)



Splicing for *Flnb* (Fig S10C)



Splicing for *Arhgef10l* (Fig S10C)



Splicing for *Kras* (Fig S10C)