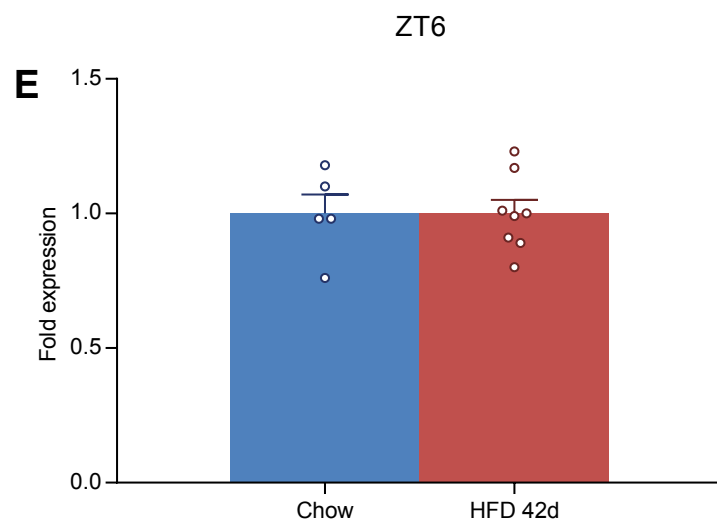
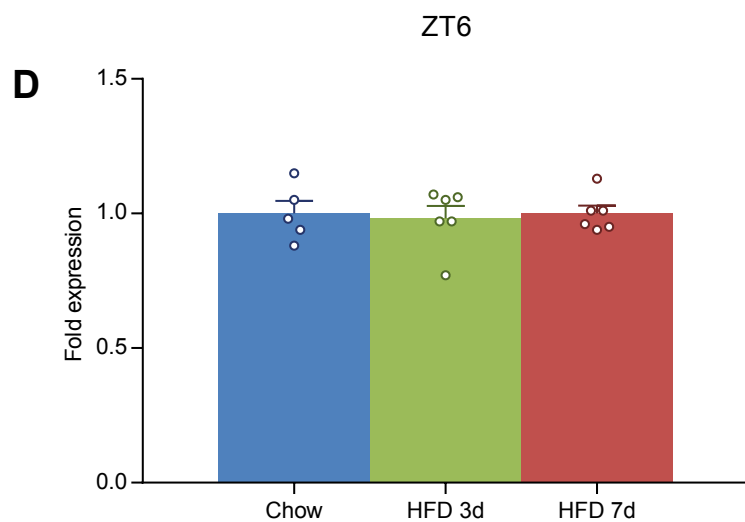
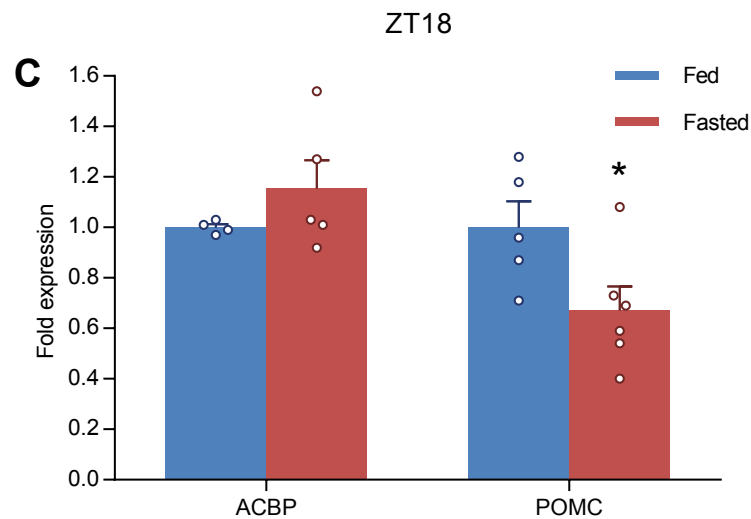
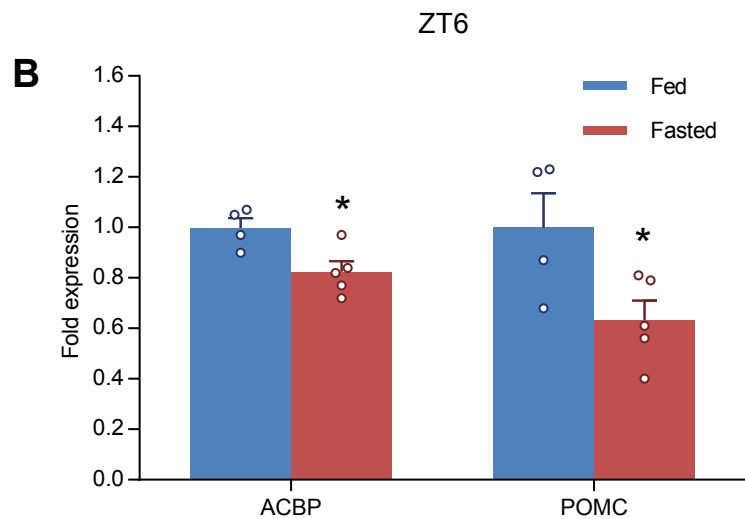
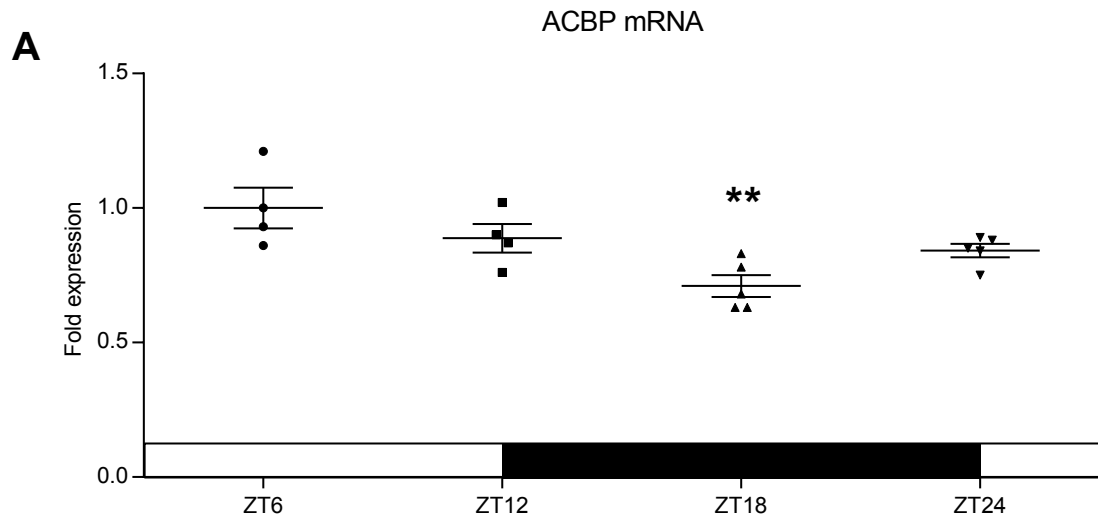
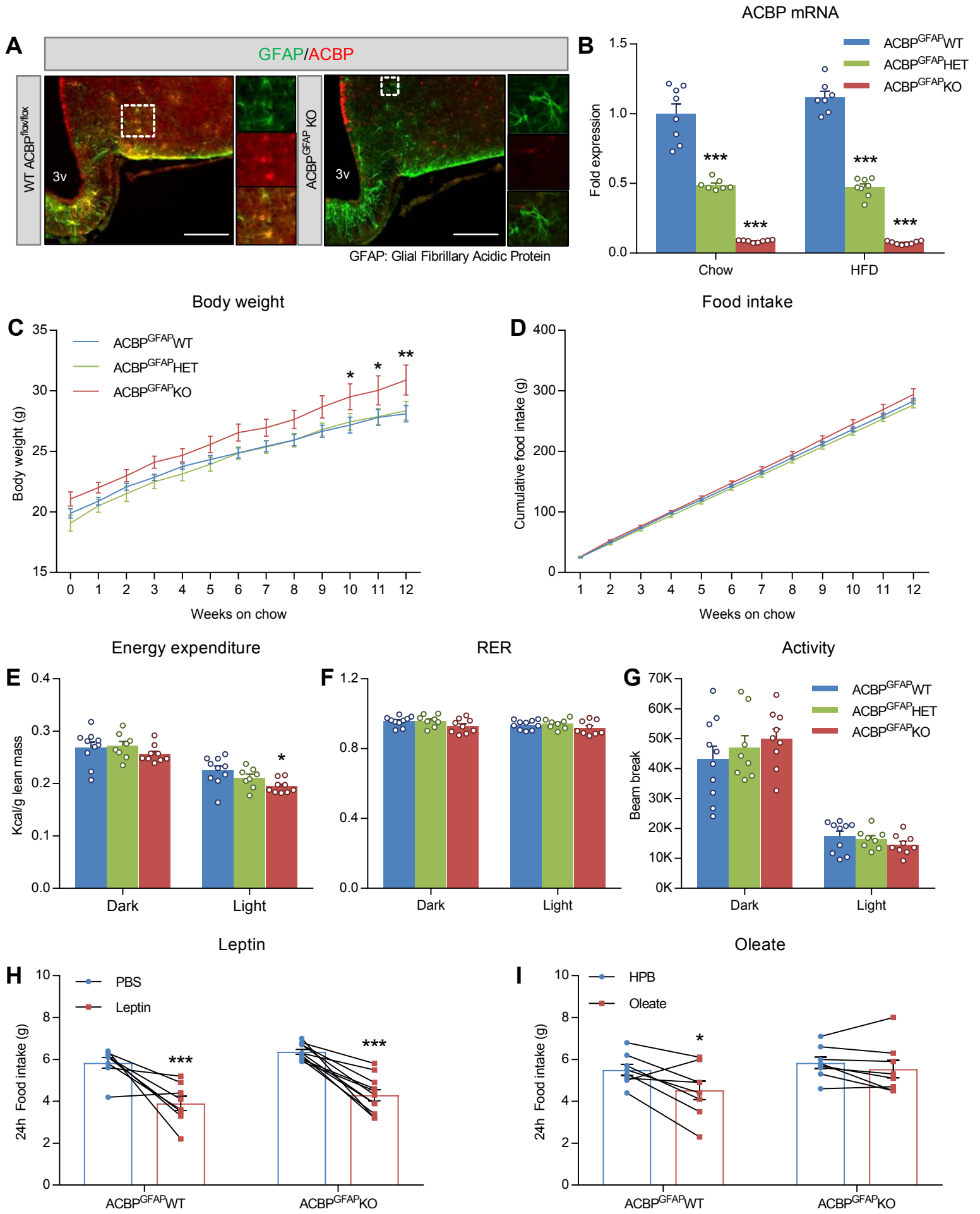


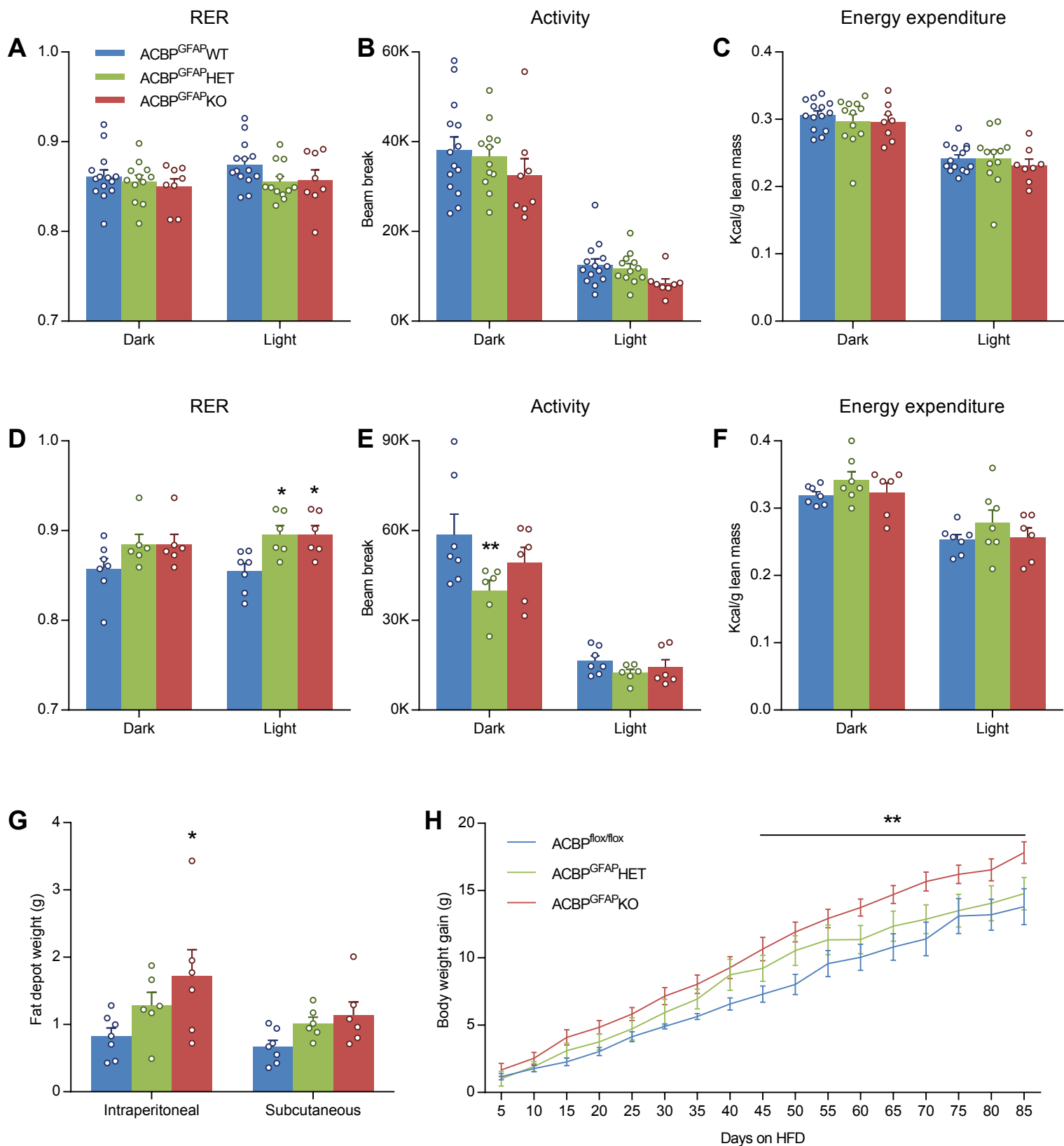
Supplemental Figure 1: *Acbp* gene expression is dependent on the circadian time and nutritional status. **A-** *Acbp* expression measured by qPCR in ARC microdissections of C57BL/6 WT male mice at different Zeitgeber times (ZT) (ZT0 corresponds to the onset of the light cycle in a 12 h light/dark cycle), ** $p < 0.01$ compared to ZT6, One-way ANOVA with Bonferroni post hoc test, N=4-5. *Acbp* and *pomc* expression in ARC microdissections of fed or 16 h-fasted C57BL/6 WT male mice at **B-** ZT6 or **C-** ZT18, * $p < 0.05$ Student's *t*-test compared to controls, N=4-5. *Acbp* expression in ARC microdissections (ZT6) of WT male mice fed with a chow or high fat diet during **D-** 3 or 7 days or **E-** 42 days. N=4-8.



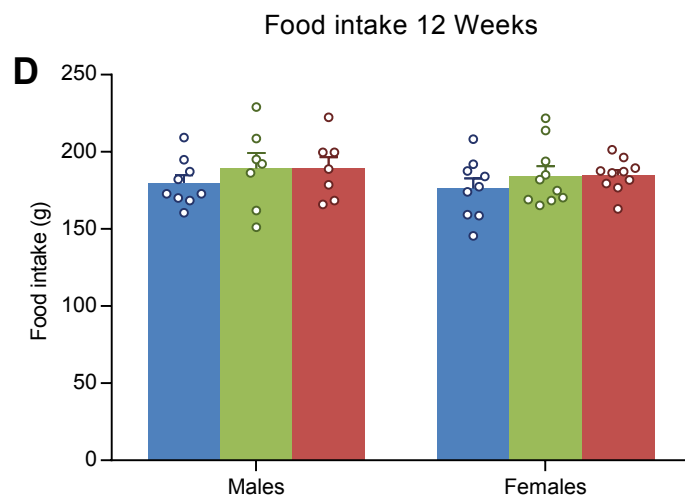
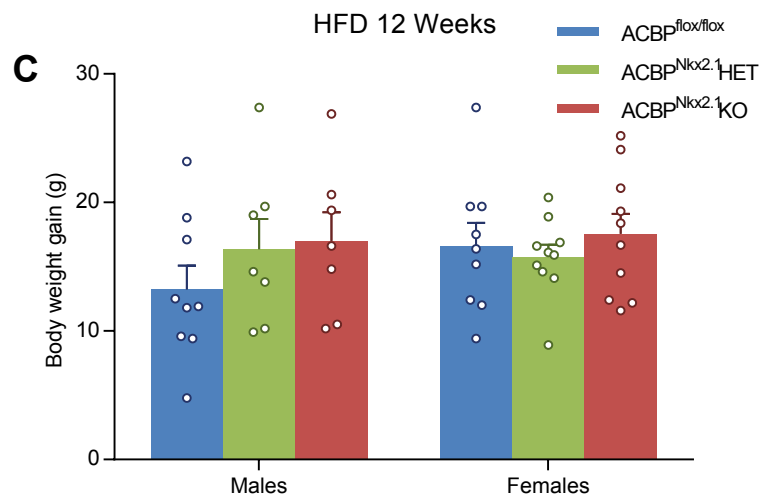
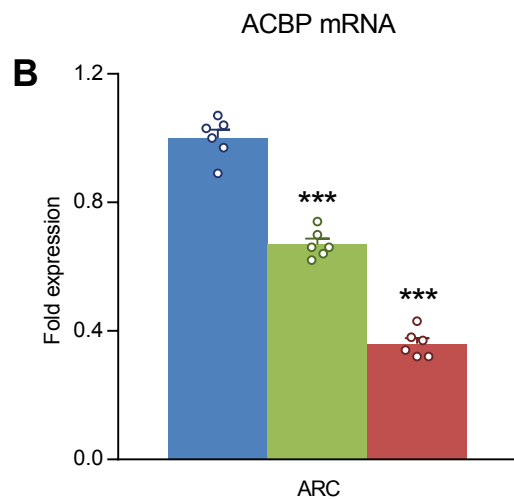
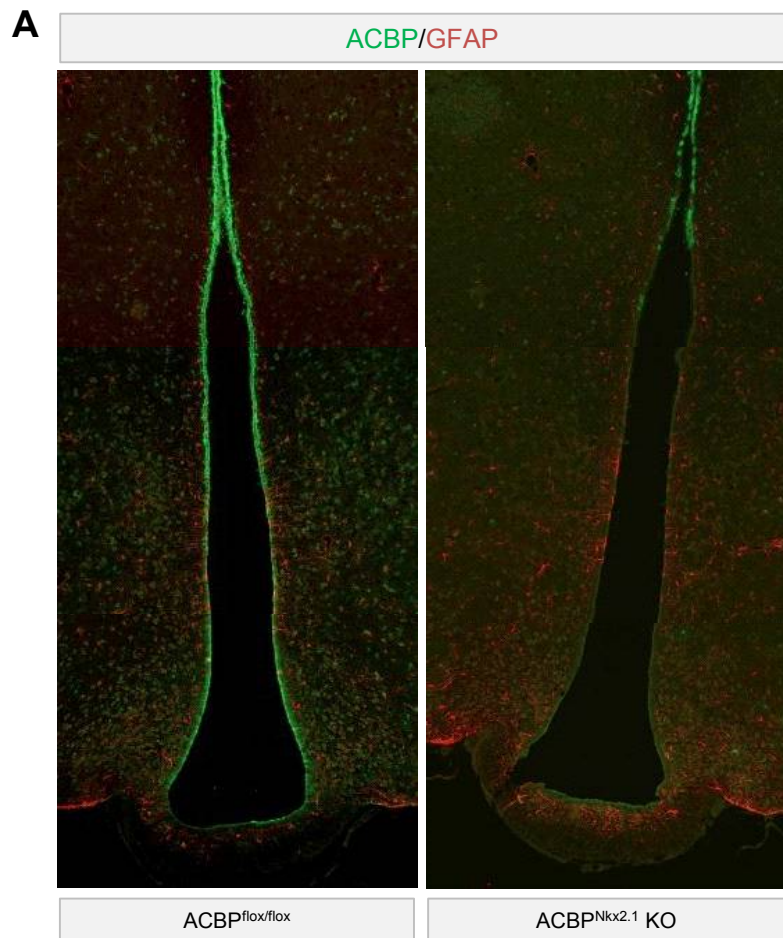
Supplemental Figure 2: Pan-brain astroglial ACBP invalidation in male mice. **A-** Immunostaining of ACBP (red) and GFAP (green) in *ACBP^{fllox/fllox}* (WT) and *ACBP^{GFAP}* KO mice. Scale bar represents 100 μ m. **B-** *Acbp* expression measured by qPCR in ARC microdissections from *ACBP^{GFAP}* WT, HET and KO male mice fed with either chow (12 weeks) or HFD (16 weeks). *** $p < 0.001$ compared to control littermates, One-way ANOVA followed by Bonferroni post hoc test, N=8. **C-** Body weight and **D-** cumulative food intake of *ACBP^{GFAP}* WT, HET and KO male mice fed with chow. **E-** Energy expenditure (normalized to lean mass), **F-** RER and **G-** Locomotor activity measured in CLAMS metabolic cages during 48 h following 24 h acclimation, * $p < 0.05$, ** $p < 0.01$ compared to controls, Two-way ANOVA with Bonferroni post hoc test, N=8-10. **H-** 24 h food intake after a 16 h fast in *ACBP^{GFAP}* WT and KO male mice receiving an ICV injection of PBS on day 1 and Leptin (1 μ g/2 μ l) on day 10, N=8-11. **I-** 24 h food intake in 2 h-fasted *ACBP^{GFAP}* WT and KO male mice receiving an ICV injection of 4.5 % cyclodextrin HPB on day 1 and oleate (6 nmol/2 μ l) on day 10, N=8. * $p < 0.05$, *** $p < 0.001$ compared to vehicle injection, Two-way ANOVA repeated measures with Bonferroni post hoc test (each animal is used as its own control).



Supplemental Figure 3: Pan-brain astroglial ACBP invalidation in male and female mice on a high-fat diet. A and D- RER, B and E- locomotor activity and, C and F- energy expenditure (normalized to lean mass) in male (N=8-15) (A-C) or female (N=6-7) (D-F) $ACBP^{GFAP}$ WT, HET and KO mice (backcrossed BL/6J) fed with a HFD during 16 weeks. G- Weight of intraperitoneal and subcutaneous fat pads of $ACBP^{GFAP}$ WT, HET and KO female mice, N=6-7 (backcrossed BL/6J). H- Body weight gain of $ACBP^{flox/flox}$, $ACBP^{GFAP}$ HET and KO female mice on a mixed BL/6J-Bom genetic background fed with a HFD during 12 weeks. * $p < 0.05$, ** $p < 0.01$ compared to control littermates, One-Way ANOVA with Bonferroni post hoc test (A-G). ** $p < 0.01$ Two-way ANOVA with Bonferroni post hoc test compared to controls, N=6-9 (H).



Supplemental Figure 4: ACBP invalidation in Nkx2.1 neural cells. **A-** Immunostaining of ACBP (green) and GFAP (red) in the hypothalamus of *ACBP^{flox/flox}* (WT) and *ACBP^{Nkx2.1}* KO male mice. **B-** *Acbp* expression measured by qPCR in ARC microdissections from *ACBP^{flox/flox}*, *ACBP^{Nkx2.1}* HET and KO mice. *** $p < 0.001$ compared to controls, One-Way ANOVA with Bonferroni post hoc test, N=6. **C-** Body weight gain and **D-** cumulative food intake in *ACBP^{flox/flox}*, *ACBP^{Nkx2.1}* HET and KO male and female mice (mixed BL/6J-Bom background) fed with a HFD during 12 weeks. N=7-9.



Supplemental Figure 5: Modulation of neuronal activity by ODN and ODN GPCR. Representative cell-attached recording of **A-** POMC or **B-** non-POMC neuron in response to 2 nM cOP. **C-** Quantification of AP frequency of non-POMC neurons in response to 2 nM cOP, N=12 neurons from 3 mice. **D-** Representative bright-field image of dissociated mediobasal hypothalamic cells in culture, scale bar represents 40 μ m. Representative Ca^{2+} imaging traces from dissociated neurons treated with 1 nM ODN in **E-** absence or **F-** presence of cdLOP ODN receptor antagonist (10 nM). **G-** Quantification of ODN response intensity (AUC) with or without cdLOP (ODN: N=36 ODN responsive / 425 total cells; ODN + cdLOP : N=17 ODN responsive/ 455 total cells), 3 independent cultures from 6 animals, * $p < 0.05$, Student's t -test. **H-** Representative trace and **I-** quantification of AP frequency of POMC neurons in the presence of GABA_A inhibitors (gabazine 5 μ M and picrotoxin 100 μ M) in response to ODN application (1 nM) with or without cdLOP (10 nM). * $p < 0.05$ compared to control (GZ + PTX) and cdLOP , One-way ANOVA repeated measures with Bonferroni post hoc test, N=10 neurons from 3 mice.

Methods: Panel **A**, POMC-neurons were recorded from POMC-eGFP mice as detailed in the method section. Non-POMC (panels **B** and **C**) and POMC neurons (panels **H** and **I**) were recorded in brain slices from 6 weeks old POMC-Cre male mice (Tg(Pomc1-cre)16Low/J, JAX Stock #005965, Jackson Laboratory, USA) bilaterally injected in the ARC (stereotaxic coordinates: antero-posterior -1.1 mm from bregma; lateral \pm 0.3 mm; dorso-ventral -5.8 mm) with an AAV-DIO-mCherry (350 nL rAVV8-hSyn-DIO-mCherry, 3.7×10^9 GC/ μ L, UNC Vector, Lot #AV4981E), 2-3 weeks before performing electrophysiological recordings as described in the method section. To decrease the chance of recording false negative cells (POMC neurons not infected by the AAV), we recorded non-fluorescent neurons (N=12) surrounded by POMC-mCherry positive neurons (panels **B** and **C**).

