Familial Congenital Hypothyroidism Due to Inactivating Mutation of the Thyrotropin Receptor Causing Profound Hypoplasia of the Thyroid Gland

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Abstract

Thyroid gland agenesis is the most common cause of congenital hypothyroidism and is usually sporadic. We investigated a brother and sister from consanguineous parents, ascertained through systematic newborn screening, and initially diagnosed with thyroid agenesis. Careful cervical ultrasonography in both patients revealed a very hypoplastic thyroid gland. By direct sequencing of the thyrotropin receptor gene, we identified the substitution of threonine in place of a highly conserved alanine at position 553, in the fourth predicted transmembrane domain. The mutation was found homozygous in the affected siblings, and heterozygous in both parents and two unaffected siblings. Functional analysis in transfected COS-7 cells showed that it resulted in extremely low expression at the cell surface as compared with the wild-type receptor, in spite of an apparently normal intracellular synthesis. The small amount of mutated receptor expressed at the surface of transfected cells bound thyrotropin with normal affinity and responded in terms of cAMP production, but the in vivo significance of these data from overexpressed receptor in transfected cells is unclear. Of note, blood thyroglobulin was unexpectedly elevated in the patients at the time of diagnosis, a finding that might prove useful in refining etiologies of congenital hypothyroidism. (J. Clin. Invest. 1997; 99:3018–3024.) Key words: thyrotropin receptor • congenital hypothyroidism • thyroglobulin • thyroid agenesis

Introduction

The pituitary glycoprotein hormone thyrotropin (TSH) stimulates growth, differentiation, and function of the thyroid gland, and these effects are mimicked by cAMP agonists (1). TSH acts on the thyroid follicular cell through a specific receptor coupled to G proteins, whose hydropathy profile (2–5) dis-

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Introduction

The pituitary glycoprotein hormone thyrotropin (TSH) stimulates growth, differentiation, and function of the thyroid gland, and these effects are mimicked by cAMP agonists (1). TSH acts on the thyroid follicular cell through a specific receptor coupled to G proteins, whose hydropathy profile (2–5) displays a characteristic serpentine structure with seven transmembrane domains. Together with the other glycoprotein hormone receptors, LH/CG and FSH, the TSH receptor (TSHr) also presents, in its aminoterminal half, a large extracellular domain involved in hormone binding. The TSHr gene contains nine introns, with exon 10 coding for the entire serpentine structure (6). Besides stimulation of the cAMP cascade, the TSHr is capable of stimulating the phospholipase C–diacylglycerol–phosphatidylinositol pathway at higher concentrations of TSH, at least in man, and these effects are mediated by Gs and Gq proteins, respectively (1). In functional studies, the TSHr displays a significant level of spontaneous activity (i.e., stimulation of second messenger production in the absence of ligand) in experimental conditions where other G protein–coupled receptors are essentially silent (7–10). Mutations causing constitutive activation of the receptor have been found at the somatic level in hyperfunctioning thyroid adenomas (hot nodules) (7, 8), and at the germline level in familial nonautoimmune thyroid hyperplasia, a very rare condition (9). Reciprocally, inactivating mutations of the receptor have recently been described in patients with TSH resistance (11, 12) (see below).

Congenital hypothyroidism affects 1 in 4,000 newborns (13). Early thyroid hormone replacement therapy prevents the growth and development delay that develop postnatally in untreated patients, and systematic newborn blood screens for high TSH and/or low thyroxine (T4) have been implemented in most western countries. The majority of cases of congenital hypothyroidism are sporadic and are due to thyroid gland agenesis (athyrosis), or dysgenesis (ectopic and hypoplastic gland). These cases are attributed to a deficient embryogenesis of the thyroid; they do not show a clear tendency to recur within families of affected children and are believed to result from nongenetic causes, like environmental insults or stochastic events early in embryogenesis. A minority of congenital hypothyroidism cases, perhaps 10%, result from inborn errors of thyroid hormonogenesis, and show Mendelian inheritance, almost always of the autosomal recessive type (1). Homozygous or compound heterozygous mutations of the thyroglobulin (TG) gene, and of the thyroid peroxidase (TPO) gene, have been described in such families. In untreated patients, the increase in blood TSH activity often results in goiter, which may be present at birth (14).

It is expected that one subtype of inborn errors, if inactivating the TSHr, will not result in goiter, and instead resemble thyroid agenesis, except for showing autosomal recessive inheritance. A mouse strain called hyt/hyt presents with inherited autosomal recessive hypothyroidism and a hypoplastic gland (15). The affected mice were shown to harbor a homozygous mutation of the murine Tshr gene (16) causing defective TSH binding (17). In man, the concept of TSH unresponsiveness has been validated by clinical descriptions of patients (18, 19) and families (20) that fit the expected phenotype of low triiodothyronine and T4, and thyroid gland hypoplasia, in spite of

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high circulating levels of bioactive TSH. With the cloning of the human TSHr gene (3–5), molecular analysis of such cases became possible, but the search for mutations remained unsuccessful in previous studies (21). Recently, one patient with sporadic congenital hypothyroidism was shown to be a compound heterozygote for presumably inactivating mutations of the TSHr gene (22). On the other hand, compound heterozygosity for functionally tested, inactivating TSHr mutations was identified in siblings with resistance (i.e., partial unresponsiveness) to TSH without overt hypothyroidism (11, 12).

In this report, we describe a familial case of TSH unresponsiveness with congenital hypothyroidism due to a homozygous inactivating mutation causing very low expression of the TSHr at the cell surface, and profound thyroid hypoplasia.

Methods

Patients. Congenital hypothyroidism was identified through systematic screening on the fourth day of life in a boy whose blood spot TSH was > 130 mU/liter, and serum T4 was low at 1.2 μg/dl. The pregnancy had lasted 41 wk, and a cesarean section was performed because of transverse presentation. The birth weight was 4.30 kg, length 51 cm, and head circumference 41.7 cm. Physical examination and 99mTc scintigraphy failed to demonstrate the presence of thyroid tissue, and the patient was given a diagnosis of athyrosis. T4 therapy was initiated on the 9th d of life, and the neonatal course was otherwise uneventful. The family history (Fig. 1) revealed that the parents were second cousins, and that congenital hypothyroidism was previously identified in siblings with resistance (i.e., partial unresponsiveness) to TSH without overt hypothyroidism (11, 12).

Inconsistent with the diagnosis of athyrosis (23), serum TG was in the high normal range at the time of diagnosis in both patients (66 ng/ml in propositus, 84 ng/ml in affected sister). Careful ultrasonography in the propositus revealed a hypoechogenic gland in normal position below the thyroid cartilage, whose transverse diameters were 5.2 and 4.9 mm for the left and right lobes, respectively, which corresponds to a very hypoplastic thyroid (24). Serum TG decreased over a 6-mo period while under T4 therapy. Treatment was interrupted for 6 wk in the sister at age 3.7 yr for investigative purposes. Physical examination revealed no palpable thyroid, whereas serum-free T4 became very low, < 0.2 ng/dl. TSH increased to 186 mU/liter, and T4 remained unchanged at 2.9 ng/ml. Like in her brother, ultrasonography showed a very hypoplastic gland in normal position, with transverse diameters of 5.6 and 4.6 mm for the left and right lobes, respectively.

Serum TG was found elevated at 53.7 ng/ml in the father, but was in the normal range at 17.1 ng/ml in the mother. TG levels in the unaffected brothers were 31.4 and 41 ng/ml, which is in the normal range for children in our population (13). Serum TSH and free T4 were within normal limits in both parents and both unaffected brothers. Autoantibodies directed against TG, TPO/microsomal antigen, or the TSHr were not detected in the patients, nor in either parent or unaffected sibling. The parents consented to these studies. These data are summarized in Fig. 1.

Sequencing of the TSHr gene. DNA was extracted from peripheral blood using standard procedures. PCR amplification was designed to produce five overlapping fragments that cover the whole length of exon 10. Exons 1 through 9 were amplified individually using couples of intronic primers. The fragment encompassing the mutation identified in this family was used to generate the following primers (positions 1567–1960): forward primer: 5’-tggcatcttggtatgccatcacttc-3’; reverse primer: 5’-caggaaaaagatctgactgagggctgttcagaatt-3’. The fragment was sequenced using couples of intronic primers and 99mTc isotopic scan.

Blood tests were performed in the parents and four children. Values in unaffected siblings are at the time of diagnosis, and all measures are in the absence of thyroid hormone treatment. Normal values are given in brackets, except for serum TG, for which normal values in our population with borderline iodine intake are as follows (ng/ml): newborns, 2–10; < 10 yr, 2–65; adults, 2–35. AutoAb, serum autoantibodies against TPO, TG, and the TSHr; −, below level of clinical significance; wt, wild-type TSHr sequence; M, Ala553→Thr (A553T) mutation as in Fig. 2.

PCR was performed in a 100 μl final volume containing 1 μg DNA, 50 mM KCl, 10 mM Tris-HCl, pH 8.3, 2 mM MgCl₂, 0.01% DMSO, 0.5 μM each primer, 0.2 mM dNTPs, 0.01% Tween, and 0.8 U Taq DNA polymerase. The amplification conditions were 94°C for 2 min, followed by 35 cycles of 94°C for 15 s, 60°C for 15 s, and 72°C for 1 min. The final extension step was 72°C for 7 min. The amplified PCR products were separated by electrophoresis in a 2% agarose gel and visualized by ethidium bromide staining.

Figure 1. Thyroid blood tests and genotypes in the family. The parents are second cousins and the propositus is indicated by the arrow. Solid symbols indicate congenital hypothyroidism with apparently absent thyroid gland on physical examination and 99mTc isotopic scan. Blood tests were performed in the parents and four children. Values in affected siblings are at the time of diagnosis, and all measures are in the absence of thyroid hormone treatment. Normal values are given in brackets, except for serum TG, for which normal values in our population with borderline iodine intake are as follows (ng/ml): newborns, 2–10; < 10 yr, 2–65; adults, 2–35. AutoAb, serum autoantibodies against TPO, TG, and the TSHr; −, below level of clinical significance; wt, wild-type TSHr sequence; M, Ala553→Thr (A553T) mutation as in Fig. 2.
gelatine, 0.2 mM dNTP, 5 U Taq Polymerase (GIBCO-BRL, Gaithersburg, MD), and 100 nM of each primer. The annealing temperature was 50°C. The PCR products were sequenced on both strands using the Thermosequenase fluorescent labeled primer cycle sequencing kit (RPN 2436; Amersham Corp., Arlington Heights, IL) according to the manufacturer’s recommendations. Samples were loaded on a 373 Stretch Sequencing Instrument (Applied Biosystems, Inc., Foster City, CA) and mutations identified using the Fasta and Sequence Navigator Software (ABI Advanced Biotechnologies, Inc., Columbia, MD).

Cloning and expression of the mutated TSHr. A CmI-BstElI fragment (1604–2169) in the wild-type TSH receptor–pSVL construct was substituted by the mutant counterpart as previously described (7). Pfu enzyme was used to generate this fragment, in a 100 µl final volume containing 200 ng genomic DNA, 10 mM KCl, 20 mM Tris-HCl, pH 8.75, 10 mM (NH4)2SO4, 2 mM MgSO4, 0.1% Triton X-100, 0.1 mg/ml BSA, 0.2 mM dNTP, 5% DMSO, 2.5 U Pfu DNA Polymerase (Stratagene Inc., La Jolla, CA), and 0.5 µM of each primer. The annealing temperature was 52°C. The resulting constructs were verified by double stranded sequencing as described above.

COS-7 cells were grown in Dulbecco’s modified Eagle’s medium supplemented with 10% fetal bovine serum, 1 mM sodium pyruvate, 100 IU/ml penicillin, 100 µg/ml streptomycin, and 2.5 µg/ml fungizone, and seeded at a density of 150,000 or 300,000 cells/cm² dish. 1 d later, they were transfected (250 or 500 ng DNA/150,000 or 300,000 cells, respectively) by the DEAE-Dextran method followed by a dimethylsulfoxide shock (25). To obtain a “standard curve” of expression, various amounts of the wild-type construct were mixed with pSVL vector or with the mutated construct at 500 ng final DNA amount and transfected in parallel in COS-7 cells.

2 d after transfection, triplicate dishes were used for flow immunocytofluorometry, cAMP determinations, and 125I-TSH binding studies; duplicate dishes were used for inositol phosphate determinations. Each experiment was repeated at least three times.

Binding and flow immunocytofluorometry studies. 2 d after transfection, cells were washed twice with NaCl-free Hank’s buffer supplemented with 280 mM sucrose, 0.2% BSA, and 2.5% low fat milk, and were incubated at room temperature in fresh medium supplemented with 100,000 counts/ml 125I-TSH (35 µCi/µg, 40 U/mg TRAK assays; BRAHMS diagnostica, Berlin, Germany) and various concentrations of unlabeled TSH. 4 h later, the cells were rinsed twice with chilled Hank’s buffer and solubilized with 1 M NaOH. Bound radioactivity was determined in a gamma scintillation counter. Under these conditions, the radioactivity nonspecifically bound to COS-7 cells expressing the TSH receptor (defined as the radioactivity bound to the dish in the presence of 100 nM unlabeled TSH) was identical to that bound to mock-transfected cells. In the absence of a consensus about the bioactivity of pure bovine TSH, we have expressed all TSH or TSH receptor concentrations in milliunits/milliliter, assuming a 1:1 stoichiometry and an Rmax of 4.6 × 10^6 M–1 l final volume of 0.1% PBS-BSA containing the 2C11 mAb (10 µg/ml) in the same buffer supplemented with 100 µM Rolipram (cAMP phosphodiesterase inhibitor, a gift from the Laboratory J. Logeais, Paris, France), in the absence of added agonist, or in the presence of various concentrations of bovine TSH (Sigma Chemical Co.) or 10 µM forskolin (Calbiochem Corp., La Jolla, CA). The medium was removed after 60 min and 0.1 M HCl was added to the cells. The samples were dried overnight in a vacuum concentrator and intracellular cAMP, expressed as picomoles per dish, was measured in duplicate for each dish by 125I-cAMP assay system (code 432, TRK; Amersham Corp.). For inositol phosphates (IPs) determination, cells were incubated with 20 µCi 3H-inositol (code TRK-911; Amersham Corp.) per ml DMEM for 24 h. The cells were then washed twice with KRH. After a preincubation in KRH supplemented with 10 mM LiCl at 37°C for 30 min, cells were incubated at 37°C for 15 min within the same medium in the absence of added agonist, or in the presence of 100 µM/ml bovine TSH or 0.1 mM ATP. The medium was removed and the reaction was stopped by addition of ice cold 3% HClO4. 125I-labeled IPs were isolated and assayed by stepwise chromatography on AG1X8 resin (28). Cellular fragments on the dishes were dissolved with 1 M NaOH and counted as phosphatidylinositols. Results were expressed as a percentage of radioactivity incorporated in IPs (IP1 + IP2 + IP3) over the sum of radioactivity incorporated in IPs and phosphatidylinositols.

Computation of transfection efficiency and specific constitutive activity. Constitutive activity was determined for each construct using the following formula: 

\[
C = \frac{A_0 - Z}{Z}
\]

where \(A_0\) is the basal (no ligand added) cAMP value as measured experimentally, \(Z\) is the basal cAMP value in mock (pSVL)-transfected cells, and \(e\) is the transfection efficiency as determined by FACS® Cytofluorometry assay, allowing for subtraction of ambient cAMP production by untransfected cells. \(C\) is thus the ratio between the cAMP level in cells expressing the receptor constructs over the cAMP level in the same number of cells transfected with the pSVL vector alone. \(C/R\) normalizes \(C\) for baseline cAMP production and allows for a measure of the relative constitutivity of the mutant receptor over the wild type is given by \(F = C/R\) mutant/C wild type. This value does not account for the differences in expression of the mutants at the cell surface. It gives a measure of relative constitutivity, at the actual level of expression achieved by individual receptor constructs. An estimation of specific constitutive activity (SCA) (i.e., constitutive activity normalized to equal level of expression) is given by SCA = \(F/R\), where \(R\) is a measure of the expression of the mutant receptor relative to the wild type with \(R = (\text{Mut-pSVL})/\text{(WT-pSVL)}, the ratio between the fluorescence of cells transfected with the mutant and the wild-type receptor constructs, after subtraction of the background fluorescence displayed by mock transfected cells. The comparison of SCAs computed for each construct determines the relative constitutivity of molar amounts of the mutant over the wild type.

Statistics. Levels of significance were evaluated by a two-tailed unpaired Student’s t test, and \(P < 0.05\) was considered significant.

Results

Familial mutation of the TSH receptor. Considering the clinical picture consistent with TSH unresponsiveness and the fa-
Familial Congenital Hypothyroidism Due to Thyrotropin Receptor Mutation

As sole anomaly, we identified a GCC→ACC mutation (Fig. 2 B) affecting the alanine residue at position 553, which was replaced by a threonine in the fourth putative transmembrane domain of the TSHr protein as schematized in A. Also represented in A is the location of the P556L mutation identified in the hyt/hty mouse (see text). The genotypes of our patients and family are mentioned in Fig. 1.

Expression of the mutated receptor. The functional characteristics of the mutated receptor were studied by transient expression in COS-7 cells. Cells transfected with a cDNA construct encoding the wild-type receptor or the empty pSVL vector were used as controls. Levels of expression at the cell surface were assessed by flow cytometry using a monoclonal antibody recognizing the linear epitope VFFEEQ (residues 354–359) (27, 29) of the extracellular amino-terminal domain. It demonstrated an extremely low level of expression for the mutant (Fig. 3). The comparison of saponin-permeabilized (Fig. 3 B) and nonpermeabilized (Fig. 3 A) cells showed that synthesis of the different constructs was quantitatively similar, which indicated that the mutant and wild-type receptors were equally recognized by the monoclonal antibody and that the low level...
of expression at the cell surface resulted from a posttranslational event, perhaps a defect in the folding of the receptor resulting in abnormal routing to the plasma membrane. The expression of the mutant was identical when transfected cells were incubated at 30°C for 48 h before the functional assays (data not shown).

Binding studies were congruent with the FACS\textsuperscript{®} analysis, and showed very low expression of TSH binding sites at the plasma membrane of cells transfected with the mutant construct, with an $R_t$ value of 0.03 mU/ml vs. 1.58 mU/ml for wild type in a representative experiment (Fig. 4). The observed dissociation constant of the mutant receptor was significantly smaller than wild type (0.21 mU/ml vs. 1.26 mU/ml, Fig. 4), which was confirmed in three independent experiments yielding values of 0.22±0.01 for $K_d$ and 0.018±0.01 mU/ml for $R_t$ (mean±SD). This may simply reflect the decrease in apparent $K_d$ that parallels the decrease in TSHr expression, as recently observed as compared with the wild-type receptor for both EC_{50} (~0.3 mU/ml) and maximally active TSH concentration (~10 mU/ml) (data not shown). In the absence of added agonist, we observed no significant stimulation of IP accumulation with the mutated as compared with the wild-type receptor. Stimulation by 100 mU/ml TSH elicited no response from the mutated construct as compared with the mock (pSVL)-transfected cells.

**Discussion**

Athyrosis is rarely, if ever, familial, and its cause has remained elusive in spite of its relatively high frequency. We show here that a genetic defect of the TSHr can cause a syndrome of apparent athyrosis that is transmitted as autosomal recessive.

The Ala553→Thr missense mutation we identified in the TSHr gene affects a highly conserved amino acid residue, and cosegregates with the disease in this family. It is located in the fourth putative transmembrane domain of the receptor (Fig. 2 A), and is reminiscent of the closely located Pro556→Leu (P556L) mutation of the murine TSHr, described in the hypothalamic hyt/hyt mouse, an animal model for TSH unresponsiveness (16). This mouse strain exhibits autosomal recessive congenital hypothyroidism with a normally located hypoplastic thyroid gland, as found in our patients. TSH unresponsiveness in these mice was found to result from defective TSH binding to a mutated receptor, which was apparently present at the plasma membrane (17). Unlike the mouse model however, TSH unresponsiveness in our patients is explained by the extremely low expression at the cell surface (Fig. 3 A) of the A553T mutant receptor, in spite of synthesis in normal amounts (Fig. 3 B).

Expression of TSH receptor constructs in COS-7 cells dif-
Familial Congenital Hypothyroidism Due to Thyrotropin Receptor Mutation

were significant at \( P = 0.0012 \) (*) between mutant and wild-type constructs, and at \( P = 0.01 \) (o) between mutant and pSVL control (mock-transfected cells). (B) Accumulation of IPs determined in unsupplemented control medium (open), under stimulation by 100 mU/ml TSH (gray) or 100 \( \mu \)M ATP (hatched gray). Results are expressed as the percentage of radioactivity incorporated in IPs over the sum of radioactivity in IPs and phosphatidylinositols ([IP1 + IP2 + IP3]/[IPs + PI]) \(
\times 100\). Values are means ± range from a representative experiment performed in duplicate dishes. No significant IP production was observed from the mutant in control medium nor under maximal TSH stimulation. *\( P = 0.477\).

fers from the natural expression in thyrocytes in many aspects: (a) the level of expression is probably two to three orders of magnitude higher, as the consequence of transcription from a high number of copies containing a strong promoter; (b) the complement and concentrations of G proteins are likely to be different; (c) the characteristics of the membrane system and routing of membrane proteins are also different in a nonpolarized cell as opposed to the polarized thyrocyte. Despite these limitations, and perhaps because of some of them, overexpression of the A553T mutant in COS-7 cells provides pertinent information on structure/function relationships that add to the current molecular understanding of the TSHr, but whose in vivo relevance is unclear. The small number of receptors reaching the membrane seem to behave quite normally in terms of TSH binding and sensitivity to hormone for adenylyl cyclase stimulation. The apparent decrease in \( K_d \) is likely to result from the small number of receptors inserted correctly in the membrane (30), as this parameter normalizes when the wild-type and mutant receptors are expressed at similar levels (Fig. 4). The mechanism responsible for this effect is not understood, but it has also been observed for the PTH/PTHrP receptor (31). That the A553T receptor is competent for normal coupling to Gs is further indicated by its specific constitutive activity for cAMP generation, which is comparable with the wild type. Interestingly, the P556L mutation of the hyt/hyt mouse also displays constitutive activity. However, in this case, unresponsiveness to TSH must result from a structural defect since the receptor has been shown to reach the plasma membrane of transfected COS cells (17). As stated above, overexpression in COS-7 cells may nevertheless not reliably reflect the behavior of the TSHr in the thyrocyte, and the significance of the relatively robust residual cAMP production in COS-7 cells is unclear. If it participates in the observed phenotype, our findings may indicate that the blood levels of TSH sustained by our patients are limiting with regard to thyroid growth, as they remained comparable with the EC_{50} of the mutant (0.3 mU/ml), which is much smaller than the pharmacological concentrations used on COS-7 cells (10 or 100 mU/ml) for maximal stimulation, and as human TSH is 5–10 times less potent on the human TSHr than the bovine TSH used in this study (32).

Although competent for coupling to Gs, the A553T mutant appears to have lost the capability to activate the phospholipase C–dependent regulatory pathway (Fig. 5 B). Dissociation of coupling to Gs and Gq has been observed previously in spontaneous TSHr mutants (8) and site-directed mutagenesis experiments (10). It agrees with a current model for G protein–coupled receptor activation, implying the existence of multiple discrete activated conformations.

The high TG in the patients at the time of diagnosis is surprising and not easily explained in the context of a very hypoplastic gland and congenital hypothyroidism (23). Apart from trivial causes of error and cross-reactivity with an unsuspected antigen in the TG assay, our patients show that significant expression of the TG gene can take place in vivo in spite of complete or near-complete TSH unresponsiveness. These findings may be confronted with the current knowledge of thyroid organogenesis and thyroid-specific gene expression. From the available evidence, it seems that development and migration of the embryonic thyroid in front of the trachea is independent of stimulation by TSH. Indeed, expression of thyroid-specific genes, including TG, TPO, and the TSHr, is subsequent to the observation of an in-place thyroid rudiment (33), which is believed to result from the interplay of at least three transcription factors, TTF1, TTF2, and Pax8 (34). However, it is tempting to suggest that normal TSHr function may be required to achieve a fully differentiated thyroid phenotype. The observation of cytological abnormalities in hyt/hyt mice, including overdistension of the endoplasmic reticulum, is compatible with such hypotheses (35). The disproportionately high plasma TG levels (considering the small size of the patients’ thyroids) could result from incomplete polarization of thyrocytes, abnormal routing of TG due to accumulation of misfolded mutant TSHr in the rough endoplasmic reticulum, or from intercellular leakage from dysplastic follicles. Testing of these hypotheses would require a biopsy of the thyroid from our patients, which we consider unethical.

The observed decrease and normalization of TG during \( T_4 \)
therapy is even more difficult to explain. It may simply correspond to the age-related decrease of serum TG observed in normal children from our population with borderline iodine intake (13). Alternatively, it is conceivable that irreversible full maturation of the thyroid could be normally dependent on autostimulation by thyroid hormones. Of note, TG levels did not increase when T$_{3}$ treatment was interrupted in the affected sister, in contrast with TSH that reached very high levels.

Unresponsiveness to TSH due to TSHr defects may represent a sizable subset of congenital hypothyroidism. It is probably rare even in the subgroup with familial occurrences of apparently absent thyroid tissue, as shown by the absence of any bly rare even in the subgroup with familial unresponsiveness in previous studies (21). This leaves room for other, still unidentified genetic defects. The unexpectedly high serum TG levels in our patients suggest that measuring unresponsiveness in previous studies (21). This leaves room for other, still unidentified genetic defects. The unexpectedly high serum TG levels in our patients suggest that measuring unresponsiveness in previous studies (21). This leaves room for other, still unidentified genetic defects.

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