

Proprotein-processing Endoprotease Furin Controls the Growth and Differentiation of Gastric Surface Mucous Cells

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Abstract

Gastric surface mucous cells originate from progenitor cells at the isthmus of the gastric gland, from where the cells migrate to the luminal surface. With migration they form secretory granules and express TGF α . We found that proprotein-processing endoprotease furin-positive cells were layered around the upper one fourth of the gastric glands of adult rats, whereas they were distributed along an outer epithelial layer in fetal rats. Because the furin-positive cell layer was localized from the upper cell proliferating zone to the less proliferating pit-cell region in the gastric gland unit, we examined the role of furin in the growth and differentiation of surface mucous cells by using the cell line, GSM06. This cell line is derived from the gastric surface mucous cells of transgenic mice harboring the temperature-sensitive simian virus 40 T antigen. At T antigen-active temperature (33°C), the cells grew to confluency, whereas at T antigen-inactive temperature (39°C), the cells ceased growing. At 33°C, the cells exhibited a high level of furin expression with a negligible level of periodic acid Schiff (PAS)-positive materials and a low level of TGF α . In contrast, at 39°C the cells produced a high level of PAS-positive materials, TGF α , and secretory granules, with a negligible level of furin expression. To further examine the role of furin, we established a GSM06 cell line introduced with either a sense or an antisense furin cDNA. The cells with sense furin expression produced fewer PAS-positive materials and a low level of TGF α even at 39°C, whereas the cells with antisense furin expression exhibited more PAS-positive materials and TGF α even at 33°C. When furin expression was suppressed by its antisense oligonucleotide, the cell growth was retarded with enhanced expression of the differentiated characteristics. Thus, we conclude that furin is instrumental in controlling the growth of the surface mucous cells. (*J. Clin. Invest.* 1997; 99:1842–1851.) Key words: gastric surface mucous cells • furin • periodic acid Schiff-positive materials • transforming growth factor- α • cell growth

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Introduction

Gastric surface mucous cells form a tubular structure by lining the wall of the gastric foveolar region (also called gastric pit) and the luminal surface region. The cells originate from the granule-free progenitor cells located at the isthmus of the gastric gland unit (1). From the isthmus they move upwards and develop to maturity through three stages: a pre-pit cell stage, a mature pit cell stage, and a final top pit cell stage with a turnover life of ~ 3 d in mouse gastric glands (2). The maturation of the cells is well characterized by the appearance of secretory granules, which are initially absent in the progenitor cells (thus called granule-free cells). Secretory granules begin to appear at a pre-pit cell stage and increase in number as the cells develop to mature pit cells. The mature cells contain a number of circular secretory granules full of mucin-like materials (2). In addition to the morphological characteristics, the surface mucous cells exhibit specific biochemical characteristics with periodic acid Schiff (PAS)¹ staining: synthesis and secretion of the two major components, macromolecular mucin and surface-active phospholipids (3, 4). With the morphological feature of secretory granules and the biochemical characteristic of PAS-positive staining, the cells are distinct from other mucosal cell subtypes such as parietal cells and chief cells (4, 5).

From a granule-free to a top pit cell stage, the differentiation of the surface mucous cells is regulated in an orchestrated manner by a variety of growth and differentiation factors, and cell-to-cell and/or cell-to-matrix interaction signals (6, 7). Such growth and differentiation factors may include epidermal growth factor (7, 8), TGF α (9, 10), heparin-binding EGF-like growth factor (11), hepatocyte growth factor (12), insulin-like growth factor-1 (10), and gastrin (6). Among these factors, TGF α is known to be produced in surface mucous cells, and increases when the cells move up to the upper pit wall of the gastric gland (13, 14). TGF α is also produced in greater amounts in parietal and chief cells (14–16). TGF α , EGF, and heparin-binding EGF-like growth factor share a common receptor that is highly expressed in parietal cells and moderately expressed in surface mucous cells, but not in a proliferating progenitor cell zone at the isthmus (11, 17). Recently, EGF/TGF α receptors were found to be localized to the basolateral region of luminal surface mucous cells (18). Because the surface mucous cells near the gastric lumen lose proliferative capacity, TGF α may act not only for cell growth but also for cell differentiation. Thus, TGF α as well as PAS-stained materials are expressed in mature gastric surface mucous cells.

1. Abbreviations used in this paper: BrdU, bromodeoxyuridine; PAS, periodic acid Schiff; PCNA, proliferating cell nuclear antigen; SV40, simian virus 40.

A primary culture of gastric surface epithelial cells has been well established by many investigators using the fundic mucosa from 1–2-wk-old rats (19), 25 gestational-d-old rabbits (20), adult dogs (10), and 16.5 gestational-d-old rats (21). These cultured cells exhibited a variety of characteristic features of surface mucous cells including TGF α responsiveness (10), production of PAS-positive materials (19, 20) and prostaglandins (20), and expression of cathepsin E (21). Although the primary culture method has many advantages over the cultured cell line method in gastric cell research, two major limitations of the primary cell culture method include an inability of cell passage and an inefficiency of gene transfer for expression studies. A cultured cell line accepts gene transfer more readily and is useful in cell differentiation studies, but highly differentiated functions tend to diminish with passage (22). To overcome this drawback of a cultured cell line, a novel method of establishing a culture cell line was devised using a temperature-sensitive simian virus 40 (SV40) large T antigen as a transforming gene (23, 24). A cultured cell line transformed with this temperature-sensitive T antigen is known to be useful in studying the differentiation process of cells (23–25). At the permissive temperature of 33°C, the cells are able to grow as usual because the T antigen is active at this temperature. When placed at the nonpermissive temperature of 39°C, which inactivates the T antigen, the cells cease growing and begin to exhibit their differentiated characteristics. The GSM06 cell line is derived from the gastric surface mucous cells of a transgenic mouse that was transformed with the temperature-sensitive SV40 T antigen DNA (25). Morphologically, GSM06 cells are rounded or polyhedral as is typical of primary cultured gastric surface mucous cells (10, 19–21). Electron microscopy revealed that the cells contained a small number of secretory granules (26). The PAS-positive staining of the GSM06 cells appeared to be due to glycoproteins and glycolipids, but not to mucin molecules (27). In a confluent state at 33°C, or even in a nonconfluent state at 39°C, the cells begin to produce PAS-positive materials that form a mucous sheet as seen in the gastric surface mucous cells (25, 27).

Many growth-related proteins are generated as a precursor, and their activation may be regulated by cleavage enzymes. As we have demonstrated using the pancreatic β cell line MIN6 (22), the proprotein processing enzyme, furin may be one of the enzymes involved in the regulation of cell growth and differentiation. Furin belongs to a member of the yeast Kex2-like family and converts many precursors to their bioactive forms at the *trans*-Golgi network (28). Cleavage by furin is specific for a unique amino acid sequence Arg⁻⁴-X-X-Arg⁻¹. This motif is found in many growth-related peptides or proteins (29). In the gastric mucosa there are a number of such candidates: a procadherin family of adhesion molecules (30); matrix metalloproteinases such as meprin A, stromelysin-3, and membrane-type matrix metalloproteinases (31–33); several growth factor precursors including PDGF and TGF- β (34, 35), and some growth factor proreceptors such as the insulin receptor, IGF-1 receptor, and hepatocyte growth factor receptor (oncoprotein *MET*) (36, 37). These furin-cleavable precursors may function for the growth and differentiation of gastric mucosal cells.

We investigated the role of furin in the growth and differentiation of surface mucous cells using the cell line GSM06. By changing the temperature and cell density of the culture, we measured the growth and differentiation of the surface mu-

cous cells with three markers: PAS staining-positive materials, TGF α , and furin expression. The present study demonstrates that the expression of furin in the GSM06 cells produces an immature state of the surface mucous cells, whereas the suppression of furin expression induces their differentiated characteristics, even in a nonconfluent state at 33°C.

Methods

Cell culture. As a culture model of the surface mucous cells, we used a GSM06 cell line. This cell line was established from the gastric surface mucous cells of transgenic mice harboring temperature-sensitive simian virus 40 large T antigen gene (25). The cells were cultured on a collagen type I-coated plastic plate (Iwaki, Tokyo, Japan) in Dulbecco's Modified Eagle's medium/Ham's F-12 medium (Sigma Chemical Co., St. Louis, MO) supplemented with 10% FBS (Gibco Laboratories, Grand Island, NY), 1% ITES (100% ITES; insulin 2 mg/liter, transferrin 2 mg/liter, ethanolamine 0.122 mg/liter, sodium selenite 9.14 μ g/liter), and 10 ng/ml EGF in a humidified 5% CO₂ atmosphere at 33°C unless otherwise indicated. The cells were grown at this temperature until a confluent monolayer was formed. Culture at 39°C started when the cells reached the cell density described below. The cells were cultured in high or low cell density. High density culture was defined as the condition in which the cells were confluent, low cell density was defined as 10% confluent.

Histologic studies. For examining the proliferation of the fetal or adult rat gastric mucosa, we used two methods, the thymidine analogue, bromodeoxyuridine (BrdU) incorporation method (38–40), and the staining of proliferating cell nuclear antigen (PCNA) (40). BrdU (80 mg/kg body wt; Sigma Chemical Co.) was injected intraperitoneally into pregnant Wistar rats at gestational day 18, or Wistar rats at neonatal day 35 (Japan SLC, Hamamatsu, Japan) 6 h before killing. Stomach tissues were removed from rats at gestational day 18 or at postnatal day 35. After mincing, the tissue was fixed in 4% paraformaldehyde in 0.1 M phosphate buffer, pH 7.4. Small pieces of the sample were incubated consecutively with 10% and then 20% saccharose in 0.1 M phosphate buffer, pH 7.4. After saccharose replacement, the samples were frozen, and then sectioned by a microtome. The sections were placed on a glass slide. The mouse anti-BrdU monoclonal antibody (BioMeda, Foster City, CA) recognizes BrdU molecules in denatured single strand DNA; thus, before staining, the sections were partially denatured by the incubation in 95% formaldehyde in 0.15 M trisodium citrate at 70°C for 45 min. Then the sections were processed using an LSAB2 staining kit (DAKO Japan, Kyoto, Japan) according to the manufacturer's instruction.

Immunostaining of PCNA was performed on cryostat microtome-sectioned tissues using a mouse monoclonal anti-PCNA antibody (PC10; DAKO SA, Glostrup, Denmark) and an LSAB2 staining kit (DAKO Japan) (40).

Immunostaining of furin was performed by a previously described method (39). Stomach tissue sections were prepared by the same procedure as used for the BrdU and PCNA stainings. GSM06 cells were cultured on a chamber slide (Nunc Inc., Naperville, IL). After fixation in 4% paraformaldehyde in 0.1 M phosphate buffer, pH 7.4, tissue sections or GSM06 cells were probed with rabbit antifurin serum (ST-73). They were then incubated with indocarbocyanine (Cy3)-conjugated affinity-purified donkey anti-rabbit IgG (Jackson ImmunoResearch Labs., Inc., West Grove, PA). The specimens were examined using a microscope with an incident illuminator (BX50; Olympus Corp., Lake Success, NY).

For PAS staining, stomach tissue sections or GSM06 cells on a chamber slide were fixed in 10% formaldehyde for 3 h at 4°C and stained with PAS using the standard method (41). Briefly, samples were oxidized for 15 min with 0.5% periodic acid. After washing, they were immersed for 15 min in Schiff's reagent, and then for 4 min two times in 1% NaHSO₃ and 0.1% HCl. After washing the samples with tap water, they were stained with hematoxylin.

For examination via electron microscopy (EM), GSM06 cells were scraped from the plastic plate and packed in 4% agarose. The cells were then fixed in OsO_4 and processed for EM.

Northern blot analysis. For isolation of the total RNA from GSM06 cells, the cells were cultured in a 10- or 15-cm plastic plate, and rinsed with ice-cold Ca^{2+} - Mg^{2+} -free PBS(-). Total RNA isolation was performed as previously described (22). The total RNA was electrophoresed on a 1.0% agarose gel and transferred to a nylon membrane (Amersham Japan, Tokyo, Japan). Hybridization was performed with a priming procedure as previously described (22). The membrane was exposed to an x-ray film (Eastman Kodak Co., Rochester, NY) with an intensifying screen at -80°C .

Immunoblotting for furin and TGF α . For immunodetection of furin and TGF α , GSM06 cells were cultured as described above. The cells were harvested with ice-cold 10% trichloroacetic acid and left for 1 h on ice. Cell precipitates were collected and centrifuged at 10,000 g for 10 min at 4°C . After removing supernatant, cell pellets were lysed with 80 μl of Urea TX solution (9 M urea, 2% Triton X-100, 5% 2-mercaptoethanol), and then sonicated shortly. To the sonicated samples, 20 μl of LDS solution (10% lithium dodecylsulfate, 0.1% bromophenol blue) was added for cell lysate preparation. Cell lysates were sonicated again, and then adjusted to pH 7.4 with 2 M Tris-HCl. Cell lysates were electrophoresed on a 20% polyacrylamide gel under a reducing condition. Separated proteins were blotted onto a nitrocellulose membrane and probed with rabbit antifurin antiserum (ST-73) or with anti-TGF α monoclonal antibody (Santa Cruz Biotechnology, Santa Cruz, CA). Antibody-reacted bands were detected using an ECL detection system (Amersham International, Little Chalfont, UK).

Measurement of cell growth. We used a cell counting kit based on the action of WST-1 [2-(4-iodophenyl)-3-(4-nitrophenyl)-5-(2,4-disulphophenyl)-2H-tetrazolium, monosodium salt] (Dojindo Lab, Kumamoto, Japan), and the cell number was evaluated with the absorbance of optical density at 450 nm (42). Briefly, cells were cultured at 3×10^3 cells/well on a collagen I-coated 96-well plate. After removing the culture medium and washing with PBS, the cells were further incubated for 3 h in the assay solution with WST-1. Then OD 450 nm was measured with a spectrophotometer. For each experiment, an average of eight sample OD values were calculated and at least three experiments were performed for each assay point. The value obtained from the cell culture under a variety of ZnCl_2 concentrations was expressed as a percentage of the control value without ZnCl_2 . Values with this method were linearly correlated with the corresponding cell numbers by counting with a hemocytometer ($r^2 = 0.995$).

Establishment of sense and antisense furin cDNA-expressing GSM06 cells. To investigate the role of furin in the growth of GSM06 cells, we constructed the two furin expression vectors using pMEP4 (Invitrogen Corp., San Diego, CA), which carries a heavy metal-inducible metallothionein promoter (22). One vector contained a sense strand of the full length furin cDNA, the other contained an antisense strand. The antisense furin cDNA was made by inserting the cDNA covering the first 364 amino acids of the mouse furin precursor sequence into the pMEP4 in a reverse direction (43). GSM06 cells were transformed with these vectors or with an empty pMEP4 by the lipofection method using DOTAP (Boehringer Mannheim, Indianapolis, IN). Transformed cells were selected to a single colony by maintaining the culture in a medium containing hygromycin B (0.1 mg/ml, Sigma Chemical Co.). To evaluate the effect of furin expression on the growth of GSM06 cells, ZnCl_2 was added to the culture medium to activate the metallothionein promoter that controls the expression of sense or antisense furin cDNA. ZnCl_2 alone, up to 100 μM , did not cause any change in cell viability, cell growth, PAS staining, or expression of TGF α , as long as the cells were cultured in the medium containing 10% FBS. However, when ZnCl_2 was added to the culture medium without FBS, the cells began to detach from the bottom of the plastic dish.

Measurement of cell growth with antisense furin oligonucleotide-treated conditioned medium. To determine the effect of furin expres-

sion on the growth of GSM cells, we used an antisense oligonucleotide to suppress furin in addition to the above described experiment. Because we did not need to use ZnCl_2 in this experiment, we were able to culture the cells in a serum-free medium. Antisense oligonucleotide (5'-CACCCATAGCAACCAGGATCTCAGCTCCAT-3') and sense oligonucleotide (5'-ATGGAGCTGAGATCCTGGTTGCTATGGGTG-3') were designed to cover the start codon for furin cDNA (43). The oligonucleotides were synthesized using a nucleotide phosphorothioate method to prevent the degradation from nucleases (44). The antisense oligonucleotide was used to suppress furin expression, while the sense oligonucleotide was used as a control. For culturing GSM06 cells, we used serum-free medium PM-1000 (Eiken Chemical Co., Tokyo, Japan) with 0.1% BSA (Sigma Chemical Co.). The cells were cultured in the presence or absence of various concentrations of the sense or antisense oligonucleotide for 48 h before the cells were counted.

We investigated the effect of the conditioned medium prepared in the presence or absence of the sense or antisense oligonucleotide on the growth of the intact GSM06 cells. Briefly, the cells were cultured in 10-cm collagen-coated plates in a serum-free P-1000 medium with 0.6 μM sense or antisense oligonucleotide for 24 h. The medium was collected from each dish and concentrated to 1/100 vol using the Centricon 3 microconcentrator (Amicon Inc., Beverly, MA). To culture the intact GSM06 cells, concentrated conditioned medium was added to a fresh PM-1000 medium consisting of 1% of the total volume. After culturing the cells for 24 h, the cell number was measured with the WST-1 method. Each experiment was performed at least three times and the values were expressed as mean \pm SEM.

Data analysis. Comparisons between multiple groups were made using repeated-measures analysis of variance; when significant differences were found, further comparisons were made using the paired *t* test. Differences were considered significant at $P < 0.05$.

Results

Characteristics of gastric mucosal cells of fetal and adult rats. The gastric epithelium of fetal rats at gestational days 16 and 17 is covered mostly by a simple layer, and partly by a stratified layer of cuboidal cells (21, 45, 46). Towards birth, gastric epithelium becomes a single layer of columnar cells and begins to invaginate and form a pit and glandular crypt structure. After birth, gastric epithelium develops into a complex gastric gland composed of several functionally distinct cell subtypes, until the weaning period (47). We compared the characteristics of gastric epithelial cells between gestational day 18 and postnatal day 35 using three markers: cell proliferating capacity by BrdU incorporation or by PCNA staining, formation of PAS-positive materials, and expression of furin. The epithelium at gestational day 18 was made up of a stratified layer of round-to-oblong-shaped cells (Fig. 1 A). Many nuclei in both mucosal and submucosal cells were stained dark brown by BrdU incorporation, while nonstained nuclei remained blue. In epithelium, the BrdU-positive cells were observed mostly along the basal epithelial layer, but a few in the upper epithelium. These findings indicate that there is extensive cell division at this stage. With PAS staining (Fig. 1 B), the mucosal and submucosal cells, but not muscular cells, were lightly positive, indicating that the capacity to synthesize glycoconjugates is slightly developed at this stage. Furin immunostaining was strongly positive along an outer epithelial cell layer where a few BrdU-positive cells were scattered, but negative along the basal epithelial cell layer where BrdU-positive cells were enriched (Fig. 1 C). There are some furin-positive cells scattering in a submucosal layer. Although the epithelial, subepithelial, and muscu-

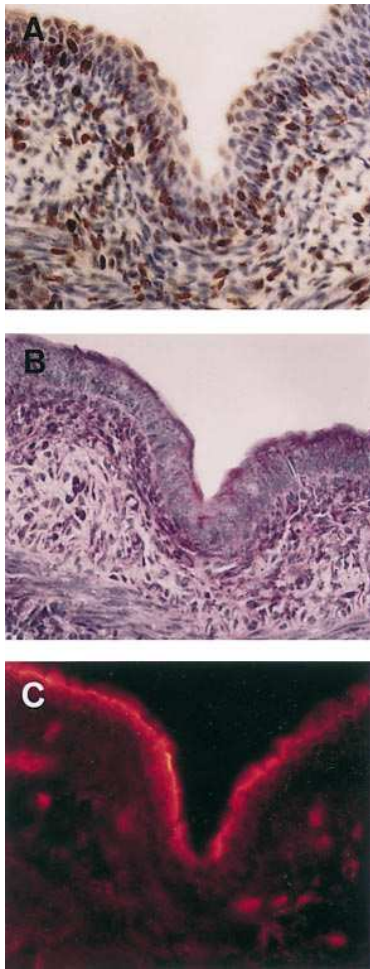


Figure 1. Characterization of rat fetal gastric mucosa. Gastric mucosa was obtained from a rat fetus at gestational day 18. (A) Immunostaining of BrdU. BrdU-incorporated nuclei were stained dark brown due to the horseradish peroxidase enzyme reaction. Sections were counterstained with hematoxylin so that nuclei without BrdU incorporation were visualized blue by hematoxylin. (B) PAS staining. PAS was faintly stained in the mucosal layer and upper submucosal layer, but not in the lower submucosal layer and smooth muscular layer. (C) Immunostaining of furin. Immunostaining was visible along the outer epithelial layer. Several furin-positive cells were scattered in the submucosal layer.

lar layers were formed, the cells in the epithelial layer looked homogeneous, but evidently were not differentiated as they lacked the distinct subtype features present in the adult gastric epithelium.

At postneonatal day 35, the gastric gland unit has already matured morphologically and functionally, and forms a pit and crypt structure (47). Cell division takes place at the isthmus between the pit and glandular crypt of the gastric unit (1). The cells with BrdU-incorporated nuclei were localized to an isthmus between the pit and neck cell regions, which was approximately a third level of the gastric gland unit (Fig. 2 A), as was similarly shown by the incorporation of [3 H]thymidine (1). For BrdU staining, we treated formaldehyde to denature double-stranded DNA to a single one so that gastric glands look thinner than the other stainings (Fig. 2 A vs. 2, B–D). PCNA is known as a DNA-tracking protein required for the rapid and processive replication of DNA by the polymerases δ and ϵ (48). PCNA is highly expressed at the S phase of the cell cycle. PCNA-positive cells were mostly localized to a third level of the gastric glands, as were BrdU-positive cells, but the PCNA-positive cell layer was distributed wider than the BrdU-positive cell layer (Fig. 2, A and B). PAS staining revealed a strong dark-red pattern at the top pit cells and a bright-red pattern at the cells along the middle portion of the gastric unit (Fig. 2 C). The stained cells may represent surface mucous cells at the top, and mucous neck cells at the middle portion (1). This PAS

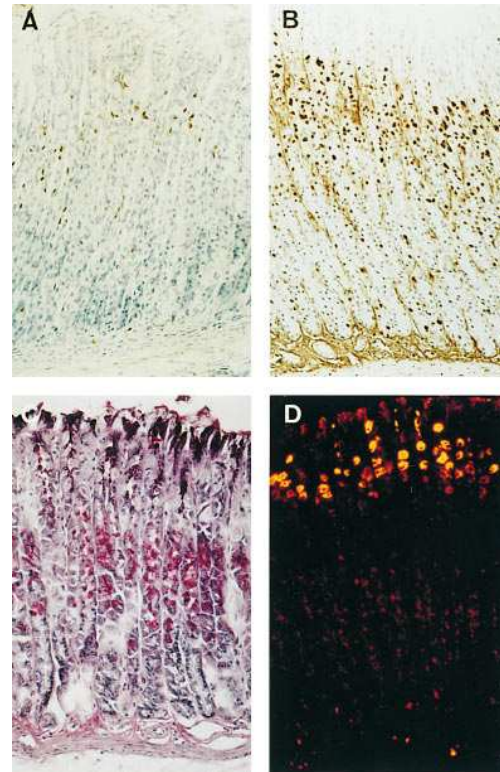


Figure 2. Characterization of adult rat gastric mucosa. Gastric mucosa was obtained from a young adult rat at postnatal day 35. The same staining procedures as in Fig. 1 were performed. (A) Immunostaining of BrdU. BrdU-incorporated cells were mostly distributed at a third level of the gastric unit. A few positive cells were scattered at the neck region of the gastric unit. (B) Immunostaining of PCNA. The cells with PCNA-stained nuclei were distributed as a wider layer than those with BrdU-positive nuclei. (C) PAS staining. PAS was strongly stained at the top pit cells along the surface epithelial lumen of the gastric glands and stained bright red along the middle portion of the gastric unit. (D) Immunostaining of furin. Furin-positive cells were layered at the upper one fourth of the gastric glands. Small furin-positive cells were scattered at the subglandular region over the muscularis mucosae.

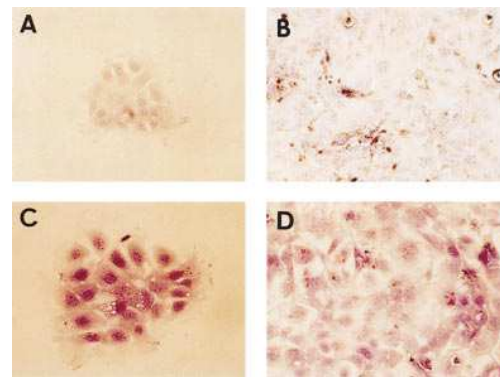


Figure 3. PAS staining of GSM06 cells cultured under various conditions. (A) 33°C, low cell density. (B) 33°C, high cell density. (C) 39°C, low cell density. (D) 39°C, high cell density. PAS staining was positive at 39°C in both low and high cell densities. The staining covered whole cytoplasm and nuclei, but was not specifically localized in the secretory granules.

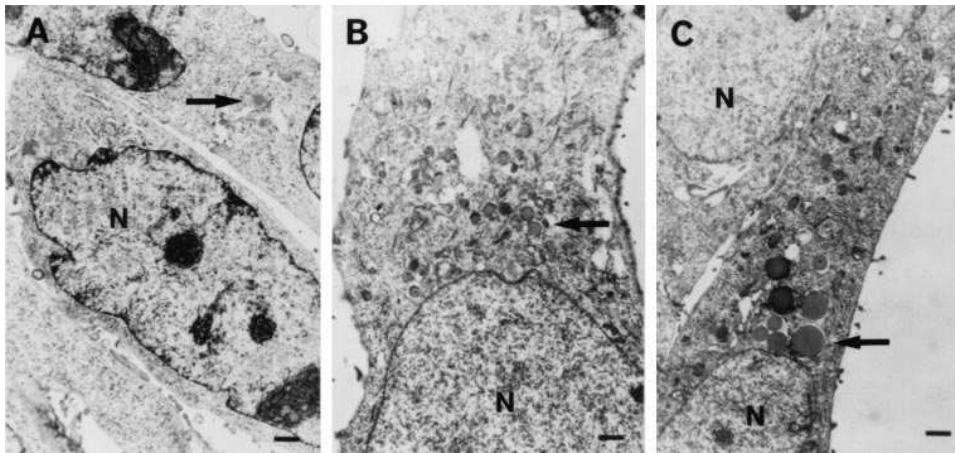


Figure 4. Electron microscopy of GSM06 cells. GSM06 cells were observed by electron microscopy under the two distinct conditions. (A) 33°C. (B and C) 39°C. N, nucleus. Bars indicate 1 μ m. At 33°C only a few granules were visible (A, arrow). In contrast, at 39°C, 500–600-nm granules appeared in many cells (B, arrow). Some cells contained larger granules up to 1,250 nm (C, arrow). Most granules contained homogeneous light materials in the center with dark materials around the outer circle.

staining pattern did not change even after diastase treatment. Furin was stained as a distinct zonal layer around the upper one fourth of the gastric glands, but unlike PAS staining, the top pit cell region was not stained (Fig. 2D). The furin-positive cell layer was located from the higher cell-proliferative zone to less proliferative pit cell region. Some cells appeared to be located in both the furin-positive layer and the cell proliferating zone. Thus, furin seemed to be expressed in the late stage or immediately poststage of cell division in embryonal and adult rat gastric mucosa.

PAS staining of GSM06 cells. A GSM06 cell line was established by the transformation with temperature-sensitive SV40 T antigen (25). The cells grow to confluency at 33°C, whereas the cells stop growing at 39°C. We characterized the cells with PAS staining by placing them at 33 or 39°C; cells were stained either at high density (almost confluent) or low density (1/10 of a confluent density). At 33°C and low cell density, the cells were not positive for PAS staining (Fig. 3A). When the cells became confluent at 33°C, PAS-positive materials were not visible yet (Fig. 3B). However, when cells were kept confluent over 3 d, PAS-positive materials appeared as described previously (25, 26). In contrast, when the cells were transferred from 33 to 39°C for 48 h, the cells became clearly positive for PAS staining in low or high cell density (Fig. 3, C and D). Thus, we noted that the higher temperature induced the expression of PAS-positive materials in the GSM06 cells both at low and high cell density.

Electron microscopy of GSM06 cells. Development of progenitor cells to mature pit cells via a pre-pit cell stage is charac-

terized by the appearance and increase of mucous granules (2). These granules are positive for PAS staining (19, 20). We examined the presence of mucous granules in the GSM06 cells cultured at 33 and 39°C. The cells at 33°C contained a few secretory granules, indicating that the cells are probably in an early pre-pit cell stage, but not in a granule-free progenitor cell stage, as Karam and Leblond described previously (Fig. 4A) (1, 2). When the cells were placed at 39°C, a fair number of secretory granules appeared in the cytoplasm (Fig. 4, B and C). Granules were various in diameter up to \sim 1,250 nm. At 39°C most cells contained 500–600-nm granules (Fig. 4B, arrow). Some cells contained much larger granules, up to 1,250 nm (Fig. 4C, arrow). Some granules contained dark materials at their outer circle with light materials in the center (Fig. 4B, arrow), others displayed homogeneous light materials (Fig. 4C, arrow). However, even at 39°C no cells displayed as many granules as seen in the mature pit cells (2). Thus, the number of secretory granules suggests that the cells appeared to develop from an early pre-pit cell stage to a more advanced pre-pit stage and exhibited embryonic features characteristic of rat gastric mucosa at gestational days 18–21 (45, 46).

Expression of furin. We examined the expression of furin in the GSM06 cells by Northern and Western blotting, and immunocytochemistry. In the Northern blot of furin, the message was at high levels at 33°C in both low and high cell densities, and the message at low density was a little stronger than that at high density (Fig. 5A, lanes 1 and 2). In contrast, the message was at low levels at 39°C in both low and high densities (Fig. 5A, lanes 3 and 4). In the Western blot of furin, the band ap-

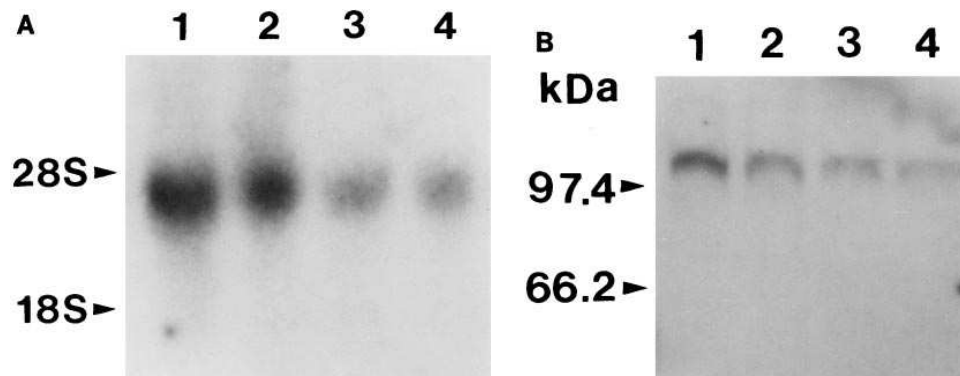


Figure 5. Expression of furin. (A) Northern blot. Lane 1, 33°C, low cell density. Lane 2, 33°C, high cell density. Lane 3, 39°C, low cell density. Lane 4, 39°C, high cell density. (B) Western blot. Lane 1, 33°C, low cell density. Lane 2, 33°C, high cell density. Lane 3, 39°C, low cell density. Lane 4, 39°C, high cell density. By the two different blotting methods, the blot at 33°C in low cell density was the strongest, while the blot at 39°C in high cell density was the weakest.

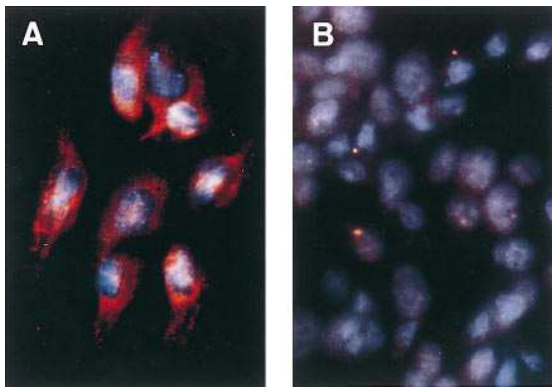


Figure 6. Immunostaining of furin in GSM06 cells. (A) 33°C, low cell density. (B) 39°C, high cell density. Strong immunostaining of furin was observed in the cells at 33°C in low cell density. Note that the staining was markedly decreased in the cells cultured at 39°C in high cell density. Nuclei were stained as blue with 4',6'-diamidine-2-phenylindole-dihydrochloride.

peared at a little over 97.4 kD. Furin was strongest in the cells cultured at 33°C in low density, followed by conditions at 33°C in high density, and lastly at 39°C in both low and high densities (Fig. 5 B). Thus, the tendency of furin expression was almost similar between the mRNA and protein. The immunocytochemical study showed that the cells at 33°C in low density displayed a strong staining of furin with a localization pattern of the Golgi stacks as reported before (22, 28) (Fig. 6 A). On the other hand, negligible staining of furin was observed in the cells at 39°C in high cell density (Fig. 6 B). Thus, furin expression was significantly different between these distinct conditions.

Immunoblot of TGF α . We then examined the immunoblot of TGF α under the same conditions used for the furin immunoblotting. The immunoblot of TGF α appeared at a little under 6.5 kD, which may indicate the active form of 50 amino

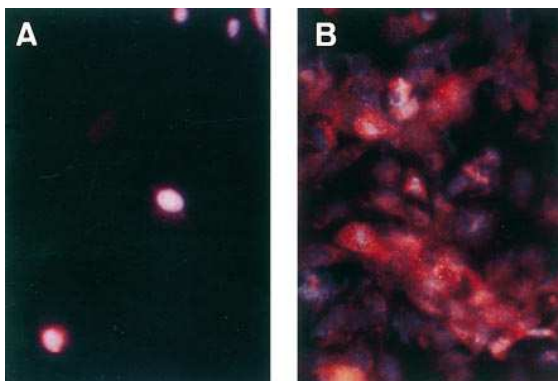


Figure 8. Immunostaining of furin in the GSM06 cells expressing sense or antisense furin cDNA. Sense or antisense furin cDNA was expressed by placing the cells for 48 h in culture with or without 100 μ M ZnCl₂. The cells were stained after culture with ZnCl₂. (A) GSM06 cells expressing an antisense furin DNA in low cell density at 33°C. Note that the furin staining is negligible compared with that in Fig. 6 A. (B) GSM06 cells expressing a sense furin DNA in high cell density at 39°C. Note that the furin staining is clearly visible in almost half the population of cells compared with that in Fig. 6 B.

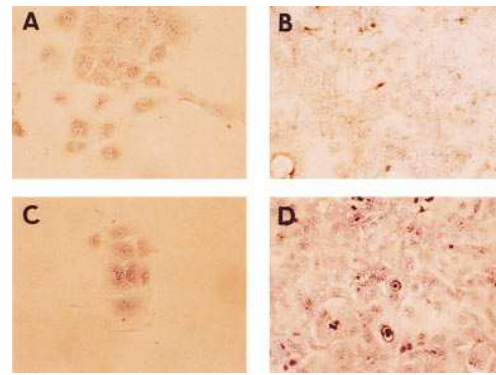


Figure 9. PAS staining of GSM06 cells expressing sense furin cDNA. Furin-expressing GSM06 cells were cultured for PAS staining under the same conditions as in Fig. 3. (A) 33°C, low cell density. (B) 33°C, high cell density. (C) 39°C, low cell density. (D) 39°C, high cell density. PAS staining was only faint even at 39°C in both low and high cell densities. Note the difference between Figs. 9 and 3.

acid TGF α . TGF α is known to be formed as the 20–22 kD precursor, and then processed to a 6-kD form by way of several intermediate precursors (49). Although we observed faint bands at precursor-size positions by longer x-ray film exposure, the major bands appeared close to a 6-kD position. As to immunostaining, the strongest signals were present in the high density cells at 39°C, and then the high density cells at 33°C, followed closely by the low density cells at 39°C, and lastly the low density cells at 33°C (Fig. 7). We observed one more extra band under the major one at 39°C in high density (Fig. 7, lane 4). This extra band was observed also in the previous work (49), and may reveal a more processed form of the major band. TGF α expression in these cell conditions appeared to be ranked in an opposite order from the furin expression and PAS-positive material formation.

Effect of sense or antisense furin cDNA on the expression of furin, PAS-positive materials, and TGF α . We used GSM06 cells transfected with a sense or antisense furin cDNA expression vector to examine the appearance of PAS-positive materi-

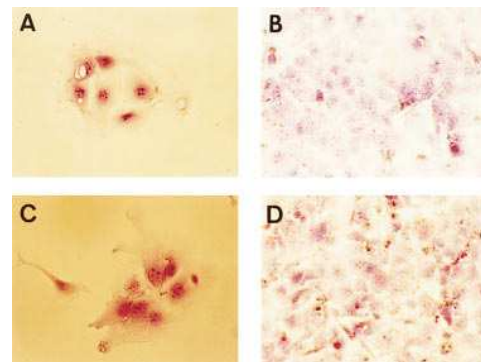
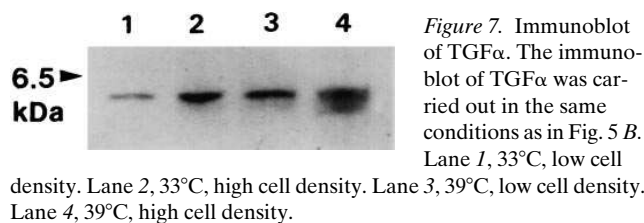
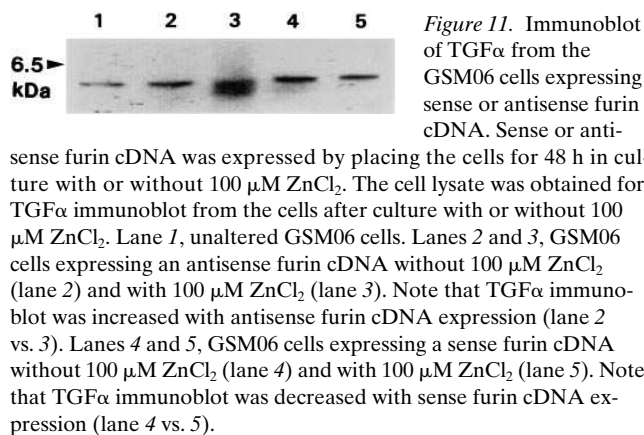


Figure 10. PAS staining of GSM06 cells expressing antisense furin cDNA. Antisense furin-expressing GSM06 cells were cultured for PAS staining under the same conditions as in Fig. 3. (A) 33°C, low cell density. (B) 33°C, high cell density. (C) 39°C, low cell density. (D) 39°C, high cell density. PAS staining was positive even at 33°C in both low and high cell densities. Note the difference between Figs. 10 and 3.



als and the expression of TGF α . The sense or antisense furin cDNA was placed under a mouse metallothionein promoter that can be induced by the addition of ZnCl₂ in the culture medium. With this regulatable expression system, the expression of antisense furin cDNA may suppress endogenous furin, whereas the expression of sense furin cDNA may result in the additive increase of furin. The Zn²⁺-inducible expression of furin was confirmed by immunocytochemistry. As shown in Fig. 8 B, furin was immunostained when the sense furin cDNA was expressed in the cells with 100 μ M ZnCl₂, even at high cell density at 39°C, where furin was not expressed in unaltered GSM06 cells (Fig. 6 B). Overexpressed furin was colocalized with the TGN marker protein TGN38 (data not shown). Almost half of the cells became furin positive in their condition. In contrast, in the cells with the induction of the antisense furin cDNA, furin was only faintly immunostained even at low cell density at 33°C (Fig. 8 A), where furin was strongly stained in unaltered cells (Fig. 6 A). Thus, we could control furin expression by the induction of sense or antisense furin cDNA.

With the expression of sense furin RNA, the cells exhibited virtually no PAS-positive staining at 33°C either at low or high density (Fig. 9, A and B); unaltered GSM06 cells also did not show PAS-positive staining (Fig. 3, A and B). At 39°C, PAS-positive materials appeared only faintly in low and high density cells (Fig. 9, C and D), unlike the contrasting positive staining in the corresponding culture conditions shown in Fig. 3, C and D. On the contrary, expression of antisense furin DNA induced the appearance of PAS staining in both the low and high density cells at 33°C (Fig. 10, A and B), whereas unaltered cells did not stain PAS-positive (Fig. 3, A and B). Furthermore, the expression of PAS-positive materials appeared to be augmented at 39°C in both low and high cell densities (Fig. 10, C and D) compared with the corresponding conditions of the cells shown in Fig. 3, C and D. Thus, judging from the formation of PAS-positive materials, furin expression



maintained the cells in a less differentiated state, whereas furin suppression facilitated the differentiation of the cells.

The immunoblot of TGF α showed that TGF α expression increased in the cells in which antisense furin cDNA was induced with ZnCl₂ (Fig. 11, lane 3). On the contrary, with the induction of the sense furin cDNA, the TGF α expression was slightly diminished (Fig. 11, lane 5). Thus, the appearance of the PAS-positive materials, as well as the expression of TGF α , were negatively correlated with furin expression.

Effect of furin expression on the growth of GSM06 cells. Since furin expression is instrumental in the differentiation process of GSM cells, we examined whether furin expression affects the growth of GSM06 cells. We used two methods to regulate furin expression: the first was the GSM cell line expressing the sense or antisense furin cDNA used in the above experiments, the other was a nuclease-resistant sense or antisense furin oligonucleotide. In the first regulation system, cell growth was compared between the cells expressing sense or antisense furin cDNA 48 h after the addition of ZnCl₂. We initially confirmed that both unaltered GSM06 cells and empty vector pMEP4-introduced GSM06 cells did not exhibit a significant change in their growth rate with or without ZnCl₂ up to a 100- μ M concentration. The cells proliferated with a similar growth rate as unaltered GSM06 cells (Fig. 12), although we expected an enhanced growth of the cells due to increased furin expression. In this experiment, we had to plate cells less confluent to observe the cell growth, which resulted in the extensive furin expression. This additional expression may have induced no further effect on cell growth. On the other hand, when the antisense cDNA was induced, the growth was significantly retarded at 80 to 100 μ M ZnCl₂ (Fig. 12).

In the second regulation system using oligonucleotides, we used a serum-free medium for cell culture so that we could exclude the serum effect on cell growth. In a Zn²⁺-inducible expression system, we had to add serum to the cell culture to alleviate Zn²⁺ cell toxicity. The growth was evaluated 48 h after the addition of sense or antisense oligonucleotide. When up to

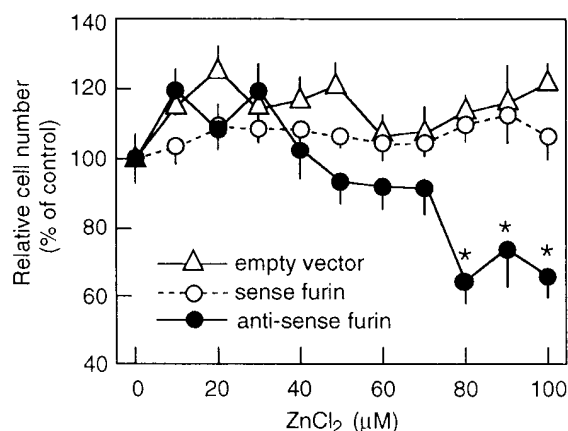


Figure 12. Growth of sense or antisense furin cDNA-expressing GSM06 cells. Cell growth was measured by using the dye WST-1 as described in Methods. The cells were cultured for 48 h in the presence of 100 mM ZnCl₂. Open circle, the cells expressing a sense furin cDNA. Closed circle, the cells expressing an antisense furin cDNA. Open triangle, the cells expressing an empty vector pMEP4 alone. The growth was retarded only when the antisense furin cDNA was expressed. **P* < 0.01.

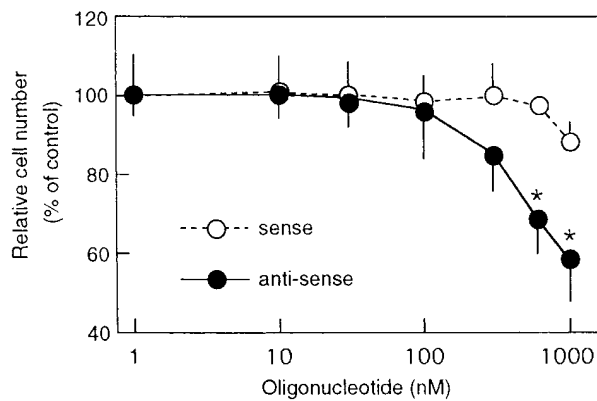


Figure 13. Effect of a sense or antisense oligonucleotide encoding a part of the furin-coding sequence on the growth of GSM06 cells. The GSM06 cells were cultured in a serum-free medium PM-1000 with a sense or antisense furin oligonucleotide (1, 10, 30, 100, 300, 600, 1,000 nM). Cell growth was measured by using the dye WST-1. Open circle, the culture in the presence of a sense furin oligonucleotide. Closed circle, the culture in the presence of an antisense furin oligonucleotide. The growth was more retarded in the culture with an antisense oligonucleotide than with a sense oligonucleotide (300–1,000 nM). * $P < 0.01$.

600 nM of the sense oligonucleotide was added to the culture, the growth was not retarded, but growth did decrease with 1,000 nM (Fig. 13). In contrast, when the antisense oligonucleotide was used, growth slowed slightly at 300 nM, and then was significantly retarded with increased oligonucleotide concentrations (Fig. 13). Thus, with the two regulation systems, we confirmed that furin suppression decreased the growth rate of the cells.

Growth-promoting effect of the culture media conditioned with the sense and antisense furin oligonucleotides. Because the growing GSM06 cells expressed a high level of furin and because furin suppression resulted in the retardation of the cell growth, we hypothesized that the cells might secrete bioactive growth factors that are converted from their precursors by furin cleavage. To examine this possibility, we prepared the conditioned media by culturing the cells with the sense or antisense furin oligonucleotide, expecting that the cells release such growth-promoting factors in the medium. We used serum-free PM-1000 medium because serum itself contains a high level of growth-promoting factors and may obscure the effect of newly generated growth factors. We prepared the three conditioned media: C, S, and AS. C was prepared without addition of oligonucleotides, while the S and AS media were prepared by adding the sense or antisense furin oligonucleotide to the GSM06 cell culture, respectively, at a final concentration of 600 nM and incubated for 24 h. Each medium was concentrated with an Amicon membrane filter concentration system. With this concentration process, most oligonucleotides passed through a membrane with a continuous flow of PM-1000 serum-free medium because we could not detect any leftover oligonucleotide by an ethidium bromide detection method. When conditioned medium C or S was used for the GSM06 culture, the cells grew more than those in the cell culture using nonconditioned PM-1000 (Fig. 14, NC). As predicted from Fig. 13, the conditioned medium prepared with the sense oligonucleotide had a similar effect on the cell growth to

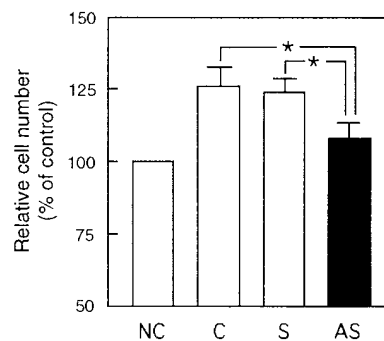


Figure 14. Effect of conditioned media from the culture with the sense or antisense furin oligonucleotide on the growth of GSM06 cells. Conditioned media were prepared by culturing the GSM06 cells in the absence of both oligonucleotides (C), in the presence of 600 nM

sense oligonucleotide (S), or in the presence of antisense oligonucleotide (AS). Unaltered GSM06 cells were cultured with nonconditioned PM-1000 medium NC, the conditioned media C, S, or AS. The cell number was assessed with the dye WST-1. NC, culture of the nonconditioned assay medium. C, culture with the conditioned medium without oligonucleotides. S, culture with the conditioned medium in the presence of the sense furin oligonucleotide. AS, culture with the conditioned medium in the presence of the antisense furin oligonucleotide. The growth of GSM06 cells was significantly retarded by the conditioned medium prepared with the antisense oligonucleotide compared with the conditioned medium with or without the sense oligonucleotide. * $P < 0.01$.

that prepared without the oligonucleotide. When the AS-conditioned medium was used, however, cell growth was retarded (Fig. 14). This result suggests that suppression of furin may have reduced the conversion of growth factor precursors to their bioactive forms.

Discussion

The present study demonstrated that at the permissive temperature (33°C) of a temperature-sensitive SV40 large T antigen, the gastric surface mucous cell line GSM06 cells had undifferentiated features with high expression of the proprotein processing endoprotease furin, negligible production of PAS-positive materials, poor formation of secretory granules, and a low expression of TGF α . At the nonpermissive temperature (39°C), on the other hand, they exhibited differentiated features with furin suppression, production of PAS-positive materials, appearance of many secretory granules, and increased expression of TGF α . When furin was induced in the GSM06 cells, the cells produced no PAS-positive materials and less TGF α , even in a differentiated condition (confluent state at 39°C). Furin-suppressed cells, however, produced PAS-positive materials and a higher amount of TGF α , even in a less-differentiated condition (nonconfluent state at 33°C). Furthermore, furin suppression retarded cell growth, suggesting that furin may be instrumental in the growth of gastric surface mucous cells.

At gestational day 18, fetal rat furin is localized along an outer layer of the epithelial cells that cover the gastric mucosa (Fig. 1 C). At this stage, gastric epithelial cells are remarkably active in cell proliferation (Fig. 1 A), but are poor in the production of PAS-positive materials (Fig. 1 B). Surface mucous cells contain a trace of secretory granules at gestational days 18 and 19, but at gestational days 20 and 21, many granules appeared in the cytoplasm (45, 46). Coincidentally, TGF α appeared faintly after the gestational day 20 in rat surface epithe-

lial cells (49). In the gastric mucosa of adult rats (postnatal day 35), furin-positive cells were layered at the upper one fourth of the gastric glands (Fig. 2 D). The cell proliferating zone identified by BrdU incorporation and PCNA staining was located a little lower than a furin-positive cell layer, but some cells appeared to stay in the proliferative zone with furin-positive staining (Fig. 2, A, B, and D). PAS-positive cells were distributed at both upper and lower sides of the furin-positive area (Fig. 2, C and D). TGF α is expressed along a pit cell region from the pit region to the lumen (13, 14, 49). Since furin-positive cells were localized to the upper cell proliferating zone and less proliferating pit cell region, furin may activate precursors for growth-related proteins.

A great amount of information has been obtained from culture cell studies since the primary culture of gastric surface mucous cells has been established (10, 16, 19–21). Most of the information obtained has been important for interpreting the in vivo function of gastric mucous cells in the fundic mucosa. Caution should be used, however, when extrapolating information from the cell culture studies to the in vivo function of individual cells localized in the gastric glands. In gastric glands, surface mucous cells are composed of heterogeneous populations in their differentiation process and only a small fraction of the cells divide at the proliferative zone of the isthmus (1, 2). When the surface mucous cells are placed in primary culture, however, $\sim 50\%$ of the cells are dividing by thymidine or BrdU incorporation, and many cells exhibit a decrease in PAS-positive materials (10). In general, culture cells tend to lose their differentiated characteristics and instead acquire fetal stage-specific characteristics (22). Indeed, GSM06 cells at 33°C and fetal epithelial cells exhibit similar characteristics: a high level of furin expression, a negligible level of PAS-positive materials, a low level of TGF α production, and only a few secretory granules (45–47). Thus, we assume that GSM06 cells in a nonconfluent state at 33°C may correspond to the in vivo surface epithelial cells in the fetal stage around gestational day 18. In contrast, the cells in a confluent state at 39°C showed a negligible level of furin, the appearance of PAS-positive materials, a high level of TGF α production, and a considerable number of secretory granules. These characteristics indicate that the cells in this state may correspond to the in vivo cells at gestational days 20 and 21, rather than the gastric epithelial cells in the adult stage.

With these precautions in mind, we used the GSM06 cell line as a model for gastric surface mucous cells. We examined the role of furin in growth and differentiation by the use of PAS-positive materials and TGF α as a differentiation marker. For this purpose, we generated two modified cell lines, one was transformed with a mouse furin cDNA vector, and the other with an antisense furin cDNA vector that was designed to suppress native sense furin RNA-directed protein synthesis. The expression of these vectors was controlled under a metallothionein promoter, which was activated by zinc ions (22). With the sense furin RNA expression, GSM06 cells exhibited undifferentiated characteristics, virtually no appearance of PAS-positive materials, and a low level of TGF α expression. As long as furin was expressed in the cells, PAS-positive materials did not appear, even in the differentiation state, in high cell density and at the nonpermissive temperature of 39°C. On the contrary, expression of antisense furin RNA in GSM06 cells caused the induction of PAS-positive materials and TGF α production even in low cell density and at the permissive tempera-

ture of 33°C. Moreover, expression of antisense furin RNA decreased the growth rate of GSM06 cells. These findings indicate that furin expression keeps GSM06 cells in an undifferentiated state. However, the overexpression of furin did not increase cell growth any further. In growing cells, endogenous furin may have been expressed enough to induce the full capacity of cell growth.

Since furin is known to convert precursors for growth-related proteins to their active forms (28–33, 35–37), we determined whether the furin-suppressing cells produce fewer bioactive forms of growth factors. We examined the effect of conditioned medium from the furin-suppressing cells on intact GSM06 cells. Conditioned medium from the cell culture in the presence of the antisense oligonucleotide caused decreased DNA synthesis in GSM06 cells as compared with the control conditioned medium from the cell culture in the presence or absence of the sense oligonucleotide, suggesting that furin suppression may generate less conversion of growth factor precursors. Recently, it was demonstrated that furin recycles between the TGN and plasma membrane (50). Furin possesses an important signal for this recycling in its 58 amino acid cytoplasmic tail (51). When furin is increased, the conversion of growth factor precursors may be facilitated. In addition, the constitutive pathway may enlarge due to more active furin recycling.

In fetal gastric mucosa, furin appears to convert growth-promoting factor precursors to their active forms, but such growth factor precursors with a furin-cleavable processing site remain to be identified. The processing of TGF α precursor to the small 50 amino acid peptide is an important step for growth control (49), but the TGF α precursor does not possess furin-cleavable sites (52). Furthermore, TGF α appears after gestational day 20 in the rat fetal gastric epithelial cells (53). Thus, it is likely that some other growth-promoting peptides with a furin-cleavable site are involved in the active proliferation of the fetal gastric epithelial cells. Although the substrates for furin are uncertain, we suggest that furin is important in the growth of the gastric mucosa in both fetal and adult stages by converting yet unknown precursors to their active forms.

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