RNA Metabolism in Myotonic Dystrophy

Patient Muscle Shows Decreased Insulin Receptor RNA and Protein Consistent with Abnormal Insulin Resistance

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Abstract

Myotonic dystrophy is a dominantly inherited clinically variable multisystemic disorder, and has been found to be caused by heterozygosity for a trinucleotide repeat expansion mutation in the 3’ untranslated region of a protein kinase gene (DM kinase). The mechanisms by which the expanded repeat in DNA results in a dominant biochemical defect and the varied clinical phenotype, is not known. We have recently proposed a model where disease pathogenesis may occur at the RNA level in myotonic dystrophy: the mutant DM kinase RNA with the expansion mutation may disrupt cellular RNA metabolism in some general manner, as evidenced by defects in RNA processing of the normal DM kinase gene in heterozygous patients (dominant negative RNA mutation). Here we further test this hypothesis by measuring RNA metabolism of other genes in patient muscle biopsies (nine adult onset myotonic dystrophy patients, two congenital muscular dystrophy patients, four normal controls, and four myopathic controls). We focused on the insulin receptor gene because of the documented insulin resistance of DM patients. We show that there is a significant decrease in insulin receptor RNA in both total RNA and RNA polyA+ pools relative to normal and myopathic control muscles (P < 0.002), measured relative to both dystrophin RNA and muscle sodium channel RNA. We also show reductions in insulin receptor protein. Our results reinforce the concept of a generalized RNA metabolism defect in myotonic dystrophy, and offer a possible molecular mechanism for the increased insulin resistance observed in many myotonic dystrophy patients. (J. Clin. Invest. 1997. 99:1691–1698.) Key words: myotonic dystrophy • insulin resistance • trinucleotide repeat expansion disorder • insulin receptor • DM-kinase

Introduction

Myotonic dystrophy (DM)‡ is an autosomal dominant neuromuscular disease with an estimated incidence in adults of about 1 in 8,500 individuals (1). Presenting clinical symptoms range from very mild (cataacts in middle age), to very severe (neonatal lethal hypotonia). The classical presentation includes myotonia and progressive weakness and wasting of distal skeletal muscles. Many organ systems are involved, however, with patients often showing cardiac conduction defects, smooth muscle involvement, hypersomnia, endocrine problems (increased insulin resistance), and premature balding and testicular atrophy in males (2). The disease is caused by an unstable CTG trinucleotide repeat in the 3’ untranslated region of the DM kinase gene on chromosome 19 q 13.3 (3, 4, 5, 6, 7).

The effect of the expanded trinucleotide repeat on gene and protein expression is actively being investigated for myotonic dystrophy, and many of the other trinucleotide repeat disorders (Fragile X, bulbospinal muscular atrophy, Huntington’s disease, spinocerebellar ataxias, Friedreich’s ataxia, etc.). In recessively inherited Fragile X and Friedreich’s ataxia, gene expression data is consistent with loss of function of the protein product of the corresponding gene, as expected in all recessively inherited disorders. In Fragile X, the trinucleotide repeat is at the 5’ end of the gene, where it appears to decrease transcription via hypermethylation of the promoter (8). In Friedreich’s ataxia, the trinucleotide repeat is in an intron, where it interferes with correct formation of RNA (9). All of the other trinucleotide repeat disorders except Myotonic dystrophy show the expanded repeat in the coding sequence of the gene, leading to extensions of a polyglutamine tract in the protein product. This alteration of the protein product likely leads to a gain or a change of function, consistent with the dominant inheritance pattern of all of these disorders (10). The expansion in the DM kinase gene causing myotonic dystrophy is unique in its location in the 3’ untranslated region of the gene. The location of the repeat is difficult to reconcile with the dominant inheritance pattern of myotonic dystrophy. Most dominantly inherited disorders show expression of abnormal protein products (change of function or dominant negative). The DM kinase protein encoded by the abnormal gene, however, should show no sequence variations, and hence should not show a change of function or dominant negative effect at the protein level.

Early studies of transcription patterns of the normal and mutant DM kinase gene in affected heterozygous patients were contradictory, some showing increased (11), (but most
showing decreased) expression from the mutant gene (12, 13, 14, 15). Using patient muscle biopsies, we recently showed that the DM kinase gene containing the expansion mutation is indeed transcribed at levels similar to the normal gene. Both the normal and the abnormal RNAs accumulate in total RNA pools, however, the normal and mutant RNAs showed defects in RNA processing (16). Based on these results, we hypothesized that myotonic dystrophy may represent the first dominant RNA disease, where generalized disruptions of RNA metabolism mediated by accumulation of the abnormal RNA could cause the varied clinical symptoms. This model could offer an explanation for the dominant inheritance pattern and dramatic clinical variability of myotonic dystrophy: different subpopulations of RNAs could be altered in a cell type and in a developmentally specific pattern. If this hypothesis is true, then it should be possible to identify abnormal process and/or accumulation of RNAs from other specific genes in patient muscle. The most likely candidates for abnormalities would be those genes whose gene products may be involved in the disease pathogenesis. It is well documented that myotonic dystrophy patients show increased insulin resistance (17, 18, 19, 20, 21). Some endocrinological studies of myotonic dystrophy patients have suggested that the insulin resistance could be a consequence of decreased receptor number (22). We hypothesized that the insulin resistance could be the result of abnormal metabolism of insulin receptor RNA in patient cells (dominant-negative RNA). To test this hypothesis, we studied expression of the insulin receptor gene in myotonic dystrophy patient muscle biopsies.

Methods

Muscle biopsies. We used preexisting muscle biopsies collected for diagnostic purposes from 17 patients. The muscles had been flash frozen after biopsy in isopentane cooled in liquid nitrogen, and were stored in airtight containers at −80°C until analysis. Nine biopsies were from patients with classical adult onset myotonic dystrophy, and two were from congenital muscular dystrophy patients. The presence of a CTG trinucleotide repeat expansion ranging from 83–2300 repeats was determined by DNA studies of muscle and/or blood. Four biopsies (myopathic controls) were from patients referred for dystrophin protein testing. All had myopathic histopathology and clinical biopsies (myopathic controls) were from patients referred for dystrophin or sodium channel, creatine kinase, transferrin, and DM kinase were designed to vary a relatively small amount in size. For each set, the forward primer was synthesized with an amino link residue at the 5’ end, and the primer was covalently coupled via an ester linkage to carboxyl-fluorescein (Molecular Probes, Inc., Eugene, OR) as previously described (29). For each PCR product, the reverse primer was designed upstream of the sequence-specific reverse primer used for cDNA synthesis. Primer sequences were as follows: adult skeletal muscle sodium channel (Na+): 4702F: 5’-GGT GAC TGC GGC AAC CCC TCC A, 4820R: 5’-GTG GCC ACA TTG AAC TTC TCC; insulin receptor (InR): 4090F: 5’-GAG ATG GAT TTT GAG GAC ATG, 4223R: 5’-GTG TAA GGG AGG AGG TGT TGC TCG TCG TAG; dystrophin (Dys): 5369F: 5’-AGG CAC AGC TGA ATG ACG TAC G, 5533R: 5’-CTT GCC AGT CTT AAT TCT GTG T; DM kinase (DMK): 1014F: 5’-ACT TGT CGG AGC TTA GCC GAG GT; 1155R: 5’-TGG CTC AAG CAG CTG CTC GG; creatine kinase M (CKM): 997F: 5’-AGTTCG AGG AGA TCC TCA CCC GC, 1196R: 5’-ATG GAC TGG CTT TTC TAC AAC TCC; and transferrin receptor (TfR): 2151F: 5’-CTG TCT TTT GCC TAC GTC TCT CC, 2305R: 5’-AGA GCC AAC TGG TTT CTT AGC A. Hemi-nested multiplex fluorescent RT-PCR with six different products to check each total RNA and polyA+ RNA sample was done (see Fig. 1). A mix of 70 ng of each primer set (above), and 2 µl of cDNA corresponding to about 20 ng of total RNA, was amplified with 2.5 U AmpliTaq DNA polymerase (Perkin Elmer Cetus Instruments, Branchburg, NJ), 25 mM of dNTPs, and 1× PCR reaction buffer in a total volume of 25 µl. Cycling conditions for these six primers sets were 94°C for 1 min, 60°C for 1 min, and 72°C for 1 min.

Quantitation of insulin receptor RNA by multiplex fluorescent RT-PCR. Multiplex RT-PCR and quantitation of fluorescent PCR products of insulin receptor RNA related to dystrophin, Na+ channel, creatine kinase, or transferrin RNA, was done as previously described (30). Standardized mixes of primers were made, and cDNA corresponding to ~20 ng of total RNA amplified with primer mixes for 22 cycles using standard conditions (denaturation at 94°C for 3 min, 94°C for 1 min, 50°C for 1 min, and 72°C for 1 min for 10 cycles; and then an additional 12 cycles at 94°C for 1 min, 60°C for 1 min, and 72°C for 1 min (total 22 cycles)).

Fluorescent RT-PCR products corresponding to 2 ng of total input RNA were denatured and loaded on an ABI 373A DNA automated sequencer (6% acrylamide, 8 M urea, 0.5× TBE), using a ROX/fluorescein matrix standard (both from Applied Biosystems, Inc. Foster City, CA). ROX-labeled markers (Applied Biosystems) were included as internal size standards. Computer lane tracking was done manually based on gel images, and tracked lanes integrated using the GeneScan software (Applied Biosystems). Peak areas corresponding to expected RT-PCR product sizes were determined, and peak area ratios of insulin receptor (relative to internal control dystrophin or sodium channel) were calculated. Peak area ratios were normalized to the average of control biopsy values, and were presented with standard errors. Three to six separate RT-PCR measurements were done per patient biopsy.
Statistical significance was done using ANOVA analysis of variance using all calculated ratios.

Quantitative immunoblotting. Cryosections of muscle biopsies were solubilized in sample buffer (10% SDS, 0.1 M Tris pH 8, 5 mM EDTA, and 10 mM DTT), and proteins were electrophoretically separated on 3.5–12.5% gradient SDS-PAGE gels and transferred to nitrocellulose as previously described (31). Quadruplicate immunoblots were incubated with the following antibodies: rabbit α-human insulin receptor polyclonal (Signal Transduction, Inc., Lexington, KY), and monoclonals-directed fast-twitch myosin heavy chain (F58) (32), and fast-twitch Ca^{2+} Mg^{2+} ATPase (D2) (33). Second antibodies were conjugated to horseradish peroxidase, and immuno complexes were visualized using chemiluminescence (ECL; Amersham).

Quantitations were done using Bioscan. Statistical comparisons were done using Student’s t-test.

Immunofluorescence. Cryosections (4 μm) were thawed on Superfrost slides (Fisher Scientific Co., Pittsburgh, PA) and processed both unfixed, and fixed with cold acetone. Blocking and washes were done in PBS with 10% horse serum. Antibodies used were for fast-twitch Ca^{2+} Mg^{2+} ATPase (D2) (38), and for the α subunit of the insulin receptor (34). Visualization was done with species-specific antibodies bound by Cy3 (Jackson Laboratories, Bar Harbor, ME). Photodocumentation was done using a fluorescent microscope (FXA; Nikon Inc., Melville, NY).

**Results**

**RNA studies.** RNA was isolated from 17 muscle biopsies (nine adult onset myotonic dystrophy patients, two congenital myotonic dystrophy patients, four normal controls, and four myopathic disease controls) (Table I). For each RNA sample, 100 ng of total RNA, and polyA+ RNA corresponding to 100 ng of total RNA, were reverse-transcribed into cDNA using six different sequence-specific primers simultaneously (see Methods). Approximately 20 ng of each total and corresponding polyA+ cDNA was amplified using internal sequence-specific primers in three separate mixtures. One qualitative multiplex amplification contained six different PCR products (Na+ channel

<table>
<thead>
<tr>
<th>Patient No.</th>
<th>Age of biopsy</th>
<th>Age of onset</th>
<th>Muscle</th>
<th>Endocrine</th>
<th>Brain</th>
<th>Heart</th>
<th>Eye</th>
<th>CTG repeats</th>
</tr>
</thead>
<tbody>
<tr>
<td>DM1</td>
<td>41 yr.</td>
<td>23</td>
<td>Myotonia, distal atrophy</td>
<td>Normal</td>
<td>School difficulties</td>
<td>Normal</td>
<td>Lens opacity</td>
<td>ND</td>
</tr>
<tr>
<td>DM2</td>
<td>29 yr.</td>
<td>12</td>
<td>Myotonic facies, myotonia, distal atrophy</td>
<td>Thyroid goiter</td>
<td>Normal</td>
<td>Mitral prolapse</td>
<td>Lens opacity</td>
<td>715</td>
</tr>
<tr>
<td>DM3</td>
<td>22 yr.</td>
<td>15</td>
<td>Myotonia, distal atrophy</td>
<td>Marked testicular atrophy, hypospermatogenesis</td>
<td>Normal</td>
<td>Slight right ventricle dilatation</td>
<td>Bilateral cataract</td>
<td>638</td>
</tr>
<tr>
<td>DM4</td>
<td>58 yr.</td>
<td>50</td>
<td>Myotonia, slight distal atrophy</td>
<td>Normal</td>
<td>Normal</td>
<td>Slight mitral and tricuspid reflux</td>
<td>Normal</td>
<td>205</td>
</tr>
<tr>
<td>DM5</td>
<td>26 yr.</td>
<td>20</td>
<td>Distal atrophy</td>
<td>Normal</td>
<td>Normal</td>
<td>Conduction defect</td>
<td>Cataracts</td>
<td>180</td>
</tr>
<tr>
<td>DM6</td>
<td>27 yr.</td>
<td>24</td>
<td>Myotonia, distal atrophy, weakness</td>
<td>Normal</td>
<td>Q.I. = 73 (WAIS test)</td>
<td>Normal</td>
<td>Lens opacity</td>
<td>1550</td>
</tr>
<tr>
<td>DM7</td>
<td>46 yr.</td>
<td>42</td>
<td>Myotonia</td>
<td>Marked hypospermatogenesis</td>
<td>Normal</td>
<td>Abnormal ventricular potentials</td>
<td>Normal</td>
<td>83</td>
</tr>
<tr>
<td>DM8</td>
<td>15 yr.</td>
<td>11</td>
<td>Myotonia facies, myotonia, mild distal atrophy</td>
<td>Normal</td>
<td>Normal</td>
<td>Mild right ventricle dilatation</td>
<td>Normal</td>
<td>564</td>
</tr>
<tr>
<td>DM9</td>
<td>42 yr.</td>
<td>38</td>
<td>Myotonia, distal atrophy, weakness</td>
<td>Hypospermatogenesis</td>
<td>Q.I. = 108 (WAIS test)</td>
<td>I+ Heart block</td>
<td>Bilateral cataract</td>
<td>265</td>
</tr>
<tr>
<td>cDM1</td>
<td>2 yr.</td>
<td>–</td>
<td>Talipes, severe hypotonia, weakness, atrophy</td>
<td>Normal</td>
<td>Severe mental retardation</td>
<td>Normal</td>
<td>Normal</td>
<td>2300</td>
</tr>
<tr>
<td>cDM2</td>
<td>10 mo.</td>
<td>–</td>
<td>Talipes, severe hypotonia, weakness, atrophy</td>
<td>Normal</td>
<td>Severe mental retardation</td>
<td>Normal</td>
<td>Normal</td>
<td>1328</td>
</tr>
<tr>
<td>MC1</td>
<td>10 yr.</td>
<td>–</td>
<td>Mild weakness</td>
<td>Normal</td>
<td>Normal</td>
<td>Normal</td>
<td>Normal</td>
<td>ND</td>
</tr>
<tr>
<td>MC2</td>
<td>16 yr.</td>
<td>–</td>
<td>Proximal weakness</td>
<td>Normal</td>
<td>Slight mental retardation</td>
<td>Normal</td>
<td>Normal</td>
<td>ND</td>
</tr>
<tr>
<td>MC3</td>
<td>22 yr.</td>
<td>–</td>
<td>Progressive myopathy</td>
<td>Normal</td>
<td>Normal</td>
<td>Normal</td>
<td>Normal</td>
<td>ND</td>
</tr>
<tr>
<td>MC4</td>
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<td>6</td>
<td>Proximal weakness</td>
<td>Normal</td>
<td>Normal</td>
<td>Normal</td>
<td>Normal</td>
<td>ND</td>
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</tbody>
</table>

ND = not determined.
nel 118 bp, InR 133 bp, DMK 140 bp, TIR 155 bp, CKM 199 bp). This assay was used to check the integrity of the RNAs (data not shown).

Two quantitative mixes were used. One contained both insulin receptor (InR) and dystrophin (Dys) (exon 37) primers. The second quantitative amplification contained both insulin receptor (InR) and sodium channel (Na+/H+) primers. RT-PCR products (22 cycles) corresponding to 2 ng of input total RNA were electrophoresed on an ABI automatic sequencer (373A) to quantitate amplification products using peak area. From two different cDNA syntheses, 3–6 different RT-PCR amplifications of InR and Dys were quantitated for each data point (Fig. 1) (Table II). A similar assay was done for insulin receptor (InR) and sodium channel (Na+/H+) (Fig. 1) (Table II).

Adult onset myotonic dystrophy and congenital myotonic dystrophy patient muscle biopsies showed a statistically significant decrease in insulin receptor RNA in the total RNA pools relative to dystrophin (exon 37) (Fig. 2 A). Both adult and congenital DM patients showed 50% of insulin receptor RNA levels seen in controls (P = 0.0000014). In polyA+ pools, the average insulin receptor RNA levels in DM patients also showed a decrease in InR polyA+ RNA, to levels ~ 65% of normal (P = 0.018) (Fig. 2 A).

The observed statistically significant decrease in insulin receptor in both total RNA and in polyA+ RNA pools presumed that dystrophin RNA levels were invariant between the different groups of patients. To rule out alterations of dystrophin RNA levels seen in controls (P = 0.0000014). In polyA+ pools, the average insulin receptor RNA levels in DM patients also showed a decrease in InR polyA+ RNA, to levels ~ 65% of normal (P = 0.018) (Fig. 2 A).

Table II. Quantitative Studies of Insulin Receptor Expression in Total RNA and PolyA+ RNA in DM Patients and Myopathic Controls

<table>
<thead>
<tr>
<th>Patient No.</th>
<th>Average total RNA InR/Dys ratio</th>
<th>Average PolyA+ InR/Dys ratio</th>
<th>Average total RNA of InR/Na+ ratio</th>
<th>Average PolyA+ of InR/Na+ ratio</th>
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<tbody>
<tr>
<td></td>
<td>%</td>
<td>%</td>
<td>%</td>
<td>%</td>
</tr>
<tr>
<td>DM1</td>
<td>40</td>
<td>33</td>
<td>15</td>
<td>16</td>
</tr>
<tr>
<td>DM2</td>
<td>32</td>
<td>87</td>
<td>24</td>
<td>33</td>
</tr>
<tr>
<td>DM3</td>
<td>38</td>
<td>80</td>
<td>21</td>
<td>39</td>
</tr>
<tr>
<td>DM4</td>
<td>53</td>
<td>91</td>
<td>56</td>
<td>46</td>
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<tr>
<td>DM5</td>
<td>68</td>
<td>89</td>
<td>45</td>
<td>66</td>
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<tr>
<td>DM6</td>
<td>64</td>
<td>24</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>DM7</td>
<td>47</td>
<td>75</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>DM8</td>
<td>61</td>
<td>33</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>DM9</td>
<td>41</td>
<td>58</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>cDM1</td>
<td>53</td>
<td>70</td>
<td>34</td>
<td>73</td>
</tr>
<tr>
<td>cDM2</td>
<td>55</td>
<td>48</td>
<td>46</td>
<td>57</td>
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<tr>
<td>MC1</td>
<td>114</td>
<td>119</td>
<td>111</td>
<td>55</td>
</tr>
<tr>
<td>MC2</td>
<td>101</td>
<td>74</td>
<td>112</td>
<td>114</td>
</tr>
<tr>
<td>MC3</td>
<td>–</td>
<td>–</td>
<td>84</td>
<td>64</td>
</tr>
<tr>
<td>MC4</td>
<td>–</td>
<td>–</td>
<td>113</td>
<td>131</td>
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</table>
sodium channel RNAs. Dystrophic dystrophy patients showed statistically significant decreases in insulin receptor RNA compared to controls (Fig. 2A). The posttransfer gel corresponding to the insulin receptor immunoblot was stained for residual myosin heavy chain as previously described (31) as a control for muscle tissue loading (Fig. 2B). All myotonic dystrophy biopsies showed a decrease in insulin receptor protein relative to adjacent controls, and this was not a consequence of differences in protein loaded on the gel (Fig. 2A and B).

It is well documented that some myotonic dystrophy patients show selective loss of slow-twitch (type I) myofibers, leading to fiber type II predominance (1). Some studies have shown decreased insulin binding to rodent fast-twitch fibers relative to slow-twitch fibers, however, in higher vertebrates such correlations have not been confirmed (35). We were nevertheless concerned that the decrease in insulin receptor levels in myotonic dystrophy could be a consequence of preferential loss of type I fibers. To test this concern, we studied five myotonic dystrophy biopsies for fiber type content, both by immunoblotting (Fig. 3, C and D), and by immunofluorescence (data not shown). Patient DM showed dramatic loss of type I fibers, while the other patients had less extensive fiber type predominance. Insulin receptor levels were decreased in each sample irrespective of the extent of fiber type predominance (Fig. 3). Immunofluorescence staining of cryosections also showed a decrease of signal intensity in myotonic dystrophy patients relative to controls, although the signal was very weak (data not shown).

The insulin receptor immunoblot was quantitated by computer densitometry, and the levels of insulin receptor protein were normalized to total myosin content and expressed as a percentage of the average levels seen in controls (Fig. 3E). Myotonic dystrophy patients showed an ~30% decrease in protein levels (P = 0.054).

Discussion

The molecular pathogenesis of myotonic dystrophy is controversial. It is clear that the disease process is initiated by the pathological expansion of a trinucleotide GTG repeat in the 3' untranslated region of a protein kinase gene (DM kinase). The mechanisms by which the expanded repeat causes the highly variable and multisystemic clinical features enigmatic of myotonic dystrophy, however, are not yet clear. There are currently three pathogenetic hypotheses with data that support each: (a) loss of function/haploinsufficiency; (b) chromatin changes with multigene transcription changes; (c) generalized RNA metabolism defect mediated by the mutant RNA.

The loss of function hypothesis suggests that the expanded repeat leads to a loss of transcription, translation, or both. Each patient, however, is heterozygous, so this change should only affect transcription or translation from the mutant allele (haploinsufficiency). Consistent with this model, early publications showed that RNA levels of the DM kinase were reduced in both patient muscle and in cell cultures (12, 13, 14, 15). The
corresponding protein also showed deficiency in patient cells (36). More recent studies have documented transcriptional activity from the mutant DM kinase gene resulting in accumulation of mutant RNA in total RNA pools, although the abnormal RNA is not polyadenylated and is not available for translation (16, 17, 18). While all these data are consistent with haploinsufficiency of the cAMP kinase, it is difficult to understand how such mild changes in protein levels could result in the dramatic clinical variability seen in myotonic dystrophy patients. Moreover, transgenic mice that have no functional DM kinase (homozygous recessive, complete loss of function) show only a mild, late onset muscle phenotype, and share few of the other symptoms of the human disease (37, 38).

The second hypothesis for disease pathogenesis is that the presence of the expanded repeat in chromatin leads to altered chromatin structure (39) and abnormal nucleosomes binding (40, 41). The altered chromatin structure could result in long-range effects on gene transcription, perhaps altering expression of both the DM kinase gene and other neighboring genes. If gene transcription were in fact decreased from a series of neighboring genes, then a contiguous gene haploinsufficiency syndrome could result, as is seen in contiguous gene deletion syndromes. The clinical variability of the disease could be explained by a relationship between the size of the CTG expansion mutation and the number of neighboring genes affected. If this mechanism is true, then it might be expected that DM kinase transcription from the mutant allele is turned off; however, this is clearly not the case (16, 17, 18). A series of novel genes have been identified neighboring the DM kinase gene (42, 43, 44), however, it is not yet clear if transcription from any of these genes is affected by expansion of the DM kinase CTG repeat. It is important to note that the insulin receptor gene is in fact on chromosome 19, as is the DM kinase gene—long range transcriptional inactivation of chromosome 19 could conceivably explain the decrease in insulin receptor RNA we describe here.

The third hypothesis suggests that the presence of the CTG expansion in RNA could have a deleterious effect on the cell, altering RNA metabolism of a number of genes. This dominant negative RNA hypothesis is supported by four publications from three laboratories using different experimental systems, and by data presented in this paper. In situ nucleic acid hybridization experiments done by Singer’s group (45) found that relatively large amounts of RNA containing the expansion mutation could be found in nuclei of muscle and cultured fibroblasts from myotonic dystrophy patients. This RNA was not polyadenylated, appeared as aggregates in the nucleus, and was not physically associated with the mutant copy of the gene (DNA). In parallel, both we (16) and Krahe et al. (46) showed that the DM kinase RNA containing the expansion mu-

Identical immunoblots were also incubated with monoclonal antibodies directed against fast-twitch isoforms of myosin heavy chain (C), and Ca\(^{2+}\)-Mg\(^{2+}\) ATPase (D) to investigate the extent of fiber type II (fast-twitch) predominance. All muscle biopsies from myotonic dystrophy patients show a reduction in levels of insulin receptor protein (A). These reductions are not explained by reduced protein content of the lanes (B), or fiber-type predominance (C and D). (E) Quantification of immunoblots, which documents the decrease in insulin receptor protein.

Figure 3. Immunoblot analysis shows decreased insulin receptor protein in muscle biopsies from myotonic dystrophy patients. Shown is immunoblot analysis of the insulin receptor (A), and corresponding posttransfer Coomassie blue staining of the myosin heavy chain (B).
It has been well documented that myotonic dystrophy patients have increased insulin resistance, and that this endocrine abnormality is possibly the consequence of decreased numbers of insulin receptors in muscle. We therefore tested the hypothesis that the RNA and protein corresponding to the insulin receptor gene is decreased in patient muscle. If true, then this data would lend support to the dominant negative RNA hypothesis.

We quantitated insulin receptor RNA and protein levels in muscle biopsies from a series of adult onset myotonic dystrophy patients, neonatal onset congenital myotonic dystrophy patients, myopathic disease controls, and normal controls. Insulin receptor RNA levels were measured in both total and polyA+ RNA pools, and quantitations were done relative to both dystrophin RNA and sodium channel RNA using an assay we have previously described (QMF-PCR). We found statistically significant reductions in insulin receptor RNA in myotonic dystrophy muscle. The reductions were disease-specific, and were statistically significant in both total and polyA+ pools. Similar reductions were present when normalizing to either dystrophin or the sodium channel RNAs. Consistent with the RNA data, we found a disease-specific decrease in insulin receptor protein in DM patient muscle.

These data taken together show that the mutant DM kinase gene is transcribed into RNA containing the expansion mutation, and that the mutant RNA is improperly processed, and forms aggregates within the patient nuclei where it sequencers CUG RNA binding proteins, making them unavailable for their normal cytosolic functions. Loss of CUG binding proteins from the cytoplasm then has a dominant effect on RNA metabolism, leading to disregulation of multiple RNAs, including the insulin receptor RNA. This hypothesis could explain the dominant inheritance, dramatic clinical variability, and multisystemic nature of the disease through alterations of specific RNA molecules of other genes, in a developmental- and tissue-specific manner.

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References

dependent receptor redistribution in HIRcB cells.


