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### Research Article

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# Lipoprotein Lipase Modulates Net Secretory Output of Apolipoprotein B In Vitro

## A Possible Pathophysiologic Explanation for Familial Combined Hyperlipidemia

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### Abstract

We showed previously that net secretory output of apolipoprotein B (apo B) from cultured human hepatoma cells (HepG2) is regulated by rapid reuptake of nascent lipoproteins before they have diffused away from the vicinity of the cells. We now sought to determine if the nascent lipoproteins could be remodeled to enhance or impede reuptake. We found that lipoprotein lipase (LpL), an enzyme that hydrolyzes lipoprotein triglyceride, reduced HepG2 output of apo B to one-quarter to one-half of control. The reduction was apparent during co-incubations as short as 2 h and as long as 24 h. Heparin, which blocks receptor-mediated binding of lipoproteins, abolished the effect of LpL on apo B output, without causing enzyme inhibition. To assess uptake directly, we prepared labeled nascent lipoproteins. LpL tripled the cellular uptake of labeled nascent lipoproteins, from  $15.2\% \pm 0.7\%$  to  $48.7\% \pm 0.3\%$  of the total applied to the cells. Cellular uptake of  $^{125}\text{I}$ -labeled anti-LDL receptor IgG was unaffected by LpL; thus, LpL enhanced reuptake by altering lipoproteins, not receptors. Because LpL is present in the space of Disse in the liver, we conclude that LpL may act on newly secreted lipoproteins to enhance reuptake in vivo. LpL deficiency would reduce local reuptake of apo B, which would appear as overproduction, thereby providing a mechanistic link between partial LpL deficiency and familial combined hyperlipidemia. (*J. Clin. Invest.* 1991. 88:1300–1306.) Key words: fibrin acid • futile cycling • heparin • hepatoma G2 cell • space of Disse

### Introduction

Apolipoprotein B-100 (apo B) is a 550-kD glycoprotein that is an essential component of low-density lipoproteins (LDL) and very low-density lipoproteins (VLDL) and has been implicated in the development of coronary artery disease. In humans, apo B is made by the liver, but mechanisms of secretory control have been poorly understood.

We recently showed that net secretory output of apo B from human hepatoma G2 (HepG2)<sup>1</sup> cells in culture can be regu-

A preliminary report of this study was presented at the 63rd Scientific Sessions of the American Heart Association, Dallas, TX, 13 November 1990 (1).

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1. Abbreviations used in this paper: HepG2, human hepatocellular carcinoma G2; LpL, lipoprotein lipase; PTA, phosphotungstic acids.

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lated by rapid reuptake of nascent lipoproteins shortly after their export across the plasma membrane (2). Thus, a variety of stimuli affects the net output of apo B, such as altering the number of cellular LDL receptors (2, 3) or altering the rate of diffusion out of the unstirred water layer by changing the viscosity of the culture medium (2, 4).

We now sought additional factors that might modulate net cellular output of apo B by enhancing or impeding reuptake. Several factors have been reported to affect the binding of mature plasma lipoproteins to the cell surface, and might therefore affect the binding and reuptake of apo B-rich nascent particles. These factors include lipoprotein lipase (LpL) (5–10), hepatic lipase (5, 10, 11), phospholipase A<sub>2</sub> (12, 13), cholesteryl ester transfer protein (11, 14), lecithin: cholesterol acyl transferase (15), lysophosphatidylcholine (16), apo E (17–19), apo AIV (20), and the C-apoproteins (21, 22).

For several reasons, we began our investigations with LpL. First, it is present in and near the space of Disse in adult mammalian liver (23). Thus, it would have physical access to newly secreted hepatic lipoproteins. Second, its enzymatic action on mature, triglyceride-rich lipoproteins is rapid (24), suggesting that it could substantially remodel nascent particles before they have diffused away from the vicinity of the hepatocytes. Third, LpL has been documented to enhance receptor binding and cellular uptake of a very broad range of apo B-containing lipoproteins, including LDL (5), VLDL (6, 7),  $\beta$ -VLDL (8), and chylomicrons (9, 10). Finally, we hypothesized a physiologic link between LpL and apo B secretion (1), based on the recent finding of partial LpL deficiency in some cases of familial combined hyperlipidemia (25, 26), a disease of hepatic apo B overproduction (27).

### Methods

**Purification of LpL.** LpL (EC 3.1.1.34) was purified from fresh cow's milk by a modification of the method of Posner et al. (28). The cream, which contains no detectable LpL activity, was removed from fresh milk by centrifugation (22,000 g for 75 min) and discarded. 10 ml of Intralipid (Kabi-Vitrum, Alameda, CA) was added per 210 ml of skim milk, and the mixture was incubated for 30 min at 37°C on a shaker. The Intralipid, which binds LpL, was recovered and washed by two centrifugations at 3,300 g for 60 min at 4°C in a SW 28 rotor (Beckman Instruments, Inc., Somerset, NJ). The final floating fat cakes were recovered with the use of a spatula and suspended in cold acetone (80 ml per cake), filtered on a medium-porosity sintered glass funnel, and delipidated with diethyl ether washes (50 ml per cake). The resulting powder is a partially purified preparation of LpL that is stable at –20°C for several months (28).

Before each experiment, a fresh batch of highly purified LpL was prepared by subjecting a portion of the acetone powder to heparin-agarose chromatography (Affi-gel heparin, Bio-Rad Laboratories, Richmond, CA) (28). The acetone powder was suspended by end-over-end rotation in 0.1% Triton X-100 in standard buffer (50 mM NH<sub>4</sub>OH-HCl, pH 8.5) at a concentration of 6.25 mg of powder per ml. It was

then applied to the column equilibrated in standard buffer, at 4.0 ml of suspension per ml of wet gel. The column was washed with 0.3 M NaCl, 0.2 M sodium citrate in standard buffer, followed by 0.72 M NaCl in standard buffer. Pure LpL was eluted with 2.0 M NaCl in standard buffer, then simultaneously concentrated and dialyzed into 10 mM sodium phosphate, 150 mM NaCl, pH 7.40, using a YM30 membrane (Amicon Corp., Lexington, MA). The final enzyme preparation was adjusted to 30% glycerol and stored at  $-20^{\circ}\text{C}$  until use.

By polyacrylamide gel electrophoresis in sodium dodecyl sulfate and mercaptoethanol (29), there was a single band at 58,000 D. Lipase activity was measured using Intralipid labeled with glycerol tri [9,10(n)- $^3\text{H}$ ]oleate as the substrate (Amersham Corp., Arlington Heights, IL) (30) and apo-HDL as a source of apo CII. The reaction mixture (200  $\mu\text{l}$ ) was incubated at for 15 min at  $25^{\circ}\text{C}$ , and the reaction was terminated by the addition of 3.25 ml of methanol/chloroform/heptane (1.41:1.25:1.00, vol/vol/vol), followed by 1.05 ml of potassium carbonate buffer (pH 10 Buffer Standard Solution, Fisher Scientific Co., Malvern, PA) (31). Release of [ $^3\text{H}$ ]oleate was determined by liquid scintillation counting of the top (aqueous) phase. Specific activities at  $25^{\circ}\text{C}$  for our preparations were  $118.0 \pm 16.6$  U/mg (mean  $\pm$  SEM,  $n = 5$ ), where 1 U was defined as the amount of LpL that released 1  $\mu\text{mol}$  of free fatty acid per minute.

**Apo B secretion experiments.** In our secretion experiments, HepG2 cells grown to near-confluency in 35-mm wells were incubated overnight in a selenium-supplemented, serum-free medium that sustains their usual rate of growth, as described previously (2). The next morning, the cells received fresh medium supplemented with L-[4,5- $^3\text{H}$ ]leucine (120–190 Ci/mmol, Amersham Corp., 75  $\mu\text{Ci}/\text{ml}$  of medium, 1.0 ml of medium per 35-mm well), with or without the highly purified bovine milk LpL. Buffer with 30% glycerol was added to all control wells, to match the volume of LpL. After incubation for 2–24 h at  $37^{\circ}\text{C}$ , media and cells were harvested.

Tritiated apo B was purified from media either by immunoprecipitation or by ultracentrifugation followed by precipitation in 50% isopropanol. Immunoprecipitation was performed by an adaptation of the procedures of Harlow and Lane (32), using a monospecific rabbit anti-human apo B antiserum that was generously supplied by Dr. Charles L. Bisgaier, Columbia University (33). Samples of media (300–500  $\mu\text{l}$ ) were mixed with a detergent solution to achieve final concentrations of 1.25% Triton X-100 and 0.625% sodium dodecyl sulfate, then incubated overnight at  $4^{\circ}\text{C}$  to solubilize the apo B completely. A mock immunoprecipitation was performed by adding 5  $\mu\text{l}$  of nonimmune serum and 40  $\mu\text{l}$  of protein A-Sepharose beads (catalogue no. P-3391, Chemical Co., St. Louis, MO) to each sample and incubating for 1 h at room temperature. After removing the protein A beads by centrifugation, a true immunoprecipitation was performed by adding 5  $\mu\text{l}$  of immune serum, followed by a 1-h incubation at room temperature, addition of another 40  $\mu\text{l}$  of protein A beads, then another 1-h incubation. The second set of protein A beads was washed 10 times with a solution of Triton X-100 and sodium dodecyl sulfate, and near-absence of radioactivity in the final wash was verified by scintillation counting ( $< 20$  cpm above background). Bound immune complexes were released from the beads by adding 500  $\mu\text{l}$  of release buffer (2% sodium dodecyl sulfate, 60 mM Tris, 10% glycerol, pH 6.8) and incubating for 10 min at  $80^{\circ}\text{C}$  (32). Released radioactivity was measured by liquid scintillation counting.

Quantitative and specific immunoprecipitation was verified in several ways. First, samples of unlabeled conditioned medium were mixed with  $^{125}\text{I}$ -LDL, then subjected to immunoprecipitation, to verify that antiserum was added in excess of antigen, and that protein A beads were added in excess of antiserum (32). We achieved  $92.0 \pm 0.6\%$  (mean  $\pm$  SEM,  $n = 10$ ) recovery of  $^{125}\text{I}$ -LDL. Second, samples of labeled conditioned medium were removed from cells, incubated at  $37^{\circ}\text{C}$  for 2–24 h with or without LpL, and then immunoprecipitated. These studies showed that LpL had no effect on [ $^3\text{H}$ ]apo B recovery (decreased by  $3.3 \pm 2.0\%$ ,  $n = 3$ , NS). Incubation with LpL also had no effect on immunoprecipitation of  $^{125}\text{I}$ -LDL that had been added to unlabeled conditioned medium (increased by  $1.6 \pm 0.7\%$ ,  $n = 4$ , NS).

Finally, selected immunoprecipitates from experimental incubations were subjected to polyacrylamide gel electrophoresis and autoradiography, to verify the identity of the precipitated radioactivity as [ $^3\text{H}$ ]apo B (see Results section).

Purification of tritiated apo B from media by ultracentrifugation and isopropanol precipitation was performed by the method of Egusa et al. (34), with minor modifications as previously described (2). Specifically, media samples were each supplemented with  $\sim 200$   $\mu\text{g}$  of carrier LDL protein. The ultracentrifugation was performed at a density of 1.21 g/ml, to ensure total recovery of all lipoproteins. The ultracentrifugation supernatants were then mixed with distilled water, to lower the density to 1.063 g/ml before addition of isopropanol to precipitate apo B at  $4^{\circ}\text{C}$  (cf. reference 34).

Total secreted  $^3\text{H}$ -protein was measured by liquid scintillation counting of the washed pellet obtained from precipitation of media with 10% (wt/vol) trichloroacetic acid and 1.0% (wt/vol) phosphotungstic acid (TCA/PTA) (35). Total cellular protein was determined by modified Lowry (36). Data for apo B output by the cells were expressed either as the percentage of total labeled secreted protein that was labeled apo B (i.e., 100 times [ $^3\text{H}$ ]apo B radioactivity divided by  $^3\text{H}$ -protein radioactivity), or as total [ $^3\text{H}$ ]apo B radioactivity secreted per well.

In one secretion experiment, heparin (20 mg/ml; catalogue no. H-3125, lot no. 29F-0314; Sigma Chemical Co.) was added simultaneously with the [ $^3\text{H}$ ]leucine and LpL, to block receptor-mediated reuptake of nascent lipoproteins (37, 38). Media were harvested 2 h later for determination of secreted [ $^3\text{H}$ ]apo B and total  $^3\text{H}$ -protein.

**Apo B uptake experiments.** In our experiments to examine uptake of lipoproteins by HepG2 cells directly, we prepared  $^3\text{H}$ -labeled nascent lipoproteins,  $^{125}\text{I}$ -labeled LDL, and  $^{125}\text{I}$ -labeled monoclonal anti-LDL receptor IgG.  $^3\text{H}$ -labeled nascent lipoproteins were prepared by incubating one set of HepG2 cells with [ $^3\text{H}$ ]leucine in serum-free medium overnight. Media were harvested, pooled, and the  $d < 1.063$  g/ml fraction was isolated (39), without the addition of carrier lipoproteins. Excess salt and [ $^3\text{H}$ ]leucine were removed by dialysis. By electrophoresis in sodium dodecyl sulfate, autoradiography, and laser densitometry, the distribution of label among the apoproteins was 73.2% apo B, 3.7% apo E, and 23.1% apo AI. These labeled nascent lipoproteins were placed onto a second set of HepG2 cells at 5  $\mu\text{g}$  of lipoprotein protein/ml, with or without LpL, and incubated for 5 h. Incubations were performed in the presence of 10 mM unlabeled leucine, to reduce recycling of label (40). Cellular removal of labeled nascent lipoproteins from the media was assessed by measuring TCA-precipitable  $^3\text{H}$  (i.e.,  $^3\text{H}$ -apoproteins) remaining in the media and by measuring total  $^3\text{H}$ -radioactivity accumulated by the cells.

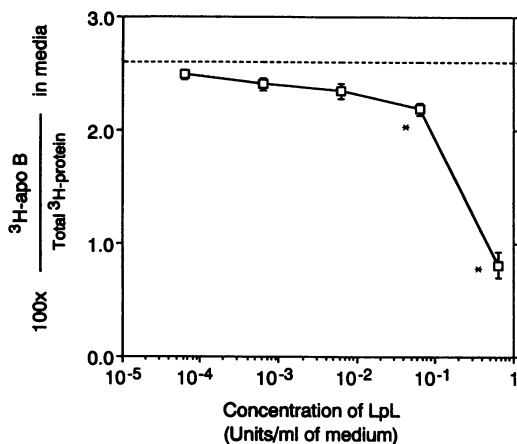
$^{125}\text{I}$ -Labeled LDL and  $^{125}\text{I}$ -labeled anti-LDL receptor IgG were prepared by radioiodination by the iodine monochloride method (41, 42). The LDL was from a fasted, normolipidemic woman. The anti-LDL receptor IgG was produced from a hybridoma obtained from the American Type Culture Collection, Rockville, MD (catalogue no. CRL 1691) (43, 44). The two labeled proteins were placed onto HepG2 cells at a concentration of 18 nM, and incubated for 5 h with or without LpL. Incubations in wells without cells were conducted in parallel. Cellular uptake of  $^{125}\text{I}$ -proteins was assessed as  $^{125}\text{I}$ -radioactivity in cells after scraping and rinsing. Cellular degradation of  $^{125}\text{I}$ -proteins was assessed as TCA-soluble, chloroform-insoluble  $^{125}\text{I}$ -radioactivity in the media (42). Degradation in cell-free wells was  $< 3\%$  of the degradation in the presence of cells, and was unaffected by LpL.

**Statistics.** Results are given as mean  $\pm$  SEM. Statistical comparisons were performed by Student's two-tailed  $t$  test.

## Results

Our preliminary experiments indicated that LpL reduced the net secretory output of [ $^3\text{H}$ ]apo B from HepG2 cells to  $22.5 \pm 0.6\%$  of control ( $n = 4$ ,  $P < 0.00005$ ) (1). We sought to characterize this effect more completely.

Fig. 1 demonstrates that the output of [ $^3\text{H}$ ]apo B from

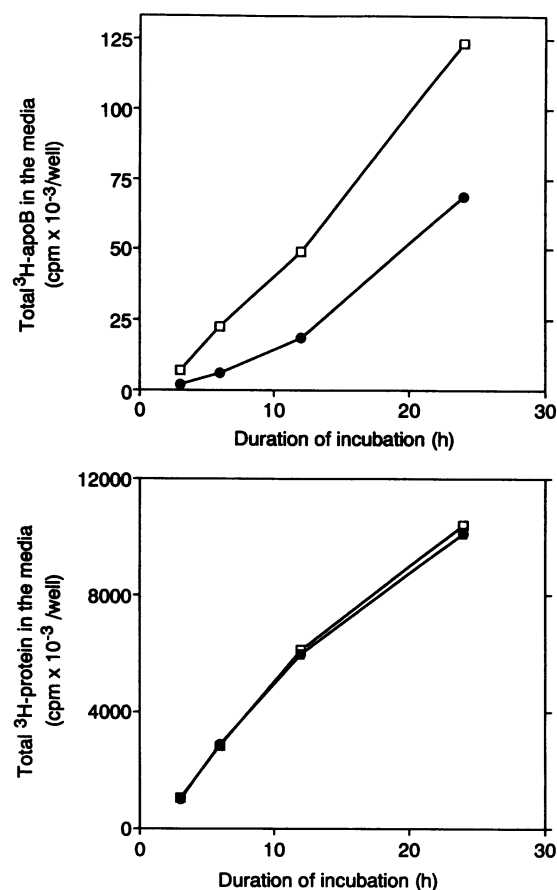


**Figure 1.** Dose response of LpL-mediated reduction in [ $^3\text{H}$ ]apo B output by HepG2 cells. HepG2 cells were incubated for 2 h in media supplemented with 75  $\mu\text{Ci}$  of [ $^3\text{H}$ ]leucine/ml, 0–0.640 U of bovine milk LpL per ml, and buffer to equalize volumes. Secreted [ $^3\text{H}$ ]apo B was purified from the  $d < 1.21$  mg/ml fraction of media by precipitation in 50% isopropanol. Total secreted  $^3\text{H}$ -protein was purified from media by TCA/PTA precipitation. The dashed line indicates cellular output of apo B in the absence of added LpL. The solid line indicates output in the presence of the indicated concentrations of LpL activity. Displayed are means  $\pm$  SEM,  $n = 4$ . Asterisks indicate outputs significantly different from the no-LpL control ( $P < 0.01$ ). Cellular output of total  $^3\text{H}$ -protein was not significantly affected by LpL.

HepG2 cells during a 2-h incubation is dose responsive to the addition of increasing amounts of LpL. At 0.064–0.640 U LpL/ml, the reduction in [ $^3\text{H}$ ]apo B output was large and statistically significant. At low concentrations of added LpL, there was a consistent trend toward reduced [ $^3\text{H}$ ]apo B output. Cellular output of total  $^3\text{H}$ -protein was not significantly affected at any LpL concentration.

The top panel of Fig. 2 shows the time course of [ $^3\text{H}$ ]apo B output in the presence and absence of added LpL. The reduction in output caused by addition of LpL was rapid in onset and sustained throughout the 24-h incubation. The degree of reduction was fairly constant for the first 12 h after addition of LpL. Between 12 and 24 h, however, the effect of LpL on the apo B secretory rate was somewhat less pronounced. This might be the result of loss of enzymatic activity with time or suppression of cellular LDL receptors. As before, LpL had no effect on the secretory output of total  $^3\text{H}$ -protein (Fig. 2, bottom).

To determine the mechanism of the reduction in apo B output, we directly blocked reuptake by the HepG2 cells of their own secreted particles (Fig. 3). Blockage of receptor-mediated uptake with heparin abolished the ability of LpL to reduce apo B output. In the absence of heparin, the output of apo B from LpL-supplemented cells was  $56.3\% \pm 1.5\%$  less than the output from nonsupplemented cells. In the presence of heparin, however, the output from LpL-supplemented cells was slightly higher than the output from nonsupplemented cells, though not significantly. In separate studies, we found that this concentration of heparin had no inhibitory effect on the enzymatic activity of our preparations of LpL, consistent with several prior reports (reviewed in reference 24). Thus, these results indicate that essentially all of the difference in apo B output caused by LpL depends on receptor-mediated reuptake. With

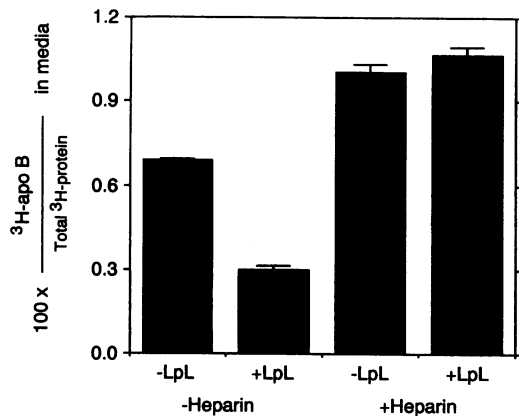


**Figure 2.** Time course of cellular output of [ $^3\text{H}$ ]apo B and total  $^3\text{H}$ -protein in the presence and absence of LpL. At  $t = 0$  h, HepG2 cells were given [ $^3\text{H}$ ]leucine, with (solid symbols) or without (open symbols) 0.640 U LpL/ml. Media and cells were harvested at the indicated times. The top panel displays the outputs of [ $^3\text{H}$ ]apo B, which was purified from media by immunoprecipitation, and its identity verified by polyacrylamide gel electrophoresis and autoradiography. The bottom panel displays the outputs of total secreted  $^3\text{H}$ -protein, which was purified from media by TCA/PTA precipitation. Means are shown,  $n = 4$ . SEMs were smaller than the graphic symbols. The output of [ $^3\text{H}$ ]apo B at each time point was significantly reduced by LpL ( $P < 0.00005$ ). Outputs of  $^3\text{H}$ -protein were unaffected by LpL.

or without LpL, heparin raised the net cellular output of [ $^3\text{H}$ ]apo B to 46–55% above the no-LpL, no-heparin control. This result confirms that there is substantial reuptake of nascent lipoproteins even in the absence of added LpL, consistent with previous results (2). In the presence of LpL, our results indicate that most apo B exported across the plasma membrane is taken back up (compare second and fourth columns in Fig. 3).

To verify directly that LpL enhances uptake of nascent lipoproteins by HepG2 cells, we incubated HepG2 cells for 5 h with  $^3\text{H}$ -labeled nascent lipoproteins, in the presence or absence of added LpL. The loss of TCA-precipitable radioactivity from the medium tripled, from  $15.2\% \pm 0.7\%$  without LpL to  $48.7\% \pm 0.3\%$  with LpL ( $n = 5$ ,  $P < 0.00005$ ) (Fig. 4, top). Lipoprotein lipase caused a corresponding threefold increase in the cellular accumulation of radioactivity (Fig. 4, bottom).

Finally, we sought to determine if LpL-induced enhancement of cellular uptake of lipoproteins results from alterations



**Figure 3.** Effect of heparin and LpL on the output of newly synthesized apo B from HepG2 cells. HepG2 cells were incubated for 2 h with [ $^3\text{H}$ ]leucine, with or without LpL (0.640 U/ml), with or without heparin (20 mg/ml), as indicated. [ $^3\text{H}$ ]apo B was then isolated from media by isopropanol precipitation; essentially identical quantitative results were obtained in a separate experiment using apo B immunoprecipitation.  $^3\text{H}$ -protein was isolated by TCA/PTA precipitation. Displayed are outputs of labeled apo B as a percentage of total labeled secreted protein. Means  $\pm$  SEM are shown,  $n = 5$ . The three columns to the right are each significantly different from the leftmost column ( $P < 0.00005$ ). The two columns labeled "+ Heparin" are not significantly different from each other.

in the lipoproteins or in cellular LDL receptors. Two ligands for the LDL receptor were used: mature plasma LDL and a monoclonal anti-LDL receptor IgG (43, 44). LDL is a substrate for LpL (5), whereas the IgG is not. Both ligands were radioiodinated, then separately incubated with HepG2 cells for 5 h, with and without LpL.

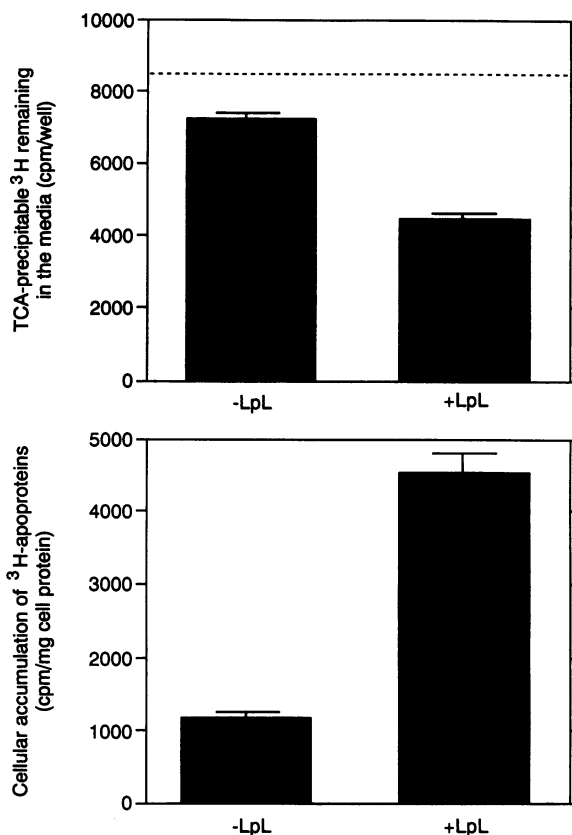
Lipoprotein lipase substantially increased uptake and degradation of  $^{125}\text{I}$ -LDL by HepG2 cells (cf. reference 5), but had no effect on uptake or degradation of the  $^{125}\text{I}$ -labeled anti-LDL receptor IgG (Fig. 5). Thus, LpL enhances uptake of lipoproteins by altering the lipoproteins, without altering cellular receptors, consistent with previous reports (5, 11).

## Discussion

Our results indicate that LpL substantially reduces the net secretory output of apo B from HepG2 cells. This reduction is primarily due to an enhancement of cellular reuptake of newly secreted lipoproteins. In the presence of LpL, the vast majority of apo B exported out of the cell is taken back up. The enhancement of reuptake is the result of alterations in the lipoproteins that lead to enhanced cell-surface binding, similar to effects previously described for apo B-rich lipoproteins from plasma (5–10).

The direct physiologic importance of our findings depends on the presence of LpL in the liver, particularly near sites of lipoprotein secretion and LDL receptor expression. Although LpL has generally been regarded as an extrahepatic enzyme, recent work indicates that it can be present in the liver in large amounts (reviewed in reference 23), including in adult mammalian liver (23, 30, 45–48).

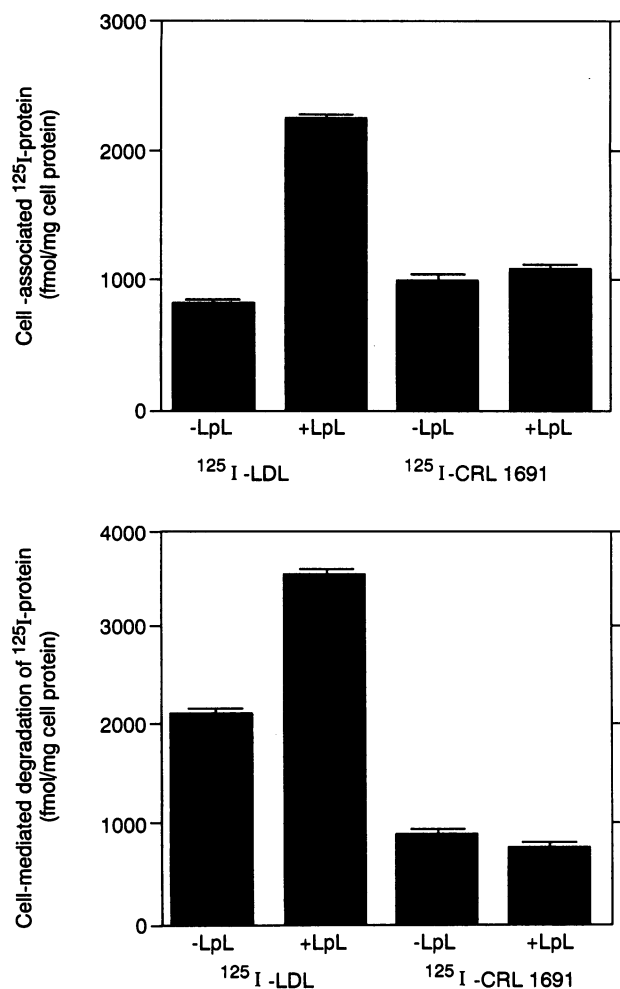
There are two possible sources for intrahepatic LpL. First, it could be synthesized locally, either by hepatic parenchymal cells (49–51) or by macrophages (52, 53) within the liver. The second possibility, for which more evidence exists in the adult



**Figure 4.** Effect of LpL on cellular uptake of nascent lipoproteins from the bulk fluid phase of media. Labeled, nascent lipoproteins (5  $\mu\text{g/ml}$ ) were incubated with HepG2 cells, with or without LpL (0.640 U/ml), as indicated. After 5 h, media and cells were harvested. The top panel displays TCA/PTA-precipitable radioactivity remaining in the media. The dashed line indicates TCA/PTA-precipitable radioactivity in the media before the 5-h incubation. The bottom panel displays cellular accumulation of  $^3\text{H}$  from labeled apoproteins. Means  $\pm$  SEM are shown,  $n = 5$ . Average cellular protein mass was 0.973 mg per 35-mm well.

mammal, is that LpL is transported from its major sites of synthesis in extrahepatic tissues to the liver. Hepatic uptake of LpL from the circulation has been directly demonstrated (30, 54), and this LpL remains enzymatically active intrahepatically (55). Transport from extrahepatic tissues to the liver can be enhanced by a fatty meal (30) or by injection of Intralipid (23, 30, 48). Most importantly, intrahepatic LpL is located along sinusoidal endothelial cells and on the microvillus extensions of hepatocytes into the space of Disse (23). Both sites are in intimate contact with nascent lipoproteins, which are secreted from the basal surface of hepatocytes into the space of Disse (56). Both sites are also near LDL receptors, which are expressed on the basal surface of hepatocytes (57) and on endothelial cells (58, 59). Some LpL was also found within hepatocytes (23), raising the speculative possibility of intracellular remodeling of nascent lipoproteins (cf. reference 60).

The physiologic importance of our findings also requires that LpL have similar effects on nascent lipoproteins from HepG2 cells and from human livers. HepG2 cells secrete apo B primarily on a triglyceride-rich particle that has the same density as LDL (61, 62), whereas human livers secrete primarily VLDL, with some "direct" secretion of LDL (63, 64). All of



**Figure 5.** LpL increases uptake and degradation of mature plasma LDL, but not of a monoclonal anti-LDL receptor antibody (CRL 1691). Both ligands were radioiodinated, then separately incubated with HepG2 cells for 5 h, with or without added LpL (0.640 U/ml), as indicated. The top panel displays cellular uptake of  $^{125}\text{I}$ -protein. The bottom panel displays cell-specific degradation of  $^{125}\text{I}$ -protein. Means  $\pm$  SEM are shown,  $n = 4$ . Uptake and degradation of  $^{125}\text{I}$ -LDL were significantly increased by LpL ( $P < 0.001$  and  $P < 0.0001$ , respectively). Uptake and degradation of  $^{125}\text{I}$ -CRL 1691 were unaffected by LpL. Average cellular protein mass was 0.963 mg per 35-mm well.

these apo B-rich lipoproteins show large increases in their uptake by cells in the presence of LpL (references 5 and 6; Figs. 4 and 5). Because LpL has been shown to enhance cellular uptake of virtually every apo B-rich lipoprotein (5–10), we expect that net hepatic secretion of almost any type of apo B-rich particle could be reduced by LpL in vivo.

Partial LpL deficiency has been associated with some cases of familial combined hyperlipidemia (25, 26), a disease of hepatic apo B overproduction. It was hypothesized that the hyperlipidemia may arise in these patients because of impaired lipoprotein removal from the circulation (25), contrary to the usual finding of overproduction in kinetic studies (27). Our results suggest an alternative explanation: partial LpL deficiency would result in incomplete remodeling of nascent lipoproteins in and near the space of Disse and therefore cause impaired reuptake, which would appear in kinetic studies as overproduction (1). Thus, remodeling of nascent lipoproteins by LpL

within the space of Disse to enhance reuptake may be a normal physiologic mechanism for regulating secretory output of apo B. It may also be subject to pharmacologic manipulation. For example, fibric acid compounds induce LpL (65–68) and decrease VLDL apo B secretion (69–72). To discern between net hepatic overproduction (1) and reduced peripheral clearance (25) as explanations for hyperlipidemia in patients with partial LpL deficiency, it will be important to measure their net hepatic apo B production.

Lipoprotein lipase deficiency was reported to account for only one-third of cases of familial combined hyperlipidemia (26). The other two-thirds may have abnormalities in other factors known to affect lipoprotein binding to receptors (5–22) or in factors known to affect LpL (24, 73). Of particular interest are cholesteryl ester transfer protein, which can change the neutral lipid composition of nascent lipoproteins (38) and might therefore alter receptor affinity (11, 14), and apoproteins CIII and AIV, which inhibit apo E-mediated binding (20, 21) and modulate LpL activity (73, 74). An association between apo AI restriction fragment length polymorphisms and familial combined hyperlipidemia has been reported (75, 76), which could reflect abnormalities in the closely linked genes for apo CIII and apo AIV (75–77).

The adaptive value of rapid remodeling and reuptake of nascent lipoproteins may, like other so-called futile cycles, involve regulation. Cholesterol enrichment of HepG2 cells results in increased secretion of apo B-rich particles (2, 3) through receptor suppression and inhibition of reuptake (2). Delivery of LpL and lipids to the liver during postprandial hyperlipidemia could alter hepatic output of apo B-rich lipoproteins through altered reuptake. Additionally, transfer and exchange of material onto nascent lipoproteins before reuptake may provide a means for importation of nutrients (cf. reference 38).

Overall, our findings indicate that reuptake of newly exported material can be affected by local, extracellular processing of the secreted ligand (cf. reference 2). There are other ligands besides apo B-rich lipoproteins that can be remodeled to alter cellular uptake. For example, if specific secreted glycoproteins are desialated within the space of Disse, re-uptake by the hepatic asialoglycoprotein receptor (78, 79) could occur, thereby affecting net output. Local reuptake, especially following local remodeling, may be a widespread phenomenon.

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This article is dedicated to the memory of Charles B. Hesler, Ph.D. (1954–1990), our able colleague and friend.

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