Cerebroventricular Calcitonin Gene–related Peptide Inhibits Rat Duodenal Bicarbonate Secretion by Release of Norepinephrine and Vasopressin

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Abstract

Proximal duodenal bicarbonate secretion is an important factor in humans and animals protecting the mucosa against acid-peptic damage. This study examined the mechanisms responsible for the central nervous system regulation of duodenal bicarbonate secretion by calcitonin gene–related peptide (CGRP) in unrestrained rats. Cerebroventricular administration of rat CGRP significantly inhibited basal duodenal bicarbonate secretion as well as the stimulatory effects of vasoactive intestinal peptide, neuropepsia, a luminal PGE1 analogue, misoprostol, and hydrochloric acid. The inhibitory effects of cerbroventricular CGRP were abolished by ganglionic blockade with chlorisondamine, significantly attenuated by noradrenergic blockade with bretylium, and enhanced by vagotomy. Inhibition of duodenal bicarbonate secretion induced by CGRP coincided with significant increases in plasma norepinephrine (NE) and vasopressin concentrations. The alpha adrenergic receptor antagonist, phentolamine, and the vasopressin V1 receptor antagonist, (1-deaminopenicillamine, 2-[O-methyl][Tyr, 8-Arg]-vasopressin, given intravenously reversed the central inhibitory effect of CGRP by ~50% each. Pretreatment of the animals with both phentolamine and the vasopressin antagonist completely abolished the central inhibitory effect of CGRP. Peripheral vasopressin and NE significantly decreased duodenal bicarbonate secretion, and their inhibitory effects were additive and prevented by phentolamine and the vasopressin antagonist, respectively. We conclude that cerebroventricular CGRP inhibits rat duodenal bicarbonate secretion by activation of sympathetic efferents and subsequent release of NE and vasopressin that act on alpha adrenergic and vasopressin receptors, respectively. (J. Clin. Invest. 1990. 85:25–32.)

Introduction

Duodenal bicarbonate secretion protects the mucosa against acid-peptic damage in humans and animals (1–4). Diminished proximal duodenal bicarbonate secretion at rest and in response to hydrochloric acid appears to be an important factor in the pathogenesis of duodenal ulcer disease (5). Like most gut secretory processes, duodenal bicarbonate secretion is controlled by the autonomic nervous system. Stimulation of parasympathetic (vagal) efferents increases while stimulation of sympathetic (splanchnic) efferents decreases duodenal bicarbonate secretion (6, 7). Corticotropin-releasing factor is an endogenous central nervous system transmitter that mediates stimulation of duodenal bicarbonate secretion during stress by release of β-endorphin from the pituitary (8). Thyrotropin-releasing hormone is a candidate central nervous system transmitter that stimulates gastric acid and duodenal bicarbonate secretion by increasing vagal outflow (9). The central nervous system transmitter(s) that mediates inhibition of duodenal bicarbonate secretion in situations of increased noradrenergic activity is unknown.

We have recently screened 10 neuropeptides and tested their central nervous system effects on proximal duodenal bicarbonate secretion in awake, nonrestrained rats (10). Calcitonin gene–related peptide (CGRP; 11) was the only peptide that significantly inhibited proximal duodenal bicarbonate secretion (10). This peptide selectively stimulates noradrenergic sympathetic outflow (12) and potently inhibits gastric acid secretion and gastrointestinal transit in rats and dogs (13–19). The purposes of the present investigation were threefold: (a) to examine the pharmacological properties of CGRP to inhibit resting and stimulated duodenal bicarbonate secretion in awake, freely moving rats; (b) to determine the pathways that are involved in mediating the central nervous system effects of CGRP; and (c) to study the peripheral effects of presumed transmitters released by CGRP. Our results indicate that cerebroventricular CGRP inhibits resting and stimulated duodenal bicarbonate secretion by stimulation of noradrenergic sympathetic outflow and vasopressin release. They also demonstrate that the inhibitory effects of peripheral NE and vasopressin on bicarbonate secretion are synergistic and mediated by specific alpha adrenergic and vasopressin receptors.

Methods

Male Sprague-Dawley rats (250–300 g) were used. The animals were housed in separate cages under illumination-, temperature-, and humidity-controlled conditions, and fed a standard rat diet before and a liquid diet after surgery. 4 to 7 d before the first experiment the proximal duodenum was isolated under general anesthesia (ketamine-HCl, 50 mg/kg; Ketanest [Parke Davis & Co., Berlin, FRG]; and xylazine, 6 mg/kg [Rompun, Bayer AG, Leverkusen, FRG]). Paying particular attention to the integrity of surrounding pancreatic tissue and blood supply, the first 2 cm of the duodenum (proximal to the pancreatic duct) were isolated and the pylorus anastomosed to the distal duodenum by seven to eight single sutures. The proximal and distal ends of the isolated duodenal segment were connected with polyethylene catheters (PE-90) and secured by purse string sutures. The catheters then were routed subcutaneously to exit in the interscapular region of

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1. Abbreviations used in this paper: CGRP, calcitonin gene–related peptide; VIP, vasoactive intestinal peptide.
the animal’s neck. They were closed with cut needle ends that served as connectors for catheters used to perfuse the duodenal segment and to collect duodenal perfusates. After surgery the segment was flushed every 8 h with 5 ml of 0.15 M NaCl to remove duodenal mucus and maintain patency of the segment and catheters (8).

During a second surgery (2–4 d before the experiment and 3–5 d after the intestinal surgery), a 22-gauge stainless steel cannula (Plastic Products Co., Roanoke, VA) was implanted using a stereotaxic instrument (David Kopf Instruments, Tujunga, CA) so that its tip was inside the right cerebral ventricle (20). In each animal, correct cannula placement was verified by injection of methylene blue (20). The animals were also fitted with a right jugular venous catheter (PE-50) that was routed subcutaneously to exit in the interscapular region of the animal’s neck, where it was closed by a cut needle end and secured with adhesive tape (20). In some animals, bilateral, truncal, subdiaphragmatic vagotomy, or adrenalectomy were performed as previously described (20). Hypophysectomized animals were commercially obtained (Wiga, Sulzfeld, FRG). Completeness of adrenalectomy and hypophysectomy were documented by macroscopic inspection. To demonstrate completeness of vagotomy, a 2-deoxy-D-glucose test (100 mg/kg; 3 min; Sigma Chemie GmbH, Deisenhofen, FRG) was performed (20). Adrenalectomized and hypophysectomized animals had free access to 0.15 M NaCl.

On the day of the experiment the animals were placed into 15-liter buckets to which they were accustomed. They were fasted for 16 h but had free access to tap water until the beginning of the experiment. The cerebroventricular cannula was connected with a polyethylene catheter (PE-10), the jugular vein catheter with PE-50, and the duodenal catheters with PE-90. The duodenumal segment was constantly perfused with 0.15 M NaCl (39°C, 30 ml/h) and the effluent collected at 15-min intervals in graduated cone vials. To clear the segment from residual mucus and secretions, the first four collections were discarded (8). Rat CGRP and calcitonin (16) were injected cerebroventricularly or intravenously over 1 min in 10-μl and 0.5-ml volumes, respectively (16). Peptides were kindly provided by Dr. Jean E. Rivier (Salk Institute, La Jolla, CA), stored in lyophilized form, and freshly dissolved in sterile water before each experiment. The vehicle, sterile water, served as control.

Bicarbonate concentrations of the duodenal effluent were measured by a validated back-titration method for low bicarbonate concentrations (2, 8). Measurements were performed in triplicate. A 2-ml aliquot of the effluent was added to 5 ml of CO2-free H2O and acidified with 125 μl of 100 mM H2SO4. To remove any residual CO2, the solution was gassed for 5 min with N2 and washed with Ba(OH)2. The sample then was titrated by the addition of 15 mM NaOH under continuous gassing with N2 to pH 8.4 on an automated titration system (Autoburette ABU SO, Titratom TTT 80, pH meter PHM 82; Radiometer, Copenhagen, Denmark). On each test day the perfusates that served as blank and bicarbonate standards (0.65, 1.3, and 2.6 mM) were measured to assure reproducibility (8).

To ascertain that CGRP did not increase mucosal permeability, recovery studies with exogenous bicarbonate were performed. The duodenal segment was perfused with 22 mM HCO3 (30 ml/h) for 2 h (1,320 amol/2 h). CGRP (1 nmol) or control (sterile water) were given cerebroventricularly at the zero time point. The mean ±SEM bicarbonate outputs in control and CGRP-treated animals (n = 8) were 1.364 ± 1.4 and 1.331 ± 1.6 amol/2 h, respectively, and were not significantly different from one another. The amount of recovered bicarbonate that was greater than the amount of exogenously infused bicarbonate reflects active bicarbonate secretion in the presence and absence of central CGRP.

In dose-response studies the cerebroventricular effects of CGRP were tested on basal (resting) and stimulated duodenal bicarbonate secretion. Each dose (0, 0.01, 0.1, and 1 nmol/animal) was administered on a separate day and in random order. Bicarbonate secretion after intravenous injection of 0.15 M NaCl (0.5 ml) served as control. Stimulated bicarbonate secretion was measured in response to intravenous bolus injection of neurotensin or vasoactive intestinal peptide (VIP; 10 nmol/kg each) and in response to a 5-min duodenal perfusion with 0.15 M HCl or during continuous perfusion of the duodenal segment with a PG E2 analogue, (±) methyl(11α,13E)-11,16-dihydroxy-16-methyl-9-oxoprost-13-en-1-olate (misoprostol, 10−5 M; G. D. Searle, Skokie, IL). CGRP was given at the same time the intravenous or duodenal substances were administered. Pilot studies indicated that the greatest bicarbonate responses occurred between 30 and 45 min after injection of the intravenous or intraduodenal stimulants. This interval was used for dose-response studies.

To delineate the pathways that may be involved in mediating the central nervous system effects of CGRP on duodenal bicarbonate secretion, pharmacological approaches were used in addition to surgical vagotomy, adrenalectomy, and hypophysectomy. To determine the role of peripheral vasopressin, the V1 vasopressin receptor antagonist, (1-deamino-2-[[[4-arginine]-vasopressin (100 nmol·kg−1·h−1), was used (21). This antagonist was provided by Dr. Jean E. Rivier (Salk Institute). Alpha adrenergic receptor blockade was performed with phentolamine (1 μmol·kg−1·h−1; Regitin, Ciba-Geigy GmbH, Wehr, FRG). Preventing the release of NE from noradrenergic nerve endings was achieved with beryllium tosylate (25 mg·kg−1·h−1; Bertylom, American Critical Care, American Hospital Supply Corp., McGaw Park, IL) (17). Ganglion blockade of neuronal effectors was performed with chlorisondamine (3 mg·kg−1·h−1; Ecolid, Ciba-Geigy, Basel, Switzerland) (20). Opiate receptor blockade was achieved with naloxone (1 mg·kg−1·h−1; Sigma Chemie GmbH) (8). Intravenous infusion of these agents was begun 1 h before bicarbonate measurements and 2 h before cerebroventricular injection of CGRP. Except for alpha adrenergic blockade with phentolamine, these pharmacological approaches were previously validated (8, 9, 15, 17, 19, 20). The effectiveness of phentolamine in blocking the actions of NE on duodenal bicarbonate secretion was documented in this study.

Dose-response studies were performed with NE (5, 25, and 50 nmol/kg; Arterenol, Hoechst AG, Frankfurt, FRG) and (8-arginine)-vasopressin (0.05, 0.25, and 0.5 nmol/kg; provided by Dr. Jean E. Rivier). Each dose was given intraperitoneally on a separate day and in random order and the 30–45 min interval after injection (interval of greatest response) was used for analysis. In some experiments NE and vasopressin were given concomitantly. In other experiments phenolamine or the vasopressin antagonist were given intravenously as constant infusions before intraperitoneal administration of NE or vasopressin.

Venous blood was aspirated at times 0, 15, and 30 min for determination of plasma concentrations of epinephrine and NE (0.6 ml/sample) and vasopressin (1.2 ml/sample) in separate groups of animals. In additional experiments vasopressin (0.5 nmol/kg) and NE (50 nmol/kg) were injected intraperitoneally and plasma concentrations of vasopressin and NE were determined before and 30 min after injection. Blood was replaced by equal volumes of 0.15 M NaCl. Blood was collected in chilled tubes, centrifuged at 3,000 g (4°C), and stored at −20°C. Plasma catecholamine concentrations were determined by a single isotope radioenzymatic method (22). [35S]-methionyl-T-methionine was purchased from New England Nuclear (Boston, MA). Plasma vasopressin concentrations were determined by RIA using the antiserum kindly provided by Dr. John D. Fernstrom (University of Pittsburgh, Pittsburgh, PA) (23). Plasma was extracted as previously described (24). The minimal detection of this assay was 250 pg/tube and the intraassay variation was 3%. Catecholamine and vasopressin measurements were performed in duplicate and in single assays.

The data were subjected to analysis of variance and differences between treatment groups were determined by the Neuman-Keuls multiple range test (25). Results obtained from six to eight animals in each group were expressed as means ±SE and considered significant if P < 0.05.

**Results**

Intravenous bolus injection of neurotensin and VIP, as well as 5-min duodenal perfusion with the PG E2 analogue, miso-
prostaglandin E (PGE), analogues, misoprostol (dmpG-E), was given intraduodenally as a constant infusion, and VIP, neuropeptide Y (NPY), and control (0.15 M NaCl) were given intravenously. CGRP or the vehicle (zero dose) were given cerebroventricularly at the time intraduodenal or intravenous stimuli were administered. The bicarbonate response was measured during the 30–45-min interval (interval of maximal response) after cerebroventricular injection. Doses of CGRP were given in random order and on separate days. *P < 0.05; **P < 0.01 compared with the zero dose.

**Table 1. Peripheral Effects of CGRP and Calcitonin on Duodenal Bicarbonate Secretion**

<table>
<thead>
<tr>
<th>HCO₃⁻</th>
<th>0 min</th>
<th>15 min</th>
<th>30 min</th>
<th>45 min</th>
<th>60 min</th>
<th>75 min</th>
<th>90 min</th>
<th>105 min</th>
<th>120 min</th>
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<tbody>
<tr>
<td></td>
<td>μmol/cm h</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Control</td>
<td>11.5±0.4</td>
<td>11.5±0.5</td>
<td>11.9±0.6</td>
<td>12.0±0.6</td>
<td>12.1±1.0</td>
<td>12.1±1.0</td>
<td>12.6±0.8</td>
<td>12.4±0.7</td>
<td>12.2±0.8</td>
</tr>
<tr>
<td>CGRP</td>
<td>11.7±0.6</td>
<td>8.3±0.8*</td>
<td>10.7±0.6</td>
<td>10.9±0.6</td>
<td>11.4±0.8</td>
<td>11.8±0.8</td>
<td>11.2±0.7</td>
<td>11.9±0.9</td>
<td>11.7±0.9</td>
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<tr>
<td>Calcitonin</td>
<td>11.4±0.7</td>
<td>11.3±0.9</td>
<td>11.3±0.8</td>
<td>11.3±0.9</td>
<td>11.6±0.9</td>
<td>11.9±0.9</td>
<td>11.3±0.5</td>
<td>10.4±0.5</td>
<td>10.1±0.6</td>
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CGRP and calcitonin (1 nmol each) were given intravenously at time zero. *P < 0.01 compared with the corresponding time point of control.

**Figure 1.** Central nervous system effects of CGRP on proximal duodenal bicarbonate secretion in freely moving rats. HCl was given intraduodenally for 5 min, a synthetic PG E, analogues, misoprostol (dmpG-E), was given intraduodenally as a constant infusion, and VIP, neuropeptide Y (NPY), and control (0.15 M NaCl) were given intravenously. CGRP or the vehicle (zero dose) were given cerebroventricularly at the time intraduodenal or intravenous stimuli were administered. The bicarbonate response was measured during the 30–45-min interval (interval of maximal response) after cerebroventricular injection. Doses of CGRP were given in random order and on separate days. *P < 0.05; **P < 0.01 compared with the zero dose.
tylum did not (Fig. 3). Chlorisondamine completely abolished and bretylium significantly ($P < 0.01$) attenuated inhibition of duodenal bicarbonate secretion induced by cerebroventricular CGRP (Fig. 3, A and B). In contrast, truncal vagotomy did not attenuate the inhibitory effect of CGRP (Fig. 3 C). The 2-h total bicarbonate outputs in response to CGRP expressed as a percentage of the total 2-h bicarbonate outputs in response to the respective controls were: bretylium 84±5% and vagotomy 46±3%. These responses were significantly ($P < 0.01$) different from the response produced by CGRP in NaCl-treated animals (63±5%) (Fig. 2 A).

As depicted in Figs. 2 and 3, phentolamine, chlorisondamine, and truncal vagotomy altered the baseline of resting duodenal bicarbonate secretion. To compare the relative effects of these pharmacological and surgical procedures on CGRP-induced inhibition of duodenal bicarbonate secretion, the effects of CGRP were expressed as a percentage of the bicarbonate responses of the appropriate control experiments. The maximal inhibitory effect of CGRP was observed during the third collection interval (30–45 min) after cerebroventricular administration. Therefore, this interval was chosen for comparison. In control animals (no pharmacological pretreatment or surgery) CGRP decreased the bicarbonate response to 37±5% (Fig. 4). Vagotomy significantly ($P < 0.05$) enhanced

![Figure 2](https://doi.org/10.1172/JCI114420)

**Figure 2.** Effects of a vasopressin receptor antagonist (AVP-Ant) and adrenergic receptor blockade (phentolamine) on CGRP-induced inhibition of duodenal bicarbonate secretion. Intravenous infusions with 0.15 M NaCl (A), AVP-Ant (100 nmol/kg·h) (B), phentolamine (1 µmol/kg·h) (C), and AVP-Ant + phentolamine (D) were begun 2 h before cerebroventricular administration of CGRP (1 nmol) or control, indicated by the arrow. Note the differences in the scale of the vertical ordinates between the upper and lower panels. *$P < 0.05$; **$P < 0.01$ compared with the corresponding time points of control.

![Figure 3](https://doi.org/10.1172/JCI114420)

**Figure 3.** Effects of ganglionic blockade (chlorisondamine), noradrenergic blockade (bretylium), and truncal vagotomy on CGRP-induced inhibition of duodenal bicarbonate secretion. Intravenous infusions with chlorisondamine (3 mg/kg·h) (A) and bretylium (25 mg/kg·h) (B) were begun 2 h before cerebroventricular administration of CGRP (1 nmol) or control indicated by the arrow. *$P < 0.05$; **$P < 0.01$ compared with the corresponding time points of control.

![Figure 4](https://doi.org/10.1172/JCI114420)

**Figure 4.** Relative inhibitory effects of cerebroventricular CGRP (1 nmol) on duodenal bicarbonate secretion in control experiments (0.15 M NaCl infusion) and during autonomic nervous system and vasopressin receptor blockade. CGRP, control, and blocking agents were administered at times indicated in the legends to Figs. 2 and 3. The bicarbonate responses measured at the 30–45 min interval in response to CGRP were expressed as a percentage of the bicarbonate responses measured at the 30–45 min interval in response to the vehicle, sterile water. *$P < 0.05$; **$P < 0.01$ compared with control.

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the inhibitory effect of CGRP. Bretylium, phenolamine, and the vasopressin receptor antagonist significantly ($P < 0.01$) reversed the inhibitory effect of CGRP (Fig. 4). Combined treatment with phenolamine and the vasopressin receptor antagonist or ganglionic blockade with chlorisondamine reversed the inhibitory effect of cerebroventricular CGRP on duodenal bicarbonate secretion (Fig. 4).

Hypophysectomy, adrenalectomy, and opiate blockade with naloxone did not significantly alter resting bicarbonate secretion or attenuate the inhibitory effects of CGRP (Table II). Cerebroventricular administration of CGRP (1 nmol) significantly increased plasma concentrations of vasopressin and NE (Fig. 5) but not of epinephrine (data not shown).

Intraperitoneal administration of increasing doses of vasopressin and NE produced dose-dependent inhibition of duodenal bicarbonate secretion (Fig. 6). Of note, on a molar basis vasopressin appeared to be ~ 100 times more potent and effective than NE in inhibiting bicarbonate secretion in vivo. The vasopressin receptor antagonist and the alpha adrenergic antagonist, phenolamine, abolished the effects of exogenous vasopressin and NE on bicarbonate secretion, respectively (Fig. 6). Intraperitoneal administration of vasopressin (0.25 nmol/kg) and NE (25 nmol/kg) given concomitantly inhibited duodenal bicarbonate secretion by 63±4%, while vasopressin and NE given separately inhibited the bicarbonate response by 33±2 and 29±3%, respectively (Fig. 6). Combined treatment with intraperitoneal vasopressin and NE produced inhibition of duodenal bicarbonate secretion that was similar to the response observed after a maximal dose of cerebroventricular CGRP (5 nmol/kg) (Fig. 6). Pretreatment of the animals with the vasopressin receptor antagonist and phenolamine completely reversed the inhibitory effect produced by intraperitoneal administration of vasopressin and NE (Fig. 6).

To evaluate if the exogenously administered doses of vasopressin and NE produced similar plasma concentrations that were observed after cerebroventricular administration of CGRP, the highest doses that were used in the bioassay were given intraperitoneally. The highest doses that were used in the bioassay were given intraperitoneally. The highest doses that were used in the bioassay were given intraperitoneally. The highest doses that were used in the bioassay were given intraperitoneally. The highest doses that were used in the bioassay were given intraperitoneally.

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<th>Table II. Inhibition of Duodenal Bicarbonate Secretion by CGRP: Role of the Pituitary, Adrenals, and Opiate Receptors</th>
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<tr>
<td></td>
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<tr>
<td>HCO3⁻</td>
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<tr>
<td>NaCl</td>
</tr>
<tr>
<td>Hypophysectomy</td>
</tr>
<tr>
<td>Adrenalectomy</td>
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<tr>
<td>Naloxone</td>
</tr>
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</table>

Bicarbonate secretion was measured 30–45 min after cerebroventricular administration of CGRP (1 nmol) or control. Naloxone infusion (1 mg/kg·h) was begun 2 h before cerebroventricular injection. *P < 0.01 compared with control.

Figure 5. Effects of cerebroventricular administration of CGRP (1 nmol) and control (vehicle) on plasma concentrations of NE and vasopressin. *P < 0.05; **P < 0.01 compared with the corresponding time points of control.

Discussion

Like most gut secretory processes, duodenal mucosal bicarbonate secretion is under the control of the autonomic nervous system. Stimulation of parasympathetic (vagal) efferents increases, while stimulation of sympathetic (splanchnic) efferents decreases duodenal bicarbonate secretion in a number of species (6, 7, 26, 27). The central nervous system transmitters that promote these responses are largely unknown. We have recently provided evidence that thyrotropin-releasing hormone is a candidate neurotransmitter that stimulates duodenal bicarbonate secretion by increasing vagal outflow (9). To the best of our knowledge, CGRP is the only oligopeptide to date that acts centrally to inhibit duodenal bicarbonate secretion (10). This study examined the central nervous system effects of CGRP on proximal duodenal bicarbonate secretion in a newly developed animal model that permits measurement of duodenal bicarbonate secretion in freely moving animals (8).

Intravenous administration of the gastrointestinal peptides neurotensin and VIP and intraluminal perfusion of the duodenum with a PGE₁ analogue and HCl increased bicarbonate secretion, extending previous observations in restrained or anesthetized rats (2, 28–30), conscious dogs (4), and humans (1). Intraluminal PGs and circulating neurotensin and VIP stimulate duodenal bicarbonate secretion by distinct mechanisms (4, 28–30) and intraluminal HCl stimulates bicarbonate secretion via a PG- and VIP-dependent mechanism (2, 30). Therefore, the peripheral transmitters (i.e., NE and vasopressin) released in response to cerebroventricular CGRP probably interfere with each of these mechanisms.

Cerebroventricular administration of CGRP produced dose-dependent and long-lasting inhibition of resting (basal) bicarbonate secretion, while intravenous administration of CGRP exhibited a transient inhibitory effect. The peripheral effect of CGRP on bicarbonate secretion may be secondary to the peptide's pronounced peripheral hypotensive effect since the marked decrease in mean arterial pressure in an earlier study (12) preceded the transient decrease in bicarbonate secretion observed in this study. Differences in the time course of
peptide action after central and peripheral administration also suggest differences in site of action; i.e., cerebroventricular CGRP acts within the central nervous system, while intravenous calcitonin acts at a peripheral, as yet unidentified site (15).

While the central nervous system effects of CGRP and calcitonin on gastric acid secretion and gastrointestinal transit are similar (16, 17, 19, 31), unlike CGRP, calcitonin does not inhibit duodenal bicarbonate secretion. This observation suggests that rat calcitonin is not a ligand for the cerebral CGRP receptor (32, 33) involved in mediating the peptide's effect on duodenal bicarbonate secretion. Distinct receptors for CGRP and calcitonin have also been implicated in mediating the inhibitory effects of these two peptides on gastric acid secretion and gastrointestinal transit (19, 31). However, the precise cerebral site of action of CGRP to inhibit duodenal bicarbonate secretion remains to be elucidated.

To determine the pathways that mediate the inhibitory effects of cerebroventricular CGRP surgical and pharmacological procedures were used. Ganglionic blockade and truncal vagotomy significantly decreased resting bicarbonate secretion, suggesting that parasympathetic efferents not only stimulate (6, 26, 27) but also exert a tonic effect on proximal duodenal bicarbonate secretion. In contrast, blockade of vasopressin and opiate receptors using specific antagonists, and preventing the release of NE from adrenergic nerve endings with bretylium, as well as adrenalectomy and hypophysectomy, did not significantly alter resting duodenal bicarbonate secretion, suggesting that none of these pathways is involved in the tonic control of duodenal bicarbonate secretion (8, 9). Of note, the nonselective alpha adrenoceptor antagonist phentolamine significantly increased resting bicarbonate secretion in this and in a previous study by an as yet unidentified mechanism (34). It is possible that phentolamine stimulates bicarbonate secretion either by acting directly on receptors of the mucosal cells or by vasodilation and increased splanchnic blood flow (35).

Most importantly, the results of this study demonstrate that CGRP given cerebroventricularly increases plasma concentrations of NE (12) and vasopressin, and that the release of both NE and vasopressin coincides with the inhibitory effect of CGRP on duodenal bicarbonate secretion. Phentolamine and the vasopressin antagonist reversed CGRP-induced inhibition of bicarbonate secretion by ~ 50% each, and both agents together abolished the inhibitory effect of CGRP. Furthermore, exogenous administration of vasopressin and NE resulting in plasma concentrations that were similar to or less than those observed after central administration of CGRP produced dose-dependent inhibition of bicarbonate secretion. Their combined inhibitory effects were additive and abolished by the vasopressin and alpha adrenergic receptor antagonists, respectively. These observations indicate that cerebroventricular CGRP inhibits duodenal bicarbonate secretion by release of vasopressin and NE, which exert their peripheral inhibitory effects on bicarbonate secretion via specific vasopressin and alpha adrenergic receptors, respectively. The results also imply synergism between vasopressin and NE as reflected by their additive inhibitory effects. Further studies using isolated organ or cell preparations are needed to determine the precise localization of vasopressin and noradrenergic receptors and their cellular interactions that are involved in regulating duodenal bicarbonate secretion.

In contrast to our findings in awake, nonrestrained rats, NE did not significantly alter duodenal bicarbonate secretion in anesthetized rats, but attenuated the stimulatory effect of phentolamine (34). The experimental design (i.e., state of anesthesia and in vitro back-titration in this study versus continuous pH stat titration and ~ 20 times lower doses of NE used in our experiments) may account for these different observations (34). The present findings demonstrating that noradrenergic blockade with bretylium attenuates inhibition of bicarbonate secretion produced by CGRP-induced increase in sympathetic noradrenergic outflow concurs with previous findings.

Table III. Plasma Vasopressin and NE Concentrations

<table>
<thead>
<tr>
<th>Vasopressin</th>
<th>NE</th>
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<tbody>
<tr>
<td></td>
<td>0 min</td>
</tr>
<tr>
<td>AVP + NE</td>
<td>8.0±0.7</td>
</tr>
<tr>
<td>Control</td>
<td>7.0±1.9</td>
</tr>
</tbody>
</table>

Effects of concomitant intraperitoneal administration of vasopressin (AVP; 0.5 nmol/kg) and NE (50 nmol/kg) or control (0.15 M NaCl) on plasma vasopressin and NE concentrations. Agents were given at 0 min. * P < 0.05 compared with the corresponding control.
that noradrenergic blockade with guanethidine abolished the inhibitory effect of splanchnic nerve stimulation (7).

The observation that ganglionic blockade with chlorisondamine abolished the inhibitory effect of CGRP indicates that autonomic efferents are involved. This finding also suggests that ganglionic blockade affects both the noradrenergic and vasopressin-sensitive pathways that mediate the inhibitory effects of CGRP. Of interest, hypophysectomy did not prevent the inhibitory effects of CGRP on bicarbonate secretion, suggesting that nonpituitary vasopressin is released by cerebroventricular administration of CGRP. A vasopressin-like peptide has been identified in the mammalian sympathetic nervous system, including in the principal noradrenergic neurones of ganglia and in nerve fibers innervating peripheral tissues (36). These findings support the contention that CGRP may corelease NE and vasopressin from sympathetic neuronal fibers rather than from the adrenal glands, since adrenalectomy did not alter the duodenal inhibitory effect of CGRP. However, the precise site of CGRP-induced vasopressin release remains to be determined.

Inhibition of gastric acid secretion and gastrointestinal transit induced by cerebroventricular administration of CGRP appears to be mediated by parasympathetic (vagal) efferent fibers and not by noradrenergic efferents (13, 17, 19). In this study vagotomy did not attenuate the inhibitory effect of CGRP on duodenal bicarbonate secretion. In addition, loss of vagal control further accentuated the inhibitory effect of CGRP, suggesting that removal of vagal transmitters facilitated inhibition of bicarbonate secretion induced by increased noradrenergic drive. Conversely, cerebroventricular corticotropin-releasing factor inhibits gastric acid secretion predominantly by increasing sympathetic outflow (20, 37), but modulates intestinal transit by altering parasympathetic outflow (38).

The physiological importance of CGRP increasing sympathetic outflow and thereby regulating duodenal bicarbonate secretion is unknown. Hemorrhagic shock decreases duodenal bicarbonate secretion and produces mucosal damage (39). Furthermore, hemorrhage-induced hyposecretion of duodenal bicarbonate is mediated by activation of sympathetic nerves (40). Therefore, it is tempting to speculate that under these conditions CGRP may serve as an endogenous central nervous system mediator.

In summary, the results of this study indicate that CGRP acts within the central nervous system to inhibit resting and stimulated proximal duodenal bicarbonate secretion in awake, nonrestained rats. CGRP exerts this action by enhancing sympathetic, noradrenergic outflow that results in release of NE and possibly nonpituitary vasopressin released from adrenergic nerve endings. NE and vasopressin inhibit duodenal bicarbonate secretion via specific noradrenergic and vasopressin receptors, respectively, and their inhibitory effects are additive.

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