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## Research Article

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# Relationship of Urinary and Blood Carbon Dioxide Tension during Hypercapnia in the Rat

Its Significance in the Evaluation of Collecting Duct Hydrogen Ion Secretion

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## **Abstract**

This study was designed to establish the relationship between urinary pCO2 and systemic blood pCO2 during acute hypercapnia and to investigate the significance of this relationship to collecting duct hydrogen ion (H+) secretion when the urine is acid and when it is highly alkaline. In rats excreting a highly alkaline urine, an acute increase in blood pCO2 (from 42±0.8 to 87±0.8 mmHg) resulted in a significant fall in urine minus blood (U-B) pCO<sub>2</sub> (from  $31\pm2.0$  to  $16\pm4.2$  mmHg, P < 0.005), a finding which could be interpreted to indicate inhibition of collecting duct H<sup>+</sup> secretion by hypercapnia. The urinary pCO<sub>2</sub> of rats with hypercapnia, unlike that of normocapnic controls, was significantly lower than that of blood when the urine was acid (58 $\pm$ 6.3 and 86 $\pm$ 1.7 mmHg, P < 0.001) and when it was alkalinized in the face of accelerated carbonic acid dehydration by infusion of carbonic anhydrase (78±2.7 and  $87\pm1.8$  mmHg, P < 0.02). The finding of a urinary pCO<sub>2</sub> lower than systemic blood pCO<sub>2</sub> during hypercapnia suggested that the urine pCO<sub>2</sub> prevailing before bicarbonate loading should be known and the blood pCO2 kept constant to evaluate collecting duct H<sup>+</sup> secretion using the urinary pCO<sub>2</sub> technique.

In experiments performed under these conditions, sodium bicarbonate infusion resulted in an increment in urinary pCO<sub>2</sub> (i.e., a ΔpCO<sub>2</sub>) which was comparable in hypercapnic and normocapnic rats ( $40\pm7.2$  and  $42\pm4.6$  mmHg, respectively) that were alkalemic (blood pH  $7.53\pm0.02$  and  $7.69\pm0.01$ , respectively). The U-B pCO<sub>2</sub>, however, was again lower in hypercapnic than in normocapnic rats (15 $\pm$ 4.0 and 39 $\pm$ 2.5 mmHg, respectively, P < 0.001). In hypercapnic rats in which blood pH during bicarbonate infusion was not allowed to become alkalemic (7.38 $\pm$ 0.01), the  $\Delta$ pCO<sub>2</sub> was higher than that of normocapnic rats which were alkalemic (70±5.6 and 42 $\pm$ 4.6 mmHg, respectively, P < 0.005) while the U-B pCO<sub>2</sub> was about the same (39±3.7 and 39±2.5 mmHg). We further examined urine pCO<sub>2</sub> generation by measuring the difference between the urine pCO<sub>2</sub> of a highly alkaline urine not containing carbonic anhydrase and that of an equally alkaline urine containing this enzyme. Carbonic anhydrase infusion to hypercapnic rats that were not alkalemic resulted in a fall in urine

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pCO<sub>2</sub> (from 122 $\pm$ 5.7 to 77 $\pm$ 2.2 mmHg) which was greater (P < 0.02) than that seen in alkalemic normocapnic controls (from 73 $\pm$ 1.9 to 43 $\pm$ 1.3 mmHg) with a comparable urine bicarbonate concentration and urine nonbicarbonate buffer capacity. CO<sub>2</sub> generation, therefore, from collecting duct H<sup>+</sup> secretion and titration of bicarbonate, was higher in hypercapnic rats than in normocapnic controls.

We conclude that in rats with acute hypercapnia, the U-B pCO<sub>2</sub> achieved during bicarbonate loading greatly underestimates collecting duct  $H^+$  secretion because it is artificially influenced by systemic blood pCO<sub>2</sub>. The  $\Delta$ pCO<sub>2</sub> is a better qualitative index of collecting duct  $H^+$  secretion than the U-B pCO<sub>2</sub>, because it is not artificially influenced by systemic blood pCO<sub>2</sub> and it takes into account the urine pCO<sub>2</sub> prevailing before bicarbonate loading.

#### Introduction

The carbon dioxide tension (pCO<sub>2</sub>) of a highly alkaline urine reaches a value substantially higher than that of systemic blood (1–8). Pitts and Lotspeich (1) proposed that the existence of an appreciable gradient between urine and blood pCO<sub>2</sub> was the consequence of distal H<sup>+</sup> secretion into bicarbonate-rich tubular urine in the face of delayed carbonic acid dehydration. They reasoned that, in the absence of carbonic anhydrase in the lumen of the distal nephron, carbonic acid would not completely dehydrate to carbon dioxide during the time that tubular fluid is in transit through the distal nephron. Completion of carbonic acid dehydration in the renal pelvis would ultimately increase urinary pCO<sub>2</sub> because the surface to volume relationship in this region is not favorable for CO<sub>2</sub> back diffusion (1).

It is now known from recent direct intrarenal pCO<sub>2</sub> determinations that during alkalinization of the urine a near maximal urinary pCO2 is achieved in the papillary collecting duct (9, 10). That the rise in urinary pCO<sub>2</sub> is an intrarenal rather than a postpapillary event, however, does not invalidate the classic notion that distal H<sup>+</sup> secretion is largely responsible for the high urinary pCO<sub>2</sub> achieved in a highly alkaline urine. Although other explanations have been offered to account for the development of a urine to blood pCO<sub>2</sub> gradient (3, 6, 7, 11-17), it is generally accepted that distal H<sup>+</sup> secretion largely accounts for this phenomenon (18-29). The recent finding of a significant disequilibrium pH in the papillary collecting duct under conditions of bicarbonate loading has been proposed as proof of the notion that H<sup>+</sup> secretion by the collecting duct is the most likely determinant of the urine minus blood (U-B)<sup>1</sup> pCO<sub>2</sub> gradient observed in an alkaline urine (9).

The U-B pCO<sub>2</sub> gradient achieved in response to bicarbon-

<sup>1.</sup> Abbreviations used in this paper: GFR, glomerular filtration rate; U-B, urine minus blood.

ate loading, however, may not be the ideal way to express the relationship between urinary pCO<sub>2</sub> generation and distal H<sup>+</sup> secretion (30). The use of the U-B pCO<sub>2</sub> as a marker of distal H<sup>+</sup> secretion assumes that blood pCO<sub>2</sub> and urine pCO<sub>2</sub> are about the same before alkalinization of the urine. There are situations, however, where the urinary pCO<sub>2</sub> can be lower than blood pCO<sub>2</sub> as in metabolic and respiratory acidosis (3, 6, 31-33). It also assumes that during bicarbonate loading medullary blood pCO<sub>2</sub> is similar to that of systemic blood. The latter assumption was supported by the findings of Uhlich et al. (34), which showed that during bicarbonate loading vasa recta blood pCO<sub>2</sub> was not substantially different from that prevailing in systemic blood (as determined by renal artery pCO<sub>2</sub>). Using a recently developed pCO<sub>2</sub> microelectrode (35– 39), DuBose (9), however, found that during bicarbonate loading the pCO<sub>2</sub> of the papillary collecting duct is equal to that of its adjacent vasa recta but is markedly higher than systemic blood pCO<sub>2</sub>. He suggested a role for trapping of CO<sub>2</sub> in the medullary countercurrent system to explain the similar values for pCO<sub>2</sub> in the papillary collecting duct and its adjacent vasa recta (9).

Although the contribution of CO<sub>2</sub> trapping (if any) to urinary pCO<sub>2</sub> generation during hypercapnia is unknown, there is no a priori reason to believe that systemic blood pCO<sub>2</sub> would reflect vasa recta pCO<sub>2</sub> in hypercapnic subjects excreting an alkaline urine. Moreover, it is now known that in rats with acute respiratory acidosis, inner collecting duct pCO<sub>2</sub> is lower than that of the blood (38, 39). Based on these considerations, we reasoned that the use of the U-B pCO<sub>2</sub> achieved during bicarbonate loading could be a poor marker of collecting duct H<sup>+</sup> secretion under conditions where systemic blood pCO<sub>2</sub> is altered. Accordingly, we examined urinary pCO<sub>2</sub> generation and U-B pCO<sub>2</sub> gradients in hypercapnic rats studied under conditions where the urine was acid and when it was alkalinized in the presence and in the absence of accelerated carbonic acid dehydration.

# **Methods**

Adult Sprague-Dawley rats (250-380 g) were allowed free access to food (Purina Lab Chow, Ralston Purina Co., St. Louis, MO) and water until the morning of each experiment. Studies using conventional clearance techniques were performed in anesthetized rats. Anesthesia was accomplished by the intraperitoneal administration of Inactin (100 mg/kg body weight). Tracheostomy was performed and one carotid artery and jugular vein were cannulated. Rats were connected to a rodent respirator (model 680, Harvard Apparatus Co., Inc., S. Natick, MA) to sustain a blood pCO<sub>2</sub> at the desired level. At the start of the experiment, <sup>125</sup>I-iothalamate diluted in saline (0.75 μCi/ml) was infused by an infusion pump at a rate of 0.024 ml/min throughout the course of the experiment as a marker of glomerular filtration rate (GFR). An equilibration period of 60 min was allowed before any collection was started. Urine samples were collected via a suprapubic catheter and drained into preweighed glass vials containing mineral oil; urine volume was determined gravimetrically. Blood samples were collected from the carotid artery during the middle portion of each clearance period.

Timed urine collections of  $\sim 20-30$  min duration were obtained in the following groups of rats: in group Ia, eight normocapnic rats were infused with sodium bicarbonate (0.9-M solution) at a rate of 6 ml/h. At least two clearance collections were obtained from each rat after a urine pH of 7.8 or higher was achieved. The first urine collection with a urine pH of 7.8 or higher was discarded to eliminate mixing of urine of different pH's. After completion of this protocol, acute hypercapnia was induced in the eight rats by making them inhale a mixture of 10% CO<sub>2</sub>, balance air via tracheal intubation. After 2 h of

stable hypercapnia (blood pCO<sub>2</sub> range, 80–100 mmHg), urine collections were obtained while sodium bicarbonate infusion was continued throughout the experiment to keep urinary pH above 7.8.

In group Ib, nine rats were studied during normocapnia in response to a sodium bicarbonate infusion as described above. After obtaining at least two control clearance collections at a urinary pH of 7.8 or higher, moderate hypercapnia was induced using a mixture of 5% CO<sub>2</sub>, balance air. Blood pCO<sub>2</sub> was kept constant between 68 and 78 mmHg for 2 h. Thereafter, experimental urine collections were completed while sodium bicarbonate infusion was administered as in group Ia.

In group IIa, seven rats were studied before and after induction of severe hypercapnia (blood pCO<sub>2</sub>, 80-100 mmHg). Urine collections were obtained during the first and the second hour of sustained respiratory acidosis to determine the effect of this acid-base alteration on urinary pCO<sub>2</sub> and renal acid excretion.

In group IIb, seven normocapnic rats excreting a urine of low bicarbonate concentration (<3 meq/liter) were studied during 2 h and served as controls for hypercapnic rats excreting an acid urine (group IIa).

In group IIIa, seven rats were studied during severe hypercapnia as in group IIa except that a bolus dose of 5 mg of carbonic anhydrase followed by a continuous infusion (5 mg/h) was administered before induction of severe hypercapnia. Thereafter, carbonic anhydrase was infused at a rate of 5 mg/h during 2 h of sustained hypercapnia to examine urinary pCO<sub>2</sub> generation in the face of accelerated carbonic acid dehydration.

In group IIIb, six normocapnic rats were studied during the infusion of carbonic anhydrase as in group IIIb and served as normocapnic controls.

After completion of the above protocols, groups IIa, IIb, IIIa, and IIIb were infused with sodium bicarbonate. In groups IIIa and IIIb, the infusion of carbonic anhydrase was continued throughout the experiment. At least three urine collections were completed during the infusion of sodium bicarbonate (0.9-M solution) at a rate of 6 ml/h while blood pCO<sub>2</sub> was held constant.

In group IV, eleven rats were studied during 2 h of acute respiratory acidosis as in group IIa. Thereafter, they were infused with sodium bicarbonate at a slower rate (3 ml/h) in an effort to prevent a rise in blood pH to the alkalemic range that occurred in all previous groups studied during the course of sodium bicarbonate infusion.

In group IVa, a subset of six rats of group IV were further studied during carbonic anhydrase infusion (5 mg/h for 2 h after an initial bolus dose of 5 mg). Three collections of  $\sim$ 30 min duration were obtained while the rate of sodium bicarbonate infusion was maintained at  $\sim$ 3 ml/h.

In groups Va and Vb, ten normocapnic rats served as controls for group IVa hypercapnic rats. After completion of three urine collections during the infusion of sodium bicarbonate, five of them were given carbonic anhydrase (group Va) and five were not (group Vb, time controls). The rate of sodium bicarbonate infusion was maintained at  $\sim$ 3 ml/h throughout the protocol.

Analytical methods. The urine and arterial blood samples for pH and pCO<sub>2</sub> were measured on a Radiometer system (Radiometer America, Inc., Westlake, OH) immediately after collection. Bicarbonate concentration was calculated from the pH and pCO<sub>2</sub> using the Henderson-Hasselbalch equation as previously described (40, 41). Sodium and potassium were measured by flame photometry. Titratable acidity was calculated from the amount of 0.1 NaOH used to titrate a 1-ml sample (0.1 ml urine plus 0.9 ml distilled  $H_2O$ ) up to a pH of 7.4. Ammonium was measured by a formulin titrimetric method (42).

Nonbicarbonate buffer capacity was measured using an automatic titrator (Radiometer America, Inc., model TTT 60, ABU 12) as previously described by Stinebaugh et al. (20). Briefly, HCl was added in sufficient amounts as to lower the pH of each urine specimen to below 2.5. The specimens were agitated for 30 min to ensure carbon dioxide loss. Thereafter, a titration curve from 2.5 to 8.5 was obtained by adding NaOH (0.1 M) to each of the urine samples. The nonbicarbonate buffer capacity was calculated from the pH 7.0-8.5 portion of

the titration curve as well as from 3.0 to 8.5 pH U. Either calculation yielded similar results, owing to the linearity of the titration curve. Carbonic anhydrase B from bovine erythrocytes was obtained from Sigma Chemical Co. (St. Louis, MO). Carbonic anhydrase activity in the urine was measured by the method of Maren et al. (43). This technique measures carbonic anhydrase activity by the rate of carbon dioxide hydration. Statistical analyses were performed using the t test for paired and unpaired data when appropriate. Values are means  $\pm$  SE.

#### Results

Effect of an acute elevation of blood pCO<sub>2</sub> on urine pCO<sub>2</sub> in rats excreting a highly alkaline urine (Table I, Fig. 1) Effect of severe hypercapnia (group Ia). The data of rats infused with sodium bicarbonate before induction of hypercapnia are presented in Table I. In these rats, urine pCO<sub>2</sub> after alkalinization of the urine was 75±1.6 mmHg and U-B pCO<sub>2</sub> was 31±2 mmHg. Induction of acute hypercapnia in the face of continuous sodium bicarbonate infusion resulted in a fall in blood pH and a rise in plasma bicarbonate. The increase in blood pCO<sub>2</sub> (from 42±0.8 to 87±0.8 mmHg) was associated with an increase in urine pCO<sub>2</sub> (from 75±1.6 to 103±4.2 mmHg, P < 0.001) and a decrease in U-B pCO<sub>2</sub> (from  $31\pm2$ to  $16\pm4.2$  mmHg, P < 0.005). The rise in urine pCO<sub>2</sub> was of a greater magnitude than the rise in urine bicarbonate concentration (from 163±5.2 to 196±12 meg/liter); consequently, urinary pH fell (from  $7.93\pm0.009$  to  $7.87\pm0.02$ , P < 0.01).

Effect of moderate hypercapnia (group Ib). In these rats, blood pCO<sub>2</sub> was increased to a lesser degree in order to investigate whether the low U-B pCO<sub>2</sub> observed in rats with severe hypercapnia had been, at least in part, the consequence of their higher blood pCO<sub>2</sub> per se, rather than failure to generate a sufficiently large urinary pCO<sub>2</sub>.

The increase in blood pCO<sub>2</sub> (from  $40\pm0.6$  to  $73\pm0.06$  mmHg) resulted in a fall in blood pH despite an increase in plasma bicarbonate secondary to continuous sodium bicarbonate infusion. Urinary pCO<sub>2</sub> increased from  $79\pm2.8$  to  $103\pm5.2$  mmHg, P < 0.005; that is, to a value virtually identical to that observed in rats with severe hypercapnia (group Ia). This increase in urine pCO<sub>2</sub> occurred in the face of a modest rise in urine bicarbonate concentration (from  $145\pm4.5$  to  $199\pm11$  meq/liter) which was also virtually identical to that observed in rats with severe hypercapnia (group Ia) (Fig. 1).

In contrast to rats with severe hypercapnia, however, the U-B pCO<sub>2</sub> did not decrease significantly in response to induction of hypercapnia of moderate severity (from 39±2.5 to 30±4.9 mmHg). Hence, the U-B pCO<sub>2</sub> of rats with moderate hypercapnia was significantly higher than that of rats with severe hypercapnia (30±4.9 and 16±4.2 mmHg, respectively; P < 0.05). This difference was likely the consequence of a blood pCO<sub>2</sub> lower in rats with moderate hypercapnia than in rats with severe hypercapnia (73±0.6 and  $87\pm0.8$  mmHg, P < 0.001) because urinary pCO<sub>2</sub> was virtually identical between both groups (Fig. 1). Blood pH, plasma sodium, plasma potassium, and urine flow were not significantly different between these two groups of hypercapnic animals. After induction of hypercapnia GFR fell to a comparable level in both groups. A fall in GFR was also observed in normocapnic time control rats (group Vb) that are presented in Table V (see below).

Acid excretion during acute respiratory acidosis (Table II) Induction of severe hypercapnia (group IIa) resulted in a significant fall in blood pH, a fall in urinary pH, and an

increase in net acid excretion which were observed during the first hour of acute hypercapnia. The increase in titratable acid excretion was associated with an increase in phosphate excretion. A significant increase in ammonium excretion and a further increase in titratable acid and net acid excretion were observed during the second hour of sustained hypercapnia. The increase in ammonium excretion observed during the second hour of sustained hypercapnia was associated with a urinary pH identical to that observed during the first hour of hypercapnia. As anticipated, plasma K and plasma phosphate increased in response to hypercapnia. GFR did not fall significantly after induction of acute respiratory acidosis.

# Urinary pCO<sub>2</sub> during acute respiratory acidosis

The urinary pCO<sub>2</sub> of group IIa rats was  $57\pm9.1$  mmHg in the first hour and  $58\pm6.3$  mmHg during the second hour of sustained hypercapnia. These values of urinary pCO<sub>2</sub> were significantly lower than blood pCO<sub>2</sub> ( $57\pm9.1$  vs.  $84\pm2.2$  mmHg, P < 0.02, and  $58\pm6.3$  vs.  $86\pm1.7$  mmHg, P < 0.005). Therefore, in hypercapnic rats the U-B pCO<sub>2</sub> difference was markedly negative throughout the 2 h of observation (Fig. 2). The urinary pCO<sub>2</sub> of control rats (group IIb) was not significantly different than that of the blood ( $37\pm3.2$  and  $41\pm1.3$  mmHg, respectively).

The urinary pCO<sub>2</sub> observed in group IIa rats during the second hour of hypercapnia was compared to that of normocapnic control rats (group IIb) which were excreting an acid urine. The urinary pCO<sub>2</sub> of hypercapnic rats was higher than that of normocapnic control rats (58±6.3 and 37±3.2 mmHg, respectively, P < 0.02) with a comparably low urine bicarbonate concentration (1.1±0.3 and 1.6±0.3 meq/liter, respectively). Consequently, urinary pH was lower in hypercapnic rats than in control rats (5.78±0.06 and 6.17±0.07, respectively, P < 0.005). Hypercapnic rats had a urinary phosphate concentration higher than that of normocapnic controls (72±17 and 29±6.3 mmol/liter, respectively, P < 0.05).

Urinary pCO<sub>2</sub> during acute respiratory acidosis and carbonic anhydrase infusion

In rats with acute respiratory acidosis infused with carbonic anhydrase (group IIIa), urinary pCO2 was 50±4.3 and 55±4.7 mmHg during the first and second hour of hypercapnia, respectively. These values of urinary pCO<sub>2</sub> were significantly lower than blood pCO<sub>2</sub> (50±4.3 vs.  $80\pm1.5$  mmHg, P < 0.001, and  $55\pm4.7$  vs.  $86\pm2.2$  mmHg, P < 0.001, respectively). The urinary pCO<sub>2</sub> of these rats was also higher than that of normocapnic controls (group IIIb) infused with carbonic anhydrase (55 $\pm$ 4.7 and 30 $\pm$ 3.6 mmHg, respectively, P < 0.005) (Table III). The urinary pCO<sub>2</sub> of hypercapnic rats infused with carbonic anhydrase (group IIIa) was not significantly different from that of hypercapnic rats not receiving this enzyme (group IIa) (55±4.7 and 58±6.3 mmHg, respectively). Hence, the urinary pCO<sub>2</sub> of hypercapnic rats excreting a urine of low bicarbonate content was lower than that of the blood regardless of whether carbonic anhydrase was given or not. This resulted in a markedly negative U-B pCO<sub>2</sub> difference in hypercapnic but not in normocapnic rats (Fig. 2).

Effect of alkalinization of the urine on urinary  $pCO_2$  in rats infused with carbonic anhydrase (Table III)

In hypercapnic rats (group IIIa), sodium bicarbonate infusion resulted in a significant rise in urine pCO<sub>2</sub> (from  $55\pm4.7$  to  $78\pm2.7$  mmHg, P < 0.005). In normocapnic control rats

Table I. Effect of Hypercapnia on Urine pCO2 in Rats Excreting an Alkaline Urine

			Blood			Plasma			Fractional excretion	xcretion		Urine				:
	GFR	>	Hd	pCO <sub>2</sub>	нсо,	×	Na a	۵	Na	۵	×	PO.	нсо,	Hd	pCO <sub>2</sub>	U = B pCO <sub>2</sub>
	m/lm	m/lm		ттНв	meq/ liter	meq/ liter	meq/ liter	meq/ liter	%	8%	8%	mmol/ liter	meq/ liter		ттНв	ттНв
Severe hypercapnia (group Ia, $n = 8$ )																
NaHCO <sub>3</sub> loading	4.1 ±0.3	0.28 ±0.02	7.67 ±0.01	42 ±0.8	49 ±1.2	2.8 ±0.08	161 ±2.1	99 ±0.8	10 ±0.7	4.0 ±0.4	41 ±4.3	8.8	163 ±5.2	7.93 ±0.009	75 ±1.6	31 ±2.0
P<	0.001	SN	0.001	0.001	0.001	SN	0.001	NS	SN	SZ	0.02	0.05	0.02	0.01	0.001	0.005
NaHCO <sub>3</sub> loading + hypercapnia	2.3 ±0.2	0.24 ±0.05	7.55 ±0.02	87 ±0.8	75 ±3.9	3.0 ±0.11	181 ±2.8	98 ±1.1	14 ±2.0	3.0 ±0.9	27 ±4.2	12 ±1.5	196 ±12	7.87 ±0.02	103 ±4.2	16 ±4.2
Moderate hypercapnia (group Ib,																
(6 - 11	3.3	0.39	7.68	40	46	2.7	162	901	20	9.2	49	4.2	145	7.85	62	39
NaHCO <sub>3</sub> loading	±0.3	±0.03	±0.01	70.€	±1.9	±0.1	±1.3	70.€	±2.3	±1.9	±5.3	±1.4	±4.5	±0.1	±2.8	±2.5
P<	0.01	0.02	0.001	0.001	0.001	SN	0.001	0.001	SN	NS	0.005	0.02	0.001	SN	0.001	NS
NaHCO <sub>3</sub> loading + hypercapnia	2.1 ±0.2	0.26 ±0.09	7.58 ±0.01	73 ±0.6	68 ±1.9	2.7 ±0.4	185 ±3.4	103 ±0.6	20 ±1.3	5.9 ±0.7	28 ±2.7	6.0 ±0.6	199 ±11	7.85 ±0.1	103 ±5.2	30 ±4.9
*	SZ	SN	SN	<0.001	SZ	SN	SN	<0.001	<0.025	<0.025	NS	<0.005	NS	SN	NS	<0.05

U - B pCO<sub>2</sub>, urine minus blood pCO<sub>2</sub> difference. \* Group Ia vs. group Ib during hypercapnia.

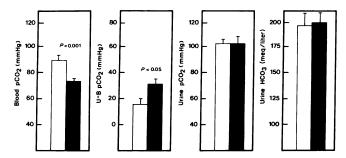


Figure 1. Blood pCO<sub>2</sub>, U-B pCO<sub>2</sub>, urine pCO<sub>2</sub>, and urine bicarbonate concentration in rats with severe hypercapnia (open bars) and in rats with moderate hypercapnia (dark bars) infused with sodium bicarbonate. The lower U-B pCO<sub>2</sub> of rats with severe hypercapnia was totally accounted by their higher blood pCO<sub>2</sub> because urine pCO<sub>2</sub> was the same as that of rats with moderate hypercapnia.

(group IIIb), sodium bicarbonate infusion also resulted in a significant rise in urine pCO<sub>2</sub> (from  $30.5\pm3.6$  to  $42\pm1.7$  mmHg, P < 0.02). The increment in urinary pCO<sub>2</sub> elicited by sodium bicarbonate loading was higher in hypercapnic than in normocapnic rats ( $24\pm5.2$  and  $12\pm3.1$  mmHg, respectively), but this difference did not achieve statistical significance (P > 0.05). During sodium bicarbonate loading, the urine pCO<sub>2</sub> of hypercapnic rats was significantly lower than that systemic blood pCO<sub>2</sub> ( $78\pm2.7$  and  $87\pm1.8$  mmHg, respectively, P < 0.02), while that of normocapnic rats was not ( $42\pm1.7$  and  $40\pm0.3$  mmHg, respectively). This resulted in a negative U-B pCO<sub>2</sub> in hypercapnic rats but not in controls (Fig. 2).

Urine pH was higher in normocapnic rats than in hypercapnic rats  $(8.08\pm0.05 \text{ and } 7.90\pm0.02, \text{ respectively, } P < 0.005)$  while urine bicarbonate concentration was not significantly different. GFR did not change significantly in either group while plasma sodium and plasma bicarbonate increased in both groups.

Effect of alkalinization of the urine on urinary  $pCO_2$  in rats not infused with carbonic anhydrase (Table IV)

In the set of experiments presented in Table I, hypercapnia was induced in normocapnic rats that were excreting an

alkaline urine and, therefore, had a high urine pCO<sub>2</sub>. Thus, the additional rise in urinary pCO<sub>2</sub> that occurred in these rats could have been the result of abruptly raising the blood pCO<sub>2</sub> rather than the consequence of alkalinizing the urine. In the following set of experiments, sodium bicarbonate was infused into hypercapnic rats that were excreting a urine with low bicarbonate content. In hypercapnic rats (group IIa) in which sodium bicarbonate infusion resulted in a high blood pH, urinary pCO<sub>2</sub> rose from  $58\pm6.3$  to  $98\pm4.9$  mmHg (P < 0.005) in the absence of any significant change in blood pCO<sub>2</sub>. This rise in urinary pCO<sub>2</sub> was therefore virtually identical to the rise in U-B pCO<sub>2</sub> (from  $-28\pm4.7$  to  $15\pm4.0$  mmHg). The urinary pCO<sub>2</sub> achieved under these conditions was comparable to that of rats with moderate (group Ia) and severe hypercapnia (group Ib) in which blood pCO<sub>2</sub> was increased after alkalinization of the urine (compare Table IV with Table I). Blood pH after bicarbonate loading was also not significantly different  $(7.55\pm0.02, 7.58\pm0.01, \text{ and } 7.53\pm0.02 \text{ in groups Ia, Ib, and})$ IIa, respectively). Plasma sodium was higher in the hypercapnic rats presented in Table I (groups Ia and Ib) than in those presented in Table IV (group IIa). GFR fell significantly in groups Ia and Ib but not in group IIa, probably because the duration of the experiment was longer in groups Ia and Ib.

In normocapnic control rats (group IIb), urinary pCO<sub>2</sub> increased from  $37\pm3.2$  to  $79\pm2.6$  mmHg. Hence, the increment in urinary pCO<sub>2</sub> elicited by bicarbonate infusion while blood pCO<sub>2</sub> was kept constant (thereafter to be referred to as  $\Delta$ pCO<sub>2</sub>) was almost identical in hypercapnic rats (group IIa) and normocapnic rats ( $40\pm7.2$  and  $42\pm4.6$  mmHg, respectively). After bicarbonate loading, urinary pCO<sub>2</sub> was higher in hypercapnic rats than in controls ( $98\pm4.9$  and  $79\pm2.6$  mmHg, P < 0.001), but the U-B pCO<sub>2</sub> was lower in hypercapnic rats than in controls ( $15\pm4.0$  and  $39\pm2.5$  mmHg, P < 0.001).

In hypercapnic rats (group IV) in which bicarbonate infusion resulted in a normal blood pH  $(7.38\pm0.01)$ , urinary pCO<sub>2</sub> increased from  $55\pm6.9$  to  $125\pm3.6$  mmHg, P < 0.001. Thus, the  $\Delta$ pCO<sub>2</sub> of these rats  $(70\pm5.6$  mmHg) was greater than that observed in normocapnic rats  $(42\pm4.6$  mmHg) (Fig. 3, *left*). It was also greater (P < 0.005) than that of hypercapnic rats  $(40\pm7.2$  mmHg) with a higher blood pH (group IIa). The

Table II. Effect of Acute Respiratory Acidosis (Group IIa) on Acid Excretion

			Blood									
	GFR	v	pН	pCO <sub>2</sub>	HCO <sub>3</sub>	Plasma K	Plasma PO₄	$UPO_4\times V$	Urine pH	TA	NH <sub>4</sub>	Net acid
	ml/min	ml/min		ттНд	meq/liter	meq/liter	mmol/liter	μmol/min		μmol/min	μmol/min	μmol/min
Prehypercapnia	3.4	0.01	7.42	40	25	3.5	2.0	0.1	6.5	0.2	0.4	0.5
(n=7)	±0.4	±0.03	±0.01	±1.5	±1.5	±0.2	±0.1	±0.05	±0.2	±0.05	±0.1	±0.2
P value	NS	NS	0.001	0.001	NS	0.01	NS	0.01	0.05	0.005	NS	0.005
Hypercapnia	2.6	0.02	7.13	84	28	4.4	2.7	1.2	5.7	1.3	0.4	1.7
(1st h)	±0.3	±0.02	±0.02	±2.2	±1.3	±0.3	±0.1	±0.3	±0.09	±0.2	±0.05	±0.2
P value	NS	0.05	NS	NS	NS	NS	NS	0.01	NS	0.005	0.05	0.005
Hypercapnia	3.0	0.04	7.14	86	29	4.4	2.7	2.7	5.8	2.5	1.0	3.4
(2nd h)	±0.2	±0.07	±0.02	±1.7	±1.2	±0.1	±0.09	±0.5	±0.06	±0.1	±0.2	±0.3
	NS	0.02	0.001	0.001	NS	0.01	0.01	0.005	0.025	0.001	0.05	0.001

<sup>\*</sup> Statistical analysis (P value) between hypercapnia (2nd h) and prehypercapnia.

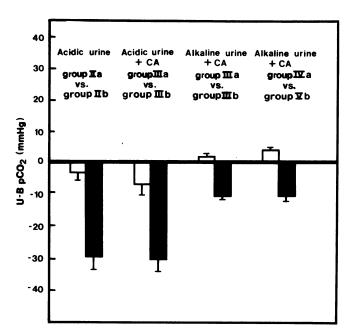


Figure 2. The U-B pCO<sub>2</sub> of hypercapnic rats (groups IIa, IIIa, and IVa) is compared with that of normocapnic control rats (groups IIb, IIIb, and Va). The U-B pCO<sub>2</sub> of hypercapnic rats was lower than that of normocapnic control rats when the urine was acid (with and without carbonic anhydrase infusion) and when it was alkalinized in the face of a continuous carbonic anhydrase (CA) infusion.  $\Box$ , control;  $\blacksquare$ , hypercapnia.

U-B pCO<sub>2</sub> of group IV hypercapnic rats (39±3.7 mmHg) was significantly higher than that of group IIa hypercapnic rats (15±4.0 mmHg) and not significantly different than that of normocapnic controls (group IIb) (39±2.5 mmHg). These differences in urine pCO<sub>2</sub> and U-B pCO<sub>2</sub> were observed in the face of a comparable increase in urine bicarbonate concentration in the three groups studied (Fig. 4). Plasma sodium also increased to a comparable level in the three groups. Plasma K was higher in hypercapnic rats than in controls.

Effect of carbonic anhydrase infusion on urine  $pCO_2$  in rats excreting an alkaline urine (Table V)

In normocapnic control rats (group Va), carbonic anhydrase infusion resulted in a fall in urine pCO<sub>2</sub> (from  $73\pm1.9$  to  $43\pm1.3$  mmHg, P<0.001) while urine bicarbonate concentration did not change significantly. Consequently, urine pH increased (from  $7.87\pm0.03$  to  $8.04\pm0.04$ , P<0.02). In hypercapnic rats (group IVa), the infusion of carbonic anhydrase also resulted in a fall in urine pCO<sub>2</sub> (from  $122\pm5.7$  to  $77\pm2.2$  mmHg, P<0.001), while urine bicarbonate concentration did not change and urine pH increased (from  $7.65\pm0.03$  to  $7.85\pm0.03$ , P<0.005). The fall in urine pCO<sub>2</sub> elicited by carbonic anhydrase infusion was greater in hypercapnic rats than in normocapnic controls ( $-45\pm4.3$  and  $-30\pm2.6$  mmHg, respectively, P<0.02) (Fig. 3, right).

Nonbicarbonate buffer capacity was not significantly different between normocapnic and hypercapnic rats either before  $(2.4\pm0.3 \text{ and } 2.3\pm0.6 \text{ meq/liter per pH unit, respectively})$  or after carbonic anhydrase infusion  $(2.5\pm0.5 \text{ and } 3.9\pm2.3 \text{ meq/liter per pH unit, respectively})$ .

Carbonic anhydrase activity in the urine was not significantly different between hypercapnic and normocapnic rats  $(275\pm96 \text{ vs. } 362\pm103 \text{ } \mu\text{g/ml}, \text{ respectively}).$  Likewise, the excretion of this enzyme was not significantly different between the two groups  $(31\pm8.3 \text{ vs. } 23\pm4.0 \text{ } \mu\text{g/min}, \text{ respectively}).$  Carbonic anhydrase infusion was associated with a significant fall in GFR and urine flow in both hypercapnic and normocapnic rats. Neither parameter, however, was significantly different between hypercapnic and normocapnic rats.

In time control rats (group Vb) GFR also fell significantly, but urine, pCO<sub>2</sub>, and U-B pCO<sub>2</sub> did not. The fall in GFR was likely the result of the prolonged duration of this experiment. Nonbicarbonate buffer capacity in the time control group was not significantly different than that of hypercapnic (group IVa) and normocapnic rats (group Va) infused with carbonic anhydrase.

After carbonic anhydrase infusion, the urine pCO<sub>2</sub> of hypercapnic rats (group IVa) fell to a level lower than that of the blood (77 $\pm$ 2.2 and 86 $\pm$ 0.9 mmHg, respectively, P < 0.025), while that of normocapnic rats (group Va) did not (43 $\pm$ 1.3 and 40 $\pm$ 1.0 mmHg, respectively). Thus, the relationship between urine pCO<sub>2</sub> and blood pCO<sub>2</sub> was virtually identical to that observed in hypercapnic (group IIIa) and normocapnic rats (group IIIb) in which the infusion of carbonic anhydrase antedated the infusion of bicarbonate (Fig. 2). As anticipated, the urinary pCO<sub>2</sub> achieved in both hypercapnic and normocapnic rats infused with sodium bicarbonate and carbonic anhydrase was markedly lower than that observed when this enzyme was not administered (compare Tables III and V with Table IV).

#### **Discussion**

This study demonstrates a clear dissociation between urine pCO<sub>2</sub> and blood pCO<sub>2</sub> during acute hypercapnia. In normocapnic rats excreting a highly alkaline urine, induction of severe hypercapnia resulted in a fall in the U-B pCO<sub>2</sub> gradient despite an increase in urine pCO<sub>2</sub> (group Ia, Table I). According to current views (9, 16-29, 36), the finding that the U-B pCO<sub>2</sub> achieved during bicarbonate loading is lower in hypercapnic rats than in normocapnic rats (Tables I and IV) requires the conclusion that collecting duct H<sup>+</sup> secretion is decreased by hypercapnia. In fact, such a conclusion was recently reached by another group of investigators who noted that the U-B pCO<sub>2</sub> gradient of hypercapnic dogs infused with sodium bicarbonate was lower than normal (23).

That the low U-B pCO<sub>2</sub> observed during bicarbonate loading reflects decreased collecting duct H<sup>+</sup> secretion during hypercapnia, however, is not likely correct. The urine pCO<sub>2</sub> of hypercapnic rats was higher than that of normocapnic rats, a finding that could be interpreted as indicative of enhanced collecting duct H<sup>+</sup> secretion in hypercapnic rats (i.e., the opposite interpretation of that portrayed by the U-B pCO<sub>2</sub> gradient). Alternatively, it could be argued that the higher urine pCO<sub>2</sub> observed in hypercapnic rats was purely the result of their higher blood pCO<sub>2</sub>. The following findings, however, mitigate against the latter possibility.

First, rats with moderate hypercapnia (group Ib) exhibited a urinary pCO<sub>2</sub> virtually identical to that of rats with severe hypercapnia (group Ia) in the face of a comparable urine bicarbonate concentration (Fig. 1). This finding indicates that the U-B pCO<sub>2</sub> was influenced by changes in systemic blood pCO<sub>2</sub> rather than by changes in urinary pCO<sub>2</sub> generation. Second, urinary pCO<sub>2</sub> did not increase to the level prevailing in the blood in hypercapnic animals excreting a urine of low

Table III. Urinary pCO2 in Rats Infused with Carbonic Anhydrase

				`													
			Blood			Plasma			Fractional excretion	excretion			Urine				
	GFR	>	핊	pC02	нсо,	Na	¥	ū	Na	ם	×	HCO,	Hd	нсо,	pC0,	U - B pCO <sub>2</sub>	∆ pC02
	ml/min	ml/min		mmHg	meq/liter	meq/liter	meq/liter	meq/liter	88	88	88	88		meq/liter	вншш	вншш	mmHg
Hypercapnia (group IIIa, $n = 7$ )																	
CA infusion	3.4	0.09	7.18	98	30	145	,	115	3.5	4.0	23	0.5	5.90	1.2	55	-30	1
	∓0.5	±0.01	∓0.02	+2.2	±1.2	<del>+</del> 1.1.	4.2 ±0.3	±3.3	∓0.4	±0.4	±1.3	±0.4	±0.07	±0.3	±4.7	±5.2	
P<	SN	0.001	0.001	SN	0.001	0.001	0.02	0.02	0.001	0.02	0.01	0.001	0.001	0.001	0.005	0.02	
CA infusion	2.9	0.37	7.51	87	0/	163	3.6	901	20	13	33	32	7.90	161	78	-10	24
+ NaHCO <sub>3</sub> loading	∓0.4	±0.05	±0.02	+1.8	±4.4	+1.1	∓0.09	±0.5	±1.7	±2.5	±1.7	±3.5	±0.02	<b>±6.1</b>	±2.7	±2.1	±5.2
Controls $(arction 111B \ n = 6)$																	
CA infusion	5.6	90.0	7.46	37	26	141	4.0	112.3	3.7	5.9	35	0.12	5.99	1.3	30	-6.7	
	±0.5	±0.01	±0.01	+0.9	∓0.8	±1.6	9.0∓	±11.7	±0.7	±1.2	±5.3	±0.04	±0.19	±0.4	±3.6	±3.0	
P <	SN	0.001	0.001	0.05	0.001	0.001	SN	SZ	0.001	NS	SN	0.001	0.001	0.001	0.02	0.0025	
CA infusion	1.8	0.19	7.73	40	53	160	3.0	110.5	20	12	35	32	808	137	42	~	2
+ NaHCO <sub>3</sub> loading	±0.3	+0.1	±0.05	±0.3	±2.6	+1.8	±0.1	±5.1	±2.4	±3.0	±2.5	±1.9	±0.05	+18	±1.7	+1.8	±3.1
*	SN	SN	0.001	0.001	0.025	SZ	SN	SN	SN	SN	SN	NS	SN	NS	0.005	0.005	ı
++	SN	NS	0.001	0.001	0.01	NS	0.001	SN	NS	SN	SZ	SN	0.005	SN	0.001	0.005	SN

ΔρCO<sub>2</sub>, denotes the rise in urine pCO<sub>2</sub> elicited by sodium bicarbonate infusion. \* Hypercapnic rats vs. normocapnic rats studied before NaHCO<sub>3</sub> loading. ‡ Hypercapnic vs. normocapnic control rats studied after NaHCO<sub>3</sub> loading. ‡ Hypercapnic vs. normocapnic

Table IV. Effect of Alkalinization of the Urine on Urinary pCO<sub>2</sub> Generation at a Constant Blood pCO<sub>2</sub>

3			100			Dlocmo			Fractions	Fractional excretion			Urine					
	GFR	>	Hd	pCO <sub>2</sub>	HCO,	Na Na	×	CI	Z s	ס	*	нсо,	PO.	Hd	нсо,	pCO2	U - B pCO <sub>2</sub>	Δ pCO <sub>2</sub>
	ml/min	ml/min		вншш	meq/liter	meq/liter	meq/liter	meq/liter	%	88	8%	%	mmol/liter		meq/liter	ттНв	ттНв	мтНв
Controls (groups IIb, $n = 7$ ) Base line	2.6 ±0.4	0.01 ±0.003	7.38 ±0.008	41 ±1.3	24 ±0.6	147 ±2.1	3.4 ±0.4	115 ±1.1	0.4 ±0.1	0.8	18 ±2.9	0.04 ±0.01	29 ±6.3	6.17 ±0.07	1.6 ±0.3	37 ±3.2	-3.4 ±3.6	1
P<	SN	0.001	0.001	SN	0.001	0.001	0.02	0.001	0.001	0.001	0.005	0.001	0.01	0.001	0.001	0.001	0.001	
NaHCO <sub>3</sub> loading	3.0 ±0.1	0.39 ±0.03	7.69 ±0.01	41 ±0.4	49 ±1.8	163 ±1.0	2.6 ±0.2	107 ±0.7	21 ±2.9	10 ±2.4	50 ±5.2	44 ±5.6	4.2 ±0.14	7.86 ±0.01	149 ±3.7	79 ±2.6	39 ±2.5	42 ±4.6
Hypercapnia, (group IIa, $n = 7$ ) Base line	3.0 ±0.2	0.04 ±0.007	7.14 ±0.02	86 ±1.7	29 ±1.2	143 ±0.7	4.4	121 ±1.0	2.2 ±0.3	2.9	24 ±2.4	0.3	72 ±17	5.78 ±0.06	1.1	58 ±6.3	−28 ±4.7	I
P<	SN	0.001	0.001	SN	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.01	0.001	0.001	0.005	0.001	
NaCHO <sub>3</sub> loading	3.3 ±0.3	0.49 ±0.05	7.53 ±0.02	83 ±1.3	69 ±2.7	163 ±1.7	3.0 ±0.1	109 ±0.8	23 ±2.2	12 ±1.1	43 ±1.9	39 ±4.7	5.8 ±0.9	7.82 ±0.02	171 ±11	98 ±4.9	15 ±4.0	40 ±7.2
Hypercapnia (group IV, n = 11) Base line	1.8	0.02 ±0.004	7.09 ±0.008	86 ±0.7	25 ±0.5	147 ±0.5	4.0 ±0.1	113 ±0.9	1.6 ±0.3	2.2 ±0.4	24 ±3.0	0.07 ±0.02	74 ±9.7	5.89 ±0.07	1.4 ±0.4	55 ±6.9	-31 ±6.9	1
P <	0.05	0.001	0.001	SN	0.001	0.001	0.025	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	
NaHCO, loading	2.4 ±0.2	0.25 ±0.03	7.38 ±0.01	86 ±0.8	49 ±1.3	165 ±1.6	3.4 ±0.2	105 ±1.2	18 ±1.4	8.8 ±1.3	43 ±2.4	35 ±2.3	12.4 ±1.2	7.69 ±0.02	165 ±10	125 ±3.6	39 ±3.7	70 ±5.6
*	NS	SN	0.001	0.001	0.001	NS	NS	SN	SN	SN	SN	NS	NS	SN	NS	0.005	0.001	SN
++	0.05	0.01	0.001	0.001	NS	NS	0.02	SN	SN	SN	SN	SN	0.001	SN	SN	0.001	SN	0.005
જ	0.02	0.001	0.001	SN	0.001	NS	SN	0.05	SN	NS	NS	SN	0.005	0.001	SN	0.001	0.001	0.005

pCO<sub>2</sub>, rise in urine pCO<sub>2</sub> elicited by sodium bicarbonate infusion. \* Hypercapnia (group IIa) vs. controls (group IIb) during NaHCO<sub>3</sub> loading. ‡ Hypercapnia (group IV) vs. controls (group IIb) during bicarbonate loading.

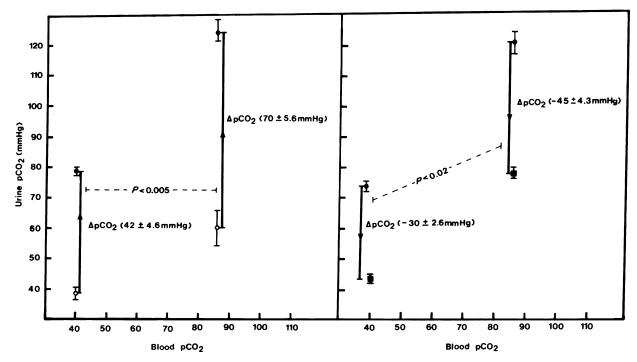


Figure 3. The relationship of urine pCO<sub>2</sub> to blood pCO<sub>2</sub> during normocapnia and hypercapnia. The *left* panel depicts the increase in urine pCO<sub>2</sub> elicited by sodium bicarbonate loading (i.e., the  $\Delta$ pCO<sub>2</sub>) in normocapnic rats (group IIb) and hypercapnic rats (group IV). Open circles denote urine pCO<sub>2</sub> when the urine was acid; closed circles denote the urine pCO<sub>2</sub> achieved during bicarbonate loading. The  $\Delta$ pCO<sub>2</sub> was significantly higher in hypercapnic rats than in normocapnic controls (P < 0.005). The *right* panel depicts the fall in

urine pCO<sub>2</sub> ( $-\Delta$ pCO<sub>2</sub>) caused by carbonic anhydrase infusion to normocapnic rats (group Va) and hypercapnic rats (group IVa). Closed circles denote urine pCO<sub>2</sub> in an alkaline urine. Closed boxes denote that urine pCO<sub>2</sub> was measured during carbonic anhydrase infusion. The fall in urine pCO<sub>2</sub> elicited by carbonic anhydrase infusion was significantly higher in hypercapnic rats than in normocapnic controls (P < 0.02).

bicarbonate content; therefore, their U-B pCO<sub>2</sub> gradient was markedly negative (Fig. 2). This finding is in agreement with recent studies in dogs with acute hypercapnia (32) and in rats with chronic hypercapnia (33). Third, in hypercapnic rats excreting a urine of low bicarbonate content, the U-B pCO<sub>2</sub> was also negative when carbonic anhydrase was given to ensure complete carbonic acid dehydration and achievement of diffusion equilibrium between tubular and peritubular CO<sub>2</sub> (Table III, Fig. 2). Thus, even in the face of accelerated carbonic acid dehydration, urinary pCO<sub>2</sub> and blood pCO<sub>2</sub> did not equilibrate during hypercapnia. This finding differs from the study of Adrogue et al. (23) in dogs with severe hypercapnia. These authors reported that during carbonic anhydrase infusion, urine pCO<sub>2</sub> before bicarbonate loading was not significantly different from arterial blood pCO<sub>2</sub> within 30 min of induction of hypercapnia (23). Accordingly, they proceeded to use the U-B pCO<sub>2</sub> achieved after bicarbonate loading as a marker of distal acidification (23). The reasons for the discrepancy between our results and those of Adrogue et al. (23) are not readily apparent except that their studies were performed in only four animals of a different species. Fourth, in hypercapnic rats infused with carbonic anhydrase and sodium bicarbonate, urinary pCO<sub>2</sub> was still significantly lower than blood pCO<sub>2</sub>  $(78\pm2.7 \text{ vs. } 87\pm1.8 \text{ mmHg}, P < 0.02)$ , which resulted in a negative U-B pCO<sub>2</sub> (Table III, Fig. 2). This finding further suggests that systemic blood pCO<sub>2</sub> does not reflect intrarenal pCO<sub>2</sub> even in the face of accelerated carbonic acid dehydration

and in the presence of large urinary bicarbonate concentrations. Thus, factors other than equilibration between systemic blood pCO<sub>2</sub> and renal pCO<sub>2</sub> determine, at least in part, the urine pCO<sub>2</sub> in hypercapnic rats (see below).

Having established that the urinary pCO<sub>2</sub> of hypercapnic rats is lower than systemic blood pCO<sub>2</sub> we sought to determine the effect of alkalinization of the urine on urinary pCO<sub>2</sub> generation when blood pCO2 was kept constant. We reasoned that if blood pCO<sub>2</sub> were not allowed to change the increment in urinary pCO<sub>2</sub> observed in response to sodium bicarbonate loading (i.e., the  $\Delta pCO_2$ ), it would provide an indirect qualitative measurement of collecting duct H<sup>+</sup> secretion. The ΔpCO<sub>2</sub> of hypercapnic rats with a high blood pH (group IIa) was comparable to that of normocapnic rats with a high blood pH (group IIb) (Table IV). The  $\Delta pCO_2$  of hypercapnic rats with a normal blood pH (group IV) was higher than that of normocapnic rats (Fig. 3, left), and than that of hypercapnic rats with a high blood pH (group IIa) (Table IV). The U-B pCO<sub>2</sub> achieved by hypercapnic rats with a high blood pH (group IIa), however, was again lower than that of normocapnic rats (group IIb) (15±4.0 and 39±1.5 mmHg, respectively), while that of hypercapnic rats with a normal blood pH (group IV) was the same (39±3.7 and 39±2.5 mmHg, respectively).

Urinary bicarbonate concentration, a major determinant of urinary pCO<sub>2</sub> generation (13-30), increased to a comparable level in both groups of hypercapnic rats and in normocapnic controls (Fig. 4). Since blood pCO<sub>2</sub> was not allowed to change,

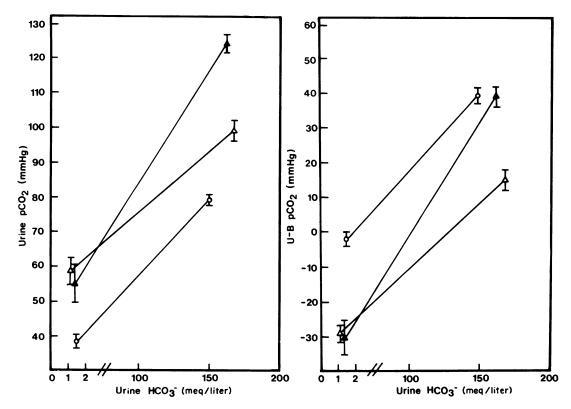


Figure 4. Urine pCO<sub>2</sub> as a function of urine bicarbonate concentration (left) and U-B pCO<sub>2</sub> as a function of urine bicarbonate concentration (right). At a comparably high urine bicarbonate concentration, rats with hypercapnia and a normal blood pH (group IV) (closed triangles) achieved a urine pCO<sub>2</sub> significantly higher than that

of hypercapnic rats (group IIa) (open triangles) and normocapnic rats (group IIb) (open circles) that were alkalemic. The U-B pCO<sub>2</sub> of group IIa rats was lower, while that of group IV was equal to that of normocapnic control rats (group IIb) with a comparably high urine bicarbonate concentration.

the rise in urine pCO<sub>2</sub> was the consequence, at least in part, of collecting duct H<sup>+</sup> secretion into a bicarbonate-rich urine and not the consequence of equilibration between intrarenal and systemic blood pCO<sub>2</sub>.

Urine pCO<sub>2</sub> and its derivatives as indices of distal acidification. The possible pitfalls of the urine pCO<sub>2</sub> and its derivatives as markers of collecting duct hydrogen ion secretion can be summarized as follows:

(a)  $U-B pCO_2$  during bicarbonate loading. The use of the U-B pCO<sub>2</sub> observed in a highly alkaline urine as an index of collecting duct H<sup>+</sup> secretion requires that during bicarbonate loading, systemic blood pCO<sub>2</sub> be similar to that of medullary blood. DuBose (9), however, has recently demonstrated that in a highly alkaline urine the pCO<sub>2</sub> in the papillary collecting duct and adjacent vasa recta is markedly higher than that of the systemic blood (9). Thus, in normocapnic subjects infused with bicarbonate, intrarenal blood pCO<sub>2</sub> and systemic blood pCO<sub>2</sub> dissociate. Since the rise in urine pCO<sub>2</sub> in an alkaline urine occurs in the collecting duct and is, therefore, associated with a rise in vasa recta pCO<sub>2</sub>, the use of the U-B pCO<sub>2</sub> serves no purpose because systemic blood pCO2 is not a reflection of intrarenal blood pCO<sub>2</sub> (30). This fact, in our view, invalidates the use of the U-B pCO2 gradient in a highly alkaline urine as an index of distal H<sup>+</sup> secretion even during normocapnia. This is not to say that the conclusions derived from numerous studies in normocapnic subjects using the U-B pCO<sub>2</sub> gradient as a marker of distal acidification need to be altered. When urinary pCO<sub>2</sub> and blood pCO<sub>2</sub> are about the same before bicarbonate loading (as is usually the case

during normocapnia), the rise in urinary pCO<sub>2</sub> elicited by sodium bicarbonate infusion is equivalent to the U-B pCO<sub>2</sub> achieved under this condition (e.g., normocapnic rats, Table IV). Therefore, both parameters yield the same information in regards to the ability to generate CO<sub>2</sub> as a result of collecting duct H<sup>+</sup> secretion. When blood pCO<sub>2</sub> and urine pCO<sub>2</sub> are widely different, however, the U-B pCO<sub>2</sub> gradient achieved during bicarbonate loading becomes an artifact that gives no physiological information about collecting duct H<sup>+</sup> secretion. The artifactual nature of the U-B pCO<sub>2</sub> gradient is illustrated by our studies during hypercapnia, where it clearly underestimates distal H<sup>+</sup> secretion (Table IV, Fig. 4). This situation is the opposite of hypocapnia, where the U-B pCO<sub>2</sub> gradient can overestimate collecting duct H<sup>+</sup> secretion (44).

(b) Urine  $pCO_2$  after bicarbonate loading. To take into account the relationship between urine  $pCO_2$  and blood  $pCO_2$  that prevailed before administration of sodium bicarbonate, we used the  $\Delta pCO_2$  as an index of collecting duct  $H^+$  secretion (Table IV, Fig. 3, left). Blood  $pCO_2$  was no longer a variable because it was not allowed to change, and the base-line urine  $pCO_2$  (i.e., the  $pCO_2$  of an acid urine) was known. Thus, the  $\Delta pCO_2$  is appropriate as a qualitative index of collecting duct  $H^+$  secretion, in that it integrates the effects of both an acid and a highly alkaline urine on intrarenal  $CO_2$  generation while it eliminates the blood  $pCO_2$  as a variable. If the  $\Delta pCO_2$  were the best way to express the relationship between urinary  $pCO_2$  generation and collecting duct  $H^+$  secretion, one would have to conclude that collecting duct  $H^+$  secretion was normal in

Table V. Effect of Carbonic Anhydrase Infusion on Urine pCO2 in Rats Excreting an Alkaline Urine

			Blood			Plasma			Fractional ex	l ex	Urine						Non-HCO,
	GFR	>	hф	pCO <sub>2</sub>	нсо,	Na	×	۵	Na	ū	P0.	Hd	нсо,	PC02	U - B pCO <sub>2</sub>	-∆ PC02	buffer capacity
	m/lm	m/lm		mmHg	meq/liter	meq/liter	meq/liter	meq/liter	88	88	mmol/liter		meq/liter	вншш	вншш	ттНв	meq/liter/ pH unit
Controls (group Va, $n = 5$ ) HCO <sub>3</sub> loading	2.3 ±0.2	0.14	7.62 ±0.02	38 ±1.5	37 ±0.6	156	3.2	108	11	5.6	13	7.87	146	73	35		4.2
P<	0.005	0.05	SN	NS	0.05	0.05	SN	SZ	SZ	SN SN	NS S	0.02	S. A.	0.001	0.001		NS NS
HCO <sub>3</sub> loading + CA	1.1 ±0.3	0.08 ±0.01	7.64 ±0.01	40 ±1.0	43 ±1.7	162 ±2.0	3.7 ±0.5	108 ±2.2	13 ±2.4	6.7 ±2.0	11 ±1.7	8.04 ±0.04	12 <b>8</b> ±12	43 ±1.3	3.1 ±1.9	−30 ±2.6	2.5 ±0.5
Hypercapnia (group IVa, n = 6) HCO <sub>3</sub> loading	2.0 ±0.3	0.27 ±0.05	7.37 ±0.02	86 ±0.9	49 ±2.4	166 ±2.9	3.5 ±0.4	107 ±1.7	20 ±2.0	12 ±1.6	10 ±1.4	7.65	144 ±7.0	122 ±5.7	36 ±5.2		2.3 ±0.6
P< HCO <sub>3</sub> loading + CA	0.02 1.0 ±0.08	0.02 0.12 ±0.02	NS 7.40 ±0.01	NS 86 ±0.9	NS 52 ±1.8	NS 172 ±2.4	NS 3.8 ±0.2	NS 108 ±1.7	NS 20 ±3.6	NS 11 ±2.9	NS 11 +1.8	0.005 7.85 ±0.03	NS 148 ±10	0.001 77 ±2.2	0.001 -9.0 ±2.8	-45 ±4.3	NS 3.9 ±2.3
Time controls (group Vb, n = 5) HCO <sub>3</sub> loading	2.4 ±0.6	0.18 ±0.02	7.63 ±0.02	40 ±0.9	41 +1.4	154 ±2.0	3.5 ±0.1	111	15 ±2.4	9.7	8.5 ±2.0	7.81 ±0.02	135 ±16	75 ±5.4	35 ±5.1		1.9 4.0+
P<	0.02	SZ	0.005	SN	0.05	0.01	SN	SN	SN	SN	SN	SN	SN	SZ	SN		SZ
HCO <sub>3</sub> loading	1.5 ±0.4	0.13 ±0.03	7.67 ±0.02	41 ±1.3	47 ±3.4	165 ±2.3	3.8 ±0.3	110 ±1.9	17 ±4.5	104 ±3.8	9.4 ±1.5	7.85 ±0.04	138 ±18	70 ±3.8	29 ±3.4	-5 ±6.9	1.8 ±0.2
* ++ ∞	S S S	NS N	0.001 NS 0.001	0.001 NS 0.001	0.01 NS NS	0.02 NS NS	SN SN SN	S S S	S S S	S S S	S S S S	0.005 0.01 NS	SN SN SN SN	0.001 0.001 NS	0.01 0.001 0.001	0.02 0.01 0.001	SN SN SN SN

\* Hypercapnia (group IVa) vs. controls (group Va). (comparing data obtained during the second phase of the experiment). ‡ Time controls (group Va) vs. controls (group Va) vs. controls (group Va) (comparing data obtained during the second phase of the experiment). § Time controls (group Vb) vs. hypercapnia (group IVa) (comparing data obtained during the second phase of the experiment).

hypercapnic rats that were alkalemic (group IIa) and enhanced in hypercapnic rats that were not alkalemic (group IV). This conclusion requires validation by measuring collecting duct pCO<sub>2</sub> and vasa recta pCO<sub>2</sub> before and after bicarbonate loading during sustained hypercapnia. While this information is currently not available, it is now known that in normocapnic rats, vasa recta pCO<sub>2</sub> and papillary collecting duct pCO<sub>2</sub> increase in a one to one ratio when the urine is highly alkaline (9). There is no a priori reason to believe that such a relationship does not exist during hypercapnia.

The  $\Delta pCO_2$  may be problematic, in that a number of variables (blood pH, plasma sodium, plasma potassium, volume status, medullary blood flow rate, etc.) change during bicarbonate diuresis as compared with base-line conditions. In this regard, the end point urine pCO<sub>2</sub> observed under identical conditions of bicarbonate diuresis, urine flow rate, and volume status is a more suitable marker of collecting duct H<sup>+</sup> secretion than the  $\Delta pCO_2$ . If one were to use the end-point urine  $pCO_2$ , one would have to conclude that collecting duct H<sup>+</sup> secretion was enhanced in both groups of hypercapnic rats because the end-point urine pCO<sub>2</sub> was higher than that of normocapnic controls (Fig. 4). It is readily apparent, however, that the urine pCO<sub>2</sub> (i.e., that observed in a highly alkaline urine) fails to take into account the fact that the pCO<sub>2</sub> prevailing in the urine of hypercapnic animals before bicarbonate loading is higher than that of normocapnic rats. Thus, during hypercapnia, the end point urine pCO<sub>2</sub> observed after bicarbonate loading is uncertain as a marker of collecting duct H+ secretion because it could overestimate it as opposed to the U-B pCO<sub>2</sub>, which underestimates it.

We attempted to equalize urine pCO<sub>2</sub> and blood pCO<sub>2</sub> in hypercapnic rats by alkalinizing the urine in the presence of carbonic anhydrase infusion. We reasoned that the rise in urinary pCO<sub>2</sub> attributable to H<sup>+</sup> secretion into a bicarbonaterich urine would be dissipated by accelerating carbonic acid dehydration, and that urinary pCO<sub>2</sub> and blood pCO<sub>2</sub> would be approximately the same. Urinary pCO<sub>2</sub> was about that of the blood in normocapnic rats but not in hypercapnic rats (Tables III and V, Fig. 2). The finding of a urinary pCO<sub>2</sub> lower than that of the blood in a highly alkaline urine rich in carbonic anhydrase deserves consideration. We think that accelerated consumption of CO<sub>2</sub> within collecting duct cells accounts for lowering of tubular CO<sub>2</sub> (and final urine CO<sub>2</sub>) below systemic blood levels during hypercapnia. CO<sub>2</sub> is likely consumed and bicarbonate regenerated because of enhanced secretion of H<sup>+</sup>, which under these conditions is titrated by bicarbonate and by nonbicarbonate buffer. The titration of H+ by bicarbonate, in the presence of carbonic anhydrase, does not increase urine pCO2 because of accelerated carbonic acid dehydration; the titration of H+ by nonbicarbonate buffer consumes CO<sub>2</sub> in the process of bicarbonate regeneration and lowers urine CO<sub>2</sub>. These two processes likely occur in both hypercapnic and normocapnic rats. That urine pCO<sub>2</sub> is lower than that prevailing in the blood in hypercapnic rats but not in normocapnic rats requires either enhanced collecting duct H<sup>+</sup> secretion or excess nonbicarbonate buffer in the hypercapnic animals. The urinary nonbicarbonate buffer capacity was comparable between hypercapnic and normocapnic rats (Table V), which suggests that enhanced collecting duct H+ secretion was, at least in part, the cause of tubular CO2 removal during hypercapnia.

The basic principle in using the urinary pCO<sub>2</sub> in a highly alkaline urine as a marker of distal acidification is to estimate

the amount of pCO<sub>2</sub> generated from H<sup>+</sup> secretion and titration of bicarbonate buffer in the face of delayed carbonic acid dehydration (i.e., in the absence of carbonic anhydrase in the lumen of the distal nephron). In this regard, the difference in urine pCO<sub>2</sub> obtained in a highly alkaline urine not containing carbonic anhydrase and in an equally alkaline urine containing carbonic anhydrase should provide the best possible estimate of pCO<sub>2</sub> generated from H<sup>+</sup> secretion and titration of bicarbonate buffer. To attack this issue, we gave carbonic anhydrase to hypercapnic and normocapnic rats that were excreting an alkaline urine (Table V). This maneuver resulted in a greater fall in urine pCO<sub>2</sub> (i.e.,  $-\Delta$ pCO<sub>2</sub>) in hypercapnic rats than in normocapnic rats (Fig. 3, right). This clearly demonstrates that the amount of pCO<sub>2</sub> generated in the face of delayed carbonic acid dehydration (i.e., before carbonic anhydrase infusion) was greater in these hypercapnic rats than in controls, a finding which denotes enhanced collecting duct H<sup>+</sup> secretion. Blood pH was normal in the hypercapnic rats and alkalemic in normocapnic rats. Since changes in serosal pH (21, 45, 46) are known to modulate H+ secretion, it seems likely that the difference in urinary pCO<sub>2</sub> generation noted between hypercapnic and normocapnic rats was determined by the prevailing blood pH rather than blood pCO<sub>2</sub>. This is further suggested by the finding of a higher  $\Delta pCO_2$  in hypercapnic rats with a normal blood pH (group IV) than in those with a high blood pH (group IIa), while blood pCO<sub>2</sub> was identical in both groups (Table IV).

(c) Urinary pCO2 before bicarbonate loading. The finding that urinary pCO<sub>2</sub> is lower than systemic blood pCO<sub>2</sub> in hypercapnic rats excreting an acid urine (Fig. 2, Tables III and IV) also deserves some comment. In an acid urine, secreted H<sup>+</sup> must react with buffers other than bicarbonate because the amount of filtered bicarbonate reaching the collecting duct is very low. Carbonic acid formation is necessarily low, thereby limiting the amount of CO<sub>2</sub> generated from the dehydration of this acid. In keeping with this notion, carbonic anhydrase infusion to hypercapnic rats in which the urine contained little bicarbonate did not result in a urinary pCO<sub>2</sub> different from that of hypercapnic rats not infused with the enzyme. In other words, the enzyme had no effect on urinary pCO<sub>2</sub> because bicarbonate was virtually absent from the lumen of the collecting duct. Collecting duct H<sup>+</sup> secretion when luminal bicarbonate is negligible results in the titration of nonbicarbonate buffer, a process that causes consumption of CO<sub>2</sub> because CO<sub>2</sub> combines with hydroxyl anions in the cell to regenerate bicarbonate.

Note, however, that the mechanism(s) whereby urine pCO<sub>2</sub> is lower than that of the blood in animals excreting an acidic urine has never been definitively elucidated. Rector (47) considered several possibilities other than collecting duct H<sup>+</sup> secretion. Enhanced bicarbonate reabsorption by a nonhydrogen ion secretory mechanism could lower tubular CO<sub>2</sub> (31, 47). This is not likely, because the amount of bicarbonate reaching the collecting duct in our experiments was minimal, as judged by the extremely low urinary bicarbonate concentration obtained during acute respiratory acidosis.

Another factor that could lower urine pCO<sub>2</sub> is mixing of urine containing excess of nonbicarbonate buffer with a poorly buffered acid urine (3, 47). Assuming that the nonbicarbonate buffer in excess during acute respiratory acidosis is phosphate, carbonic acid present in acidic urine will be consumed in the following reaction:  $H_2CO_3 + Na_2HPO_4 \rightarrow NaHCO_3 + NaH_2PO_4$ . We cannot exclude a role for excess urinary

phosphate buffer in lowering urine pCO<sub>2</sub> below systemic blood pCO<sub>2</sub>, because urine phosphate concentration was higher in rats with acute respiratory acidosis than in controls. It must be emphasized that the hypothetical role of admixture of heterogeneous urine as a mechanism of lowering tubular CO<sub>2</sub> tensions does not negate the role of enhanced H<sup>+</sup> secretion as the main mechanism responsible for this phenomenon.

It is now known that during acute respiratory acidosis, renal cortical pCO<sub>2</sub> exceeds systemic blood pCO<sub>2</sub> (36) while inner medullary collecting duct pCO<sub>2</sub> is lower than arterial blood pCO<sub>2</sub> (38, 39). Therefore, tubular CO<sub>2</sub> is lost in transit from the superficial distal tubule to the inner medullary collecting tubule. Bengele et al. (39) and Gougoux et al. (32) explained the findings of an inner medullary collecting duct pCO<sub>2</sub> lower than that of the blood by postulating a loss of luminal CO<sub>2</sub> by diffusional flux into the lining epithelium. We think that this loss of tubular CO<sub>2</sub> (and thereby final urine CO<sub>2</sub>) requires that H<sup>+</sup> secretion be enhanced and cell CO<sub>2</sub> consumed in the process of bicarbonate regeneration. As H+ is secreted into a bicarbonate-poor luminal fluid, ammonium and acid phosphate are preferentially formed, thereby resulting in cell CO<sub>2</sub> consumption. This results in the creation of a lumen-to-cell and peritubular blood-to-cell CO2 gradient and tubular CO2 removal.

The titration of secreted hydrogen ions by nonbicarbonate buffer must result in the formation of either ammonium or titratable acidity or both. That urinary pH fell while titratable acidity and ammonium excretion increased in response to hypercapnia (Table II) is also consistent with the notion that the low U-B pCO<sub>2</sub> of these rats was achieved in association with enhanced collecting duct H+ secretion. The increase in ammonium excretion observed in the second hour of sustained hypercapnia was unassociated with a fall in urine pH (Table II), which suggests that ammonia production increased. This is in agreement with several studies showing that overall kidney acidification is increased during hypercapnia (48-56) and with recent work showing a prompt adaptive increase in ammoniagenesis in response to acute hypercapnia (57). While proximal nephron segments are the major site of ammonia production, it is now known that a large portion of ammonium and net acid excreted in the final urine are added to the medullary collecting during respiratory acidosis (39) as well as metabolic acidosis (58, 59).

Based on all the above considerations, we believe that there is no evidence for the notion that distal acidification is decreased during acute hypercapnia (23). In view of recent direct studies showing that hypercapnia does not increase acidification above normal in either the superficial distal tubule (60) or in the inner medullary collecting duct (39), it seems likely that during hypercapnia acidification is either normal or enhanced only in the outer portion of the medullary collecting duct and/or the cortical collecting duct.

In conclusion, this study shows that hypercapnic rats have a urinary pCO<sub>2</sub> lower than systemic blood pCO<sub>2</sub> when the urine is acid and when it is alkalinized in the face of accelerated carbonic acid dehydration. These findings suggest that during acute hypercapnia, CO<sub>2</sub> is removed from the collecting duct owing to consumption of cell CO<sub>2</sub> in the process of bicarbonate regeneration via collecting duct H<sup>+</sup> secretion and titration of nonbicarbonate buffer. During bicarbonate loading, rats with hypercapnia have a U-B pCO<sub>2</sub> lower than or equal to that of normocapnic control rats, but the rise in urine pCO<sub>2</sub> elicited by this maneuver when blood pCO<sub>2</sub> is kept constant is equal

or higher in hypercapnic rats than in normocapnic rats, depending on the prevailing blood pH. In hypercapnic rats, the U-B pCO<sub>2</sub> underestimates collecting duct H<sup>+</sup> secretion because this parameter is artificially influenced by systemic blood pCO<sub>2</sub>, a variable that does not reflect vasa recta pCO<sub>2</sub> during bicarbonate loading. The  $\Delta$ pCO<sub>2</sub> is a more appropriate qualitative index of collecting duct H<sup>+</sup> secretion than the U-B pCO<sub>2</sub>, because it is not influenced by systemic blood pCO<sub>2</sub> and takes into account the urine pCO<sub>2</sub> prevailing before bicarbonate loading. A more precise evaluation of collecting duct H<sup>+</sup> secretion requires the infusion of carbonic anhydrase to estimate the amount of pCO<sub>2</sub> generated from H<sup>+</sup> secretion and titration of bicarbonate buffer when carbonic acid dehydration is accelerated and when it is delayed.

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