

Human platelet-derived growth factor stimulates prostaglandin synthesis by activation and by rapid de novo synthesis of cyclooxygenase.

A J Habenicht, ... , B Kommerell, R Ross

J Clin Invest. 1985;75(4):1381-1387. <https://doi.org/10.1172/JCI111839>.

Research Article

Human platelet-derived growth factor (PDGF) stimulated prostaglandin (PG) E₂ synthesis in the cell cycle of Swiss 3T3 cells at two distinct time intervals, with a first plateau within 10 min and a second plateau within 2-4 h after addition of PDGF. At 4 h, the concentration of PGE₂ in PDGF-stimulated cultures exceeded the quiescent control cells by a factor of 10-15. Quiescent cells incubated with up to 16 microM exogenous arachidonic acid (AA) synthesized only small amounts of PGE₂. In contrast, 4 h after addition of PDGF, the concentration of PGE₂ synthesized from exogenous AA exceeded that in quiescent cultures by a factor of 28. The effect of PDGF stimulation on PG synthesis from exogenous AA could not be explained by growth factor-mediated increase in the cellular free AA pool as shown in experiments using [¹⁴C]AA. PDGF also stimulated synthesis of PGI₂ (prostacyclin), thromboxane, and PGF₂ alpha from exogenous AA. While inhibition of protein synthesis by 10 micrograms/ml cycloheximide had no effect on the early increase in PGE₂ synthesis, the second increase was completely prevented. Additionally, cycloheximide treatment at 6 h after PDGF stimulation resulted in rapid decline of PGE₂ synthesis from exogenous AA. Quiescent cultures pretreated with 100 microM aspirin and stimulated by PDGF thereafter recovered from cyclooxygenase inhibition within 180 min. Our results suggest that phospholipase activation and [...]

Find the latest version:

<https://jci.me/111839/pdf>



Human Platelet-derived Growth Factor Stimulates Prostaglandin Synthesis by Activation and by Rapid *De Novo* Synthesis of Cyclooxygenase

Andreas J. R. Habenicht, Matthias Goerig, Jürgen Grulich, Dietrich Rothe, Rainer Gronwald, Ulrike Loth, Gotthard Schettler, Burghard Kommerell, and Russell Ross

University of Heidelberg, Medical School, D-6900 Heidelberg, Federal Republic of Germany; University of Washington, School of Medicine, Department of Pathology, Seattle, Washington 98195

Abstract

Human platelet-derived growth factor (PDGF) stimulated prostaglandin (PG) E₂ synthesis in the cell cycle of Swiss 3T3 cells at two distinct time intervals, with a first plateau within 10 min and a second plateau within 2–4 h after addition of PDGF. At 4 h, the concentration of PGE₂ in PDGF-stimulated cultures exceeded the quiescent control cells by a factor of 10–15. Quiescent cells incubated with up to 16 μM exogenous arachidonic acid (AA) synthesized only small amounts of PGE₂. In contrast, 4 h after addition of PDGF, the concentration of PGE₂ synthesized from exogenous AA exceeded that in quiescent cultures by a factor of 28. The effect of PDGF stimulation on PG synthesis from exogenous AA could not be explained by growth factor-mediated increase in the cellular free AA pool as shown in experiments using [¹⁴C]AA. PDGF also stimulated synthesis of PGI₂ (prostacyclin), thromboxane, and PGF_{2α} from exogenous AA. While inhibition of protein synthesis by 10 μg/ml cycloheximide had no effect on the early increase in PGE₂ synthesis, the second increase was completely prevented. Additionally, cycloheximide treatment at 6 h after PDGF stimulation resulted in rapid decline of PGE₂ synthesis from exogenous AA. Quiescent cultures pretreated with 100 μM aspirin and stimulated by PDGF thereafter recovered from cyclooxygenase inhibition within 180 min. Our results suggest that phospholipase activation and resultant AA release is not sufficient to induce the burst of PG synthesis observed in PDGF-stimulated cells. Instead, PDGF stimulates PG synthesis by direct effects on the PG-synthesizing enzyme system, one involving a protein synthesis-independent mechanism and another that requires rapid translation of cyclooxygenase.

Introduction

The relation between phospholipase activation resulting in liberation of arachidonic acid (AA)¹ from complex glycerolipids and the cyclooxygenase pathway generating prostaglandins

(PGs) and thromboxanes (TXs) is not entirely clear. Upon binding of hormones or other agonists to their respective cell surface receptor, AA is released from phospholipids through activation of phospholipases (1–9). In most if not all such instances, activation of phospholipases is associated with production of PGs and TXs (1, 2, 5, 9, 10). Therefore, it is generally believed that phospholipases control formation of PGs through release of AA. Since the intracellular pool of AA is low in mammalian cells (3, 6), there is little doubt that eicosanoid biosynthesis is dependent on phospholipase activity. However, it is less clear whether liberation of AA is sufficient to induce the burst of PG, TX, and leukotriene formation observed in many agonist response systems.

We have used quiescent Swiss 3T3 cells stimulated to divide by platelet-derived growth factor (PDGF), the principal mitogen for connective tissue-derived cells (11–13), as model systems to study the relation between growth factor-induced changes in phospholipid metabolism and AA release (14–16). Cells maintained in plasma-derived serum (PDS) become arrested in the G₀/G₁ phase of the cell cycle and can be stimulated to undergo cell cycle traverse after stimulation by PDGF in a partially synchronized fashion (17). We proposed that PDGF and epidermal growth factor stimulate AA release from phosphatidylinositol, a minor acidic phospholipid in mammalian cells, by activation of the phospholipase C/diglyceride lipase pathway (14, 15). This reaction sequence was first described by Bell et al. (18) and Majerus et al. (19) in thrombin-stimulated platelets.

The question we have addressed in the present report has been whether PDGF has direct effects on the PG synthesis pathway. It was of special interest to us whether PDGF has direct stimulatory effects on the key enzyme, the cyclooxygenase. Our results show that AA release from glycerolipids is not sufficient to induce the burst in PG synthesis in PDGF-stimulated cells (15, 20, 21). Instead, the growth factor greatly activates the PG synthetic enzyme system itself. This effect of PDGF appears to be mediated by both activation and rapid *de novo* synthesis of the cyclooxygenase.

Methods

Materials. [¹⁴C]AA (sp act, 56.9 mCi/mmol), [³H]6-keto-PGF_{1α} (sp act, 120 Ci/mmol), [³H]PGF_{2α} (sp act, 150 Ci/mmol), [³H]PGE₂ (sp act, 160 Ci/mmol), [³H]TXB₂ (sp act, 139 Ci/mmol) was obtained from New England Nuclear, Boston, MA; Dulbecco's modification of Eagle's medium from Gibco Laboratories, Grand Island, NY; methanol, ethanol, ethylacetate, benzene, acetone, dioxane, acetic acid, 200-μm silica gel-coated aluminum thin-layer chromatograms from Merck Inc., Darmstadt, Federal Republic of Germany; standard PGs were purchased from Seragen, Boston, MA; antibodies against PGE₂, 6-keto-PGF_{1α}, TXB₂, and PGF_{2α} from the Institute Pasteur, Paris; cycloheximide from Sigma Chemie, Munich, Federal Republic of Germany; AA from P. L. Biochemicals, St. Goar, Federal Republic of Germany.

This work was presented in part at the International Congress of Inflammation, Paris, 3–7 September 1984.

Address reprint requests to Dr. Habenicht, University of Heidelberg, Bergheimer Str. 58, 6900 Heidelberg, Federal Republic of Germany.

Received for publication 18 September 1984 and in revised form 18 December 1984.

1. *Abbreviations used in this paper:* AA, arachidonic acid; PDS, plasma-derived serum; PG, prostaglandin; PDGF, platelet-derived growth factor; TX, thromboxane.

J. Clin. Invest.

© The American Society for Clinical Investigation, Inc.

0021-9738/85/04/1381/07 \$1.00

Volume 75, April 1985, 1381–1387

Plasma proteins and PDGF. Human or calf PDS lacking platelet-derived growth promoting activity was prepared as described (17). PDGF was prepared as described by Raines and Ross (22). PDGF at 12 ng/ml maximally stimulated DNA synthesis in Swiss 3T3 cells.

Radioimmunoassay (RIA). The concentration of PGs was determined by RIA according to Granstrom and Kindahl (23) after reversed-phase extraction of the culture medium by octadecyl C18 silica gel column chromatography according to Powell (24) and thin-layer chromatography. Briefly, 1 ml culture medium was removed from the dish and acidified to pH 3.6. 900 μ l of the culture medium was applied to an octadecyl C18 silica gel column equilibrated with methanol. The column was washed twice with 3 ml double distilled H₂O, and the PGs were eluted with a total volume of 1.5 ml ethylacetate. The ethylacetate was dried down under a stream of N₂, taken up in 1 ml methanol, and stored until further analysis at -26°C. Before thin-layer chromatography, the methanol was dried down and the extract taken up in 150 μ l ethanol, applied to a 200 μ m silica gel-coated aluminum thin-layer chromatogram, and developed in solvent system of benzene/acetone/methanol/dioxane/H₂O, 120:80:6:3:2; vol/vol. The spot containing the respective PG was scraped from the plate and extracted with 2 ml in methanol, dried down under a stream of N₂, and taken up in phosphate-buffered saline. Procedural losses were monitored by adding tracer [³H]PGs to each sample. Recoveries of PGE₂ ranged from 34–61%, for 6-keto-PGF_{1 α} from 54–68%, and for TXB₂ from 48–72%. Cross-reactivity at 50% binding of PGE₂ was 10.1% with PGE₁; <0.2% with PGA₁, PGA₂, PGB₁, PGB₂, PGD₂, PGF_{1 α} , PGF_{2 α} , 6-keto-PGF_{1 α} , and TXB₂; and <0.03% with AA. Cross-reactivity at 50% binding of 6-keto-PGF_{1 α} was 6.8% with 6-keto-PGE₁, 2.2% with PGF_{2 α} , 0.7% with PGE₁, 0.5% with PGE₂, <0.1% with all other PGs tested, and <0.3% with AA. Water samples subjected to exactly the same treatment as the biological samples ranged between 0 and 7 pg/ml for PGE₂ and <5–15 pg/ml for 6-keto-PGF_{1 α} . Intraassay variation for PGE₂ was 2–9% and for 6-keto-PGF_{1 α} was 7–11%. Interassay variation for PGE₂ was 5–12% and for 6-keto-PGF_{1 α} was 8–22%.

Results

Kinetics of PGE₂ synthesis in the cell cycle of Swiss 3T3 cells stimulated by PDGF. The synthesis kinetics of PGE₂ in quiescent and PDGF-stimulated cells is shown in Fig. 1, A and B. We consistently observed in five experiments an early burst of PGE₂ formation that was complete within 10 min. At 2 min, PDGF caused a statistically significant increase in PGE₂ formation (Fig. 1 B). No further increase was observed until 2–4 h (Figs. 1 and 3). At that time, another sharp rise in PGE₂ levels was observed. No additional increase was detected for the next 18 h. Thus, PDGF stimulated PGE₂ synthesis in the cell cycle during two distinct time intervals: a first early increase that was complete within 10 min, and a second increase between 2 and 4 h.

PDGF stimulates PGE₂ synthesis from exogenous AA. To test the ability of the cells to metabolize exogenous AA to PGE₂, we incubated quiescent and PDGF-stimulated cells with increasing concentrations of AA (Fig. 2). Though the quiescent cultures were receiving concentrations of AA as high as 16 μ M, they synthesized only small amounts of PGE₂. In contrast, cells stimulated by PDGF synthesized up to 100 ng PGE₂/10⁶ cells in the first 60 min of the cell cycle in all of five experiments. Concentrations between 4 and 16 μ M AA did not further increase PGE₂ synthesis both in quiescent and PDGF-stimulated cells. Therefore, a concentration of 10 μ M AA was chosen for all future experiments. The ability of PDGF to stimulate PGE₂ biosynthesis from exogenous AA might have been due to a PDGF-mediated increase in the

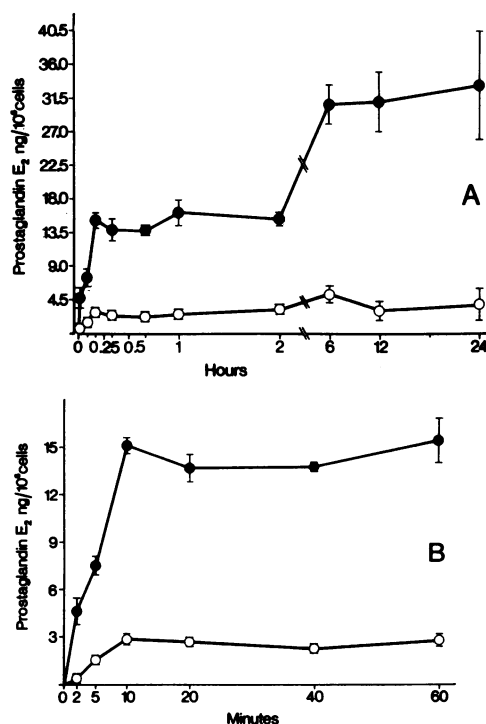


Figure 1. Kinetics of PGE₂ synthesis in the cell cycle of Swiss 3T3 cells stimulated by PDGF. Swiss 3T3 cells were plated at a density of 1.25×10^5 cells in 35-mm Costar plates (Costar, Cambridge, MA) containing 1.5 ml of Dulbecco's modification of Eagle's medium (Gibco Laboratories) supplemented with 100 U/liter penicillin, 100 μ g/ml streptomycin, and 1.25 mg/ml PDS protein. On day 3, the cultures were fed with the same medium and divided into two groups, and received either 50 μ l 10 mM acetic acid (○) or 12 ng/ml PDGF in a concentrated solution of 50 μ l 10 mM acetic acid (●). 12 ng/ml PDGF stimulated 1.25×10^5 cells to double within 36 h. The same amount of PDGF stimulated incorporation of [*methyl*-³H]thymidine into trichloroacetic acid-precipitable material by a factor of 12–35 when assayed between 23 and 24 h. At the time points indicated in the figure (A, hours; B, minutes), 1 ml culture medium was removed from the dish and acidified to pH 3.6. The concentration of PGE₂ was determined by RIA as described under Methods. Treatment of the cultures using 100 μ M aspirin completely prevented formation of PGE₂. The points indicated represent the means of three parallel cultures \pm SD.

cellular free AA pool, thus making more AA available to the cyclooxygenase. Therefore, we determined the effects of PDGF on this pool after various times of stimulation with the growth factor (Table I). PDGF had only minor effects on free [¹⁴C]AA. Analysis of individual phospholipids by two-dimensional thin-layer chromatography indicated that PDGF stimulated [¹⁴C]AA incorporation into phosphatidic acid, phosphatidylinositol, and triacylglycerol (16).

We next determined the cell cycle kinetics of PGE₂ synthesis from exogenous AA. As shown in Fig. 3, the second rise in endogenous PGE₂ synthesis which occurred between 2 and 4 h was associated with a second sharp increase in the formation of PGE₂ from exogenous AA. The concentration of PGE₂ in PDGF-stimulated cultures incubated with 10 μ M AA for 60 min rose from 60 to 100 ng/10⁶ cells in the first 2 h up to 280 ng/10⁶ cells at 4 h in all of four experiments. Cells that had been stimulated by PDGF at time 0 and were incubated with 10 μ M AA from 5 to 6 h synthesized up to 90 times more

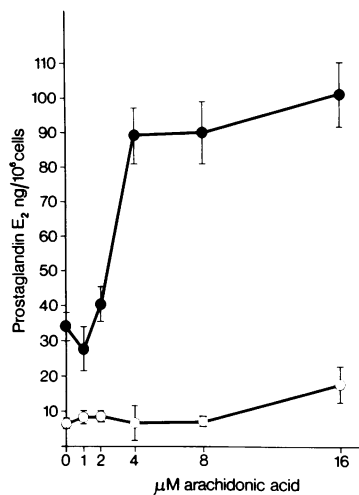


Figure 2. Synthesis of PGE₂ from exogenous AA in quiescent and PDGF-stimulated Swiss 3T3 cells. Swiss 3T3 cells were cultured as described in Fig. 1. On day 3, the cultures were fed with medium containing 1.25 mg/ml PDS protein and divided into two groups; one group received 12 ng/ml PDGF in a concentrated solution of 50 μ l 10 mM acetic acid and increasing concentrations of AA dissolved in 10 μ l ethanol (●), to give a final concentration of AA as indicated. Control cultures received 50 μ l 10 mM

acetic acid and increasing concentrations of AA (○). Cultures that were not incubated with AA received 10 μ l ethanol instead. AA was obtained from P. L. Biochemicals (St. Goar, Federal Republic of Germany), and multiple aliquots of a 10-mM stock solution were stored in ethanol under argon at -85°C . After 60 min, upon addition of PDGF and AA/ethanol, PGE₂ levels were determined as described under Methods. The data points represent the means of three parallel cultures \pm SD.

PGE₂ when compared with quiescent cells and up to 28 times when compared with quiescent cells incubated with 10 μ M AA.

PDGF stimulates prostacyclin and TXB₂ synthesis from exogenous AA. Since PDGF greatly stimulated PGE₂ synthesis from exogenous AA without comparable stimulation of the intracellular [¹⁴]AA pool, it seemed that the growth factor had direct stimulatory effects on one or more enzymes involved in PG synthesis. To test this hypothesis further, we determined the levels of 6-keto-PGF_{1 α} (the stable hydrolysis product of prostacyclin), another major cyclooxygenase-derived product

Table I. Effect of PDGF on Radioactivity of the Cellular Free Fatty Acid Pool Using [¹⁴C]AA

Additions	1 h	2 h	16 h
	dpm \times 10 ³ /dish	dpm \times 10 ³ /dish	dpm \times 10 ³ /dish
Control	4.8 \pm 0.6	6.6 \pm 2.2	10.2 \pm 1.2
+PDGF	5.8 \pm 0.6	10.4 \pm 0.8	11.6 \pm 0.6

Swiss 3T3 cells were cultured as described in Fig. 1. On day 3, the cultures were divided into two groups and either received 50 μ l 10 mM acetic acid (control) or 4 μ g/ml partially purified PDGF dissolved in a concentrated solution of 50 μ l 10 mM acetic acid. The amount of PDGF present in this preparation is equivalent to the amounts of highly purified PDGF used in the experiments in Figs. 1–3 and 5–7. At time 0, the cultures were incubated with 11.7 μ M [¹⁴C]AA dissolved in 10 μ l ethanol. At the time points indicated, the culture medium was removed and the cell monolayer washed three times with ice-cold phosphate-buffered saline. Free fatty acids were extracted and separated by thin-layer chromatography as described (14). The silica gel was transferred to a scintillation vial and radioactivity was determined in a PW 4700 Philips Scintillation Spectrometer (Philips, Eindhoven, The Netherlands). The numbers represent the means of three parallel cultures \pm SD.

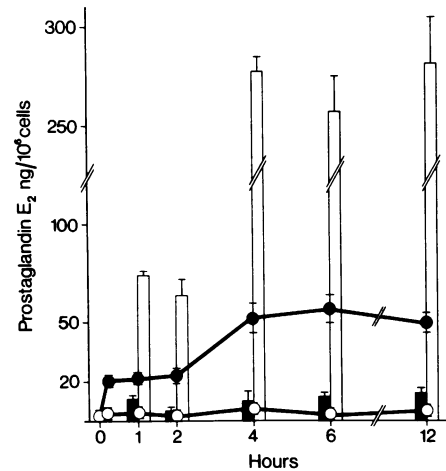


Figure 3. Kinetics of PGE₂ synthesis from exogenous AA in quiescent and PDGF-stimulated Swiss 3T3 cells. Swiss 3T3 cells were cultured as described in Fig. 1. On day 3, the cultures were fed with medium containing 1.25 mg/ml PDS protein, divided into two groups, and received either 50 μ l 10 mM acetic acid (○) or 12 ng/ml PDGF in a concentrated solution of 50 μ l 10 mM acetic acid (●). Closed bars represent cultures that received 50 μ l 10 mM acetic acid at time 0 and 10 μ M AA dissolved in a concentrated solution of 10 μ l ethanol 60 min before each time point indicated. Open bars represent cultures that received 12 ng/ml PDGF in a concentrated solution of 50 μ l 10 mM acetic acid at time 0 and 10 μ M AA dissolved in a concentrated solution of 10 μ l ethanol 60 min before the bars indicated in the figure. PGE₂ levels were determined as described under Methods. Cultures that did not receive AA were incubated with 10 μ l ethanol instead. The data represent the means of three parallel cultures \pm SD.

of AA in Swiss 3T3 cells (25), over a period of 12 h (Fig. 4). 6-keto-PGF_{1 α} levels increased up to 2 h upon stimulation by PDGF and then remained relatively constant or increased slightly for the next 10 h. In addition, PDGF-stimulated 6-keto-PGF_{1 α} formation from exogenous AA (Fig. 4). When PDGF stimulated cells incubated with AA were compared with quiescent cells incubated with AA, it became apparent that the growth factor stimulated 6-keto-PGF_{1 α} synthesis by a factor of 27–35 in two experiments. Thus, the degree of stimulation of 6-keto-PGF_{1 α} formation from exogenous AA was similar to that of PGE₂ synthesis (Fig. 3). Other experiments demonstrated comparable effects of PDGF on synthesis of TXB₂ and PGF_{2 α} from exogenous AA (results not shown).

PDGF stimulates PG synthesis by two distinct mechanisms. To obtain further information on the mechanism of PG synthesis stimulation by the growth factor, we examined the effect of the protein synthesis inhibitor cycloheximide on synthesis of PGE₂ (Table II). The concentration of cycloheximide used inhibited protein synthesis >90% as determined by [¹⁴C]leucine incorporation into trichloroacetic acid-precipitable material. While cycloheximide did not inhibit the early increase of endogenous PGE₂ synthesis, the second increase was completely prevented. Since this effect of cycloheximide might have been due to protein synthesis-sensitive phospholipase activation and AA release, we determined PGE₂ synthesis after cycloheximide treatment from exogenous AA (Table II). Cycloheximide did not alter the early increase of PGE₂ synthesis from exogenous AA while it completely prevented the second increase. In other experiments, PDGF-stimulated cells were

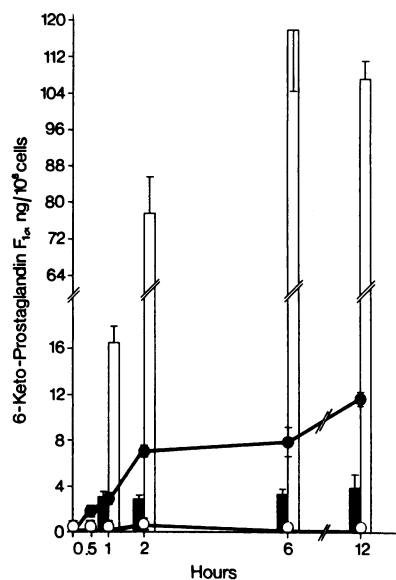


Figure 4. Kinetics of PGI₂ synthesis in the cell cycle of Swiss 3T3 cells stimulated by PDGF. Swiss 3T3 cells were cultured as described in Fig. 1. On day 3, the cultures were fed with medium containing 1.25 mg/ml PDS protein, divided into two groups, and received either 50 μ l 10 mM acetic acid (o) or 4 μ g/ml partially purified PDGF dissolved in 50 μ l 10 mM acetic acid (●). This amount of partially purified PDGF is equivalent in PDGF content to the amounts of highly purified PDGF used in the experiments detailed in Figs. 1–3 and 5–7. Closed bars represent cultures that received 50 μ l 10 mM acetic acid at time 0 and, 60 min before the bars, 10 μ M AA dissolved in 10 μ l ethanol. Cultures that did not receive AA were incubated with 10 μ l ethanol instead. PGI₂ levels were determined by RIA of 6-keto-PGF_{1 α} , the stable degradation product of PGI₂ as described under Methods. Culture medium was treated as described for PGE₂. However, before thin-layer chromatography, PG extracts from two cultures were pooled to obtain sufficient amounts of 6-keto-PGF_{1 α} for determination by RIA. The data represent the means of three parallel pooled duplicate cultures \pm SD.

incubated with cycloheximide at 6 h. A rapid decline in PGE₂ synthesis from exogenous AA was observed (Fig. 5). These results showed that the growth factor stimulated the PG synthetic enzyme system by two independent mechanisms: one that was insensitive to protein synthesis inhibition and another requiring translational activity. It is noteworthy that

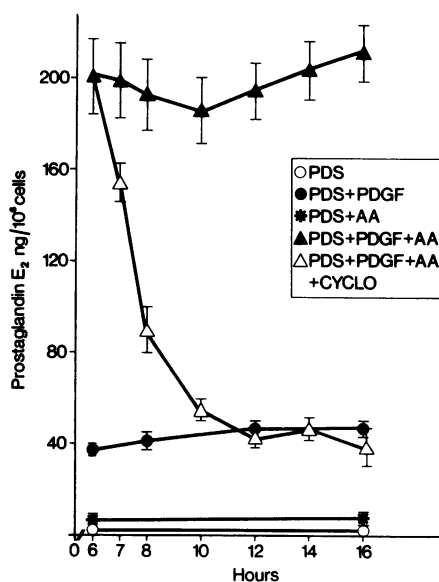


Figure 5. Kinetics of PGE₂ synthesis from exogenous AA in PDGF-stimulated Swiss 3T3 cells incubated with cycloheximide. Swiss 3T3 cells were cultured as described in Fig. 1. On day 4, the cultures were fed with medium containing 1.25 mg/ml PDS protein, divided into two groups, and received either 50 μ l 10 mM acetic acid or 12 ng/ml PDGF in a concentrated solution of 50 μ l 10 mM acetic acid. 60 min before the time points indicated, parallel cultures were incubated with 10 μ l ethanol or 10 μ M AA dissolved in a concentrated solution of 10 μ l ethanol. 6 h after stimulation with PDGF, parallel cultures received 10 μ g/ml cycloheximide. PGE₂ was determined as described under Methods. The data represent the means of three parallel cultures \pm SD.

cycloheximide did not compromise the viability of the cells as judged by trypan exclusion under our experimental conditions.

PDGF stimulates activation and rapid de novo synthesis of cyclooxygenase. To determine whether PDGF had direct effects on cyclooxygenase, we preincubated quiescent cells with aspirin (26). We then determined the kinetics of recovery by measurement of PGE₂ synthesis from exogenous AA upon removal of aspirin and stimulation with PDGF (Fig. 6). The stimulated cells largely recovered between 2 and 4 h. Furthermore, aspirin treatment resulted in up to 45% enhanced PGE₂ synthesis from exogenous AA when compared with PDGF-stimulated

Table II. Effects of Cycloheximide on PGE₂ Synthesis from Endogenous and Exogenous AA in Quiescent and PDGF-stimulated Swiss 3T3 Cells

Hours after addition of PDGF	PDS	PDS plus PDGF	PDS plus cycloheximide	PDS plus PDGF plus cycloheximide	PDS plus AA	PDS plus PDGF plus AA	PDS plus AA plus cycloheximide	PDS plus AA PDGF plus cycloheximide
<i>Prostaglandin E₂ ng/10⁶ cells</i>								
1	2.1 \pm 0.3	24.0 \pm 3.5	2.8 \pm 0.1	22.0 \pm 1.1	7.9 \pm 1.5	109.0 \pm 5.9	6.4 \pm 0.8	104.2 \pm 6.9
3	3.8 \pm 0.3	47.2 \pm 1.4	3.6 \pm 1.7	26.4 \pm 3.2	7.2 \pm 0.8	216.0 \pm 17.3	6.2 \pm 2.8	102.4 \pm 20.8

Swiss 3T3 cells were cultured as described in Fig. 1. On day 4, the cultures were fed with medium containing 1.25 mg/ml PDS protein, divided into two groups, and received either 50 μ l 10 mM acetic acid or 12 ng/ml PDGF dissolved in a concentrated solution of 50 μ l 10 mM acetic acid. 90 min before the time points indicated in the table, parallel cultures were incubated with 10 μ g/ml cycloheximide dissolved in 50 μ l phosphate-buffered saline. 60 min before the time points indicated, parallel cultures were incubated with 10 μ M AA dissolved in 10 μ l ethanol. Control cultures received 10 μ l ethanol instead. PGE₂ levels were determined as described in Fig. 1. The data represent the means of three parallel cultures \pm 1 SD.

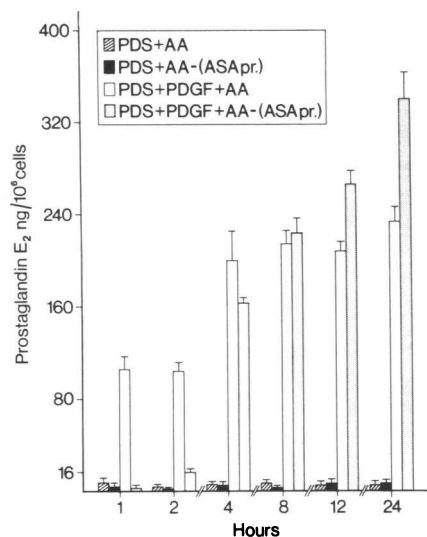


Figure 6. Recovery of PGE₂ synthesis from exogenous AA after inhibition of cyclooxygenase by aspirin in quiescent Swiss 3T3 cells stimulated by PDGF. Swiss 3T3 cells were cultured as described in Fig. 1. On day 4, the cultures were fed with medium containing 1.25 mg/ml PDS protein. 30 min before the medium was changed, parallel cultures were incubated with 50 μ l phosphate-buffered saline or 100 μ M aspirin (ASA pr.) dissolved in 50 μ l phosphate-buffered saline. Before replacement of the medium containing PDS protein, the monolayers were washed twice with 2 ml phosphate-buffered saline to remove residual aspirin. At the time of the medium change, the cultures were divided into two groups and received either 50 μ l 10 mM acetic acid or 12 ng/ml PDGF in a concentrated solution of 50 μ l 10 mM acetic acid. 60 min before each time point indicated in the figure, the cultures were incubated with 10 μ M AA dissolved in a concentrated solution of 10 μ l ethanol. PGE₂ levels were determined as described under Methods. The data represent the means of three parallel cultures \pm SD.

cells in three experiments. No significant recovery from aspirin treatment was observed within the first 2 h after PDGF stimulation.

We next examined the kinetics of recovery from cyclooxygenase inhibition in PDGF-treated cells (Fig. 7). Complete recovery was observed within 180 min in PDGF-stimulated cells when cyclooxygenase inhibition was initiated between 5.5 and 7 h. The kinetics of recovery from cyclooxygenase inhibition (Fig. 7) was similar to the kinetics of inhibition of PGE₂ synthesis from exogenous AA after protein synthesis inhibition (Fig. 5). In addition, recovery from cyclooxygenase inhibition by aspirin was completely prevented by cycloheximide (results not shown).

Discussion

The experiments performed in this investigation focused on the possibility that human PDGF stimulated synthesis of PGs by direct activation of the cyclooxygenase pathway.

The generally accepted view of PG synthesis regulation has been that AA hydrolysis from complex glycerolipids represents the major control mechanism of PG biosynthesis (1–3, 6, 9, 18). However, this concept is based on circumstantial evidence; the intracellular pool of free AA, the substrate for the cyclooxygenase, is low in mammalian cells (2, 3, 5, 6, 18) and inhibitors of phospholipase activity have been shown to suppress

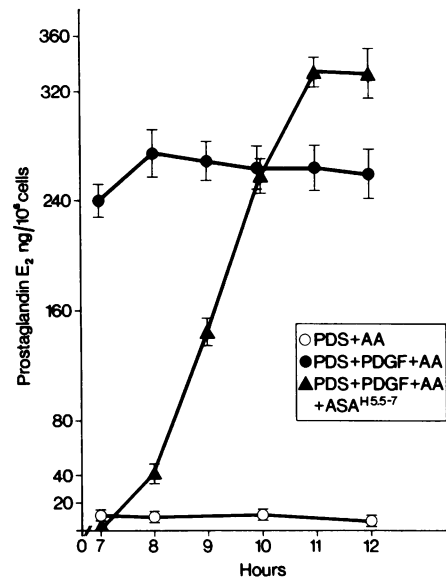


Figure 7. Kinetics of recovery from cyclooxygenase inhibition by aspirin 7 h after stimulation of quiescent Swiss 3T3 cells with PDGF. Swiss 3T3 cells were cultured as described in Fig. 1. On day 4, the cultures were fed with medium containing 1.25 mg/ml PDS protein, divided into two groups, and received either 50 μ l 10 mM acetic acid or 12 ng/ml PDGF in a concentrated solution of 50 μ l 10 mM acetic acid. Before the time points indicated, the cultures received 10 mM AA dissolved in a concentrated solution of ethanol. Between 5.5 and 7 h, parallel cultures were incubated with 50 μ l phosphate-buffered saline or 100 μ M aspirin (ASA h 5.5–7) dissolved in 50 μ l phosphate-buffered saline. Before addition of aspirin, the medium of the cultures was replaced by fresh medium containing 1.24 mg/ml PDS protein to remove PGs that had accumulated during the 5.5 h after PDGF stimulation. PGE₂ levels were determined as described under Methods. The data represent the means of three parallel cultures \pm SD.

PG synthesis from endogenous but not exogenous AA (27). Furthermore most if not all agonists eliciting increased PG synthesis concomitantly induce release of AA from phospholipids (3, 9, 14, 15, 20, 21, 28–30). While these studies are consistent with a critical role of phospholipase activity for PG synthesis, they do not provide information on several important questions: First, is AA release induced by phospholipases sufficient to induce the burst of PG synthesis observed in many agonist response systems? Second, what are the determining factors of regulation of the PG synthetic enzymes, and particularly, the cyclooxygenase? Work on mechanisms of regulation of the cyclooxygenase on the other hand has been difficult because this enzyme is subject to rapid inactivation by endoperoxide intermediates in cultured cells (31).

Several lines of published evidence suggest, however, that PG synthesis regulation is not simply the result of increased phospholipase activity. Thus, Bonser et al. (32) obtained evidence in cultured Swiss 3T3 cells that both bradykinin and thrombin stimulate PG synthesis from exogenous AA and Hyman et al. (25) showed that quiescent Swiss 3T3 cells form less PGs from exogenous AA when compared with their rapidly proliferating counterparts maintained in high serum concentrations. Furthermore, Ogekawa et al. (33) demonstrated that both the perfused hydronephrotic kidney and microsomes prepared from the renal cortex of ureter-obstructed kidneys

show greatly enhanced PGE₂ synthesis from exogenous AA. Enhanced PGE₂ synthesis from exogenous AA appears to correlate best with the proliferation of fibroblast-like cells in this system. These results are consistent with the possibility that PG synthesis regulation involves activation of the PG synthesis enzyme system independent of phospholipase activity.

In the first series of experiments presented above we showed that PDGF stimulated PGE₂ formation in the cell cycle of Swiss 3T3 cells at two distinct time intervals: a first increase that was complete within 10 min and a second increase that occurred between 2 and 4 h. These results confirmed and extended the findings of Shier (28) and Coughlin et al. (20) who showed that PDGF stimulates PG synthesis in cultured cells.

Our results using exogenous AA (Figs. 2 and 3, Table II) suggested that PDGF stimulated PG synthesis by mechanisms that are independent of phospholipase; the effect of PDGF on PGE₂ synthesis from exogenous AA did not appear to be due to the PDGF-dependent increase in the AA pool (Table I). Furthermore, quiescent cells incubated with concentrations of AA as high as 16 μM (Fig. 2) were unable to synthesize significant amounts of PGE₂ although AA was taken up by the cells.

To gain further insight into the mechanism of PDGF-dependent PGE₂ synthesis stimulation from both endogenous and exogenous AA, we next determined whether other cyclooxygenase products were stimulated by PDGF. The results presented in Fig. 4 demonstrate that PDGF stimulated 6-keto-PGF_{1α} synthesis from both endogenous and exogenous AA and other experiments revealed similar effects of the growth factor on TXB₂ and PGF_{2α} synthesis (results not shown). Since the effect of PDGF on stimulation of PG synthesis from exogenous AA included all of the major cyclooxygenase-derived products, i.e., PGE₂, 6-keto-PGF_{1α}, TXB₂, and PGF_{2α}, these results suggest that the effect of PDGF was mediated by activation of the key enzyme of PG and TX synthesis, the cyclooxygenase.

In experiments using protein synthesis inhibition, we showed that the first increase in PGE₂ synthesis from both endogenous and exogenous AA was protein synthesis independent while the second increase was completely suppressed in the presence of cycloheximide (Table II). The protein synthesis insensitivity of the first increase in PGE₂ formation might, therefore, reflect rapid activation of the cyclooxygenase pathway by posttranslational modification mechanisms. Although these results are consistent with activation of the cyclooxygenase pathway, the mechanism of activation remains to be elucidated. It is possible that phosphorylation reactions are involved since PDGF has been shown to induce, within minutes, phosphorylation of several proteins (34).

The second increase of PGE₂ synthesis observed between 2 and 4 h after stimulation of the cells was totally dependent on protein synthesis (Table II). To test whether protein synthesis was required for maintaining high PG synthesis rates from exogenous AA, we determined the kinetics of decline of PGE₂ synthesis from exogenous AA in PDGF-stimulated cells (Fig. 5). The results show that protein synthesis inhibition resulted in rapid inactivation of the cyclooxygenase pathway with approximate half-maximal inhibition between 90 and 120 min. This decline might reflect rapid turnover of enzymes involved in PG synthesis. The results are also consistent with rapid inactivation of the cyclooxygenase pathway as has been observed

by Brotherton and Hoak (31) in cultured endothelial cells. In both cases, maintenance of high PG synthesis rates required translational activity.

The requirement for protein synthesis appeared to be closely associated with cyclooxygenase activity. Thus, quiescent cells pretreated with aspirin (Fig. 6) and stimulated by PDGF thereafter largely recovered during the protein synthesis-sensitive time period, i.e., between 2 and 4 h. No significant recovery was observed during the first 2 h of the cell cycle. Furthermore, PDGF-stimulated cells that were incubated with aspirin, upon removal of aspirin, recovered from cyclooxygenase inhibition within 180 min. In three independent experiments we observed that aspirin-treated cells, after recovery from cyclooxygenase inhibition, synthesized up to 45% more PGE₂ from endogenous (not shown) and exogenous AA as compared with the control PDGF-stimulated cultures (Figs. 6 and 7). Since recovery from aspirin treatment was totally dependent on protein synthesis (results not shown), the enhanced PG synthesis rate after short-term inhibition with aspirin seemed to be due to enhanced *de novo* synthesis of cyclooxygenase after aspirin treatment. The mechanism underlying this phenomenon is presently unclear. Taken together, the results presented in Figs. 5–7 show that the PDGF-dependent stimulation of PGE₂ occurring between 2 and 4 h is mediated by rapid *de novo* synthesis of cyclooxygenase.

In conclusion, our results disclose that PDGF has profound direct stimulatory effects on the PG synthesis system in addition to the earlier observed activation of the phospholipase C/diglyceride lipase pathway (14–16).

Recently, Bailey et al. (35) reported that epidermal growth factor shares some of the activities of PDGF. It will be of major general interest to determine whether activation of cyclooxygenase by agonists represents a general mechanism of PG biosynthesis regulation. If so, our results would have important implications for a variety of physiological as well as pathophysiological conditions. These include the response of tissues to hormones and the regulation of PG, leukotriene, and lipoxin biosynthesis in inflammatory diseases.

Acknowledgments

We are indebted to Mrs. A. Röttger for technical assistance in this study and to Mrs. B. Ungemach for typing of the manuscript.

This work was supported by the Deutsche Forschungsgemeinschaft grant HA 1083/2-2 and the Forschungsrat Rauchen und Gesundheit, Hamburg.

References

1. Hammarstrom, S. 1977. Prostaglandin production by normal and transformed 3T3 fibroblasts in culture. *Eur. J. Biochem.* 74:7–12.
2. Samuelsson, B., M. Goldyne, E. Granstrom, M. Hamberg, S. Hammarstrom, and C. Malmsten. 1978. Prostaglandins and thromboxanes. *Ann. Rev. Biochem.* 47:997–1029.
3. Rittenhouse-Simmons, S., and D. Deykin. 1981. Release and metabolism of arachidonate in human platelets. In *Platelets in Biology and Pathology*, Volume 2, J. C. Gordon, editor, Elsevier/North Holland Biomedical Press, Amsterdam. 349–440.
4. Michell, R. H. 1982. Inositol lipid metabolism in dividing and differentiating cells. *Cell Calcium.* 3:429–440.
5. Majerus, P. W. 1983. Arachidonate metabolism in vascular disorders. *J. Clin. Invest.* 72:1521–1525.
6. Irvine, R. F. 1982. How is the level of free arachidonic acid controlled in mammalian cells? *Biochem. J.* 204:3–16.

7. Billah, M. M., E. G. Lapetina, and P. Cuatrecasas. 1981. Phospholipase A₂ activity specific for phosphatidic acid—a possible mechanism for the production of arachidonic acid in platelets. *J. Biol. Chem.* 256:5399–5403.
8. Bills, T. K., J. B. Smith, and M. Silver. 1977. Selective release of arachidonic acid from the phospholipids of human platelets in response to thrombin. *J. Clin. Invest.* 60:1–6.
9. Hong, S. L., and D. Deykin. 1981. The activation of phosphatidylinositol-hydrolyzing phospholipase A₂ during prostaglandin synthesis in transformed mouse BALB/3T3 cells. *J. Biol. Chem.* 256:5215–5219.
10. Michell, R. H., and C. J. Kirk. 1981. Why is phosphatidylinositol degraded in response to stimulation of certain receptors? *Trends Pharmacol. Sci.* 2:86–89.
11. Antoniadis, H. N., and A. J. Queen. 1982. Growth factors and the regulation of cell growth. *Ann. Rev. Med.* 33:445–463.
12. Westermarck, B., C. H. Heldin, B. Ek, A. Johnsson, K. Hellström, M. Nister, and A. Wasteson. 1983. Biochemistry and biology of platelet-derived growth factor. In *Growth and Maturation Factors*. H. Guroff, editors. Wiley, New York. 73–115.
13. Ross, R., and J. A. Glomset. 1979. Atherosclerosis and the arterial smooth muscle cell. *Science (Wash. DC)*. 180:1332–1339.
14. Habenicht, A. J. R., J. A. Glomset, W. C. King, C. D. Mitchell, and R. Ross. 1981. Early changes in phosphatidylinositol and arachidonic acid metabolism in quiescent Swiss 3T3 cells stimulated to divide by platelet-derived growth factor. *J. Biol. Chem.* 256:12329–12335.
15. Habenicht, A. J. R., M. Goerig, J. Grulich, D. Rothe, R. Gronwald, G. Schettler, and B. Kommerell. 1984. The phospholipase C/diglyceride lipase pathway contributes to arachidonic acid (AA) release and prostaglandin (PG) E₂ formation in platelet-derived growth factor (PDGF) stimulated Swiss 3T3 cells. *Adv. Prostaglandin Thromboxane Leukotriene Res.* In press.
16. Habenicht, A. J. R., J. A. Glomset, M. Goerig, R. Gronwald, J. Grulich, U. Loth, and G. Schettler. 1985. Cell cycle-dependent changes in arachidonic acid and glycerol metabolism in Swiss 3T3 cells stimulated by platelet-derived growth factor. *J. Biol. Chem.* 260:1370–1373.
17. Habenicht, A. J. R., J. A. Glomset, and R. Ross. 1980. Relation of cholesterol and mevalonic acid to the cell cycle in smooth muscle and Swiss 3T3 cells stimulated to divide by Platelet-Derived Growth Factor. *J. Biol. Chem.* 255:5134–5140.
18. Bell, R. L., D. A. Kennerly, N. Stanford, and P. W. Majerus. 1979. Diglyceride lipase: a pathway for arachidonate release from human platelets. *Proc. Natl. Acad. Sci. USA.* 76:3238–3241.
19. Majerus, P. W., S. M. Prescott, S. L. Hofman, E. J. Neufeld, and D. B. Wilson. 1983. Uptake and release of arachidonate by platelets. *Adv. Prostaglandin Thromboxane Leukotriene Res.* 11:45–52.
20. Coughlin, S. R., M. A. Moskowitz, B. R. Zetter, H. N. Antoniadis, and L. Levine. 1980. Platelet-dependent stimulation of prostacyclin synthesis by Platelet-Derived Growth Factor. *Nature (Lond.)*. 288:600–602.
21. Rozengurt, E., P. Stroobant, M. D. Waterfield, T. F. Deuel, and M. Keehan. 1983. Platelet-Derived Growth Factor elicits cyclic AMP accumulation in Swiss 3T3 cells: role of prostaglandin production. *Cell.* 34:265–272.
22. Raines, E. W., and R. Ross. 1982. Platelet-derived growth factor. I. High yield purification and evidence for multiple forms. *J. Biol. Chem.* 257:5154–5160.
23. Granström, E., and H. Kindahl. 1978. Radioimmunoassay of prostaglandins and thromboxanes. *Adv. Prostaglandin Thromboxane Res.* 5:119–210.
24. Powell, W. S. 1982. Rapid extraction of arachidonic acid metabolites from biological samples using octadecylsilyl silica. *Methods Enzymol.* 86:467–477.
25. Hyman, B. T., L. L. Stoll, and A. A. Spector. 1982. Prostaglandin production by 3T3-L1 cells in culture. *Biochim. Biophys. Acta.* 713:375–385.
26. Roth, G. J., N. Stanford, and P. W. Majerus. 1975. Acetylation of prostaglandin synthetase by aspirin. *Proc. Natl. Acad. Sci. USA.* 72:3073–3076.
27. Flower, R. J., and G. J. Blackwell. 1979. Antiinflammatory steroids induce biosynthesis of a phospholipase A₂ inhibitor which prevents prostaglandin generation. *Nature (Lond.)*. 278:456–459.
28. Shier, W. T. 1980. Serum stimulation of phospholipase A₂ and prostaglandin release in 3T3 cells is associated with platelet-derived growth promoting activity. *Proc. Natl. Acad. Sci. USA.* 77:137–141.
29. Michell, R. H. 1975. Inositol phospholipids and cell surface receptor function. *Biochim. Biophys. Acta.* 415:81–147.
30. Sawyer, S. T., and S. Cohen. 1981. Enhancement of calcium uptake and phosphatidylinositol turnover by Epidermal Growth Factor in A-431 cells. *Biochemistry.* 20:6280–6286.
31. Brotherton, A., and J. C. Hoak. 1983. Prostacyclin biosynthesis in cultured vascular endothelium is limited by deactivation of cyclooxygenase. *J. Clin. Invest.* 72:1255–1261.
32. Bonser, R. W., K. A. Chandrabose, and P. Cuatrecasas. 1980. Thrombin and bradykinin modulate prostaglandin synthetase independently of phospholipase. *Adv. Prostaglandin Thromboxane Res.* 6:259–262.
33. Ogekawa, T., P. E. Jonas, K. De Schryver, A. Kawasaki, and P. Needleman. 1983. Metabolic and cellular alterations underlying the exaggerated renal prostaglandin and thromboxane synthesis in ureter obstruction in rabbits. Inflammatory response involving fibroblasts and mononuclear cells. *J. Clin. Invest.* 71:81–90.
34. Cooper, J. A., D. F. Bowen-Pope, E. Raines, R. Ross, and T. Hunter. 1982. Similar effects of platelet-derived growth factor and epidermal growth factor on the phosphorylation of tyrosine in cellular proteins. *Cell.* 31:263–273.
35. Bailey, J. M., B. Muza, T. Hla, and J. Pash. 1984. Role of epidermal growth factor (EGF) in synthesis of cyclooxygenase. Kyoto Conference on Prostaglandins, Kyoto, Japan. 52–3. (Abstr.)