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Research Article

The relationship between alveolar PO₂ and the rate of O-demethylation of p-nitroanisole, a model substrate for cytochrome P-450 -linked mixed-function oxidation, was evaluated in the isolated rabbit lung perfused with Krebs-Ringer bicarbonate buffer. The appearance of the product, p-nitrophenol, in the pulmonary perfusate was measured spectrophotometrically. The PO₂ of the ventilating gas was varied with an accurate gas mixing pump and measured with an electrochemical O₂ analyzer. In control lungs ventilated with 5% CO₂ in air, the rate of p-nitrophenol production was approximately equal to 3.1 ± 0.04 (mean ± SE; n = 9) μmol/h per g dry wt. p-Nitrophenol production was unaltered when O₂ in the ventilating gas was decreased to 1%, but it was depressed reversibly when alveolar O₂ WAS 0.1% OR LESS AND WAS ABOLISHED DURING VENTILATION WITH 0.005% O₂. The rate of the reaction was inhibited by 50% when alveolar PO₂ was 0.3 mm Hg representing and intracellular [O₂] OF approximately equal to μM. In the presence of metyrapone (0.1--1 mM), an inhibitor of cytochrome P-450-dependent reactions, p-nitrophenol production was 0.07--0.17 μmol/h per g dry wt. Ventilation of lungs with varying CO concentration in 20% O₂ resulted in 50% inhibition of p-nitrophenol production when CO concentration was 10% (CO/O₂ = 0.5). These results indicated that O-demethylation of p-nitroanisole by the lung is a cytochrome P-450-dependent reaction [...]

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Relationship between Alveolar PO₂ and the Rate of *p*-Nitroanisole *O*-Demethylation by the Cytochrome P-450 Pathway in Isolated Rabbit Lungs

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ABSTRACT The relationship between alveolar PO₂ and the rate of *O*-demethylation of *p*-nitroanisole, a model substrate for cytochrome P-450-linked mixed-function oxidation, was evaluated in the isolated rabbit lung perfused with Krebs-Ringer bicarbonate buffer. The appearance of the product, *p*-nitrophenol, in the pulmonary perfusate was measured spectrophotometrically. The PO₂ of the ventilating gas was varied with an accurate gas mixing pump and measured with an electrochemical O₂ analyzer. In control lungs ventilated with 5% CO₂ in air, the rate of *p*-nitrophenol production was $\approx 3.1 \pm 0.04$ (mean \pm SE; $n = 9$) $\mu\text{mol/h}$ per g dry wt. *p*-Nitrophenol production was unaltered when O₂ in the ventilating gas was decreased to 1%, but it was depressed reversibly when alveolar O₂ was 0.1% or less and was abolished during ventilation with 0.005% O₂. The rate of the reaction was inhibited by 50% when alveolar PO₂ was 0.3 mm Hg representing an intracellular [O₂] of $\approx 0.4 \mu\text{M}$. In the presence of metyrapone (0.1–1 mM), an inhibitor of cytochrome P-450-dependent reactions, *p*-nitrophenol production was 0.07–0.19 $\mu\text{mol/h}$ per g dry wt. Ventilation of lungs with varying CO concentration in 20% O₂ resulted in 50% inhibition of *p*-nitrophenol production when CO concentration was 10% (CO/O₂ = 0.5). These results indicated that *O*-demethylation of *p*-nitroanisole by the lung is a cytochrome P-450-dependent reaction and that its rate is not affected until alveolar PO₂ is < 1 mm Hg.

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INTRODUCTION

The mammalian lung has the capacity to metabolize drugs and other foreign compounds by the microsomal mixed-function oxidation system (1–4). This system requires molecular oxygen and NADPH, and consists of a flavoprotein (NADPH-cytochrome P-450 reductase) and cytochrome P-450 which is the terminal oxidase (5). Although the precise physiological role of the cytochrome P-450-linked mixed-function oxidation pathway in the lung has not been defined, it may function in the biotransformation of toxic or other compounds that reach the pulmonary parenchyma through the airways or circulation. This biotransformation may inactivate some compounds (2) but in other cases may generate toxic (6) or carcinogenic species (7). It has been suggested recently that the cytochrome P-450 system may have additional functions in the lung to facilitate oxygen diffusion (8) or to mediate the pulmonary arterial response to hypoxia (9, 10). These latter potential roles would be expected to be critically dependent upon the affinity of the pulmonary cytochrome P-450 system for O₂. However, this information for the pulmonary cytochrome P-450 system is lacking. In the present study, we have used the intact rabbit lung to investigate the relationship between alveolar PO₂ and the rate of metabolism of *p*-nitroanisole to its product, *p*-nitrophenol. We have recently shown that this reaction can be used to study factors that influence the rate of mixed-function oxidation in the intact lung (3).

METHODS

Animals. Male weanling Dutch-Belt rabbits (Skippack Farms, Skippack, Pa.), weighing 0.4–0.9 kg, were allowed free access to food (Wayne Rabbit Ration, Allied Mills, Chicago, Ill.) and water. Rabbits were anesthetized with intravenous pentobarbital, 50 mg/kg body wt.

Lung perfusion. Lungs were removed from rabbits and placed in a water-jacketed incubation chamber maintained at 37°C as described previously (3). Lungs were ventilated at a frequency of 80/min and 3.5 ml tidal volume using an

animal respirator (Harvard Apparatus Co., Inc., Millis, Mass.). An end-expiratory pressure of 2–3 cm of water was maintained. Lungs were perfused using a peristaltic pump Model 1210, Harvard Apparatus Co., Inc. at 30 ml/min with Krebs-Ringer bicarbonate solution (pH 7.4) containing 5 mM glucose.

Two different perfusion systems were used in these experiments. With one system, lungs were perfused in a once-through circuit as described previously (3). The perfusion medium was maintained in large reservoirs in a water bath at 37°C. The perfusate in the reservoirs were gassed for at least 30 min before use with fritted glass gas dispersion tubes (Arthur H. Thomas, Co., Philadelphia, Pa.). After passing through the lung, the perfusate dripped from the transected left atrium into a small collection chamber of 3–5-ml volume. A sample of the lung effluent was withdrawn continuously from the chamber for analysis (see below) and the remainder was discarded. The second method for lung perfusion was a recirculating system with a 60-ml perfusate volume. The effluent from the transected left atrium was collected in the incubation chamber, passed through an aerator, and then reinfused into the lung as described previously for the rat and guinea pig isolated lung preparations (11). Mixing of the lung effluent with the recirculating perfusate was found to be virtually complete in 2 min. The pressures required for lung ventilation and perfusion were continuously monitored with strain gauges (3). The PO₂ of the gases used for ventilation and of the expired gas from the perfused lung was measured with an O₂ analyzer (model S-3A, Applied Electrochemistry, Inc., Sunnyvale, Calif.) which had been calibrated with dry room air. The accuracy of the calibration was checked with analyzed low O₂ standards of 5 and 0.1% O₂ in N₂ (Aircro Industrial Gases, Aircro Inc., Murray Hill, N. J.). The measurement error was <3% of the meter reading. It should be noted that the electrical signal from this oxygen analyzer is a function of the difference between the unknown gas and room air and, therefore, it is well-suited to measure very low PO₂ values.

The gas used for ventilation of lungs and aeration of the perfusate under control conditions was 5% CO₂ in air. Low O₂ gas mixtures were prepared "on-line" from tanks of N₂, CO₂, and 5% O₂ in N₂ using an accurate gas-mixing pump (Dial-A-Gas model M3000, Calibrated Instruments Inc., Ardsley, N. Y.). A gas mixture of 50 ppm O₂ was obtained by adsorbing the major fraction of residual O₂ in the N₂ gas with an Oxy-Trap (Alltech Associates, Inc., Arlington Heights, Ill.). Gas mixtures with varying CO content were also prepared on-line by mixing varying proportions of tanks containing 75% N₂:20% O₂:5% CO₂ and 75% CO:20% O₂:5% CO₂. All ventilating gas mixtures contained 5% CO₂ in addition to other components.

Measurement of *p*-nitrophenol production. For once-through perfusion, two separate reservoirs were maintained with and without 0.2 mM *p*-nitroanisole. Perfusion was begun with the drug-free medium. A sample of the effluent perfusate from the lung was pumped from the collection chamber through a flow cell in a spectrophotometer (model 139, Hitachi America, Ltd., San Francisco, Calif.) where the optical density at 425 nm was measured continuously (3). After a stable base line of optical density was obtained with drug-free perfusate (≈20 min), the perfusate was switched to the solution containing *p*-nitroanisole. The rate of *p*-nitrophenol production was calculated from the lung perfusion rate and the change in absorbance of the effluent using an extinction coefficient (pH 7.6) of 10.0×10^6 cm²/mol.

With the recirculating system, lungs were equilibrated for 10 min and then *p*-nitroanisole was added to the perfusate to give a concentration of 0.2 mM. Some experiments were carried out with metyrapone (CIBA Pharmaceutical Company,

Div. of CIBA-GEIGY Corp., Summit, N. J.) added to the perfusion medium. 1-ml samples of perfusate taken at 5-min intervals after addition of *p*-nitroanisole were centrifuged, the supernate was adjusted to pH 11 with NaOH, and the absorbance was measured in a spectrophotometer at 425 nm. The rate of *p*-nitrophenol production was calculated from the recirculating perfusate volume and its linear rate of increase in absorbance between 5 and 30 min of perfusion using an extinction coefficient (pH 11) of 12.4×10^6 cm²/mol.

Lungs were lyophilized to constant weight and results were expressed on the basis of dry weight.

RESULTS

Lungs perfused for the approximate 1-h duration of these experiments appeared normal with no evidence of alveolar edema. Ventilation and perfusion pressure each changed by <2 cm H₂O during perfusion. The ratio of dry weight to wet weight at the end of perfusion was >0.16. There were no differences in gross appearance, dry to wet weight ratio, ventilation pressure or perfusion pressure between experiments carried out under normoxic or hypoxic conditions, and perfusion pressure did not increase when the ventilating gas was made hypoxic. There was no detectable difference in O₂ concentration between the mixture used for ventilation and the expired gas from the isolated perfused lung even when lungs were ventilated with hypoxic gas mixtures.

Two experiments were carried out with the once-through perfusion system to determine the approximate relationship between oxygen concentration and *p*-nitrophenol production. As shown previously (3), infusion of *p*-nitroanisole into the isolated rabbit lung produced a rapid increase of optical density indicating production of *p*-nitrophenol from *p*-nitroanisole (Fig. 1). The peak rate of *p*-nitrophenol production in this experiment was 4.0 μmol/h per g dry wt. The rate of *p*-nitrophenol production did not change significantly as the O₂ concentration in the alveolar gas was decreased incrementally from 95 to 1% (Fig. 1). When the alveolar O₂ was

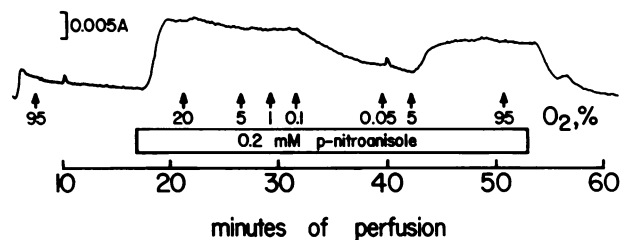


FIGURE 1 Effect of alveolar hypoxia on production of *p*-nitrophenol by isolated lungs from a Dutch-Belt rabbit perfused with a once-through system at 30 ml/min. The tracing indicates the absorbance of the effluent perfusate during perfusion. *p*-Nitroanisole was added to the perfusate as indicated by the bar. The O₂ in the ventilating gas was changed as indicated by the arrows. In all cases, the ventilating gas in addition to O₂ contained 5% CO₂ with the balance N₂.

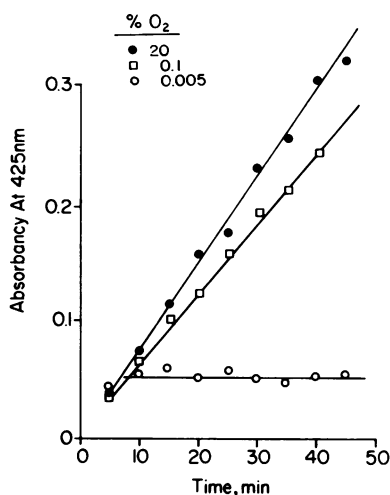


FIGURE 2 Production of *p*-nitrophenol during recirculating perfusion of isolated Dutch-Belt rabbit lungs. The absorbancy of the perfusate at 425 nm wavelength was measured in samples taken at 5-min intervals and adjusted to pH 11. Each set of points represents the result from one experiment carried out at the indicated O₂ concentration in the ventilating gas. The perfusate was equilibrated with the same gas-mixture used for ventilation. The rate of *p*-nitrophenol production was calculated from the slope of the line and the recirculating volume of perfusate (i.e., 60 ml).

TABLE I
Effect of Alveolar O₂ Concentration on *p*-Nitrophenol Production from *p*-Nitroanisole by Isolated Rabbit Lungs during Recirculating Perfusion

Alveolar O ₂	<i>p</i> -Nitrophenol production	
%	μmol/h/g dry wt	%*
20	3.1 ± 0.04 (9)	—
5	3.1 ± 0.05 (3)	100
1	3.2 ± 0.36 (4)	103
0.1	2.8 ± 0.12 (4)	90
0.05	1.8 ± 0.33‡ (3)	58
0.01	0.66 ± 0.15‡ (3)	21
0.005	0 ± 0‡ (3)	0

Results are mean standard error for number of experiments indicated in parentheses.

* Percent mean value for 20% O₂.

‡ *P* < 0.05 compared with 20% O₂ by *t* test for independent samples (12).

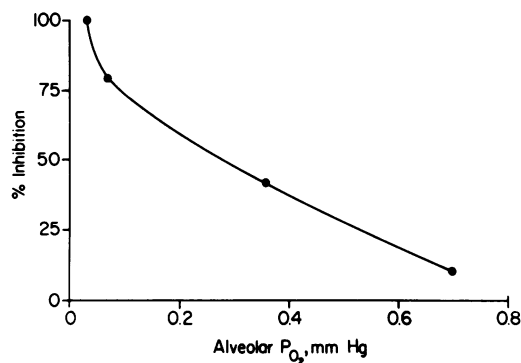


FIGURE 3 Percent inhibition of the rate of *p*-nitrophenol production as a function of the calculated PO₂ in the ventilating gas. Data points represent the mean values from Table I.

lowered to 0.1%, however, there was a rapid decrease in the absorbancy of the effluent perfusate indicating an approximate 50% decrease in the rate of *p*-nitrophenol production. A further decrement in the rate of *p*-nitrophenol production occurred with 0.05% O₂. When 5% O₂ was reintroduced, the rate of *p*-nitrophenol production increased rapidly indicating that the inhibitory effect of low O₂ concentration was reversible (Fig. 1). The results with the second experiment using once-through perfusion were similar.

29 experiments were carried out with the recirculating perfusion system. The absorbancy of the perfusate at 425 nm under both control and hypoxic conditions increased linearly with time during 45 min of perfusion (Fig. 2) indicating a constant rate of *p*-nitrophenol production. The rate of *p*-nitrophenol production was not different from control when lungs were ventilated with 5 or 1% O₂ but it decreased progressively during ventilation with O₂ concentrations below 0.1% and was inhibited totally when the ventilating gas contained 0.005% (50 ppm) O₂ (Fig. 2 and Table I). 50% inhibition of the mixed-function oxidation reaction occurred when alveolar O₂ was ≈ 0.04% (Fig. 3). This corresponds to a gas phase PO₂ of ≈ 0.3 mm Hg and an O₂ concentration in solution of ≈ 0.4 μM.

Two additional series of experiments were carried out to confirm the involvement of cytochrome P-450 in *p*-nitroanisole *O*-demethylation in the lung. In the first series, two experiments using the once-through perfusion system evaluated the relationship between alveolar CO concentration and the rate of *p*-nitrophenol production from *p*-nitroanisole. The alveolar O₂ concentration was held constant at 20% and the alveolar CO concentration was varied from 0 to 40%. The rate of *p*-nitrophenol production was inhibited by ≈ 50% when the alveolar CO was 10% (CO/O₂ = 0.5) and by 90% when the CO/O₂ was 2 (Table II). In

TABLE II

Effect of CO in Ventilating Gas on *p*-Nitrophenol Production from *p*-Nitroanisole by Isolated Rabbit Lungs during Once-Through Perfusion

CO*	CO/O ₂	<i>p</i> -Nitrophenol production [‡]		
		Expt. 1	Expt. 2	Mean
%		μmol/h/g dry wt		
0	0	6.9	6.5	6.7
5	0.25	4.7	5.1	4.9
10	0.5	3.4	3.7	3.6
20	1	1.8	1.8	1.8
40	2	0.7	0.7	0.7

* Percent CO in the ventilating gas which, in addition, contained 20% O₂, 5% CO₂, and the balance N₂.

‡ Ratio of CO to O₂ in the ventilating gas.

§ In experiment 1, the CO concentration was progressively increased, and in experiment 2, CO concentration was progressively decreased.

the second series, five experiments with the recirculating system evaluated the effect of metyrapone on *p*-nitrophenol production. In the presence of 1 mM metyrapone (three experiments), *p*-nitrophenol production was 0.10 ± 0.01 μmol/h per g dry wt. In two experiments with 0.1 mM metyrapone, *p*-nitrophenol production was 0.19 and 0.16 μmol/h per g dry wt.

DISCUSSION

The present experiments were undertaken to investigate the relationship between oxygen concentration in the alveolar gas and the rate of oxidative demethylation of *p*-nitroanisole by the pulmonary microsomal mixed-function oxidation pathway. This relationship was evaluated by determining the effect of graded alveolar hypoxia on the rate of *p*-nitrophenol production from *p*-nitroanisole by the isolated, perfused rabbit lung. A once-through perfusion system was utilized to find the approximate range of alveolar oxygen tensions that would inhibit *p*-nitrophenol production. In subsequent experiments, a recirculating system was utilized to assure steady-state conditions and to control for the duration of perfusion on the mixed-function oxidation reaction. The results for the two types of experiments were essentially the same and indicated that the rate of *p*-nitroanisole *O*-demethylation was not inhibited until very low levels of alveolar PO₂ were reached. Inhibition was not observed until alveolar PO₂ was <1 mm Hg and 50% inhibition required a calculated alveolar PO₂ of ≈ 0.3 mm Hg (intracellular [O₂] ≈ 0.4 μM).

The most likely mechanism for inhibition of *p*-nitrophenol production by low oxygen tension is the decreased availability of molecular oxygen for cyto-

chrome P-450, the terminal oxidase of the mixed-function oxidation system. It should be noted that molecular oxygen is a reactant in *p*-nitroanisole *O*-demethylation (3) as well as in other mixed-function oxidation reactions (5). Other potential mechanisms for the hypoxic effect such as reduction of ATP supply or damage to lung parenchyma are unlikely because ATP is not involved in these reactions and the effect of hypoxia on *p*-nitrophenol production was rapidly reversible.

There are several lines of evidence indicating that *p*-nitroanisole is metabolized in the lung by a cytochrome P-450-linked pathway. First, subcellular fractionation studies indicate that *p*-nitroanisole *O*-demethylase specific activity is greatest in the microsomal fraction (3). Second, activity appears to be inducible by pretreatment of animals with a phenothiazine drug (3). Third, *p*-nitroanisole *O*-demethylase activity is much greater in rabbit lungs compared with rat lungs (3), a finding which correlates with their relative cytochrome P-450 contents (13). Fourth, *p*-nitroanisole *O*-demethylation was inhibited in the presence of metyrapone, an inhibitor of cytochrome P-450 dependent reactions (14). Finally, *p*-nitroanisole *O*-demethylation was inhibited by 50% when the CO/O₂ was ≈ 0.5 (Table II); this degree of inhibition by CO is compatible with the known relative affinities of cytochrome P-450 for CO vs. O₂ of $\approx 1-2$ (15).

The relationship between PO₂ and cytochrome P-450-linked mixed-function oxidation reactions has been studied previously with isolated microsomes and with perfused liver. The *K_m* for O₂ in isolated microsomes were found to be <1 μM (16). On the other hand, the rate of *p*-nitroanisole *O*-demethylation in the isolated perfused liver was found to decrease when arterial PO₂ was lowered below 200 mm Hg (17). However, the venous outflow PO₂ in these latter experiments was zero so that the actual intracellular PO₂, which could not be determined, was probably quite low. Although the liver, because of its high rate of drug metabolism, has been widely used to study cytochrome P-450 reactions, the lung presents a unique opportunity to evaluate the relationship between oxygen and intracellular metabolic events in an intact organ since the lung cells are in close proximity to their source of oxygen (i.e., the alveolar space). In the present experiments, there should have been no uptake of O₂ from the alveolar space into the perfusate because the latter was aerated with the same gas used for lung ventilation. O₂ consumption by the normal rabbit lung parenchyma is ≈ 0.04 ml/min (13) which would represent an insignificant fraction (<2%) of O₂ delivered to the alveoli via ventilation until the O₂ content of the ventilating gas is reduced below 1%. Therefore, the approximate magnitude of the

intracellular PO₂ is well-defined at least until very low values of alveolar O₂ are reached.

The results presented in this study provide some evidence to define whether cytochrome P-450 may function physiologically as a carrier to facilitate oxygen transport or as a sensor of alveolar hypoxia as proposed by other investigators (8–10). These functions require the effects of O₂ to be manifest at PO₂ levels approximately 100-fold greater than those found to inhibit *p*-nitroanisole *O*-demethylation in the rabbit lung. If the affinity of the cytochrome P-450 system for O₂ is similar with *p*-nitroanisole and other substrates, then cytochrome P-450 could not fulfill this proposed physiological role. Therefore, the influence of PO₂ on the metabolism of other substrates for mixed-function oxidation (e.g. endogenous lipids and steroids) will have to be investigated before such a conclusion may be generalized.

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