

Interactions between Vitamin D Deficiency and Phosphorus Depletion in the Rat

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Research Article

To evaluate the role of vitamin D in the physiologic response to phosphorus depletion (P depletion) and the response to vitamin D administration in P depletion, we studied vitamin D-deficient (-D) rats, fed either a normal or low phosphorus diet and then injected intraperitoneally on alternate days with replacement vitamin D₃, 1.25 µg qod (D₃); 1.25-dihydroxy-vitamin D₃[1,25(OH)₂D₃] in physiologic, 54 ng qod (LD), and pharmacologic doses, 400 ng qod (HD); or vehicle alone (-D). The following results were obtained: (a) With P depletion, urinary excretion of inorganic phosphorus (Pi) fell to almost undetectable levels in -D rats, and two physiologic features of P depletion a calcemic effect and hypercalciuria, ensued. (b) With administration of vitamin D₃ or 1,25(OH)₂D₃ in either doses to P-depleted rats, the renal retention of Pi was unaltered despite a significant elevation of serum Pi. (c) The calcemic response to P depletion was accentuated by vitamin D sterols, and the hypercalciuria of P depletion was reduced by 1,25(OH)₂D₃, HD > LD > D₃. (d) In -D animals receiving normal Pi (+P), D₃, and 1,25(OH)₂D₃, both LD and HD produced a significant calcemic and phosphatemic effect. (e) Urinary Pi excretion in +P animals was reduced slightly by vitamin D₃ whereas 1,25(OH)₂D₃, both LD and HD, lowered urinary Pi markedly despite an increased serum Pi. (f) The serial values [...]

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ABSTRACT To evaluate the role of vitamin D in the physiologic response to phosphorus depletion (P depletion) and the response to vitamin D administration in P depletion, we studied vitamin D-deficient (-D) rats, fed either a normal or low phosphorus diet and then injected intraperitoneally on alternate days with replacement vitamin D₃, 1.25 μg qod (D₃); 1,25-dihydroxy-vitamin D₃ [1,25(OH)₂D₃] in physiologic, 54 ng qod (LD), and pharmacologic doses, 400 ng qod (HD); or vehicle alone (-D). The following results were obtained: (a) With P depletion, urinary excretion of inorganic phosphorus (Pi) fell to almost undetectable levels in -D rats, and two physiologic features of P depletion a calcemic effect and hypercalciuria, ensued. (b) With administration of vitamin D₃ or 1,25(OH)₂D₃ in either doses to P-depleted rats, the renal retention of Pi was unaltered despite a significant elevation of serum Pi. (c) The calcemic response to P depletion was accentuated by vitamin D sterols, and the hypercalciuria of P depletion was reduced by 1,25(OH)₂D₃, HD > LD > D₃. (d) In -D animals receiving normal Pi (+P), D₃, and 1,25(OH)₂D₃, both LD and HD produced a significant calcemic and phosphatemic effect. (e) Urinary Pi excretion in +P animals was reduced slightly by vitamin D₃ whereas 1,25(OH)₂D₃, both LD and HD, lowered urinary Pi markedly despite an increased serum Pi. (f) The serial values of serum Ca and Pi and urinary Ca in PD rats and the sequential values for urinary and serum Pi in +P rats indicated more rapid effects of 1,25(OH)₂D₃, both HD and LD, compared with D₃. We conclude that: (a) The renal adaptation and physiologic response to PD does not

require the presence of vitamin D. (b) 1,25(OH)₂D₃ may directly enhance the renal tubular reabsorption of Pi even as serum Pi rises. (c) A hypocalciuric action of 1,25(OH)₂D₃ in rats on low phosphorus diet could be direct or occur as a consequence of an increase in serum Pi produced by 1,25(OH)₂D₃. The different sequential renal response to D₃ compared with 1,25(OH)₂D₃ raises the possibility that other natural forms of vitamin D₃ [i.e., 25(OH)D₃, 24,25(OH)₂D₃, etc.] which may be present in vitamin D-fed rats but not those given only 1,25(OH)₂D₃, could modify the actions of 1,25(OH)₂D₃.

INTRODUCTION

With dietary phosphorus restriction and the appearance of phosphorus depletion, several physiologic events consistently occur: These include a marked reduction in urinary phosphorus excretion, the appearance of hypophosphatemia, hypercalciuria, and, in certain species, the development of hypercalcemia (1-5). There is indirect evidence in animals (3, 6) and direct measurements in humans (7) suggesting that physiologic hypoparathyroidism develops during phosphate depletion. However, it has been shown that administration of parathyroid hormone does not modify renal phosphate conservation during phosphorus depletion (8, 9) and only partially reduces the hypercalciuria (3).

It is now well established that interactions exist between phosphorus metabolism and the bioconversion and actions of vitamin D. Thus, the administration of 1,25-dihydroxy-vitamin D₃ [1,25(OH)₂D₃]¹ enhances intestinal phosphorus transport in experimental ani-

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¹Abbreviations used in this paper: +D, normal vitamin D; -D, vitamin D deficient; HD, high dose; LD, low dose; 1,25(OH)₂D₃, 1,25-dihydroxy-vitamin D₃; +P, normal phosphorus; -P, low phosphorus; Pi, inorganic phosphorus; PTH, parathyroid hormone.

mals (10) and augments phosphorus absorption in man (11). Moreover, there is potent phosphatemic action of 1,25-dihydroxy-vitamin D₃ in hypophosphatemic animals as a result of phosphorus mobilization from bone (12, 13). Furthermore, there is evidence that phosphorus depletion can stimulate the conversion of 25-hydroxy-vitamin D₃ to 1,25(OH)₂D₃ (14). Thus, an important feedback mechanism may exist whereby phosphorus restriction alters the metabolism of vitamin D, resulting in enhanced intestinal absorption of phosphorus and its mobilization from bone. These events would act to maintain phosphorus homeostasis. The possibility that alterations in vitamin D metabolism and action may play an important role in the physiologic response to phosphorus depletion has not been thoroughly examined. In the present study, the physiologic response to phosphorus depletion was evaluated in animals that were either depleted of vitamin D or replaced with known amounts of vitamin D₃ or specific quantities of 1,25(OH)₂D₃, the most active form of vitamin D.

METHODS

Weanling male Holtzman rats (Holtzman Co., Madison, Wis.) were fed an artificially prepared, vitamin D-deficient diet (15) containing 0.3% phosphorus and 0.5% calcium for 6 wk. At that time hypocalcemia was prominent (serum calcium, 1.70 ± 0.05 [SE] mM), and the animals weighed 140–160 g. The animals were then randomly separated into groups and housed in individual metabolic cages. 24-h urine samples were collected, and blood samples were obtained from the tail vein on alternate days. The animals were divided into two main groups: those receiving a low phosphorus (0.03%) diet (–P) and those receiving a normal phosphorus (0.3%) intake (+P). After the animals had received the vitamin D-deficient –P or +P diet for 3 days, each major group was then subdivided into four subgroups of 8–10 animals, each of which received either a vitamin D sterol or vehicle as indicated in Table I. The quantity of vitamin D₃ given was that considered to be a normal replacement dose (16), whereas the two levels of 1,25(OH)₂D₃ given were (a) near that needed for maintaining growth and serum calcium normal (low dose, LD), and (b) the quantity previously found to maximally stimulate intestinal calcium transport (high dose, HD).² The vitamin D sterols or vehicle were injected intraperitoneally at the end of the 24-h urinary collections for days 3, 5, 7, and 8. The vitamin D₃ was obtained from n.V. Phillips-Duphar (Weesp, The Netherlands), and the 1,25(OH)₂D₃ was provided by Dr. M. Uskokovic of Hoffmann-La Roche (Nutley, N. J.).

Food intake was measured daily, and the animals were weighed on each morning of injection. Although the animals were not pair-fed, there was no difference in dietary intake in any of the groups, except for the –P vitamin D-deficient (–D) group, which consumed less food only on days 4 and 5 ($P < 0.05$). The average daily food intake ranged from 9.0 to 11.0 g/d in the –P–D group, compared with 8.8 to 13.5 g/d in the –P groups, and 11.0 to 15.0 g/d in the +P groups. In no group was there a significant change in food intake from days 4 through 8 when the changes in excretion of calcium

² Hartenbower, D. L., M. W. Walling, and J. W. Coburn. Unpublished observations.

TABLE I
Treatment Groups and Abbreviations

Sterol treatment	Dose (every other day)	Designation	
		Dietary phosphorus	
		0.03% (–P)	0.3% (+P)
Vehicle	Ethanol:propanediol, 1:1; 0.20 ml	–P – D	+P – D
Vitamin D ₃ *	1.25 μg†	–P + D	+P + D
1,25(OH) ₂ D ₃ *	54 ng	–P + 1,25LD	+P + 1,25LD
1,25(OH) ₂ D ₃ *	400 ng	–P + 1,25HD	+P + 1,25HD

* Given in the ethanol:propanediol vehicle, 0.2 ml, by intraperitoneal injection.
† Equivalent to 50 IU.

and inorganic phosphorus (Pi) were observed. Body weight remained unchanged in all groups for days 4 through 9, although weight gain occurred in each group from days 1 to 4 when they received no vitamin D sterol. The reason for the lack of weight gain is not certain, but frequent intraperitoneal injections and tail-vein bleedings may have contributed.

Calcium was analyzed in serum by EGTA titration utilizing the Automatic Calcium Analyzer (Calcette; Precision Scientific Group, Chicago, Ill.) and in urine utilizing atomic absorption spectrometry (Perkin Elmer model 303, Perkin-Elmer Corp., Norwalk, Conn.); phosphorus was measured in serum utilizing the malachite green micromethod (17) and in urine utilizing a standard method previously reported (3). Serum urea nitrogen levels were measured in blood samples obtained on day 9 using the method of Siest et al. (18). The statistical analysis involved one-way analysis of variance, the Dunnett's tests for multiple group comparison (19), and multiple correlations carried out in a manner previously described (20). All statistical comparisons of possible physiologic importance were made, but only the more pertinent *P* values are cited in the text.

RESULTS

Serial values of serum Pi in animals fed +P or –P diets are shown in Fig. 1. In the animals receiving the normal phosphorus (P) intake, there was a significant phosphatemic effect in the groups receiving either dose of 1,25(OH)₂D₃, with mean serum Pi reaching levels of 3.25 ± 0.05 mM and 3.82 ± 0.06 mM after 24 h (day 4) in the 1,25LD and 1,25HD groups compared with 2.40 ± 0.06 mM in the –D group ($P < 0.01$). Serum Pi increased in the vitamin D-fed group (+D) but to a lesser degree ($P < 0.01$ compared with 1,25LD and 1,25HD on day 4). After day 4, serum P remained stable in 1,25LD or even fell slightly in 1,25HD; the serum Pi rose to its maximal level, 4.10 ± 0.14 mM, on day 6 in the +D group.

In animals receiving the –P diet, serum Pi decreased substantially within 48 h after initiation of –P diet. By 24 h after administration of 1,25(OH)₂D₃, serum Pi increased prominently to 1.09 ± 0.10 and 1.59 ± 0.06 mM in 1,25LD and HD groups, respectively, compared with 0.08 ± 0.012 mM in the –P animals ($P < 0.01$). Serum Pi remained low in vitamin D₃-treated animals until the end of day 6 when it reached a level similar to that

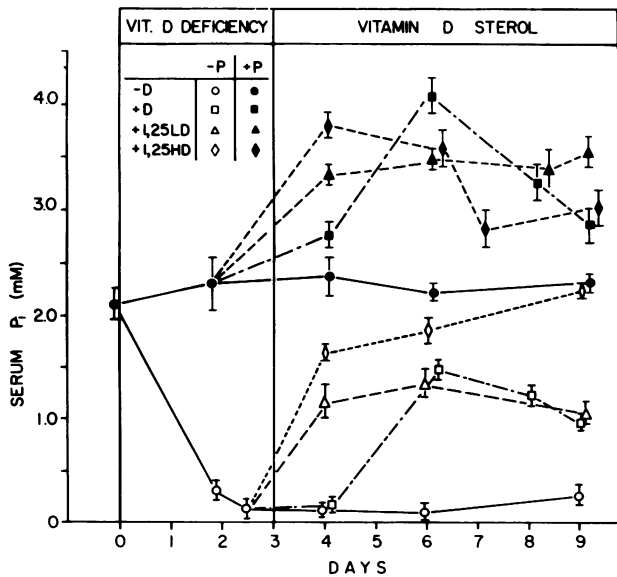


FIGURE 1 Mean values of serum phosphorus (Pi) in -D rats receiving either a +P (0.3%) or a -P (0.03%). The doses of vitamin D₃ and 1,25(OH)₂D₃, LD and HD, are given in Table I. Each point represents mean±SEM in six to eight rats.

observed in the 1,25LD animals. Serum Pi was significantly higher in the 1,25HD, -P animals than in the other -P groups throughout the study ($P < 0.01$ at all times).

The values of serum Ca in animals given the +P and -P diets are shown in Fig. 2. In the +P animals, serum Ca increased significantly to 2.20 ± 0.050 , 2.15 ± 0.052 , and 2.28 ± 0.047 mM in +D, 1,25LD, and 1,25HD, respectively, by day 4, compared with a value of 1.04 ± 0.051 mM in -D animals ($P < 0.01$). Serum

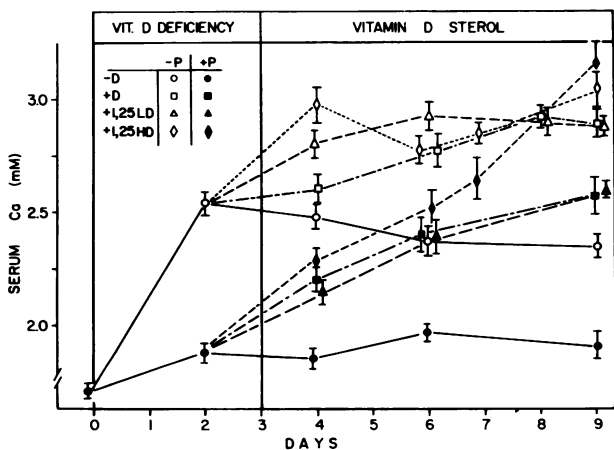


FIGURE 2 Mean±SEM values of serum Ca in -D rats continued on either a +P (0.3%) diet or given a -P (0.03%) diet and then given vitamin D₃ or different doses of 1,25(OH)₂D₃ as noted in Table I.

Ca plateaued after day 6 in the +D animals, and there was a small but statistically significant further increase in the 1,25LD rats from 2.40 ± 0.059 to 2.60 ± 0.019 mM ($P < 0.01$). Serum Ca continued to increase in 1,25HD rats, reaching 3.15 ± 0.062 mM on day 9.

In -P-D rats, serum Ca increased from 1.70 ± 0.050 to 2.62 ± 0.055 mM by day 2 of P depletion ($P < 0.01$); thereafter, serum Ca was 2.49 ± 0.068 mM on day 4 and 2.35 ± 0.025 mM on day 9 ($P = NS$). In both 1,25LD and 1,25HD rats, serum Ca increased on day 4 within 24 h of treatment ($P < 0.01$); in the +D rats, serum Ca increased more slowly, and the difference between +D and -D did not become significant until day 6 (2.75 ± 0.97 vs. 2.38 ± 0.062 , $P < 0.01$). For the 1,25HD and the 1,25LD groups, serum Ca did not show a significant change after day 4. In comparing levels of serum Ca between the -P and +P animals, the values were all significantly greater in the -P rats ($P < 0.01$) except for the values obtained in the 1,25HD group on day 9.

Rates of urinary Pi excretion are shown in Fig. 3 for both +P and -P animals. Within 24 h after administration of the -P diet, the urinary Pi fell markedly from 1.74 ± 0.12 to 0.052 ± 0.006 mmol Pi/mmol creatinine per 24 h ($P < 0.01$). During the remainder of the study, urinary Pi fell even further, and mean values were below 0.052 mmol/mmol creatinine per 24 h, regardless of the treatment given. Because of the very low quantities of Pi found in the urine, no statistical comparisons were made.

In the +P animals, urinary Pi remained relatively constant in -D rats with a small and insignificant decrease by the end of the study. In the +D rats, there

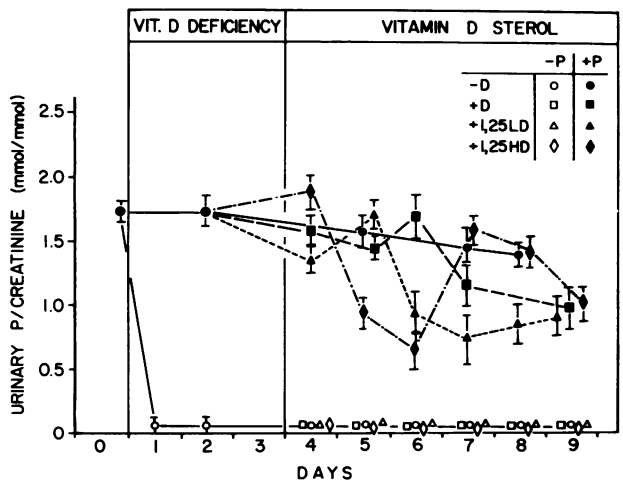


FIGURE 3 Urinary P excretion, expressed as the ratio of urinary P/urinary creatinine, in 24-h urine samples collected from -D rats fed either a +P (0.3%) or -P (0.03%) diet. The doses and regimen for treatment of vitamin sterols are given in Table I.

were no differences in urinary Pi for the first 3 d of vitamin D₃ treatment, but the values fell significantly on days 7 and 9 (compared with day 3, $P < 0.01$); however, the values did not differ from the -D group. In the 1,25HD group, urinary Pi fell significantly on days 5 and 6 ($P < 0.01$), but later increased to a level not different from that observed in the -D rats. In the 1,25LD group, urinary Pi did not fall until day 6; however, the values were lower than those on day 3 throughout the remainder of the study ($P < 0.01$). Regardless of the vitamin D sterol given, urinary Pi was higher in the +P than in the -P rats ($P < 0.01$). When one compares the relationship between urinary and serum Pi, urinary Pi was lower on day 6 in both 1,25(OH)₂D₃ groups compared with the +D group, despite higher levels of serum Pi in the former.

The values for urinary Ca for both +P and -P rats are shown in Fig. 4. In the rats receiving the +P diet, urinary Ca was low during the control period, with a mean value of 0.89 ± 0.003 mmol/mmol creatinine. There were no significant differences between urinary Ca in any +P group given any vitamin D sterol and the values in the -D+P group at any time of the study ($P > 0.05$).

In the -P rats, urinary Ca increased markedly within 24 h after initiation of the -P diet ($P < 0.01$), with a mean value of 1.41 ± 0.028 mmol/mmol creatinine per 24 h. The maximal value on the 3rd day of the -P diet was 2.61 ± 0.099 mmol/mmol creatinine per 24 h. The values fell somewhat thereafter, but urinary Ca continued to be high, and values ranged from 1.61 ± 0.12 to 2.12 ± 0.34 mmol/mmol creatinine per 24 h during the remainder of the study. In the +D group, urinary Ca did not differ from that in the -D rats except on day 8 when the mean value was marginally greater (2.65 ± 0.33 vs. 2.09 ± 0.11 mmol/mmol creatinine per 24 h, $P < 0.05$). In the 1,25LD group, mean urinary Ca fell

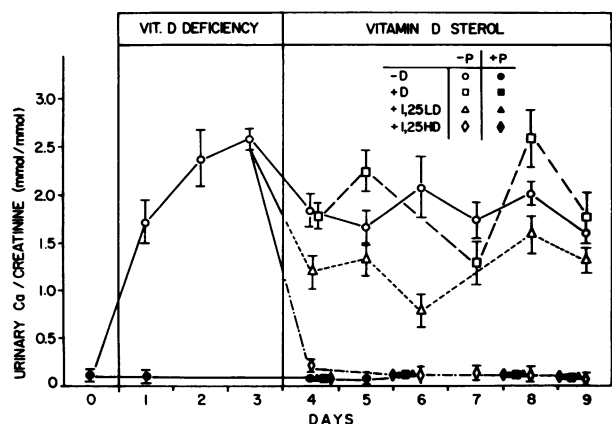


FIGURE 4 Mean values of Ca excretion, indicated as ratio of urinary Ca/creatinine in -D rats fed either a +P (0.3%) or a -P (0.03%) diet. The vitamin D sterol and dosage given are indicated in Table I.

TABLE II
Values of Serum Urea Nitrogen in Various Treatment Groups (Day 9)

Treatment group	Serum urea nitrogen	
	-P	+P
	mM	
-D	7.5±0.59	8.6±0.72
+D ₃	9.3±0.50	9.7±0.74
+1,25LD	9.1±1.2	9.4±0.86
+1,25HD	14.6±1.5*	15.3±2.0†

* Value differs from -D, $P < 0.01$, and from +D and 1,25LD, $P < 0.05$.

† Value differs from -D, +D, and +1,25LD, $P < 0.01$.

below the values observed in the -D rats on days 4 and 6 ($P < 0.01$); for the other days, urinary Ca was not significantly lower, but serum Ca was clearly higher in the 1,25LD than in -D rats throughout the study (Fig. 2). In the +1,25HD, -P rats, urinary Ca fell markedly, with values not different from those in +P animals. When urinary Ca is compared with the corresponding serum Ca, the urinary excretion was lower on the 1st and 3rd d of treatment in the 1,25(OH)₂D₃-treated groups compared with +D rats, despite a higher serum Ca in 1,25(OH)₂D₃ groups on day 4.

Levels of serum urea nitrogen obtained on day 9 are shown in Table II. The values did not differ in any of the groups except for 1,25HD, both -P and +P groups, which showed an increase.

DISCUSSION

The present results indicate that several physiologic responses to phosphate depletion, including hypophosphaturia, hypercalciuria, and a rise in serum Ca, can occur without vitamin D and develop during the administration of known quantities of 1,25(OH)₂D₃ or vitamin D₃. Steele et al. (12) have provided evidence from a short-term study that there is increased renal tubular phosphate reabsorption in -D rats given a -P diet. The results of Steele et al. (12) and observations from the present study indicate that neither vitamin D nor alterations in vitamin D metabolism are required for enhanced renal tubular reabsorption of phosphate in the face of dietary phosphate restriction. This decrease in urinary P persisted during phosphate depletion even as serum Pi was raised by an acute phosphate infusion in the study of Steele et al., and as it increased more slowly as a consequence of the continued administration of 1,25(OH)₂D₃ in the present study. These results underscore the potency of dietary phosphate restriction in augmenting tubular Pi reabsorption, observations previously made by Tröhler et al. (21) and Steele et al. (12).

The present studies demonstrate that marked hypercalciuria occurs during P depletion in the -D as well as the vitamin D₃-treated animals. Hypocalciuria is very commonly found in vitamin D deficiency in man (22); however, the present data indicate that urinary Ca may be normal or even elevated when vitamin D deficiency and P depletion coexist in the rat. Metabolic acidosis occurs in -D rats (23); this factor could influence urinary Ca; however, hypercalciuria was not reversed by vitamin D itself, nor was it seen in the +P group. Hence, any contribution of acidosis is speculative.

The initiation of phosphate restriction was associated with a calcemic effect in the -D animal, and treatment with various vitamin D sterols raised serum Ca even further. It has been shown that P depletion, per se, does not lead to increased intestinal absorption of Ca in the -D rat (24). Others found augmented release of radiocalcium from bone during P depletion produced in vivo (13) and from bone maintained in organ culture and exposed to a reduced phosphate concentration (25). Thus, bone is the most likely source of Ca that leads to both the rise in serum Ca and the hypercalciuria observed during P depletion. The present observations suggest that Ca can be mobilized from bone in response to P depletion in the absence of vitamin D. The failure of either vitamin D deficiency or the administration of small amounts of vitamin D₃ or moderate quantities of 1,25(OH)₂D₃ to modify physiologic responses to P depletion casts doubt on the suggestion that altered bioconversion of vitamin D is responsible for these events (26).

The present data demonstrate a marked decrease in urinary Ca excretion in the P-depleted, hypercalciuric rat in response to treatment with 1,25(OH)₂D₃. This effect was observed during the first 24-h period after administration of 1,25(OH)₂D₃ and was greater in rats given the large dose of 1,25(OH)₂D₃ compared with those receiving the smaller dose. Serum Ca increased at the same time; thus, there probably was an increase in tubular reabsorption of Ca. Steele et al. (12) suggested that the fraction of filtered Ca excreted was reduced by 1,25(OH)₂D₃, and Costanzo et al. (27) found lower clearance ratios of Ca/Na in rats given a single dose of vitamin D₃, 2.5 μg. Taken together, these data suggest that 1,25(OH)₂D₃ may enhance tubular Ca reabsorption when given to -D animals. However, there was an increase in serum Pi with the administration of the vitamin D sterol, and an inverse relationship existed between urinary Ca and serum Pi for all animals studied. An inverse relationship between serum Pi and urinary Ca has been previously noted in P-depleted dogs and rats (3, 12, 26), and it is possible that the alterations in serum Pi were responsible for the changes in urinary Ca observed during treatment with the vitamin D sterols. Steele et al. (12) found that the infusion of phosphate to -D rats previously fed either

0.03 or 0.1% P was associated with a substantial decrease in urinary Ca. On the other hand, studies by Rizzoli et al. (28) demonstrated that urinary Ca increased in response to the administration of 11 ng/d of 1,25-(OH)₂D₃. Moreover, the administration of 1,25(OH)₂D₃ to normal humans has generally produced an increase in urinary Ca, although this may occur because of a small increase in filtered Ca (29, 30). Thus, the action of 1,25(OH)₂D₃ may vary, depending on the Ca needs or stores of the organism.

The results of the present studies indicate that the administration 1,25(OH)₂D₃ in only moderate doses can reduce urinary Pi excretion in -D rats receiving a +P diet. This was evident by the 2nd d of 1,25(OH)₂D₃ administration (day 5) in the 1,25HD group and on day 6 in the 1,25LD group. These effects occurred earlier than observed in the vitamin D₃-treated rats. Also, urinary Pi excretion was lower in the 1,25LD or 1,25HD groups for any given level of serum Pi. Popovtzer et al. (31) and Puschett et al. (32) both reported an antiphosphaturic action of 25(OH)D₃ in rats that was dependent upon the presence of parathyroid hormone (PTH). The quantities of 25(OH)D₃ utilized in their studies were considerably greater than the amount of vitamin D₃ given here and much greater than the amount of 1,25(OH)₂D₃ employed. Costanzo et al. (27) found that a large dose of vitamin D₃, 2.5 μg, reduced urinary Pi excretion 48 h later in -D rats. The present results indicate that 1,25(OH)₂D₃, in a quantity only 2.5 times greater than that described by Bonjour et al. (33) as a "replacement" dose in parathyroidectomized rats, was associated with a definite reduction in urinary Pi excretion in -D rat. Suppression of PTH secretion could contribute to this effect, and serum levels of PTH are not available in the present study. Serum levels of Ca were no different in the +P rats receiving D₃ or the two doses of 1,25(OH)₂D₃ on days 4 through 6 of the study. Hence, one would not anticipate greater inhibition of PTH secretion with 1,25(OH)₂D₃ than with +D; also, there was no increase in urinary Ca, a finding that would be expected if there were marked suppression of PTH secretion. Bonjour et al. (33) have shown that 1,25(OH)₂D₃ may permit greater phosphate excretion in parathyroidectomized rats given a high dietary Pi intake. Thus, it is possible that 1,25(OH)₂D₃ can enhance Pi excretion in the phosphate-loaded or hyperphosphatemic state, whereas it can reduce urinary Pi excretion in a hypophosphatemic animal. Such effects could account, in part, for the effect of 1,25(OH)₂D₃ to raise serum Pi in hypophosphatemic rats and cause a fall in serum Pi in hyperphosphatemic rats (34).

The possibility that altered dietary intake or a change in renal function may have accounted for the reduction in urinary Ca or the fall in urinary Pi should be considered. Only in the -P-D rats was food intake slightly lower than in the other groups; despite this, hypercal-

ciuria was most marked in this group. Also, there was no significant day-to-day differences in food intake for any group. Thus, altered dietary intake could not account for the changes in urinary excretion. The possibility that decreased renal function may have contributed to the fall in urinary Ca and decrease in urinary P in the 1,25HD groups exists, and the serum urea nitrogen levels were slightly increased. Parallel studies carried out in our laboratory indicate that decreased renal function and nephrocalcinosis do not occur in rats treated with 1,25(OH)₂D₃ until hypercalcemia supervenes (35).³ Moreover, the fall in urinary Ca in the -P+1,25HD rats occurred as soon as day 4, and the urinary Pi fell in the +P+1,25HD group by day 5; on the other hand, the hypercalcemia only appeared on day 9 in the 1,25HD groups. The serum urea nitrogen was not increased in the +1,25LD or +D groups, which also exhibited a fall in urinary Pi and Ca. Thus, we believe it unlikely that a decrease in renal function occurred early in the +1,25HD groups when the most pronounced effect on urinary Ca and Pi were seen.

Observations of serial changes in serum Ca and Pi and urinary Ca in the -P rats and of sequential alterations in urinary Pi and serum Pi in the +P rats indicate an earlier effect of 1,25(OH)₂D₃ with both the "low" and "high" dose than observed with vitamin D₃. These findings were unexpected because the conversion of vitamin D₃ to 25(OH)D₃ and then to 1,25(OH)₂D₃ should proceed rapidly and effectively in rats with vitamin D deficiency, secondary hyperparathyroidism, and, in certain groups, P depletion as well (14). This "delayed" effect of vitamin D₃ was observed with the administration of a quantity of vitamin D₃ was observed with the administration of a quantity of vitamin D₃ shown to be highly effective in healing rickets and promoting growth (16). The reasons for the more rapid appearance of action of 1,25(OH)₂D₃ are not known, but at least two explanations may be considered. It is possible that 54 ng of 1,25(OH)₂D₃, given on alternate days, exceeds the maximal capacity for renal production of this sterol. However, this amount is only 2.5 times the quantity of 1,25(OH)₂D₃ that can restore Ca absorption to normal in parathyroidectomized rats (27). The conversion of only 4% of the administered vitamin D₃ to 1,25(OH)₂D₃ would have resulted in the generation of the quantity of 1,25(OH)₂D₃ given the +1,25LD group. Boyle et al. (36) and Norman and Wong (37) reported that as much as 25% of a single dose of vitamin D₃ was recovered as 1,25(OH)₂D₃ from various tissues of -D rat given 0.125 ng of vitamin D₃. Alternatively, it is possible that other naturally occurring vitamin D sterols, such as 25(OH)D₃ or 24,25(OH)₂D₃, etc., present in the vitamin D₃-treated

rats but not in those given 1,25(OH)₂D₃, may have actions that oppose certain isolated effects of 1,25(OH)₂D₃. Although such a suggestion is speculative, it has been shown that 25(OH)D₃ may exert effects on the skeleton different from the actions of 1,25(OH)₂D₃ (38, 39), and other data indicate that 24,25(OH)₂D₃ given in combination with 1,25(OH)₂D₃ has effects to suppress parathyroid gland weight much greater than 1,25(OH)₂D₃ alone (40). Such observations suggest that certain vitamin D sterols may have actions qualitatively different from those of other vitamin D compounds.

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REFERENCES

- Day, H. G., and E. McCollum. 1939. Mineral metabolism, growth, symptomatology of rats on diet extremely deficient in phosphorus. *J. Biol. Chem.* **130**: 269-283.
- Lotz, M., E. Zisman, and F. C. Bartter. 1966. Evidence for a phosphorus depletion syndrome in man. *N. Engl. J. Med.* **278**: 409-415.
- Coburn, J. W., and S. G. Massry. 1970. Changes in serum and urinary calcium during phosphate depletion. Studies on mechanisms. *J. Clin. Invest.* **49**: 1073-1087.
- Freeman, S., and F. C. McLean. 1941. Experimental rickets. Blood and tissue changes in puppies receiving diet very low in phosphorus with and without vitamin D. *Arch. Pathol.* **32**: 387-408.
- Aubel, C. E. 1936. The effects of low phosphorus ratios on growing pigs. *J. Agric. Res. China.* **52**: 149-159.
- Baylink, D. J., and M. Stauffer. 1971. Formation, mineralization, and resorption of bone in hypophosphatemic rats. *J. Clin. Invest.* **50**: 2519-2530.
- Dominguez, J. H., R. W. Gray, and J. Lemann, Jr. 1976. Dietary phosphate deprivation in women and men: effects on mineral and acid balances, parathyroid hormone and the metabolism of 25-OH-vitamin D. *J. Clin. Endocrinol. Metab.* **43**: 1056-1068.
- Tröhler, U., J. P. Bonjour, and H. Fleisch. 1976. Renal tubular adaptation to dietary phosphorus. *Nature (Lond.)* **261**: 145-146.
- Steele, T. H., and H. F. DeLuca. 1976. Influence of dietary phosphorus on renal phosphate, reabsorption in the parathyroidectomized rat. *J. Clin. Invest.* **57**: 867-874.
- Chen, T. C., L. Castillo, M. Korycka-Dahl, and H. F. DeLuca. 1974. Role of vitamin D metabolites in phosphate transport of rat intestine. *J. Nutr.* **104**: 1056-1060.
- Brickman, A. S., D. L. Hartenbower, A. W. Norman, and J. W. Coburn. 1977. Actions of 1 α -dihydroxy-vitamin D₃ and 1,25-dihydroxy-vitamin D₃ on mineral metabolism in man. *Am. J. Clin. Nutr.* **30**: 1064-1069.
- Steele, T. H., J. E. Engle, Y. Tanaka, R. S. Lorence, K. L. Dudgeon, and H. F. DeLuca. 1975. Phosphatemic action of 1,25-dihydroxy-vitamin D₃. *Am. J. Physiol.* **229**: 489-495.
- Castillo, L., Y. Tanaka, and H. F. DeLuca. 1975. The

³ Hartenbower, D. L., T. M. Stanley, J. W. Coburn, and A. W. Norman. Unpublished observations.

- mobilization of bone mineral by 1,25-dihydroxy-vitamin D₃ in hypophosphatemic rats. *Endocrinology*. **97**: 995–999.
14. Hughes, M. R., P. F. Brumbaugh, M. R. Haussler, J. E. Wergedal, and D. J. Baylink. 1975. Regulation of serum 1,25-dihydroxy-vitamin D₃ by calcium and phosphorus in the rat. *Science (Wash. D. C.)*. **190**: 578–584.
 15. Walling, M. W., D. L. Hartenbower, J. W. Coburn, and A. W. Norman. 1977. Effects of 1 α ,25-24R,25-, and 1 α ,24R,25-hydroxylated metabolites of vitamin D₃ on calcium and phosphate absorption by duodenum from intact and nephrectomized rats. *Arch. Biochem. Biophys.* **182**: 251–257.
 16. Steenbock, H., and C. Herting. 1955. Vitamin D and growth. *J. Nutr.* **57**: 449–468.
 17. Hohenwallner, W., and E. Wimmer. 1973. The malachite green micromethod for the determination of inorganic phosphate. *Clin. Chim. Acta.* **45**: 169–175.
 18. Siest, G., C. Vigneron, D. Palaszewski, and D. Marchal. 1968. Etude du dosage de l'uree et de la citrulline en milieu proteique. Methods manuelle et automatique a la diacetylmonoxime. *Clin. Chim. Acta.* **20**: 373.
 19. Dunnett, C. W. 1955. A multiple comparison procedure for comparing several treatments with a control. *Am. Statist.* **50**: 1096–1121.
 20. Walling, M. W., and S. S. Rothman. 1969. Phosphate independent, carrier mediated active transport of calcium by rat intestine. *Am. J. Physiol.* **217**: 1144–1148.
 21. Tröhler, U., J. P. Bonjour, and H. Fleisch. 1976. Inorganic phosphate homeostasis. Renal adaptation to the dietary intake in intact and thyroparathyroidectomized rats. *J. Clin. Invest.* **57**: 264–273.
 22. Morgan, B. 1973. The Osteomalacia Syndrome Due to Vitamin D Deficiency in Osteomalacia, Renal Osteodystrophy and Osteoporosis. N. Kugelmann, ed. Charles C Thomas publisher. Springfield, Ill. 73–144.
 23. Lee, W. S., J. Russell, and L. V. Avioli. 1977. 25-Hydroxycholecalciferol to 1,25-dihydroxycholecalciferol: Conversion impaired by systemic metabolic acidosis. *Science (Wash. D. C.)*. **195**: 994–996.
 24. Ribovich, M. L., and H. F. DeLuca. 1975. The influence of dietary calcium and phosphorus on intestinal calcium transport in rats given vitamin D metabolites. *Arch. Biochem. Biophys.* **170**: 529–535.
 25. Raisz, L. G., and I. Neiman. 1969. Effect on phosphate, calcium and magnesium on bone resorption and hormonal responses in tissue culture. *Endocrinology*. **85**: 446–452.
 26. Cuisnier-Gleizes, P., M. Thomasset, F. Saiteny-DeBove, and H. Mathieu. 1976. Phosphorus deficiency, parathyroid hormone and bone resorption in growing rats. *Calcif. Tissue Res.* **20**: 235–249.
 27. Costanzo, L. S., P. R. Sheehe, and L. M. Weiner. 1974. Renal action of vitamin D in D-deficient rats. *Am. J. Physiol.* **226**: 1490–1495.
 28. Rizzoli, R., H. Fleisch, and J. P. Bonjour. 1977. Effect of thyroparathyroidectomy on calcium metabolism in the rats: role of 1,25-dihydroxy-vitamin D₃. *Am. J. Physiol.* **233E**: 160–164.
 29. Brickman, A. S., J. W. Coburn, and S. G. Massry. 1974. 1,25-Dihydroxy-vitamin D₃ in normal man and patients with renal failure. *Ann. Intern. Med.* **80**: 161–168.
 30. Llach, F., J. W. Coburn, A. S. Brickman, K. Kurokawa, A. W. Norman, and E. Reiss. 1977. Acute actions of 1,25-dihydroxy-vitamin D₃ in normal man: effect on calcium and parathyroid status. *J. Clin. Endocrinol. Metab.* **44**: 1054–1060.
 31. Popovtzer, M. M., J. B. Robinette, H. F. DeLuca, and M. F. Holick. 1974. The acute effect of 25-hydroxycholecalciferol on renal handling of phosphorus. Evidence for a parathyroid-dependent mechanism. *J. Clin. Invest.* **53**: 913–921.
 32. Puschett, J. B., W. S. Beck, and A. Jelonek. 1975. Parathyroid hormone and 25-hydroxy-vitamin D₃: Synergistic and antagonistic effects on renal phosphate transport. *Science (Wash. D. C.)*. **190**: 473–475.
 33. Bonjour, J. P., C. Preston, and H. Fleisch. 1977. Effects of 1,25-dihydroxy-vitamin D₃ on the renal handling of Pi in thyroparathyroidectomized rats. *J. Clin. Invest.* **60**: 1419–1428.
 34. Garabedian, M., E. Pezant, L. Miravet, C. Fellot, and S. Balsan. 1976. 1,25-Dihydroxycholecalciferol effect on serum phosphorus homeostasis in rats. *Endocrinology*. **98**: 794–799.
 35. Hartenbower, D. L., T. M. Stanley, J. W. Coburn, and A. W. Norman. 1977. Serum and renal histologic changes in the rat following administration of toxic amounts of 1,25-dihydroxy-vitamin D₃. In *Vitamin D: Biochemical, Chemical and Clinical Aspects Related to Calcium Metabolism*. A. W. Norman, K. Schafer, J. W. Coburn, H. F. DeLuca, D. Fraser, H. G. Grigoleit, and D. V. Herrath, editors. Walter de Gruyter & Co., Berlin, 587–589.
 36. Boyle, I. T., R. W. Gray, and H. F. DeLuca. 1971. Regulation by calcium of in vivo synthesis in 1,25-dihydroxycholecalciferol and 21,25-dihydroxycholecalciferol. *Proc. Natl. Acad. Sci. U.S.A.* **68**: 2131–2134.
 37. Norman, A. W., and R. G. Wong. 1972. Biological activity of the vitamin D metabolite 1,25-dihydroxycholecalciferol in chickens and rats. *J. Nutr.* **102**: 1703–1718.
 38. Bordier, P., A. Ryckwaert, P. Marie, L. Miravet, A. Norman, and H. Rasmussen. 1977. Vitamin D metabolites and bone mineralization in man. In *Vitamin D: Biochemical, Chemical and Clinical Aspects Related to Calcium Metabolism*. A. W. Norman, K. Schafer, J. W. Coburn, H. F. DeLuca, D. Fraser, H. G. Grigoleit, and D. V. Herrath, editors. Walter de Gruyter & Co., Berlin. 897–909.
 39. Fournier, A., P. Bordier, J. Gueris, C. Ferriere, J. Chanard, P. Marie, J. L. Sebert, J. Bedrossian, H. F. DeLuca, and C. H. U. Amiens. 1977. Comparison of 1 α hydroxy-vitamin D₃ and 25 hydroxyvitamin D₃ in the treatment of renal osteodystrophy. Greater effect of 25 hydroxy-vitamin D₃ in mineralizing bone. In *Vitamin D: Biochemical, Chemical and Clinical Aspects Related to Calcium Metabolism*. A. W. Norman, K. Schafer, J. W. Coburn, H. F. DeLuca, D. Fraser, H. G. Grigoleit, and D. V. Herrath, editors. Walter de Gruyter & Co., Berlin. 667–669.
 40. Henry, H. L., A. N. Taylor, and A. W. Norman. 1977. Response of chick parathyroid glands to the vitamin D metabolites, 1,25-dihydroxycholecalciferol and 24,25-dihydroxycholecalciferol. *J. Nutr.* **107**: 1918–1926.