

Myeloperoxidase-Mediated Platelet Release Reaction

Robert A. Clark

J Clin Invest. 1979;63(2):177-183. <https://doi.org/10.1172/JCI109287>.

Research Article

The ability of the neutrophil myeloperoxidase-hydrogen peroxide-halide system to induce the release of human platelet constituents was examined. Both lytic and nonlytic effects on platelets were assessed by comparison of the simultaneously measured release of a dense-granule marker, [³H]serotonin, and a cytoplasmic marker, [¹⁴C]adenine. Incubation of platelets with H₂O₂ alone (20 μM H₂O₂ for 10 min) resulted in a small, although significant, release of both serotonin and adenine, suggesting some platelet lysis. Substantial release of these markers was observed only with increased H₂O₂ concentrations (>0.1 mM) or prolonged incubation (1-2 h).

Serotonin release by H₂O₂ was markedly enhanced by the addition of myeloperoxidase and a halide. Under these conditions, there was a predominance of release of serotonin (50%) vs. adenine (13%), suggesting, in part, a nonlytic mechanism. Serotonin release by the complete peroxidase system was rapid, reaching maximal levels in 2-5 min, and was active at H₂O₂ concentrations as low as 10 μM. It was blocked by agents which inhibit peroxidase (azide, cyanide), degrade H₂O₂ (catalase), chelate Mg²⁺ (EDTA, but not EGTA), or inhibit platelet metabolic activity (dinitrophenol, deoxyglucose).

These results suggest that the myeloperoxidase system initiates the release of platelet constituents primarily by a nonlytic process analogous to the platelet release reaction. Because components of the peroxidase system (myeloperoxidase, H₂O₂) are secreted by activated neutrophils, the reactions described here [...]

Find the latest version:

<https://jci.me/109287/pdf>



Myeloperoxidase-Mediated Platelet Release Reaction

ROBERT A. CLARK, *Department of Medicine, Boston University School of Medicine, and University Hospital, Boston, Massachusetts 02118*

SEYMOUR J. KLEBANOFF, *Department of Medicine, University of Washington School of Medicine, Seattle, Washington 98195*

ABSTRACT The ability of the neutrophil myeloperoxidase-hydrogen peroxide-halide system to induce the release of human platelet constituents was examined. Both lytic and nonlytic effects on platelets were assessed by comparison of the simultaneously measured release of a dense-granule marker, [^3H]serotonin, and a cytoplasmic marker, [^{14}C]adenine. Incubation of platelets with H_2O_2 alone (20 μM H_2O_2 for 10 min) resulted in a small, although significant, release of both serotonin and adenine, suggesting some platelet lysis. Substantial release of these markers was observed only with increased H_2O_2 concentrations (>0.1 mM) or prolonged incubation (1–2 h).

Serotonin release by H_2O_2 was markedly enhanced by the addition of myeloperoxidase and a halide. Under these conditions, there was a predominance of release of serotonin (50%) vs. adenine (13%), suggesting, in part, a nonlytic mechanism. Serotonin release by the complete peroxidase system was rapid, reaching maximal levels in 2–5 min, and was active at H_2O_2 concentrations as low as 10 μM . It was blocked by agents which inhibit peroxidase (azide, cyanide), degrade H_2O_2 (catalase), chelate Mg^{2+} (EDTA, but not EGTA), or inhibit platelet metabolic activity (dinitrophenol, deoxyglucose).

These results suggest that the myeloperoxidase system initiates the release of platelet constituents primarily by a nonlytic process analogous to the platelet release reaction. Because components of the peroxidase system (myeloperoxidase, H_2O_2) are secreted by activated neutrophils, the reactions described here may have implications for neutrophil-platelet interaction in sites of thrombus formation.

Portions of this work have appeared in abstract form: *Clin. Res.* 1976. 24: 305A; and *Clin. Res.* 1977. 25: 474A.

Dr. Clark is the recipient of Research Career Development Award CA00441 from the National Institutes of Health.

Received for publication 3 July 1978 and in revised form 25 September 1978.

INTRODUCTION

When the polymorphonuclear leukocyte (PMN)¹ is activated by phagocytosis, contact with nonphagocytosable immune reactant-coated surfaces, or exposure to certain soluble stimuli, it undergoes both metabolic activation and degranulation (1–3). Although the primary role of these processes is considered to be the killing of ingested microorganisms within the phagocytic vacuole, leakage or secretion of toxic agents into the extracellular fluid can occur. Thus, the metabolic burst results in the formation of several reactive species of partially reduced oxygen, including superoxide anion, hydrogen peroxide (H_2O_2), and hydroxyl radical (2, 3), and some of these agents have been detected in the extracellular fluid adjacent to the activated PMN (4–8). Degranulation can result in the emptying of granule enzymes, such as acid hydrolases and myeloperoxidase, into the extracellular fluid (9–13). The potential biological effects of these PMN secretory products have attracted considerable interest.

Myeloperoxidase (MPO) plus H_2O_2 and a halide constitute a potent microbicidal system which is operative in the PMN (2, 3, 14). This system is also toxic to several types of mammalian cells, including spermatozoa (15), erythrocytes (16), leukocytes (17), and tumor cells (18), raising the possibility that MPO and H_2O_2 released from activated PMN may affect nearby host cells. Phagocytosing PMN exert a cytotoxic effect on adjacent mammalian tumor cells which is mediated by the release of MPO and H_2O_2 (5). Extracellular activity of MPO and H_2O_2 is further indicated by the fact that protein iodination by phagocytosing PMN, a phenomenon mediated by the MPO system, occurs in both extracellular and intracellular sites (19, 20). Secreted H_2O_2

¹Abbreviations used in this paper: DNP, dinitrophenol; DOG, 2-deoxy-D-glucose; MPO, myeloperoxidase; PBA, sodium phosphate buffer, 0.1 M, pH 6.5, with human albumin 1 mg/ml; PMN, polymorphonuclear leukocyte(s).

alone may exert nonenzymatic effects on certain types of mammalian cells (4, 8).

Previous investigators have suggested the possibility of in situ interaction between PMN and platelets (8, 21–23). The studies of Levine et al. (8) demonstrated an in vitro effect of activated PMN on platelets mediated by secreted H_2O_2 . Platelets which were preincubated with phagocytosing PMN or with an H_2O_2 -generating enzyme system had impaired aggregation responses, and this effect was inhibited by catalase. Other studies have reported effects of reagent H_2O_2 on various aspects of platelet physiology (24–28). In this paper, we describe the initiation of the release reaction in human platelets by the MPO- H_2O_2 -halide system.

METHODS

Platelet preparation. All processing was performed at room temperature with plastic tubes, syringes, and pipettes. Venous blood was drawn from normal adult volunteers, with potassium EDTA (K-EDTA) as the anticoagulant (10 ml of blood plus 0.2 ml of a 10% solution of K-EDTA). After centrifugation at 200 g for 10 min, the upper one-half of the platelet-rich plasma layer (≈ 2 ml) was carefully aspirated without disturbing the lower erythrocyte layer. [3H]serotonin (hydroxytryptamine binoxalate, 5-[1,2- $^3H(N)$] in 2% ethanol, ≈ 25 Ci/mmol, New England Nuclear, Boston, Mass.) and [^{14}C]adenine (adenine [8- ^{14}C] in 0.01 N HCl, ≈ 50 mCi/mmol, New England Nuclear) were added to the platelet-rich plasma to final concentrations of $0.2 \mu M$ serotonin and $20 \mu M$ adenine. In some experiments, [^{14}C]serotonin (5-hydroxytryptamine binoxalate [2- ^{14}C] in 2% ethanol, ≈ 50 mCi/mmol, New England Nuclear) and [3H]adenine (adenine- $^3H[G]$, ≈ 20 Ci/mmol, New England Nuclear) were used in similar concentrations. The platelet-rich plasma-serotonin-adenine mixture was incubated at room temperature for 40 min with occasional gentle mixing. It was then centrifuged at 500 g for 10 min and the platelet pellet was gently washed with 5 ml of 0.1 M sodium phosphate buffer, pH 6.5, containing human albumin (Sigma Chemical Co., St. Louis, Mo.), 1 mg/ml (PBA). After centrifugation at 600 g for 10 min, the platelet pellet was again suspended in 5 ml of PBA. An aliquot of 0.01 ml was diluted in 0.99 ml of PBA, and the platelets in this suspension were counted in duplicate in a hemocytometer. The stock platelet suspension was then adjusted to contain 10^8 platelets/ml of PBA.

Release assay. Radiolabeled platelets (0.05-ml sample of stock suspension containing 5×10^6 platelets) were incubated with the system components as detailed in the legends to the tables and figures. Incubations were performed in 12×75 -mm polystyrene tubes with a total volume of 0.5 ml, in a 37°C waterbath shaker oscillating 60 times/min. Immediately after incubation, the samples were centrifuged at 550 g for 10 min at 4°C. A portion of 0.2 ml of each sample supernate was placed in a liquid scintillation counter vial, 15 ml of Aquasol (New England Nuclear) was added, and counts of 3H and ^{14}C determined in a Beckman LS-100 counter (Beckman Instruments, Inc., Spinco Div., Palo Alto, Calif.). In each experiment, all samples were run in duplicate, and the values averaged. Standards included blank tubes (0.2 ml H_2O), platelet standards (0.05-ml platelets), and maximum release standards (0.05-ml platelets plus 0.45 ml 1% Triton X-100 [Sigma Chemical Co.] which was centrifuged as above and a

0.2-ml portion counted). The blank was subtracted from all samples and the release of each isotope was calculated as a percent of the maximum release standard. The release of both serotonin and adenine by Triton X-100 (maximum release standard) was essentially equal to the total contained in the platelet standard.

Special reagents. MPO was prepared from canine PMN by the method of Agner (29) through the end of step 6. Peroxidase activity was determined by the o-dianisidine method (30); 1 U of activity is that causing the usage of $1 \mu mol$ of substrate/min at 25°C (16). MPO prepared from human PMN by the method of Olsson et al. (31) was kindly supplied by Dr. Inge Olsson. H_2O_2 (30% solution) was obtained from Fisher Scientific Co., Fair Lawn, N. J. Dinitrophenol, 2-deoxy-D-glucose, EDTA, K-EDTA, and EGTA were obtained from Sigma Chemical Co. Stock solutions of EDTA and EGTA were made at 0.1 M with the pH adjusted to 7.0; 0.05 M $MgCl_2$ was added to the stock EGTA to assure saturation of Mg^{2+} -binding capacity (Mg-EGTA). Catalase (beef liver, 6.7 mg/ml, 62,900 U/mg) was from Worthington Biochemical Corp., Freehold, N. J. 1 U of catalase is that amount which catalyzes the oxidation of $1 \mu mol$ H_2O_2 /min at 25°C, pH 7.0.

Statistics. Standard error was used throughout as an estimate of variance, and means were compared with Student's *t* test.

RESULTS

Incubation of [3H]serotonin-labeled human platelets with MPO, H_2O_2 ($20 \mu M$), and chloride resulted in rapid and substantial release of the labeled amine which reached about two-thirds of maximum levels within 2 min (Fig. 1). Very little additional release was observed

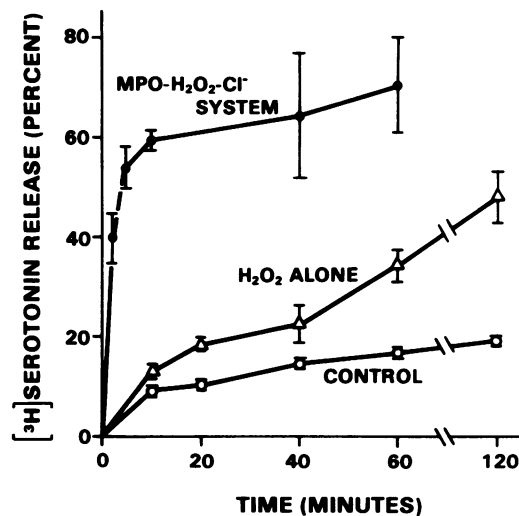


FIGURE 1 Kinetics of the release of serotonin from platelets by the MPO- H_2O_2 -chloride system and by H_2O_2 alone. Control samples (O) contained 0.03 M sodium phosphate buffer pH 6.5, $50 \mu g$ human albumin, 1.5 mM KH_2PO_4 , 1.5 mM $MgSO_4$, 0.067 M Na_2SO_4 , and 5×10^6 platelets labeled with [3H]serotonin. In the experimental samples, Na_2SO_4 was replaced by 0.1 M NaCl; either 4 mU of MPO (canine) and $20 \mu M$ H_2O_2 (●), or $20 \mu M$ H_2O_2 alone (Δ) was added. Total volume was 0.5 ml, and incubation time was varied as indicated. Data represent mean \pm SE of six experiments.

after 5–10 min. The 10-min period was used routinely as maximal levels were achieved and because sample processing was difficult when periods shorter than this were employed. There was slow leakage of serotonin throughout the incubation period from control platelets suspended in the buffer alone. In the absence of MPO, H_2O_2 (20 μM) caused minimal release above control levels at 10 min; but, with prolonged incubation, substantial release was detected. However, this always remained well below the level observed with the complete MPO system. The effect of H_2O_2 in the absence of MPO did not require halides because the same results were observed when chloride was omitted. In subsequent figures and tables, the release in control samples has been subtracted from that in experimental samples, with the mean control release indicated in the footnotes.

In the presence of MPO and chloride, H_2O_2 was effective in initiating serotonin release at concentrations of 10 μM or higher (Fig. 2). In the absence of MPO, release of serotonin by H_2O_2 during a 10-min incubation was substantial only at concentrations well above 0.1 mM. Examination of the H_2O_2 dose-response curves (Fig. 2) indicates nearly a 2-log greater potency in the presence of MPO than in its absence.

The requirements for MPO-mediated serotonin release from platelets are explored further in Table I. Several halides (chloride, iodide, bromide) and the

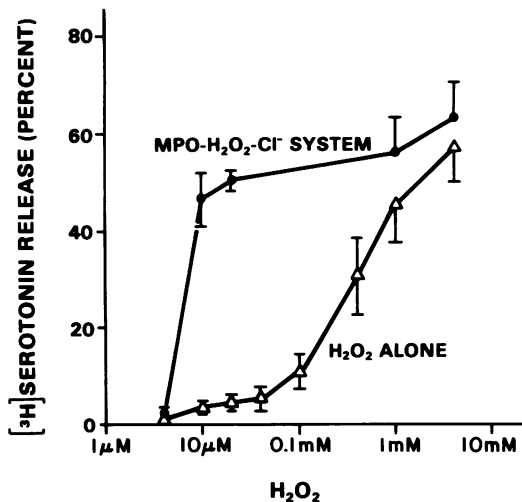


FIGURE 2 Effect of varying H_2O_2 concentrations on the release of serotonin from platelets by the MPO- H_2O_2 -chloride system. Conditions were as described in Fig. 1 except that the H_2O_2 concentration was varied as indicated. The incubation time was 10 min. Serotonin release in samples containing the complete peroxidase system (○) and in samples containing H_2O_2 without MPO (Δ) are illustrated. Data represent mean \pm SE of six experiments. Serotonin release in control samples (mean $9.2 \pm 0.6\%$) was subtracted from that in experimental samples.

pseudohalide, thiocyanate, were effective as cofactors. When each component of the system was omitted or when MPO was heated, there was a major loss of serotonin-releasing activity, indicating requirements for the active enzyme, H_2O_2 , and a cofactor. However, a small but significant ($P < 0.05$) amount of serotonin release was detected in the absence of MPO, suggesting a minor effect of H_2O_2 alone (Fig. 1). MPO isolated from both dog and human PMN was effective.

The MPO system also induced the release of [^{14}C]-adenine from labeled human platelets (Table I), although the percent release was much lower than that observed with serotonin. Results with adenine were parallel to those with serotonin, as dog and human MPO were effective; each of the cofactors could be employed (data for iodide, bromide, and thiocyanate not shown), and release was decreased when each component of the system was omitted or when the MPO was heated ($P < 0.01$ for each comparison). However, the release of adenine observed on deletion of MPO or the halide or on heating the MPO was approximately one-half of that found with the complete MPO system. Thus, it appeared that under the conditions employed, H_2O_2 alone was responsible for about one-half of the adenine release, and that the addition of MPO resulted in slight enhancement of adenine release, but a striking stimulation of serotonin release. Results in both the serotonin and adenine release systems were confirmed in parallel experiments employing different radioisotopes, i.e., [^{14}C]serotonin and [3H]adenine (data not shown).

The effect of a number of inhibitors on MPO-mediated serotonin release is shown in Table II. The peroxidase inhibitors, azide and cyanide, blocked release essentially completely. Release was also inhibited by the addition of catalase, but not heated catalase. Chelation of Ca^{2+} and Mg^{2+} by 10 mM EDTA inhibited serotonin release, but 1 mM EDTA was ineffective. Selective chelation of Ca^{2+} by 10 mM Mg-EGTA had no effect. These data suggest a requirement for Mg^{2+} , but not Ca^{2+} , for maximal MPO-induced release of serotonin from platelets. The standard reaction mixture contained 1.5 mM Mg^{2+} and no added Ca^{2+} . Dinitrophenol (DNP), an inhibitor of oxidative phosphorylation, caused a partial, but significant, decrease in serotonin release, as did 2-deoxy-D-glucose (DOG), a competitive inhibitor of glucose usage. The most marked inhibition was observed when both DNP and DOG were present ($P < 0.001$ vs. complete MPO system; $P < 0.005$ vs. DOG or DNP alone). The low level of adenine release induced by the MPO system was partially inhibited by azide, cyanide, catalase, EDTA, and DNP, but was unaffected by heated catalase, EGTA, and DOG. None of the inhibitors in the concentrations employed caused any release of

TABLE I
Release of [³H]Serotonin and [¹⁴C]Adenine from Platelets by the MPO System*

Supplements	[³ H]Serotonin release				[¹⁴ C]Adenine release
	Chloride	Iodide	Bromide	Thiocyanate	Chloride
	%				%
MPO (canine) + H ₂ O ₂					
+ cofactor	50.0†§ (24)	19.4§ (11)	25.8§ (6)	24.4§ (6)	13.4§ (22)
MPO omitted	4.0 (12)	3.5 (7)	4.2 (3)	10.4 (3)	8.3§ (12)
H ₂ O ₂ omitted	0.3 (7)	1.3 (7)	1.9 (3)	1.9 (3)	1.0 (7)
Cofactor omitted	0 (8)	0 (8)	0 (8)	0 (8)	5.9§ (8)
MPO heated	2.2 (8)	3.1 (6)	—	—	6.4§ (8)
MPO (human) + H ₂ O ₂					
+ cofactor	47.2§ (4)	21.2§ (3)	—	—	12.7§ (5)
H ₂ O ₂ omitted	3.8 (4)	3.2 (4)	—	—	1.9 (4)

* The complete reaction mixture contained 0.03 M sodium phosphate buffer pH 6.5, 50 μg human albumin, 1.5 mM KH₂PO₄, 1.5 mM MgSO₄, 5 × 10⁶ platelets labeled with [³H]serotonin and [¹⁴C]adenine, and the supplements listed below as follows: MPO (canine or human) 4 mU; H₂O₂ 20 μM; NaCl 0.1 M; NaI 0.1 mM; NaBr 0.1 mM; NaCNS 0.1 mM. Na₂SO₄ 0.067 M was added to all reaction mixtures which did not contain NaCl to maintain isomolarity. MPO was heated at 100°C for 15 min where indicated. Total volume was 0.5 ml. Incubation time was 10 min.

† Mean of experiments. No. of experiments in parentheses. Release in control samples (mean 9.2±0.6%) was subtracted from that in experimental samples.

§ P < 0.001 vs. control.

^{||} P < 0.05 vs. control.

serotonin or adenine above control levels in the absence of the peroxidase system components. EDTA, DNP, and DOG did not inhibit protein iodination by the MPO-H₂O₂-halide system (32), which suggests that

their inhibitory activity in these experiments was not a consequence of blocking the peroxidase system, but rather was mediated by an effect on the platelets.

DISCUSSION

A number of investigators have demonstrated effects of reagent H₂O₂ on certain aspects of platelet function, although these studies have employed rather high concentrations of H₂O₂, generally well in excess of 0.1 mM. Although platelet aggregation by H₂O₂ has been reported (24), Canoso et al. (25) demonstrated no aggregation by this agent alone, but enhancement of the response to other agents (e.g., ADP) added simultaneously with the H₂O₂. Under some conditions, preincubation of platelets with H₂O₂ may impair subsequent aggregation responses to ADP or other agents (25, 26). Ultrastructural studies indicated aggregation of platelets exposed to H₂O₂ in concentrations >1 mM (26). H₂O₂ has not been reported to cause serotonin release from platelets (25) except in concentrations exceeding 1 mM (26). Holmsen and colleagues (27, 28) noted that incubation of platelets in H₂O₂ (≥0.1 mM) resulted in decreases in platelet ATP, but the adenylate energy charge decreased only if ATP levels were more than two-thirds depleted. Under these conditions, H₂O₂ caused inhibition of shape change, primary aggregation, and secretion of constituents of dense granules and α-granules only if the decrease in adenylate energy charge exceeded 5%. Handin et al. (33)

TABLE II
Effect of Inhibitors on Release of [³H]Serotonin from Platelets by the MPO-H₂O₂-Chloride System*

Inhibitors	[³ H]Serotonin release
	%
None	50.0±2.5† (24)
Azide (0.1 mM)	2.5±1.0§ (8)
Cyanide (0.1 mM)	3.8±1.4§ (8)
Catalase (210 U)	1.5±0.5§ (6)
Heated catalase (210 U)	55.1±8.8 (6)
EDTA (10 mM)	9.4±0.9§ (5)
EDTA (1 mM)	44.6±3.2 (4)
Mg-EGTA (10 mM)	60.2±5.9 (5)
DNP (10 μM)	33.5±3.1§ (11)
DOG (1 mM)	29.2±3.6§ (11)
DNP (10 μM) + DOG (1 mM)	13.4±2.4§ (10)

* Conditions were the same as described for the MPO-H₂O₂-chloride system in Table I except for the addition of inhibitors as noted. Azide and cyanide were added as the sodium salts. Canine MPO was used.

† Mean±SE. No. of experiments is in parentheses. Release in control samples (mean 9.2±0.6%) was subtracted from that in experimental samples.

§ P < 0.001 vs. complete MPO system.

described the initiation of platelet aggregation and serotonin release by xanthine oxidase, a superoxide-generating enzyme. Inhibition of these effects by superoxide dismutase, but not catalase or mannitol, suggested mediation by superoxide rather than H_2O_2 or hydroxyl radical. Longer incubation of platelets with xanthine oxidase caused release of radioactive sodium chromate, a cytoplasmic label, and this was blocked by catalase, but not superoxide dismutase. It was thus suggested that superoxide caused specific release of granule components, whereas H_2O_2 (formed by superoxide dismutation) caused lytic damage to the platelet.

Our data document the release of platelet constituents by the MPO- H_2O_2 -halide system. The release of serotonin, a dense-granule constituent, was employed as an indicator of the platelet release reaction, whereas the release of adenine was considered to be a lytic or cytotoxic event (34, 35). If both serotonin and adenine are solubilized, but the former predominates, it may be inferred that both the release reaction and lysis are occurring.

Under our experimental conditions, brief exposure of platelets to H_2O_2 alone (20 μM) resulted in a small, although significant, release of adenine as well as serotonin, suggesting some platelet lysis. H_2O_2 alone initiated a large amount of release of these markers only at higher concentrations (>0.1 mM), or on prolonged exposure (1–2 h) to low concentrations. Serotonin release by H_2O_2 was markedly enhanced by the addition of MPO and a halide. The amount of adenine released by the MPO- H_2O_2 -chloride system ($\cong 13\%$) was considerably less than the serotonin release (50%), which suggests that the peroxidase system triggers the nonlytic secretion of granule constituents independent of a lytic effect. Each component of the system (MPO, H_2O_2 , halide) was required, and either chloride, bromide, iodide, or the pseudohalide, thiocyanate, could be employed. Chloride was effective in physiologic concentrations (0.1 M). Relatively low concentrations of H_2O_2 (10 μM) could be employed, and serotonin release reached maximum levels in 2–5 min. Serotonin release by the MPO system was inhibited by the heme enzyme inhibitors, azide and cyanide, as well as by catalase. Although the inhibition by azide and cyanide appears to be a result of an effect on the peroxidase, it should be noted that azide, but not cyanide, inhibits the platelet aggregation and release responses to collagen, ADP, and epinephrine by a direct effect on the platelet, the mechanism of which remains to be determined (27, 36).

Chelation of Ca^{2+} by Mg-EGTA did not inhibit MPO-induced serotonin release, whereas chelation of both Ca^{2+} and Mg^{2+} by EDTA did, suggesting a requirement for Mg^{2+} . DNP, an inhibitor of oxidative phosphorylation, partially blocked serotonin release, as did DOG, a glycolytic inhibitor, and maximal inhibition was

achieved by a combination of DNP and DOG. Other aspects of platelet function are also maximally inhibited by simultaneously blocking oxidative and glycolytic pathways (37–40). This requirement for intact platelet metabolism to observe maximal release responses to the MPO system suggests an active process with respect to the platelets, as opposed to a passive lytic or cytotoxic process.

Thus, several lines of evidence indicate that the release of serotonin from platelets by the MPO system is primarily a nonlytic event analogous to the release reaction (34). This hypothesis is supported by the rapidity of release, the marked predominance of serotonin release over adenine release, the divalent cation requirement, and the necessity for intact platelet metabolic activity. A lesser component of lytic release appears to be associated with a nonenzymatic effect of H_2O_2 alone.

In view of the ability of the MPO system to release platelet granule constituents and the known secretion of MPO and H_2O_2 by activated PMN (1, 2, 4–13), it is reasonable to suggest an interaction between PMN and platelets mediated by this system. Levine et al. (8) have demonstrated that H_2O_2 formed by phagocytosing PMN can inhibit platelet aggregation responses to ADP. Their protocol, in which the PMN and platelets were separated by a dialysis membrane, might preclude the detection of an MPO-mediated effect as the large molecular weight of this enzyme (41, 42) would not allow passage through the membrane. It would be expected that an H_2O_2 -mediated effect would be considerably enhanced by MPO in the presence of a suitable halide cofactor. Platelets contain H_2O_2 -degrading enzyme systems (e.g., catalase and glutathione peroxidase [8, 27, 43]) which may limit perturbation by low levels of H_2O_2 . The protective effect of these enzymes might be overcome by increased H_2O_2 levels, by persisting low steady-state levels, or by successful competition for H_2O_2 by MPO.

Our studies, together with those of others (8, 22, 33), suggest that activated PMN might influence platelets by several mechanisms, including superoxide anion, H_2O_2 , or the MPO- H_2O_2 -halide system. Further, the effect on platelets may be primary, i.e., initiation of release reactions, or secondary, i.e., impairment of the response to other stimuli. These observations may have important implications for hemostatic processes in vivo. The simultaneous presence of PMN and platelets in sites of thrombus formation (44, 45) raises the distinct possibility that PMN products, such as MPO and H_2O_2 , could promote the release of platelet constituents within the developing thrombus.

ACKNOWLEDGMENTS

The authors gratefully acknowledge the expert technical assistance of Coralie Baker. This work was supported by grants

REFERENCES

1. Goldstein, I. M. 1976. Polymorphonuclear leukocyte lysosomes and immune tissue injury. *Prog. Allergy*. **20**: 301-340.
2. Klebanoff, S. J., and R. A. Clark. 1978. *The Neutrophil: Function and Clinical Disorders*. Elsevier North-Holland, Inc., New York.
3. Babior, B. M. 1978. Oxygen-dependent microbial killing by phagocytes. *N. Engl. J. Med.* **298**: 659-668, 721-725.
4. Baehner, R. L., D. G. Nathan, and W. B. Castle. 1971. Oxidant injury of caucasian glucose-6-phosphate dehydrogenase-deficient red blood cells by phagocytosing leukocytes during infection. *J. Clin. Invest.* **50**: 2466-2473.
5. Clark, R. A., and S. J. Klebanoff. 1975. Neutrophil-mediated tumor cell cytotoxicity. Role of the peroxidase system. *J. Exp. Med.* **141**: 1442-1447.
6. Root, R. K., J. Metcalf, N. Oshino, and B. Chance. 1975. H₂O₂ release from human granulocytes during phagocytosis. I. Documentation, quantitation, and some regulating factors. *J. Clin. Invest.* **55**: 945-955.
7. Salin, M. L., and J. M. McCord. 1975. Free radicals and inflammation. Protection of phagocytosing leukocytes by superoxide dismutase. *J. Clin. Invest.* **56**: 1319-1323.
8. Levine, P. H., R. S. Weinger, J. Simon, K. L. Scoon, and N. I. Krinsky. 1976. Leukocyte-platelet interaction. Release of hydrogen peroxide by granulocytes as a modulator of platelet reactions. *J. Clin. Invest.* **57**: 955-963.
9. Baehner, R. L., M. J. Karnovsky, and M. L. Karnovsky. 1969. Degranulation of leukocytes in chronic granulomatous disease. *J. Clin. Invest.* **48**: 187-192.
10. Weissmann, G., R. B. Zurier, P. J. Spieler, and I. M. Goldstein. 1971. Mechanisms of lysosomal enzyme release from leukocytes exposed to immune complexes and other particles. *J. Exp. Med.* **134**: 149s-165s.
11. Henson, P. M. 1971. The immunologic release of constituents from neutrophil leukocytes. I. The role of antibody and complement on nonphagocytosable surfaces or phagocytosable particles. *J. Immunol.* **107**: 1535-1546.
12. Henson, P. M., H. B. Johnson, and H. L. Spiegelberg. 1972. The release of granule enzymes from human neutrophils stimulated by aggregated immunoglobulins of different classes and subclasses. *J. Immunol.* **109**: 1182-1192.
13. Bainton, D. F. 1973. Sequential degranulation of the two types of polymorphonuclear leukocyte granules during phagocytosis of microorganisms. *J. Cell Biol.* **58**: 249-264.
14. Klebanoff, S. J. 1975. Antimicrobial mechanisms in neutrophilic polymorphonuclear leukocytes. *Semin. Hematol.* **12**: 117-142.
15. Smith, D. C., and S. J. Klebanoff. 1970. A uterine fluid-mediated sperm-inhibitory system. *Biol. Reprod.* **3**: 229-235.
16. Klebanoff, S. J., and R. A. Clark. 1975. Hemolysis and iodination of erythrocyte components by a myeloperoxidase-mediated system. *Blood*. **45**: 699-707.
17. Clark, R. A., and S. J. Klebanoff. 1977. Myeloperoxidase-H₂O₂-halide system: Cytotoxic effect on human blood leukocytes. *Blood*. **50**: 65-70.
18. Clark, R. A., S. J. Klebanoff, A. B. Einstein, and A. Fefer. 1975. Peroxidase-H₂O₂-halide system: Cytotoxic effect on mammalian tumor cells. *Blood*. **45**: 161-170.
19. Klebanoff, S. J., and C. B. Hamon. 1972. Role of myeloperoxidase-mediated antimicrobial systems in intact leukocytes. *J. Reticuloendothel. Soc.* **12**: 170-196.
20. Odeberg, H., T. Olofsson, and I. Olsson. 1974. Myeloperoxidase-mediated extracellular iodination during phagocytosis in granulocytes. *Scand. J. Haematol.* **12**: 155-160.
21. Harrison, M. J. G., P. R. Emmons, and J. R. A. Mitchell. 1966. The effect of white cells on platelet aggregation. *Thromb. Diath. Haemorrh.* **16**: 105-121.
22. Henson, P. M. 1970. Mechanisms of release of constituents from rabbit platelets by antigen-antibody complexes and complement. II. Interaction of platelets with neutrophils. *J. Immunol.* **105**: 490-501.
23. Handin, R. I., and T. P. Stossel. 1974. Phagocytosis of antibody-coated platelets by human granulocytes. *N. Engl. J. Med.* **290**: 989-993.
24. Higashi, O., Y. Kikuchi, and K. Konno. 1972. A case of thrombasthenia with a study of platelet aggregation by hydrogen peroxide (H₂O₂). *Tohoku J. Exp. Med.* **106**: 399-409.
25. Canoso, R. T., R. Rodvien, K. Scoon, and P. H. Levine. 1974. Hydrogen peroxide and platelet function. *Blood*. **43**: 645-656.
26. Rodvien, R., J. N. Lindon, and P. H. Levine. 1976. Physiology and ultrastructure of the blood platelet following exposure to hydrogen peroxide. *Br. J. Haematol.* **33**: 19-26.
27. Holmsen, H., and L. Robkin. 1977. Hydrogen peroxide lowers ATP levels in platelets without altering adenylate energy charge and platelet function. *J. Biol. Chem.* **252**: 1752-1757.
28. Stuart, M. J., and H. Holmsen. 1977. Hydrogen peroxide, an inhibitor of platelet function: Effect on adenine nucleotide metabolism, and the release reaction. *Am. J. Hematol.* **2**: 53-63.
29. Agner, K. 1958. Crystalline myeloperoxidase. *Acta Chem. Scand.* **12**: 89-94.
30. Worthington Enzyme Manual. 1972. Worthington Biochemical Corp., Freehold, N. J. 43.
31. Olsson, I., T. Olofsson, and H. Odeberg. 1972. Myeloperoxidase-mediated iodination in granulocytes. *Scand. J. Haematol.* **9**: 483-491.
32. Klebanoff, S. J. 1967. Iodination of bacteria. A bactericidal mechanism. *J. Exp. Med.* **126**: 1063-1078.
33. Handin, R. I., R. Karabin, and G. J. Boxer. 1977. Enhancement of platelet function by superoxide anion. *J. Clin. Invest.* **59**: 959-965.
34. Day, H. J., and H. Holmsen. 1971. Concepts of the blood platelet release reaction. *Ser. Haematol.* **4**: 3-27.
35. Reimers, H. J., J. F. Mustard, and M. A. Packham. 1975. Transfer of adenine nucleotides between the releasable and nonreleasable compartments of rabbit blood platelets. *J. Cell Biol.* **67**: 61-71.
36. Stibbe, J., and H. Holmsen. 1977. Effects of sodium azide on platelet function. *Thromb. Haemostasis.* **38**: 1042-1053.
37. Ball, G., M. Fulwood, D. M. Ireland, and P. Yates. 1969. Effect of some inhibitors of platelet aggregation on platelet nucleotides. *Biochem. J.* **114**: 669-671.
38. Murer, E. H. 1969. Clot retraction and energy metabolism of platelets. Effect and mechanism of inhibitors. *Biochim. Biophys. Acta.* **172**: 266-276.
39. Holmsen, H., C. A. Setkowsky, and H. J. Day. 1974. Effects of antimycin and 2-deoxyglucose on adenine nucleotides in human platelets. Role of metabolic adenosine triphosphate in primary aggregation, secondary aggrega-

- tion and shape change of platelets. *Biochem. J.* **144**: 385-396.
40. Kattlove, H. E. 1974. Platelet ATP in ADP-induced aggregation. *Am. J. Physiol.* **226**: 325-329.
41. Ehrenberg, A., and K. Agner. 1958. The molecular weight of myeloperoxidase. *Acta Chem. Scand.* **12**: 95-100.
42. Felberg, N. T., and J. Schultz. 1972. Evidence that myeloperoxidase is composed of isoenzymes. *Arch. Biochem.* **148**: 407-413.
43. Karpatkin, S., and H. J. Weiss. 1972. Deficiency of glutathione peroxidase associated with high levels of reduced glutathione in Glanzmann's thrombasthenia. *N. Engl. J. Med.* **287**: 1062-1066.
44. Henry, R. L. 1965. Leukocytes and thrombosis. *Thromb. Diath. Haemorrh.* **13**: 35-46.
45. Jørgensen, L., H. C. Rowsell, T. Hovig, and J. F. Mustard. 1967. Resolution and organization of platelet-rich mural thrombi in carotid arteries of swine. *Am. J. Pathol.* **51**: 681-719.