

Androgenic Hormones Modulate Autoantibody Responses and Improve Survival in Murine Lupus

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ABSTRACT Antibodies to native DNA and to polyadenylic acid (Poly A) occur spontaneously and undergo a regulated switch from IgM to IgG during the course of autoimmune disease in NZB/NZW F₁ (B/W) mice. B/W females have higher titers and earlier commitment to 7S antibodies to DNA and Poly A, whereas B/W males bind DNA and Poly A primarily by 19S antibodies. We have performed castration experiments to determine the effects of sex hormones on this switch from IgM to IgG.

NZB/NZW F₁ (B/W) mice were either castrated or subjected to sham surgery at 2 wk of age and studied for immunoglobulin class of antibodies to nucleic acids at 4, 6, and 7 mo post-surgery. Prepubertal castration of males caused premature death in 60% of mice. Castrated males had a significant decline in their serum testosterone concentration, an increase in DNA and Poly A binding, and an accelerated switch from 19S to 7S antibodies to nucleic acids. Castrated females had no change in mortality. However, castrated females given maintained androgen treatment had a decreased mortality compared to castrated females receiving estrogen (14 vs. 94%). The anticipated switch to 7S antibodies to Poly A was almost eliminated in castrated females. These results suggest that sex hormones modulate immunologic regulation and that androgenic hormones are protective in murine lupus.

INTRODUCTION

Autoimmune diseases in general, and systemic lupus erythematosus (SLE)¹ in particular, are much more

common in females than in males (1). An explanation for this clinical difference is not readily apparent. The availability of the NZB/NZW F₁ (B/W) mouse as a model for SLE has permitted investigation of this question.

B/W mice spontaneously produce LE cells and antibodies to nucleic acids. They develop an immune complex glomerulonephritis associated with the deposition of DNA, immunoglobulin (particularly IgG), and complement (2). Genetic, immunologic, and viral factors are all intimately involved in the pathogenesis of their disorder (3). As in human lupus, there is a striking sex difference in the mouse disease. Female mice have an earlier onset and greater severity of immune complex nephritis compared to males. These pathologic features are associated with an earlier appearance of IgG antibodies to DNA and RNA (4).

We have observed the ordered and sequential appearance of antibodies to DNA and polyadenylic acid (Poly A) in B/W mice (5). As early as 1 mo of age, females bind DNA predominantly by 7S IgG antibodies, whereas males bind DNA predominantly by 19S IgM antibodies. As the animals age, antibody titers rise progressively and there is an enhanced formation of 7S antibodies to DNA in both sexes. Young mice of both sexes bind Poly A exclusively by 19S antibodies. Females switch to 7S antibodies to Poly A at age 6 mo, whereas this switch in males is delayed until 9 mo of age. Thus, the switch to 7S antibodies for both DNA and RNA occurs in a regular manner and earlier in female than in male B/W mice.

Neonatal thymectomy causes an initial increase in antibody response to DNA and Poly A, consistent with the removal of thymic suppressor cells (6). It also eliminates the late switch to 7S antibodies to Poly A, showing that this response is thymic dependent (6). These results suggest that the ordered sequential appearance of antibodies to nucleic acids in B/W mice is, at least in part, related to thymic regulation.

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¹Abbreviations used in this paper: B/W, NZB × NZW F₁, hybrid mice; Poly A, polyadenylic acid; SLE, systemic lupus erythematosus.

To study the effect of sex hormones on the spontaneous formation and regulation of antinucleic acid antibodies, we performed prepubertal castration experiments in young B/W mice. Our results suggest that both male and female sex hormones influence the titer and immunoglobulin class of antibodies to DNA and RNA. These hormones appear to act on thymic regulatory cells, and the predominant effect is exerted by androgens.

METHODS

Mice

B/W mice used in these experiments were derived from our colony at the University of California Vivarium, San Francisco, and were maintained at the Fort Miley Veterans Administration Hospital. Blood was collected from mice at 4, 6, and 7 mo postoperatively by orbital sinus bleeding, allowed to clot at room temperature for 1 h, and left at 4°C overnight. Serum was separated by centrifugation at 1,200 g for 10 min.

Assessment of proteinuria

Tetrabromophenol paper, (Albustix, Ames Co., Div. of Miles Lab., Inc., Elkhart, Ind.) was used to measure the protein concentration of freshly expressed urine from individual mice.

Operative techniques

Littermates were randomized and subjected to the surgical procedures or sham operated at 2 wk of age. They were anesthetized by intraperitoneal injection of nembutal.

Oophorectomy. A midline incision was made over the abdomen. Ovaries were identified and removed with electrocautery. The incision was closed with 6-0 silk suture material.

Orchiectomy. Testes were delivered through a scrotal incision. The vas deferens and spermatic vessels were transected with electrocautery, and testes and epididymis were removed. Skin was sutured with 6-0 silk suture material.

Sham operation. In age-matched sham-operated controls, ovaries or testes were identified but left intact.

Experimental animals were distinguished from sham-operated mice by tail clipping. To minimize maternal neglect, the surgical incision and maternal nasal orifices were painted with parlodion-gentian violet solution. No antibiotics were administered at any time.

Hormone replacement

To achieve a maintained hormone replacement in castrated females, a 2-cm silastic tubing containing 6–7 mg of either estrogen or androgen powder was implanted subcutaneously immediately after the castration procedure. This treatment results in a prompt establishment of an adequate hormone level reached within 12 h, which persists for at least 3 mo (Dr. P. Siiteri, personal communication).

TABLE I
Effect of Castration on Survival in B/W Mice

Procedure	No. of mice	Mortality at various ages, (in months)						
		4	6	7	8	9	10	11
%								
Sham female	14	0	43	64	78	93	100	
Castrated female	16	0	50	69	87	100		
Sham male	12	0	0	0	8	8	8	17
Castrated male	15	0	40*	47*	60*	67*	87*	100*

* $P \leq 0.05$.

Observation

Mice were observed weekly. Mortality in experimental and control groups was tabulated in 2×2 contingency tables, and evaluated by Chi square analysis with Yates' modification.

Determination of serum testosterone level

Serum samples from castrated and sham-operated male mice were pooled separately. Serum testosterone was measured using a radioimmunoassay, described in detail elsewhere (Korenbrot, C., et al. Submitted for publication). An antiserum against testosterone-3-oxime-bovine serum albumin was used. This antiserum had a 70% cross-reactivity with 5 α -dihydrotestosterone. Since no purification step was used beyond diethyl-ether extraction of the serum samples, the values given for testosterone are slightly contaminated with circulating dihydrotestosterone. This assay was kindly performed by Dr. I. Huataniemi in the Reproductive Research Laboratory of Dr. P. Siiteri.

Fractionation of serum

Serum samples from castrated and sham-operated mice were pooled separately. 200 μ l of pooled serum samples was subjected to ultracentrifugation in a 10–35% sucrose density gradient (0.15 M NaCl, pH 8.0) as previously described (7). Bovine thyroglobulin (19S), human gamma globulin (7S), and hemoglobin (4S) were used as sedimentation markers. 40 fractions were collected and each was analyzed for antibody to DNA and Poly A. Peak fractions were tested for immunoglobulin content. In the 19S region (fraction 10–20) only activity against μ -chains could be detected by Ouchterlony analysis, while in the 7S region (fractions 20–30) activity against γ -chains, but not μ -chains, was present.

Anti-DNA and anti-RNA assays

Antibodies to DNA and Poly A were determined separately with a cellulose ester filter radioimmunoassay. The radioactive nucleic acids were double-stranded DNA ($[^3\text{H}]\text{DNA}$) from KB cells (obtained from Electro-Nucleonics Laboratories, Bethesda, Md.), and $[^3\text{H}]\text{polyriboadenylic acid}$ (Miles Laboratories Inc., Miles Research Products, Elkhart,

TABLE II
Serum Testosterone Concentration of Male B/W Mice

Age of mice <i>mo</i>	Operative procedure	
	Sham	Castrated
	<i>ng/ml</i>	
4	5.77	0.92
6	4.63	0.67
7	4.71	0.69

Ind.). The radioactive antigens were incubated with decomplemented 50- μ l aliquots of each fraction for 30 min at 37°C followed by an overnight incubation at 4°C. The antigen-antibody complexes were collected onto cellulose ester filters (Millipore Corp., Bedford, Mass.). The filters were placed in counting vials and covered with 10 ml of Liquofluor-toluene scintillation medium. Radioactivity was determined in a Packard liquid scintillation counter (Packard Instrument Co., Downers Grove, Ill.) The results are expressed as corrected counts per minute retained on the filter, a value which is directly related to serum antibody concentration (8). To determine the specificity of binding, monospecific rabbit anti-mouse μ -chains and anti-mouse γ -chains were added to peak fractions from the 19S and 7S regions respectively. Anti- μ inhibited DNA binding between 77–100%, while anti- γ inhibited binding between 66–78%. Addition of goat anti-mouse albumin to these gradient fractions had no inhibitory effect.

Calculation of 7S:19S antibody binding

The radioactive binding profiles revealed clear distribution into 7S and 19S peaks of activity after sucrose gradient fractionation. The radioactivity representing the fractionated 7S and 19S peaks within a single gradient were added and compared for total binding activity, which gave a result generally comparable to results with unfractionated whole serum.

Measurement of immunoglobulin concentration

A single radial immunodiffusion method (9) was used to determine immunoglobulin concentration. Determinations were made of IgM, IgG₁, and IgG₂ with pooled sera from sham-operated and castrated B/W male and female mice. No significant difference in immunoglobulin concentrations was found.

RESULTS

Effects of castration on survival of B/W mice. Castration did not affect the life span of female B/W mice, but it increased the mortality of male B/W mice starting at 6 mo of age (Table I). By 11 mo of age, all of the castrated male mice were dead compared to only 2 of 12 sham-operated male controls. Mortality of castrated male B/W mice was not statistically different from that of sham-operated or castrated female B/W mice.

Effects of castration on serum testosterone concen-

tration of male B/W mice. Castration caused a significant decline in the concentration of serum testosterone in male B/W mice. Testosterone levels of castrated males measured at 4, 6, and 7 mo were one-sixth that of sham-operated male B/W mice (Table II).

Development of antibodies to DNA and Poly A in castrated female B/W mice. At 4 mo of age, sham-operated female B/W mice had both 19S and 7S antibodies to DNA, with the latter predominating (Table III). Binding of DNA by 7S antibodies became even more pronounced at 6 and 7 mo (Table III). Antibodies to Poly A were exclusively 19S at 4 mo, but switched to primarily 7S antibodies at 6 mo. This pattern was maintained at 7 mo (Table III).

Prepubertal castration caused a slight decrease in the magnitude of the anti-DNA response at 4 mo, but had no effect at 6 mo (Table III). At 7 mo, there was less 7S binding of DNA in castrated females, while the 19S binding was unaffected (Table III).

Poly A binding remained almost exclusively 19S post-castration. The expected 7S response to Poly A was markedly reduced at 6 mo and virtually absent at 7 mo (Table III). By contrast, the 19S response at these ages was almost unchanged.

Development of antibodies to DNA and Poly A in castrated male B/W mice. Antibodies to DNA in sham-operated male B/W mice were less than in females and predominantly 19S (Table IV). The switch to 7S antibodies to DNA occurred at 7 mo of age. Some 7S antibodies to Poly A appeared at 7 mo (Table IV). The results in sham-operated males and females were generally comparable to data obtained previously in control, nonoperated B/W mice (5, 6). The decrease in DNA binding in sham-operated males between 4 and 6 mo may be due to the formation of immune complexes, as noted previously (5).

Prepubertal castration caused a proportionate in-

TABLE III
Total Radioactivity Representing Fractionated 7S and 19S Peaks in NZB/NZW F₁ Female Mice

		DNA		Poly A	
Age	Procedure	19S	7S	19S	7S
<i>mo</i>					
4	Sham	993	1,314	439	0
	Castrated	740	872	292	0
6	Sham	348	1,449	1,004	1,326
	Castrated	351	1,467	1,096	133
7	Sham	1,327	3,702	781	2,050
	Castrated	1,294	2,271	585	37

crease in both 19S and 7S antibodies to DNA at 4 mo of age (Table IV). At 6 mo, castrated males showed an accelerated switch to 7S anti-DNA antibodies, resembling females in titer and immunoglobulin class. Markedly increased binding of DNA persisted at 7 mo (Table IV). These results are similar to the effects of neonatal thymectomy in male B/W mice (6).

Antibodies to Poly A were only slightly increased at the 4th mo post-castration. At 6 mo, there was an accelerated appearance of 7S anti-Poly A antibodies. This premature switch to 7S response to Poly A was quite marked at 7 mo, with castrated males again resembling females in this respect (Table IV).

Effects of sex hormone replacement on disease in castrated female B/W mice. Castrated female B/W mice were given subcutaneous implants containing either estrogen or androgen. Animals replaced with estrogen had a 94% mortality at 8 mo, compared to 14% in the female mice receiving androgen (Table V). The estrogen-treated mice had larger amounts of both 19S and 7S antibodies to DNA and Poly A when compared to the androgen-treated mice. The most striking difference was in the 7S response to Poly A, which was greatly reduced in the mice receiving androgen (Table V). Proteinuria ranged between 100 and 1,000 mg/100 ml in mice replaced with estrogen, and was less than 100 mg/100 ml in mice receiving androgen. In preliminary experiments, the kidneys of estrogen-treated mice showed marked lymphocytic infiltration and glomerulonephritis, whereas the kidneys in animals receiving androgen showed minimal lymphocytic infiltration and glomerular changes.

DISCUSSION

The greater severity of autoimmune disease in female B/W mice (compared to males) seems es-

TABLE V
Effects of Castration and Hormone Replacement on Survival and Autoantibody Responses in Female B/W Mice

Procedure	Mortality at 8 mo*	Anti-nucleic acid antibody responses at 7 mo†			
		DNA		Poly A	
		19S	7S	19S	7S
	%				
Castrated female receiving estrogen implant	94 (16/17)	1,552	2,583	2,138	2,427
Castrated female receiving andro- gen implant	14 (2/14)§	1,314	1,878	1,139	275

* Number of mice dead/total number of mice.

† Total radioactivity representing fractionated 7S and 19S peaks.

§ $P \leq 0.05$.

established very early in life, perhaps even at birth. There is more anti-DNA antibody formation in these female mice associated with an earlier switch to 7S antibody and accelerated appearance of severe lupus nephritis.

We undertook this study to determine whether sex hormones influenced the disease expression in B/W mice, and if so, whether the predominant effect could be attributed to male or female hormones. In a previous study, Steinberg et al. (10) observed that castration of male B/W mice at 5 wk produced a slight increase in DNA binding at 6½ mo of age. In the present study, we have obtained evidence that androgens may play a major protective role.

The alteration in hormone status as a result of prepubertal castration was shown by the marked decline in serum testosterone concentration. These castrated males developed a disease pattern that resembled sham-operated females, associated with an increase in DNA and RNA binding, and an accelerated switch to 7S antibodies. 9 of 15 castrated males were dead prematurely by 8 mo of age.

By contrast, prepubertal castration of female B/W mice had no effect on life span and less effect on anti-nucleic acid antibodies. The only change in the DNA response was a modest decrease in 7S antibodies observed at 7 mo. However, there was a striking effect on the Poly A response. The anticipated switch to 7S antibodies was virtually eliminated by prepubertal castration, whereas the 19S antibodies were unaffected.

The results of our castration experiments are consistent with the known effects of androgens and estrogens on the normal immune response. Immune

TABLE IV
Total Radioactivity Representing Fractionated 7S and 19S Peaks in NZB/NZW F₁ Male Mice

Age	Procedure	DNA		Poly A	
		19S	7S	19S	7S
mo					
4	Sham	615	324	186	0
	Castrated	1,200	881	280	0
6	Sham	136	83	815	85
	Castrated	593	1,336	722	267
7	Sham	211	788	324	137
	Castrated	1,026	2,488	928	1,304

reactivity is generally enhanced in females compared to males (11). Females have higher immunoglobulin concentrations, make greater primary and secondary antibody responses, are more resistant to the induction of immunologic tolerance, have an enhanced capacity for cell mediated immunity, are more effective in the rejection of tumors and homografts, and have enhanced responses in mixed lymphocyte culture. Eidinger and Garrett observed that castration or thymectomy each resulted in an enhanced response to polyvinylpyrrolidone in male mice (12). However, combined castration and thymectomy abolished the expected enhancement of antibody formation seen after gonadectomy alone. These results suggest that androgens exert their effect at the level of thymocytes, and that the presence of the thymus is required to demonstrate the effect of orchiectomy.

Immunologic processes localized in the thymus, therefore, seem highly sensitive to regulation by sex hormones. Our findings suggest that autoantibody responses may also be sensitive to hormonal modulation, and possibly by mechanisms involving thymocytes. Neonatal thymectomy of male B/W mice resulted in changes in anti-DNA antibodies similar to those seen in castrated male mice (6). There was a rapid and persistent increase in 19S and 7S anti-DNA antibodies which was followed by an accelerated switch to predominantly 7S antibodies. Moreover, neonatal thymectomy in both male and female B/W mice virtually abolished the late switch to 7S antibodies to Poly A, suggesting a thymic requirement for this differentiation event.

Antibodies to DNA in female B/W mice are distinguished from anti-DNA in males and from anti-Poly A in both sexes by virtue of their relative resistance to the effects of either castration or thymectomy. This could relate to a premature extra-thymic peripheralization of the anti-DNA response in females so that it becomes independent of the thymus during neonatal life. The Poly A response in females, and both DNA and Poly A responses in males, may become peripheralized more slowly and are therefore more susceptible to hormonal manipulation.

The opposite effect on the 7S response to Poly A induced by castration of male and female mice is striking. Castrated males develop an accelerated switch to 7S, whereas in the castrated females this response is abrogated. We interpret these results as reflecting an alteration in the equilibrium between helper and suppressor influences as a result of hormonal modulation. Castrated males may be deficient in suppressor regulation, whereas castrated females may lack a helper influence necessary for the full development of a 7S response to Poly A.

Our experiments offer an explanation for the

greater incidence of lupus and other autoimmune diseases in females compared to males. They further suggest that autoimmunity may arise as a consequence of disordered immunologic regulation in which physiologic control mechanisms are still operative, albeit in a defective manner. The improved survival and disease manifestations in castrated female mice given androgen suggests that hormonal modulation of murine lupus may have important therapeutic possibilities. The preliminary observations are currently being investigated in a large study involving both castrated male and female mice given the same or opposite sex hormone, as well as mice subjected to androgen therapy during the course of established disease.

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